EFFECT OF CHEMICAL AND BIOLOGICAL TREATMENTS OF SOME CROP RESIDUES ON THEIR NUTRATIVE VALUE. 2- EFFECT OF BIOLOGICAL TREATMENTS ON+CHEMICAL COMPOSITION AND *IN-VITRO* DISAPPEARANCE.

El-Ashry¹, M.A.; H.M. El-Sayed¹; M. Fadel²; H.M. Metwally¹ and M.M. Khorshed¹

- 1. Animal Production Department, Faculty of Agriculture, Ain Shams University, Cairo, Egypt
- 2. Microbial Chemistry Department, National Research Center, Dokki, Giza, Egypt.

SUMMARY

Six crop residues namely wheat straw, rice straw, cotton stalks, corn stalks and pinnae and mid ripe of date palm were subjected to biological treatments using solids state fermentation technique (SSF) to upgrade its nutritive value to be serve as a part of rummenant ration. The residues were moistend at solid : liquid ration 1 : 2 with solution composed of 4% molasses, 0.4% urea, 0.2% K₂H PO₄ and 0.03% MgSO₄.7H₂O. The moistened residues were inoculated with 10% (v / w) inoculum of *Trichoderma viride*, or *Saccharomyces cerevisiae* and 5% (v / w) of both when coculture was applied. The inoculated residues were incubated at room temperature (30°C ± 2) for 21 days.

The highest increase in crude protein contents for wheat straw, rice straw, cotton stalks and pinnae and midrib of date palme tree was achieved by coculture of *T. viride* and *S. cerevisiae* treatment. On the other hand, treatment with fungal culture alone was more suitable for corn stalks residues. The highest losses in dry matter (DM) as a result to coculture treatments was seen for corn stalks, since it was 11.0% followed by rice straw and wheat straw as it were 9.0 and 8.0% respectively. The lowest losses in dry matter were shown for pannae and mid rib of plam tree as it were 4.37 and 2.55, respectively. Biological treatments significantly decreased neutral detergent fiber (NDF) and acid detergent fiber (ADF) specially for treatments by fungus or fungus followed by yeast. Nutritive values were enhanced for all treated residues as *IVOMD* wheat straw, rice straw, cotton stalks, corn stalks, and pinnae and mid rib of date palme tree of were raised from 64.39, 62.45, 73.04, 78.10, 72.65 and 72.35 to be 81.21, 72.4, 80.87, 82.51, 78.89 and 76.02 respectively.

Keywords: Trichoderma viride, Saccharomyces cerevisiae, rise straw, wheat straw.

INTRODUCTION

The shortage of feeds in general and protein in particular attracted the attention of many research workers towards the unconventional sources of feeds crop residues are mainly fibrous materials that are by-products of crops cultivation which from a high percentage of the total volume of the feeds produced annually in Egypt. However, only 4.0 to 4.3 million tons of crop residues out of 13.7-15.2 million tons produced are used for feeding ruminants (Hathout and El-Nouby, 1990).

Residues are burned or wasted, and hence lead to environmental pollution and consequently health hazards. The primary factors limiting the utilization of crop residues are of low digestibility, low protein content and of low palatability. Thus, to increase digestibility of crop residues, it is important to destroy the linkage between cellulose, hemicellulose and lignin or to destroy the compact nature of the tissue, so that liginfied tissue is separated from non-lignified one. There have been attempts to do that by mechanical, chemical or biological treatments (Bakr et al., 1975 and Jakson, 1978).

In the recent years much interest has been evinced in new biotechniques for improving the nutritive value of lignocellulosics using biological treatments in a solid substrate fermentation (SSF) system under non sterile conditions (Han, and Andreson 1975). Although, the SSF is slower than the liquid or submerged fermentation system, due to an additional barrier from the bulk solids, the farmer has the advantage of substantially reducing the space occupied by the fermentor without seriously affecting the yield of biomass (Moo-Young et al., 1983).

