Bioconversion of Pretreated Molasses into Xanthan Gum

M.A. Abdel-Hadi

Chemistry of Natural and Microbial Products Department, National Research Centre, Dokki, Cairo, Egypt.

PRODUCTION of Xanthan gum and the growth of Xanthomonas campestris NRRL-B-1459 were studied by using pretreated sugar beet molasses and corn steep liquor (CSL) as cheap carbon and nitrogen source respectively. The gum production is highly affected by initial concentrations of beet molasses and CSL. Maximum gum production was observed by using beet molasses (40g/1) and corn steep liquor (10g/1).

Keywords: Xanthomonas campestris NRRL-B-1459, Xanthan gum, Pretreated beet molasses, Corn steep liquor.

Xanthan gum was first discribed by Rogovine et al. (1961) as a product of fermentation by Xanthomonas campestris NRRL-B- 1459. This gum has special rheological properties, so it is used in food, cosmetics, pharmaceutical, paper, textiles, adhesives and oil recovery (Linda et al., 1987). Many researchers have studied the fermentation conditions required for optimal gum production (Unashanakar et al., 1996, a,b; Amanullah et al., 1996, 199; Lo et al., 1997; Janna and Ghosh, 1997., Bruggeman et al., 1999, and Pollock et al., 999). In addition, this polymer is readily produced by fermentation of glucose (Agricultural Research Service, 1976), but it can also produced by the fermentation of some industrial wastes such as citrus waste (Michael et al., 1994), whey permeate (Yang and Silva, 1995). This work aims to production of xanthan gum by a local cheap by products such as sugar beet molasses and corn steep liquor.

Material and Methods

Microorganism and culture conditions

Xanthomonas campestris -NRRL-B-1459 was obtained from the Northern Regional Research Laboratory of the U.S. Department of Agriculture Peoria. The strain was maintained on slants with medium composed of (g/l): glucose, 10, peptone, 5, yeast extract, 3, malt extract, 3.0 and agar 15. pH was adjusted to 7.0 before autoclaving at 121°C, 20 mim. The culture was incubated at 30°C, 250 rpm for 48hr before use.

Production medium

Production medium composed of (g/l) glucose, 20; MgSO₄ 7H₂O. 0.1 K₂HPO₄, 0.5 and NH₄ Cl 8.0 (pH 7.0). The medium was autoclaved in 250 ml Erlenmeyer Flasks (50 ml of each) and cultivation with fresh slant and incubated shaken at 30°, 250 rpm for 48hr.

Corn steep liquor (CSL)

CSL 40% solids was obtained from Starch and Glucose Company, Torra, Cairo, Egypt.

Pretretment of sugar beet molasses

Curde sugar beet molasses used as cheap carbon source was kindly supplied by the Delta beet sugar factory, Egypt. This molasses was pretreated before used as follows:

Removal of suspended matter (centrifugation)

The crude molasses solution was repeatedly (3 times) centrifuged at 2000 rpm for 20 min for each. The muddy precipitate was then discarded, while the supernatant called centrifuged molasses.

Potassium ferrocyanide treatment

The crude molasses was centrifuged as mentioned above and treated with potassium ferrocyanide (0.7 g/l) while hot. The mixture was left overnight at room temperature, centrifuged, and the clear supernatant was then used in the fermentation process (Hamissa, 1970).

Sulphuric acid treatment

The crude molasses solution was acidified with 1.0N H₂SO₄ to pH 4 heated to 90°C for 30 min and finally centrifuged (Forage and Righelato, 1 978).

Egypt. J. Microbiol . 37, No. 1 (2002)

Calcium phosphate treatment

To molasses solution, calcium super phosphate (10 g/l) was added. The mixture was boiled, allowed to stand for one hour, then centrifuged. The supernatant solution was then treated with CaO to pH 7.0 and after a further 8hr standing, the sediment of calcium salts and organic matter settled at the bottom and the clarified solution was decanted for use (Allam, 1990)

Cell dry weight (CDW)

The fermentation broth was diluted two folds with distilled H₂O, and centrifuged at 2000 rpm for 20 min. The precipitate biomass was washed two times with distilled H₂O and transferred to a preweighed pyrex cup, placed in an oven (80°C) overnight, and cooled in a desiccator for 1 hr before weighing for dry weight.

