

## ANTIGENIC RELATIONSHIP BETWEEN BOVINE EPHEMERAL FEVER (BEF) AND RABIES VIRUSES

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### ABSTRACT

The results achieved in the present study explore clearly that the used techniques were unable to indicate any antigenic relationship between bovine ephemeral fever (BEF) and rabies viruses, although both viruses are of Lyssaviruses of the family Rhabdoviridae. Serum neutralization and agar gel precipitation tests were used. The results of both techniques showed that each of BEF and rabies viruses was able to generate a protective level of immunity in vaccinated mice and rats. Moreover, the challenge of vaccinated and boosted mice and rats revealed that such animals were unable to face the heterologous challenge virus while they withstood the challenge with the homologous virus resulted in protection percentage of 100 and 83.3-100, respectively.

### INTRODUCTION

Bovine ephemeral fever and rabies viruses are belonged to Lyssaviruses of the family Rhabdoviridae (Cybinski and Zakrzewski, 1983; Gard *et al.*, 1983 and Gard *et al.*, 1984).

Bovine ephemeral fever (bovine epizootic fever, three day sickness, stiff sickness) is an arthropod-borne viral disease of cattle and water buffalo characterized by an acute febrile reaction, stiffness, disinclination of movement accompanied by lameness, high morbidity but with very low mortality and short course (Burgess, 1971 and St. George, 1988).

The disease was recorded first in Central Africa 1878 (Schweinfurth, 1898) and in Egypt by Piot, (1896). It was also recorded in South Africa (Vander Westhuizen, 1967), Kenya (Davies and Walker, 1974), Australia (Doherty *et al.*, 1972), Japan (Ito *et al.*, 1969), Saudi Arabia (Abu El-Zein *et al.*, 1997) and Egypt (Hassan *et al.*, 1991; Soheir, 1994; Hassan, 2000 and Zaghawa *et al.*, 2000).

George, (1986) stated that the BEF virus adapted to laboratory animals and tissue culture were enable to develop serum glycoprotein-G which is the major neutralizing and the protective antigen of BEF virus (Kongsuwan *et al.*,

1998 and Uren *et al.*, 1994). The G-protein was found to be of a molecular weight 79 kDa (Hertig *et al.*, 1996). Also, Atanasiu *et al.*, (1974) and Cox *et al.*, (1977) reported that the glycoprotein-G of rabies virus, with a molecular weight of 80,000 dalton has been shown to be the only structural protein of the virus that induces the formation of virus-neutralizing antibodies and which confers immunity to animals.

Among the antigenic relationship between the members of the family Rhabdoviridae, Snowdon, (1970) stated that there is no evidence of antigenic or immunogenic diversity with the BEF virus population, although Cybinksin, (1988) showed that preliminary epitope mapping revealed some variations. In addition, Sodja, (1986) stated that an antigenic difference between different rabies isolates, was batch-specific and there was no correlation between protection and virus neutralization.

So, the present study aim to explore the relationship between BEF and rabies viruses using simple laboratory techniques.

## MATERIAL AND METHODS

### 1. Viruses:

#### 1.1. Bovine ephemeral fever (BEF) virus:

A local isolate of BEF virus adapted to mice brain was used in the present study as a challenge virus. It had a titre of  $10^6$  MICLD<sub>50</sub>/ml (mice intracerebral lethal dose).

#### 1.2. Rabies virus:

A virulent strain of rabies virus to mice (a challenge virus strain) "CVS" was used to challenge vaccinated mice and rats. It had a titre of  $10^7$  MICLD<sub>50</sub>/ml.

The challenge viruses were used at a final dilutions of 100 MICLD<sub>50</sub> where each mouse and rat was challenged with 0.03 ml of such dilution inoculated intracerebrally.

### 2. Vaccines:

Inactivated cell culture vaccines were locally prepared against BEF and rabies viruses at the Department of Pet Animal Vaccine Research, Veterinary Serum and Vaccine Research Institute, Abbasia, Cairo. The used dose of each vaccine was 0.1 ml injected intraperitoneally in each of mice and rats. Two doses of each vaccine were administrated on two weeks interval according to the experimental design.

### 3. Animals and experimental design:

Forty two weaned mice and forty two weaned rats were used in the present study and supplied by the Department of Pet Animal Vaccine Research, Veterinary Serum and Vaccine Research Institute, Abbasia, Cairo. They were apparently healthy and reared under hygienic measures.