The objectives of this study were to investigate. The ability of fungal yeast treatment to improve nutritive values of some crop residues.

MATERIALS AND METHODS

Microorganisms

Trichoderma viride F-516 and Saccharomyces cerevisiae AFZ-98 were obtained from Microbial Chemistry Department, National Research Centre, Dokki, Egypt. The microorganisms were mentioned on agar medium composed of (g / L) malt extract, 30.0; mycolo gical peptone, 5.0 and agar 20.0.

Crop residues

Rice straw, wheat straw, cotton stalks and corn stalks were obtained from waste department, Animal Production Institute Agriculture Research Center, Agriculture Ministry, Pinnae and mid rib of date palm tree were collected from date paim tree located in Faculty of Agricultural, Ain Shams Univ., Shoubra El-Kheima.

Chemical Composition of collected crop residues was carried out according to A.O.A.C. methods (1990) and results are shown in Table (1).

Crop residue	DM	OM	СР	Ash	Cellulose	Hemicellulose	Lignin	CF*	EE**	NFE***
Wheat straw	92.1	80.69	5.81	16.31	39.51	24.17	11.16	33.03	0.46	41.39
Rice straw	92.68	84.38	4.41	15.62	43.30	22.40	12.29	33.61	0.41	45.95
Cotton stalks	91.86	93.46	4.74	6.54	42:2	30.16	14.3	50.68	0.71	37.33
Corn stalks	91.43	80.92	4.74	19.08	30.51	. 36.42	10.5	30.67	0.69	44.77
Pannae of date palm	92.81	89.83	9:01	10.17	40.10	22.23	15.30	31.25	1.89	47.68
Mid rib of date palm	92.07	93.98	5.43	6.02	_ 43.92	26.21	18.93	40.45	0.14	<u> 47.9</u> 6
DM = dry matter	C	M = 0	rganic	matter	er CP = crude protein					

Table 1. Chemical composition of crop residues used in the work.

DM = dry matter

. . . .

ant an an an an

* Crude fiber ** Ether extract CP = crude protein

*** Nitrogen free extract

and the **44** metric of the second s

Preparation of crop residues for biological treatments

The air dried residues were chopped to about 3-5 cm, then packed till use.

Preparation of fungal inoculum

Three days old slant (20 x 200 mm) of Trichoderma viride F-516 was chrused into a flask containing 25 ml of sterilized water. The inoculum was used to inoculate 500 ml-capacity conical flasks containing 20 g of cooled sterilized residue by (autoclaving for 121°C for 30 minutemoistened by basal medium containing g / L 4% molasses, 0.4% urea, 0.2% KH_2PO_4 and 0.03 MgSO₄.7H₂O in solid liquid ratio 1:2 by 10% (v / w).

The inoculated flasks were incubated in adjusted temperature incubator at $30^{\circ}C\pm 2$ for 5 days.

Propagation of fungal biological treatment

The above prepared inocula were employed to inoculate polyethylene bags containing 200g of the demand residues moistened at solid : liquid ratio 1:2 by the above basal medium. The inoculated bages were incubated at $30^{\circ}C\pm 2$ by solid state fermentation system (SSF) for 21 days. At the end of the biological treatment period the bages were opened, oven dried at $70^{\circ}C$ and milled. The chemical analysis was performed.

Preparation of yeast inoculum

Forty eight hour old slants of *S. cere-visiae* AFZ-98 was used to inoculate 500 ml capacity conical flasks containing sterilized 100 ml of the above basal medium. The flasks were incubated at 30°C in shaker incubator for 48 h.

Propagation of biological treatment with yeast

The above prepared yeast inoculum was employed to inoculate polyethylene bags containing 200 g of the demand crop residue moistened by the basal medium in solid : liquid ratio 1 : 2 at 10% (v/w). The inoculated bages were incubated for 21 days at 30°C. At the end of incubation time the same was done as in fungal treatment for chemical analysis.