Xanthan dry weight and viscosity measurement

* Two volumes of ethanol were added slowly to one volume of cell free supernatant (prepared as mentioned above) containing 1 % KCl, and allowed to stand for one hour before centrifugation at 2000 rpm, for 20 min to precipitate xanthan gum which was separated and transferred to a preweighed pyrex cup, then placed in an oven (100°C) to constant weight (Osato et al., 1996). The dried xanthan was dissolved in 50 ml distilled H₂O before using for viscosity measurements in a BROOKFIELD DV-I + VISCOMETER at 20 rpm and spindle No 4.

Results

Effect of different concentrations of pretreated molasses

In this experiment, glucose in the basal medium was replaced by different concentrations of pretreated molasses. Data recorded in Table 1 indicated that all chemical treatments of molasses (i.e. H₂SO₄, K₄Fe(CN)₆ or Ca₃ (PO₄)₂ gave negative effect on xanthan production compared with physical treatment (centrifugation), but all these treatments (at 40 g/l) gave positive effect on xanthan production compared with crude molasses or even with the medium without molasses (basal medium). High yield of the gum was obtained with 40 g/l of molasses pretreated with only centrifugation. On the other hand, cell dry weight ranged from 3.1-3.4 g/l in all cases.

TABLE	1.	Effect	of	different	concentrations	of	pretreated	molasses	on	the
		produc	ctio	of xantha	an gum by <i>Xanth</i>	ome	onas campes	tris NRRL	-B-1	459.

Molasses		Xanthan dry	Viscosity	Cell dry weight
concentrations (g	/I)	weight (g/l)	(CP)	(g/l)
1.Basal medium *		8.7	97.1	3.192
	20	8.3	95.2	3.411
2.Crude molasses	40	15.1	158.3	3.390
	60	15.1	158.3	3.401
3. Molasses treated	20	9.1	100.2	3.403
by centrifugation	40	20.2	212.1	3.390
	60	20.1	211.8	3.410
4. Molasses treated	20	10.3	106.3	3.410
by H ₂ SO ₄	40	18.7	190.0	3.402
	60	18.8	190.7	3.421
5.Molasses treated	20	9.7	101.3	3.431
by K ₄ Fe (CN) ₆	40	15.8	162.3	3.418
	60	15.8	162.3	3.392
6.Molasses treated	20	10.5	111.8	3.218
by Ca ₃ (PO ₄) ₂	40	15.7	160.7	3.189
	60	15.8	162.3	3.330

^{*}Basal medium = (g/l): glucose, 20, K_2HPO_4 , 0.5, $MgSO_4$.7 H_2O , 0.1 and NH_4Cl , 8.0.

Effect of different concentrations of corn steep liquor (CSL)

Different concentrations of CSL as a complex organic and cheap nitrogen source (2-20 g/l) was added to molasses medium without NH₄Cl. Data presented in Table 2 showed that xanthan gum reached its maximum yield at 10 g/l of CSL, above this concentration, the gum decreased gradually, while, cell dry weight increased by increasing CSL concentrations.

Egypt. J. Microbiol . 37, No. 1 (2002)

TABLE 2. Effect of different concentrations of corn steep liquor (CSL) on Xanthan production by Xanthomonas campestris NRRL-B-1459.

| CSL concentrations | Vention day | Vigasity (CR) | Csl day | Vigasity (CR) |

CSL concentrations	Xanthan dry	Viscosity (CP)	Cell dry weight	
(g/l)	weight (g/l)		(g/l)	
Control *	18.9	187.8	3.201	
. 2	19.3	192.2	3.461	
4	19.9	193.6	3.700	
6	20.5	218.6	4.011	
8	22.6	239.8	4.203	
10	25.7	274.7	4.511	
12	23.7	241.8	4.703	
14	19.5	192.8	4.911	
16	16.3	166.2	5.231	
18	12.7	122.4	5.681	
20	8.3	91.5	5.913	

^{*}Control = (g/l):centrifuged Molasses, 40.0, K₂HPO₄, 0.5, Mg SO₄.7H₂O, 0.1 and different concentrations of corn steep liquor.