Each animal breed was divided into 4 groups as follows:

**Group (1):** Consisted of 12 animals vaccinated with rabies vaccine.

**Group (2):** Including 12 animals vaccinated with BEF vaccine.

**Group (3):** Consisted of 12 rats or mice kept unvaccinated as test control.

**Group (4):** Containing 6 animals received 2 doses of the used vaccine with 2 weeks interval and kept under observation for further 2 weeks. All of such animals (rats and mice) were sacrificed and the blood of each animal was collected aseptically and allowed to clot then the serum was separated and subjected to serological examinations.

Groups 1, 2 and 3 were challenged with homologous and heterologous viruses.

#### **4. Serum neutralization test (SNT):**

Microtitre SNT was carried out according to **Bass *et al.*, (1982)** to detect and estimate the developed antibodies in the sera of mice and rats. The antibody titres were calculated according to **Reed and Muench, (1938)**.

#### **5. Agar gel precipitation test (AGPT):**

It was carried out according to **Cowan and Graves, (1966)**.

## **RESULTS AND DISCUSSION**

Bovine ephemeral fever appeared to be as one of the viral diseases which cause economical losses due to the decline in milk production and loss of body weight (**Sharma, 1992**). Milk production was found to decrease 34-95% (**Davis *et al.*, 1984**). Abortion was recorded in the late stages of pregnancy (**Parsonson and Snowdon, 1974**).

So, vaccination against the disease could be considered a main base to control the disease. The presence of an alternative vaccine is essential and helpful especially in emergency cases.

The present studies could be considered as trials to detect an antigenic relationship between BEF, and rabies viruses.

The results of serum neutralization test (Table 1) showed that all vaccinated rats and mice exhibited very good levels of specific neutralizing antibodies according to the corresponding vaccine (1.35 and 1.2  $\log_{10}$  for rabies in rats and mice and 1.0 and 1.16  $\log_{10}$  for BEF in rats and mice, respectively) using homologous virus. On the use of heterologous viruses in SNT, a very low (0-0.1  $\log_{10}$ ) level of cross neutralization. Parallel to SNT, the results of agar gel precipitation test (Table 2) showed strong positive reactions (++++) on the use of homologous virus and antisera while negative reactions were recorded on the use of heterologous components.

Also, Table (3) showed that the protection % was 98.3-100 in case of using of homologous challenge virus according to the inoculated vaccine while this ratio was 1.6-0 in case of heterologous challenge.

The results of the three illustrated techniques, revealed that there is no antigenic relationship between BEF and rabies viruses although they are two

members of the same family. These findings come in agreement with those of **Suowdon, (1970)** who stated that there is no evidence of antigenic or immunogenic diversity with BEF virus population. In addition, **Cybinksin, (1987)** detected some variations in the preliminary epitope mapping.

Among the developed antibodies in vaccinated rats and mice, the present results agree with **George, (1986)** who reported that BEF adapted to laboratory animals and tissue culture was able to develop serum neutralization antibodies in vaccinated laboratory animals. Similar results were obtained in case of laboratory animal vaccinated with rabies vaccine (**Edries, 1994 and El-Gallad et al., 2001**).

From the recorded data, it could be concluded that there is very low or there is no antigenic relationship between BEF and rabies viruses. The study needs more investigation using more advanced techniques dealing with viral structures and genetic mapping.

#### REFERENCES

- Abu El-Zein, E.M.E.; Gameel, A. A.; Al-Afaleq, A. I.; Al-Gundi, O. and Bukhari, A. (1997):** Bovine ephemeral fever in Saudi Arabia. *Vet. Rec.*, Jun. 1997 (14), 140 (24): 630-631.
- Atanasiu, P.; Tsiang, H. and Ganet, A. (1974):** Nouveau vaccine antirabique humain de cellule primaire. *Ann. Microbiol., Paris*, 125: 419-432.
- Bass, E. P.; Gill, M. A. and Beckenhauer, W.H. (1982):** Development of a modified live, canine origin parvovirus vaccine. *J. Am. Vet. Med. Assoc.*, 181 (9): 909-913.
- Burgess, G.W. (1971):** Bovine ephemeral fever: A Review. *Vet. Bull*, 41 (11): 887-895.
- Cowan, K.M. and Graves, J.H. (1966):** A third antigenic component associated with foot and mouth disease infection. *Virology J.*, 30: 528-540.
- Cox, J.H.; Dietzschold, B. and Schneider, L.G. (1977):** Rabies virus glycoprotein. II. Biological and serological characterization. *Infect. Immun.*, 16: 743-759.
- Cybinski, D. H. (1988):** Commonwealth Scientific and Industrial Research Organization. Canberra, Australia. Unpublished data.
- Cybinski, D.H. and Zakrzewski, H. (1983):** The isolation and preliminary characterization of a rhabdovirus in Australia related to bovine ephemeral fever virus. *Vet. Microbiol.*, 8: 221-235.
- Davis, F.G. and Walker, A. R. (1974):** The isolation of bovine ephemeral fever virus cattle and culicoides midges in Kenya. *J. Hyg., Cambridge*, 75: 231-235.
- Davis, S. S.; Gibson, D.S. and Clark, R. (1984):** The effect of bovine ephemeral fever on milk production. *Aust. Vet. J.*, 61: 128-129.