Coculture treatment

The polyethylene bag containing 200 g of crop residues were inoculated with 10% (w/w) fungal culture inoculum, and after 5 days inoculated with 5% (v/w)

yeast culture and continued to ferment till 21 days at 30°C.

The biological treated residues were oven dried at 70°C to constant weight, then miled through an approximate 1 mm sieve.

Enzymes assays

Five grams of each biologically treated residues in the end of fermentation period was suspended in 50 ml 0.05 M citrate buffer pH 5.0 shaked for one hour at 150 rpm. The samples were filtered through glass wool, then centrifuged under cooling. The clear supernatants were used to determine cellulase (CMCase) according to the method described by Mandels et al. (1976) and hemicellulase (Xylanase) according to the method of Bailey et al. (1992). The enzymes units was calculated on the basis of dry weight. The enzyme unit (IU) is defined as the micro mol of basal substrate unite produced in one minute under reaction conditions

Analysis of biological treated residues

Proximate chemical analysis of raw and treated crop residues were analyzed for dry matter (DM), crude protein (CP), ether extract (EE), crude fiber (CF) and ash content according to the A.O.A.C (1990). The nitrogen free extract (NFE) was calculated by subtracting the summation percentages of CP, EE, CF and Ash content from one hundred.

Fiber fraction determinations

Dried samples were analyzed according to Goering and Van Soest (1970) to determine neutral detergent fiber (NDF), acid detergent fiber (ADF), hemicellulose and cellulose were determine by difference.

In vitro disappearance

The in vitro DM (*IVDM*) and in vitro OM (*IVOM*) disappearance were done on

samples according to Tilley and Terry (1963).

Statistical analysis

The data were analyzed according to statistical analysis system user's guide (SAS) (1998). Separation among means was carried out by using Duncan multiple test (1955).

Data of chemical composition were analyzed according to factorial design (three way classification).

RESULTS

Data obtained from biological treatments of six crop residues namely wheat straw, rice straw, cotton stalks, corn stalks, pannae and mid rib of date palm tree are presented in Tables (2-7). Generally, all treatments affect chemical composition of tested residues consequently the *IVDMD* and *IVOMD*. The differentiations of the effect of biological treatment were differ according to type of treatment and type of residue.

Promising results were achieved when coculture of T. viride F-516 and S. cerevisiae AFZ-98 was applied for wheat straw, rice straw, cotton stalks and pannae and mid rib date palm tree than other biological treatments. On the other hand treatment of corn stalks by T. viride F-516 had advantages. The crude protein increased significantly (P<0.01) and the highest increasing achieved with coculture of fungus and yeast were with wheat straw and rice straw as it were from 5.81 and 5.6 in control to be 12.95% and 13.2% respectively (Table 2, 3). Meanwhile, the crude protein was raised from 4.74 to 10.11% in cotton stalks (Table 4) and from 4.74 to 10.94 in corn stalks (Table 5). The increase in crude protein was achieved by biological treatment of pinnae of date palm tree with coculture of fungus and yeast was about 4.2% than

untreated. Slight increase in crude protein resulted when mid rib of date palm tree was the substrate for fungus and yeast as about 2.4% was achieved.

Dry matter decreased significantly (P<0.01) by fungal and coculture of fungal and yeast. The coculture treatments resulted in high losses in dry matter and the losses were differ according to the type of residues. The decreases were 8.0, 9.7, 7.9, 7.7, 4.4 and 3.9% for wheat straw rice straw, cotton stalks, corn stalks, pannae and mid rip of date palm tree respectively (Tables 2-7). No significantly changes (P<0.05) in dry matter when the residues were treated with yeast culture alone.