Effect of different concentrations of K2HPO4

Table 3 shows the role of K_2HPO_4 on the biosynthesis of xanthan gum by Xanthomonas campestris NRRL-B-1459. Data clearly indicated that the gum was completely inhibited in the basal medium without K_2HPO_4 , while the growth is very weak. On the other hand, the growth and the gum biosynthesis were not affected by the presence or absence of the added K_2HPO_4 in case of molasses or even in molasses + corn steep liquor medium.

Effect of different concentrations of Mg SO₄. 7H₂O

The effect of Mg SO₄. 7H₂O on the production of xanthan and the growth of Xanthomonas campestris NRRL-B-1459 are shown in Table 4. Data indicated that MgSO₄. 7H₂O was essential for gum production, since, when the basal medium was free from MgSO₄.7H₂O, a weak growth and gum inhibition are

observed. On the other hand, the growth and the gum biosynthesis were unaffected by the absence or presence of the added MgSO₄. 7H₂O in case of molasses medium or even in molasses + CSL medium. However the maximum gum production was obtained by using centrifuged beet molasses (40g/l) + CSL (10g/l) medium; pH, 7.0.

TABLE 3. Effect of different concentrations of K₂HPO₄ on the production of Xanthan gum by Xanthomonas campestris NRRL-B-1459.

Concentrations of (g/l)	K₂HPO₄	Xanthan duy weight (g/l)	Viscosity (CP)	Cell dry weight (g/l)
	0.0	0.00	0.00	0.105
Basal medium	0.5	8.90	95.6	3.157
	1.0	8.90	95.6	3.178
	0.0	20.30	212.7	3.188
Molasses medium ²	0.5	20.28	121.7	3.200
,	1.0	20.31	212.8	3.199
	0.0	25.40	274.7	3.210
Molasses +CSL medium³	0.5	25.41	274.7	3.210
anougan.	1.0	25.39	274.7	3.012

- 1. Basal medium (g/l):glucose, 20. Mg SO₄.7H₂O, 0.1, NH₄ Cl, 8.0 and different concentrations of K₂HPO₄.
- Molasses medium (g/l): centrifuged molasses, 40, MgSO₄. 7H₂O, 0.1, NH₄Cl, 8.0 and different concentrations of K₂HPO₄.
- Molasses + CSL medium (g/l): centrifuged molasses 40, Mg SO₄.7H₂O, 0.1, corn steep liquor 10.0, and different concentrations of K₂HPO₄.

Discussion

The above data clearly show that xanthan production is influenced by the type of molasses treatment as carbon source and initial concentration of corn steep liquor (CSL) as nitrogen source as well as the presence of K₂HPO₄ and MgSO₄.7H₂O in the medium. Physical treatment of molasses (centrifugation) with a concentration of 40g/l is the most concentration for high xanthan dry

TABLE 4. Effect of different concentrations of MgSO₄. 7H₂O on the production of Xanthan gum by Xanthomonas campestris NRRL-B-1459.

Mg SO ₄ , 7H ₂ O	(g/l)	Xanthan dry	Viscosity (CP)	Cell dry	
		weight (g/l)		weight (g/l)	
Basal medium	0.00	0.00	0.0	0.120	
	0.05	5.20	56.7	3.111	
	0.10	8.80	97.6	3.105	
	0.20	8.10	95.8	3.152	
Molasses medium ²	0.00	18.30	188.9	3.201	
	0.05	18.30	188.9	3.181	
	0.10	18.30	188.9	3.161	
	0.20	18.21	188.5	3.120	
Molasses +CSL	0.00	26.05	276.2	3.131	
medium³ 		}			
	0.05	25.93	275.9	3.193	
	0.10	25.92	275.8	3.211	
	0.20	25,93	275.8	3.195	