- Doherty, R.L.; Carley, J.G.; Skandpast, H.; Dyce, A. L. and Snowdor, W. A. (1972):** Virus strains isolated from arthropods during an epizootic of bovine ephemeral fever in Queensland. *Aust. Vet. J.*, 48: 81-86.
- Edries, S.M. (1994):** Studies on preparation of inactivated tissue culture antirabies vaccine. Ph.D. Thesis. Microbiology, Fac. Vet. Med., Cairo Univ.
- El-Gallad, S. B.; Rashwan, S.M.T.; Edries, S.M. and Attyat, M. Kotb (2001):** Comparative studies on the immune response of camels, goats and equines to ERA and HEP rabies vaccines. *J. Egypt. Vet. Med. Ass.*, 61 (1): 37-43.
- Gard, G. P.; Cybinski, D. H. and St. George, T.D. (1983):** The isolation in Australia of a new virus related to bovine ephemeral fever virus. *Aust. Vet. J.*, 60: 89-90.
- Gard, G. P.; Cybinski, D. H. and Zakrzewski, H. (1984):** The isolation of a fourth bovine ephemeral fever group virus. *Aust. Vet. J.*, 61: 332.
- George, J.C. (1986):** Bovine ephemeral fever. *Infect. Trop. Dis. Domest. Anim.*, 453-477.
- Hassan, H.Y. (2000):** An outbreak of bovine ephemeral fever in Egypt during 2000. II. Clinico-chemical and haematological alterations before and after symptomatic treatment trials. 9<sup>th</sup> Sci. Cong. 2000. Fac. Vet. Med. Assiut Univ., Egypt, 325-332.
- Hassan, H.B.; El-Danaf, N. A.; Hafez, M.A.M.; Ragab, A. A. and Fathia, M.M. (1991):** Clinico-diagnostic studies on bovine ephemeral fever with detection of its virus for the first time in Egypt. 1<sup>st</sup> Sci. Cong. Egypt. Societ. Cattle Dis. Assiut.
- Hertig, C.; Pye, A. D.; Hyatt, A. D.; Davis, S. S.; McWilliam, S.M.; Heine, H.G.; Walker, P.J. and Boyle, D.B. (1996):** Vaccinia virus-expressed bovine ephemeral fever virus G but not G (NS) glycoprotein induces neutralizing antibodies and protects against experimental infection. *J. Gen. Virol.*, 77 (4); 631-640.
- Ito, Y.; Tanaka, Y.; Inaba, Y. and Omori, T. (1969):** Bovine ephemeral fever. *Nath. Inst. Anim. Hlth.*, Q, Tokyo, 9: 35-44.
- Kongsuwam, K.; Cybinski, D. H.; Cooper, J. and Walker, P.J. (1998):** Location of neutralizing epitopes on the G protein of bovine ephemeral fever rhabdovirus. *J. Gen. Virol.*, 79 (11): 2573-2581.
- Parsonson, I. M. and Snowdon, W. A. (1974):** Experimental infection of pregnant cattle with bovine ephemeral fever virus. *Aust. Vet. J.*, 50: 329-340.
- Piot, J.B. (1896):** Epizootic of dengue fever of cattle in Egypt. Prix Monbinne. Paris, France, National Academy of Medicine.
- Reed, L.J. and Muench, H. (1938):** A simple method of estimating 50 percent end points. *Amer. J. Hyg.*, 27: 493-497.