All biological treatments were varied on its effect on organic matter. No significant differences (P>0.05) in organic matter of wheat straw when treated with fungal and coculture of fungal and yeast. On the other hand, organic matter was significantly (P<0.05) higher than that of control when the residues was treated with yeast culture under culture conditions. Significant (P<0.01) decrease in organic matter was obtained with treatments with fungal culture or coculture of fungus and yeast. The most decreases were obtained with fungal treatments for cotton and corn stalks as it were 6.0 and 5.5, respectively.

Hemicellulose, cellulose and crude fiber were significantly decreased (P<0.01) by fungal culture and coculture of fungus and yeast. Decreases were in a descending order for wheat straw, rice straw and corn stalks respectively. On the other hand, hemicellulase (Xylanase) and cellulase (CMCase) activities was obtained in a descending order accordance with the decreases in cellulose and hemicellulose for the above residues, respectively.

Ash contents of the fermented residues (Tables 2 to 7) were increased significantly (P<0.01) when the residues were treated by fungus or coculture of fungus

and yeast and it were no significant (P<0.05) when the treatment were accomplished by yeast.

Tables (2 - 7) showed that the biological treatments significantly (P<0.05) decreased NDF and ADF specially for treatment by fungus or fungus followed by yeast. The decreases in NDF and ADF were different between residues.

Data recorded in Tables (2 - 7)showed significantly (P<0.01) improvement for *IDVDMD* and *IVOMD*. The improvement rate was higher when wheat straw rice straw and cotton stalks were treated with coculture of fungus and yeast as it were 13.2 and 20.2% for wheat straw, 14.9 and 10% for rice straw and 29.1 and 7.4% cotton stalks, respectively. Fungal treatment of corn stalks improved the *IVOMD* and *IVDMD* than other two biological treatments and it were 14.6 and 4.4%, respectively. Moreover, the treatments of pinnae and mid rip of date palm tree with coculture of fungus and yeast resulted in significant (P<0.05) increased for *IVDMD* and *IVOMD* than other treatments and improvement were 14.2 and 6.2% for pannae and 22.3 and 4.2% for mid rip respectively.

Table 2. Effect of biological treatments by Trichoderma viride F-516, SaccharomycescerevisiaecerevisiaeAF-98andcoulturefor21days on the chemical composition,IVDMDandIVOMDof wheat straw.

	Biological treatment				
Item	Control	<i>T. viride</i> (1)	S. cerevisiae (2)	Coculture	±SE
Dry matter (DM)	92.10	84.91	92.60	84.10	0.57
Organic matter (OM)	80.69	74.80	80.81	73.20	0.87
Ash	16.31	17.81	16.41	17.92	0.87
Crude protein (CP)	5.81	10.75	6.90	12.95	0.10
Crude fiber (CF)	33.03	31.01	32.91	30.22	0.21
Ether extract (EE)	0.46	0.66	0.54	0.74	0.07
Nitrogen free extracted (NFF)	41.39	37.16	41.22	36.06	0.96
Fiber fraction					
NDF	68.64	63.02	68.12	62.13	0.56
ADF	48.85	44.14	48.15	43.64	0.31
Hemicellulose	24.17	17.30	24.05	16.81	0.53
Cellulose	33.51	31.16	31.31	28.02	0.53
Lignin	11.16	10.91	11.01	11.81	0.12
Enzymes (IU / g)					
CMCase		8.20		9.10	0.45
Xylanase		12.8		16.2	1.98
In vitro disappearance					
IVDMD	47.59	57.63	56.35	60.70	1.46
IVOMD	64.39	69.82	71.97	84.61	1.18