- 1. Basal medium (g/l): glucose, 20, K₂HPO₄, 0.5, NH₄ Cl, 8.0 and different concentrations of Mg SO₄.7H₂O.
- 2. Molasses medium (g/l): centrifuged molasses, 40, NH₄ Cl, 8.0 and different concentrations of Mg SO_{4.7}H₂O.
- 3. Molasses + CSL medium (g/l): centrifuged molasses 40, CSL, 10 and different concentrations of Mg SO₄.7H₂O.

weight and viscosity than other chemical treatments i.e. (sulphuric acid, calcium phosphate or potassium ferrocyanide). It is well known that the chemical treatment of molasses causes some reduction in metal ions which seems to be necessary for gum production. CSL (l0g/l) is the most convenient nitrogen source for xanthan gum production. Thonart et al. (1985) and Roseiro et al. (1992) reported on an inhibitory effect of high CSL level on xanthan production

Egypt. J. Microbiol . 37, No. 1 (2002)

and this effect may due to the drop in the C/N ratio, while Abd El-Salam et al. (1994) found that CSL can be used efficiently for xanthan gum production. At high concentrations of CSL (> 10 g/l), the carbon and nitrogen source seem to preferentially be consumed for growth, resulting in lower xanthan formation.

Though the necessity of inorganic phosphate and magnesium sulfate for xanthan production (Cadmus et al., 1978 and Souw and Demain, 1979), using molasses as carbon source and CSL as nitrogen source, the gum titter was always high even in a medium without additional phosphate and/or Mg SO₄.7H₂O. This can be explained by the fact that the K-phosphate and Mg-sulphate contents of the complex CSL and molasses is high enough to favor xanthan production.

From this study CSL (l0g/l), beet molasses pretreated by centrifugation (40g/l) can be used as a combined nitrogen, carbon and mineral sources for high xanthan gum production by *Xanthomonas campestris* NRRL-B- 1459.

References

- Abd El-Salam, M.H., Fadel, M.A. and Murad, H.A. (1994) Bioconversion of sugarcane molasses into xanthan gum. J. Biotechnol. 33, 103.
- Agricultural Research Service, US Dept. of Agricultural (1976) Polysaccharide (xanthan) of *Xanthomonas campestris* NRRL-B-1459: procedures for culture maintenance and polysaccharide production, purification, and analysis. ARS-NC-5.
- Allam, R.F. (1990) Biochemical studies on ethanol production by fermenataion. *Ph. D. Thesis*, Faculty of Science, Cairo University.
- Amanullah, A., Serrano, C-L., Galinda, E. and Nienaw, A.W. (1996) Reproducibility of pilot scale xamthan fermentation's. *Progress.* 12(4), 466.
- Amanullah, A., Serrano, C.L., Castrp, B., Galeida, E. and Nienow, A.W. (1998) The influence of impeller type in pilot scale xanthan fermentation. *Biotechnol. and Bioeng.* 57 (1), 95.
- Bruggeman, G., Smeets, D., Vandamme, E.J., Vervust, T. and Takarkat, G. (1999) Optimization of xanthan production by *Xanthomonas campestris* NRRL-B-1459 using peptone PS as sole nitrogen source. *Chem. Abst.* 130 (7), 80380.