- Schweinfurth, G. (1878):* The heart of Africa. Tr. F.E. Frewer London: Sampsonlow, Marston, Low and Searle.
- Sharma, L.K. (1992):* A report on bovine ephemeral fever in cattle. *Ind. Vet. J.*, 69: 544-546.
- Snowdon, W. A. (1970):* Bovine ephemeral fever. The reaction of cattle to different strains of bovine ephemeral fever virus and the antigenic comparison of two strains. *Aust. Vet. J.*, 46: 258-266.
- Sodja, I. (1986):* Antigenic variation in rabies virus strains. *Acta Vet.* 30 (4): 304-319.
- Soheir, M. Banoub (1994):* Ephemeral fever (three day sickness) in closed milking cow farm at Sharquia. 6<sup>th</sup> Sci.Cong., Fac.Vet.Med., Assiut University.
- St. George, T.D. (1988):* Bovine ephemeral fever: A review. *Trop. Anim. Hlth. Prod.*, 20: 194-202.
- Uren, M.F.; Walker, P.J.; Zakrzewski, H.; St. George, T.D. and Byrne, K.A. (1994):* Effective vaccination of cattle using the virion G protein of bovine ephemeral fever virus as an antigen. *Vaccine*, 12 (9): 845-850.
- Van der Westhuizen, B. (1967):* Studies on bovine ephemeral fever. I. Isolation and preliminary characterization of a virus from natural and experimental produced cases of bovine ephemeral fever. *Onderstepoort J. Vet. Res.*, 34 (1): 29-40.
- Zaghawla, A.; Akiela, M. A.; Khader, A. M. and Hassan, H.Y. (2000):* An outbreak of bovine ephemeral fever in Egypt during 2000. 9<sup>th</sup> Sci.Cong.2000, Fac.Vet.Med., Cairo University.

**Table (1): Homologous and heterologous neutralization of the induced antibodies in mice and rats vaccinated with inactivated cell culture BEF and rabies vaccine.**

Animals and vaccines	Mean neutralizing antibody titres (log <sub>10</sub> )	
	Rabies antibodies	BEF antibodies
Rats vaccinated with rabies vaccine	1.35	0.11
Rats vaccinated with BEF vaccine	0.0	1.0
Mice vaccinated with rabies vaccine	1.2	0.1
Mice vaccinated with BEF vaccine	0.12	1.16
Unvaccinated rats	0	0
Unvaccinated mice	0	0

**Table (2): Results of agar gel precipitation test using homologous and heterologous viruses.**

Immune sera	Rabies virus	BEF virns
Rabies serum from rats	++++	-
BEF serum from rats	-	++++
Rabies serum from mice	++++	-
sBEF serum from mice	-	++++
Negative rat serum	-	-
Negative mice serum	-	-

**Table (3): Potency of inactivated BEF and rabies vaccines in mice and rats challenged with the homologous or heterologous viruses.**

Animals and vaccines	Protection % in animals challenged with	
	Rabies virus	BEF virus
Rats vaccinated with rabies vaccine	100	0
Rats vaccinated with BEF vaccine	0	100
Mice vaccinated with rabies vaccine	98.3	1.6
Mice vaccinated with BEF vaccine	0	100
Unvaccinated rats	0	0
Unvaccinated mice	0	0

### الملخص العربي

## العلاقة الأنتيجينية بين فيروسى حمى الثلاث أيام وداء الكلب

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تم حقن مجموعات مختلفة من الفئران السويسرية البيضاء الصغيرة والكبيرة بكل من لقاحي حمى الثلاثة أيام وداء الكلب النسيجين المثبطين، كل على حدة، وقد تبين سيولوجيا أن هذه الفئران تكتسب مناعة جيدة ضد كل فيروس حسب اللقاح المحصنة به إلا أنه عند إجراء اختبار التحدي باستخدام الفيروس الضاري المماثل للقاح المحصن به كانت نسبة الحماية 98,30% - 100% بينما تكون هذه النسبة صفر 0% - 1,6% في حالة إجراء التحدي باستخدام الفيروس الآخر. هذا وقد أوضحت نتائج اختباري المصل المتعادل والترسيب في الأجار تفاعلات إيجابية قوية بين الأمصال المناعية والفيروسات المماثلة وتفاعلات سلبية في حالة استخدام مصل مناعي مع الفيروس المخالف. وعلى ذلك يعتقد أنه لا توجد علاقة أنتيجينية بين فيروسى حمى الثلاثة أيام وداء الكلب بالرغم من كونهما من عائلة واحدة والآخر يحتاج لمزيد من الدراسات التي تبين التشابه أو الاختلاف بين تركيب الفيروسين.