	Biological treatment				
Item	Control	T. viride	S. cerevisiae	Coculture	±SE
	Control	(1)	(2)	(1+2)	
Dry matter (DM)	92.10	84.31	92.12	83.10	0.60
Organic matter (OM)	84.38	79.21	84.42	77.60	1.11
Ash	15.62	17.71	15.92	18.11	1.11
Crude protein (CP)	4.41	9.89	4.85	11.23	0.16
Crude fiber (CF)	33.61	30.51	33.41	30.02	0.66
Ether extract (EE)	0.41				0.07
Nitrogen free extracted (NFF)	45.95	42.69	45.77	41.36	0.10
NDF	66.64	59.95	66.16	59.14	2.33
ADF	48.96	45.20	48.21	44.61	1.47
Hemicellulose	21.4	16.1	21.21	15.22	1.08
Cellulose	40.30	30.50	39.40	27.95	1.30
Lignin	12.29	11.92	12.29	13.11	0.08
Enzymes (IU / g)					
CMCase		7.18		8.25	0.56
Xylanase		11.2		14.3	1.42
In vitro disappearance					
IVDMD	42.52	54.03	49.96	56.42	1.36
IVOMD	62.45	65.67	78.08	72.43	4.88

Table 3. Effect of biological treatments by *T. viride* F-516, *S. cerevisiae* AFZ-98 and coculture for 21 days on the chemical composition, *IVDMD* and *IVOMD* of rice straw.

Table 4. Effect of biological treatments by T. viride F-516, S. cerevisiae AFZ-98 and
coculture for 21 days on the chemical composition, IVDMD and IVOMD of
cotton stalks.

···-	Biological treatment				
Item	Control	T. viride	S. cerevisiae	Coculture	±SE
	Control	(1)	(2)	(1+2)	_
Dry matter (DM)	91.86	85.56	91.91	84.01	0.56
Organic matter (OM)	93.46	96.95	93.51	87.48	0.24
Ash	6.56	6.91	6.71	7.12	0.24
Crude protein (CP)	4.74	9.05	5.20	10.11	0.21
Crude fiber (CF)	50.68	47.58	50.71	47.12	0.23
Ether extract (EE)	0.71				0.07
Nitrogen free extracted (NFF)	37.33	34.23	37.52	33.16	0.50
NDF	67.16	60.77	71.33	60.67	1.05
ADF	53.91	48.68	56.84	78.12	0.63
Hemicellulose	28.16	24.66	28.05	23.14	
Cellulose	35.21	30.12	35.11	24.21	
Lignin	14.32	14.01	14.32	13.85	
Enzymes (IU / g)					
CMCase		6.3		6.8	0.25
Xylanase		10.6		11.6	0.50
In vitro disappearance					
IVDMD	40.43	41.76	47.33	69.16	3.25
IVOMD	73.46	77.30	77.40	_80.87	2.42

Egyptian J. Nutrition and Feeds (2002)

	Biological treatment				
ltem	Cantanal	T. viride	S. cerevisiae	Coculture	±SE
	Control	(1)	(2)	(1+2)	
Dry matter (DM)	91.13	81.35	91.45	83.42	0.59
Organic matter (OM)	80.92	74.60	81.06	75.40	0.61
Ash	19.08	21.06	19.21	21.86	0.61
Crude protein (CP)	4.74	12.36	5.16	10.94	0.40
Crude fiber (CF)	30.67	27.42	30.72	27.86	0.30
Ether extract (EE)	0.69	0.89	1.16	0.79	0.01
Nitrogen free extracted (NFF)	44.77	40.57	44.82	39.05	0.77
e v					1.12
NDF	63.96	54.05	62.36	61.42	0.81
ADF	42.12	41.75	44.11	37.98	0.69
Hemicellulose	32.42	20.31	32.42	21.28	0.54
Cellulose	26.51	21.16	26.41	22.05	
Lignin	10.55	10.11	10.55	11.82	
Enzymes (IU / g)					
CMCase		8.42		9.62	0.60
Xylanase	•-	20.21		22.62	0.71
In vitro disappearance					
IVDMD	45.45	60.00	51.07	55.10	2.80
IVOMD	78.90	82.51	69.56	62.39	0.68

Table 5.	Effect of biological treatments by T. viride F-516, S. cerevisiae AFZ-98 and
	coculture for 21 days on the chemical composition, IVDMD and IVOMD of
	corn stalks.