- Cadmus, M.C., Knutson, C.A., Lagoda, A.A., Pittsley, J.E. and Burton, K.A (1978) Synthetic media for production of quality xanthan gum in 20 L. fermentor. Biotechnol. Bioeng. 20, 1003.
- Forage, A. and Righelato, R.C. (1978) Progress in Ind. Microbiol. M.J., Bull. 9ed 14, 60 Elsevier Sci. Pub. Company, Amsterdam, Oxford.
- Hamissa, F.A. (1970) Chemical studies on citric acid fermentation. Ph.D. Thesis. Faculty of Science, Cairo University, Egypt.
- Janna, A.K. and Ghosh, P. (1997) Stimulation of xanthan production by Xanthomonas campestris using acids. World J. Microbiol. Biotechnol. 13 (3), 261.
- Linda, T., Tansey, L. and Pollock, T.J. (1987) Clustering of mutations blocking synthesis of xanthan gum by Xanthomonas campestris. J. Biotechnol. 3593.
- Lo, Y.M., Yang, S.T. and Min, D.B. (1997) Effect of yeast extract and glucose on xanthan production and cell growth in batch culture of Xanthomonas campestris. Appl. Microbiol. Biotechnol. 47 (6), 689.
- Michael, G., Shelef, G. and Bilanovic, D. (1994) The effect of the various citrus waste fractions on xanthan fermentation. *The Chemical Engineering J.* 56, 37.
- Osato, M., Meiko, Y., Hiroyuki, K., Koza, N., Takashi Y., Tohru, K., Kensuke, I. and Ichiro, N. (1996) Application of shielded hot-wire viscosity sensor to monitoring cultivation of Xanthomonas campestris and Nicotiana tobacum BY-2. J. Ferment. and Bioeng. 82 (1),68.
- Oscar, M., Ricardo, F. and Nora, P. (1993). Effect of corn steep liquor on xamthan production by Xanthomonas campestris. Biotechnol. Lett. 15 (5), 495.
- Pollock, J., Mikolajiezak, M., Yamazaki, M., Thorne, L. and Armentrout, R. (1999) Production of non-native bacterial exopolysaccharide in a recombinant bacterial host. Chem. Abst. 130(5), 15415.
- Rogovine, S.P., Anderson, R.F. and Cadmus, M.C. (1961) Production of polysaccharide with Xanthomonas campestris. Biochem. Microbiol. Technol. Eng. 3, 15.
- Roseiro, J.C., Esgalhado, M.E., Collaco, M.T.A. and Emery, A.N. (1992) Medium development for xanthan production. *Process Biochem.* 27(3), 167.

Egypt. J. Microbiol . 37, No. 1 (2002)

- Souw, P. and Demain, L. A. (1979) Nutritional studies on xanthan production by Xanthomonas campestris NRRL-B- 1459. Appl. Environ. Microbiol. 37, 1186.
- Thonart, Ph., Paquot, M., and Hermans, L. (1985) xanthan production by potential measurement. Enzyme Microb. Technol. 6, 235.
- Unashanakar, H., Annadurai, G., Challapandian, M. and Krvishnan, M.R.V. (1996 a) Influence of nutrients on cell growth and xanthan production by Xanthomonas campestris. Bioprocess Engineering 14 (6), 307.
- Unashanakar, H., Annadurai, G., Challapandian, M. and Krvishnan, M.R.V. (1996 b) Xanthan producuon. Effect of agitation. *Bioprocess Engineering* 15(1), 35.
- Yang, S.T. and Silva, E.M. (1995) Novel products and new technologies for use of familiar carbohydrate, milk lactose. J. Dairy Science 78 (11), 2541.

(Received 1/11/2000; accepted 28/11/2001)

التحول البيولوجي لمولاس بنجر السكر المعامل الى الزانثان

محمد عبد النبي عبد الهادي

قسم كيسمياء المنتجبات الطبيعية والميكروبية - المركز القومي للبحوث - القاهرة - مصر.

استهدف هذا البحث استخدام المولاس المعامل لانتاج معمغ الزانثان بواسطة السلالة الميكروبية Xanthomonas campestris NRRL-B-1459 حيث أوضحت التجارب ما يلى:

١- تفرق المعاملة الميكانيكية للمولاس (الطرد المركزي) عن مثيلاتها
 الكيميائية باستخدام هامض الكبريتيك أو فوسفات الكالسيوم
 أو حديدو سيانور البوتاسيوم لانتاج صمغ الزانثان.

 ۲- اسبت خدام متحلول نقيع الذرة (الخلف من صناعة النشيا والجلوكوز) بتركيز ۱۰ جم/لتر كمصدر نتروجين بالإضافة الى المولاس (٤٠ جم/لتر) يؤدى الى زيادة انتاج الصمخ.

وأمكن استخدام هذة البيئة لانتاج الصمغ بدون اية اضافات أخرى نظرا لما تحتوية من مصادر للكربون والنيتروجين والاملاح المعدنية الضرورية لانتاج صمغ الزانثان.