Table 6. Effect of biological treatments by *T. viride* F-516, *S. cerevisiae* AFZ-98 and coculture for 21 days on the chemical composition, *IVDMD* and *IVOMD* of Pannae of date palm tree.

	Biological treatment				
Item	Control	T. viride	S. cerevisiae	Coculture	±SE
	Control	(1)	(2)	(1+2)	
Dry matter (DM)	92.81	88.62	92.82	88.40	0.53
Organic matter (OM)	89.83	84.91	90.11	84.61	0.26
Ash	10.17	10.42	10.19	10.21	0.26
Crude protein (CP)	5.75	8.44	5.65	9.95	0.27
Crude fiber (CF)	31.25	30.11	31.32	29.86	0.30
Ether extract (EE)	1.89	1.96	2.06	2.24	0.15
Nitrogen free extracted (NFF)	47.68	45.86	47.69	45.17	0.50
NDF	58.46	54.72	56. 95	56.05	0.59
ADF	45.15	46.06	46.54	45.09	0.31
Hemicellulose	26.23	23.72	26.24	22.46	0.25
Cellulose	34.10	31.65	34.27	30.60	0.25
Lignin	15.10	15.65	16.27	18.60	0.25
Enzymes (IU / g)					
CMCase	-	6.23	-	7.42	0.62
Xvlanase	-	9.7	· •	10.40	0.35
In vitro disappearance					
IVDMD	94.49	1.06	50.70	73.70	2.68
IVOMD	72.65	76.38	72.96	78.89	2.60

	Biological treatment				
Item	Control	T. viride	S. cerevisiae	Coculture	±SE
		(1)	<u>(2)</u>	(1+2)	
Dry matter (DM)	92.97	89.47	92.27	88.12	0.60
Organic matter (OM)	93.98	91.16	91.01	91.36	0.25
Ash	6.02	6.42	6.14	6.51	0.25
Crude protein (CP)	5.22	8.56	5.62	7.62	0.04
Crude fiber (CF)	40.45	39.55	40.51	39.16	0.42
Ether extract (EE)	0.14	0.18	0.19	2.01	0.01
Nitrogen free extracted (NFF)	47.96	46.27	47.98	45.96	0.44
NDF	72.68	63.51	75.13	71.72	0.39
ADF	54.24	52.40	53.27	51.06	0.38
Hemicellulose	23.21	21.32	23.22	20.86	0.25
Cellulose	39.92	35.16	39.96	34.72	0.25
Lignin	16.93	16.86	16.93	16.24	0.25
Enzymes (IU / g)					
CMCase	-	4.6	-	5.2	0.30
Xylanase	-	4.3	-	6.8	0.75
In vitro disappearance					
IVDMD	38.20	3.82	56.43	60.27	0.83
IVOMD	72.35	75.53	61.74	76.62	0.77

Table 7. Effect of biological treatments by T. viride F-516, S. cerevisiae AFZ-98 andcoculture for 21 days on the chemical composition, IVDMD and IVOMD ofmid rib of date palm tree.

DISCUSSION

Recently, the production of microbial protein from agricultural wastes products has received the attention of several workers. The organism suit able for upgrading the nutritive value of a wastes must be capable of growth in a wide range of carbon sources, have high growth rates to minimize the size of fermentation system and have a high efficiency of conversion of the substrate to biomass with high protein content. Trichoderma sp. are familiar in biological treatment of agricultural residues as it have the ability to produce sufficient amount cellulolylic enzymes namely exo, endo-gluconases and β-glucosidase (Fadel, 1983) as well as their ability to produce hemcellulase (Xylanase) Fadel (2001).

The chemical composition of the tested residues showed that it can be classified as lignocellulosic residues. Decreasing in DM, OM, CF and NFE as a result to fungal treatments are different among crop residues can be discussed on the light of the ability of T. viride to utilize cellulose and hemicellulose through enzymatic system which could degrade polypolymer in cellulose and hemicellulose to its monomer which can serve as a carbon source by the fungus to produce biomass. Thus the suitability of corn residue to stimulate the cellulose and hemicellulose production consider the limit to upgrade the nutritive value of the crop residue by biological treatment. The data obtained and presented in Tables (2-7) showed the above fact as the decreases in above mentioned parameter can be arrange in a descending order in parallel

Egyptian J. Nutrition and Feeds (2002)

with CMCase and xylanase production since, wheat straw, rice straw corn stalks, cotton stalks, pannae and mid rib of date palm produced 8.2, 7.18, 6.3, 8.42, 6.23 and 4.6 and IUCMCase and 12.8, 11.2, 10.6, 20.21, 9.7 and 4.6.

IU xylanase / g substrate respectively. The enrichment in crude protein are attributed to fungus growth on the produced fermentable sugars by the fungal enzymatic system and the available sugars basal medium employed as a moistening agent.

Promising results obtained from corn stalks treated with fungus than the treatment with coculture of fungus and yeast can be discussed on the basis of the xylanase is stimulated by corn stalks (cellulases, though *S. cerevisiae* could not utilize xylose presented in the medium resulted from hemicellulose hydrolysis (Dekkern, 1983).

Many workers used Trichoderma sp. to upgrade the nutritive value of lignocellulosic waste. Fadel, 1983 used T. harzianum for microbial protein production from rice straw and bagases and could obtained biomass of 32% crude protein. Larwence and Abada (1987) treated rice straw by T. viride or Myrothecium verrucaria and found a significant decrease in their cellulose content by 12 and 8% respectively. The treatment increased lignin content by 8 and 1% and crude protein content by 3.3 and 4.7% respectively. Kahlon and Doss, 1987 treated wheat straw with two cellulolytic non toxic fungi and reported that, decrease the average of NDF, ADF, cellulose and lignin from 89.7, 63.0, 49.1 and 5.4% in untreated wheat straw to 82.4, 56.2, 40.9 and 4.9% in fungal treated, respectively. Also, Chawla and Kundu (1987) reported the crude protein content of fermented wheat straw by T. viride was significantly (P<0.01) higher from 3.5% to 23.2% and an increase in crude protein up to 18.3%.

The increase in crude protein production and decrease in cellulose, hemeicellulose, CF and NFE by biological treatment with coculture of *T. viride* and *Saccharomyces cerevisiae* than the treatment by the *T. viride* alone is due to the *S. cerevisiae* consumed the available part of produced fermentable sugars as a result of hydolyzing enzymes of the fungus on substrates and this stimulate the fungus to produce high level of enzyme activities (Aho *et al.*, 1996).

The increase in organic matter and crude protein when the residues were treated with *S. cerevisiae* AFZ-98 not due to the action of yeast on the residue but due to the propagation of yeast biomass when consumed the sugars in basal medium used as a moistening agent as well as the available soluble sugars in the crop residues can be utilized by *S. cerevisiae*.

The above mentioned can explain why the IVDMD and IVOMD was improvement significantly (P<0.05) higher in treatment the crop residues by coculture of fungus and yeast as the higher crude protein content, fiber biodegradation and fiber biotransformation. Thus in the presence of coculture organic matter could be utilized effectively. Many authors reported an increase in IVDMD for biological treated crop residues. Rai and Mudgal (1984) reported that an increase in IVDMD from 67.9 to 77.5% when treated rice straw with T. viride as well as Shoukry et al. (1985) found that the treated sugar cane bagasse with T. viride increase IVDMD from 22.1 to 45.5%. Kornelia (1984) reported that, the In vitro digestibility of the bioconverted corn stalks increased from 16 to 22%.

In addition, different biological treatments significantly (P<0.05) decreased NDF, ADF and hemicellulose contents the control. However, yeast and combined fungi and yeast treatments showed the highest value of hemicellulose content.

Investigations of the *IVDMD* and *IVOMD* data (Table 6) indicate a significant differences between treatments (P<0.05). The highest values were obtained from combined fungi plus yeast and fungi treatments followed by yeast treatments, lowest values were those of control. It is well document that the dry matter digestibility is positively correlated to CP content and negatively correlated to CF, ADF and NDF (Sawe *et al.*, 1998). These results are in agreement with Akin *et al.* (1993).

CONCLUSION

Coculture of Trichoderma viride F-516 fungus and Saccharomyces cerviciae AFZ-98 could be used successfully to enrich chemical composition well DMD and OMD of cotton stalks and mid rib of date palm tree. However, fungus treatment was the most effective to enrich corn stalks. In addition it should be recorded that either fungus treatment or coculture treatment similar improved the chemical composition, DMD and OMD of wheat straw, rice straw and pinnae date palm tree.

REFERENCES

- Aho, S.; A. Arffman and M. Korhoda (1996). Saccharomyces cerevisiae mutants selected for increased production of Trichoderma reesei cellulases. Appl. Microbial. Biotechnol., 46, 36.
- A.O.A.C. (1990). Association of Official Analytical Chemists : Official Methods of Analysis (13th Ed.). Washington, D.C., U.S.A.
- Bailey, M.J.; P. Biely and K. Poutanen (1992). Inter laboratory testing of methods for assay of xylanase activity. J. Biotechnol., 23, 257.
- Baker, A.J.; M.A. Milett and L.D. Sattar (1975). Wood and wood based

residues in animal feed, In F. Turbak (Ed.) Cellulose Technology Research ASC Symp. series 10, American Chem. Sac., Washington, D.C.

- Chawla, A. and S.S. Kundu (1985). Chemical changes and dry matter disappearance in fungi treated wheat straw. Asian J. Dairy Research. 4(3): 137.
- Dekker, R.F.H. (1983). Bioconversion of hemicellulase : Aspects of hemicellulase production by *Trichoderma reesei* QM9414 and enzymic saccharification of hemicellulose. Biotechnol. Bioeng., 25, 1127.
- Duncan, D.B. (1955). Multiple range and multiple F test J. Biometrics. 11:1.
- Fadel, M. (1983). Microbial protein production using agro-industrial solid wastes. M.Sc. Science thesis, Faculty of Agriculture, Cairo University.
- Fadel, M. (2001). High level xylanase production from sorghum flour by a newly isolate of *Trichoderma harzianum* cultivated under solid state fermentation. Annals of Microbiology, 51, 61.
- Han, K.W. and A.W. Anderson (1975). Microbial fermentation of rice straw. Nutritive composition and in vitro digestibility of fermentation products. Appl. Microbiol., 30, 930.
- Hathout, M.K. and H. El-Nouby (1990). Practical application of crop residues treatment in Egypt. 3rd International Symp. on Feed Manufacture and Quality Control, 337.
- Jackson, M.Q. (1978). Treating wheat straw for animal feeding and health FAO, Roma.
- Kahlon, S.S. and S.K.C. Doss (1987). Biological conversion of paddy straw into feed. Biolig. Wastes, 22:11.

Egyptian J. Nutrition and Feeds (2002)

- Kornelia, Z.H. (1984). Protein enrichment of lignocellulosic agricultural wastes by Mushrooms. Biotechnol. Bioeng., 26, 389.
- Mandels, M.; T. Hontz and C. Nystron (1974). Enzymatic hydrolysis of waste cellulose. Biotechnol. Bioeng., 16, 1471.
- Moo-Young, M.; A.R. Moreivia and R.P. Tengerdy (1983). Principals of solid state fermentation. The felamentatous fungi, 4, 117.
- Rai, S.N. and V.D. Mudgal (1984). Utilization of poor quality roughages: Enzymic treatment of wheat straw. Asian. J. Dairy Res., 3(4): 193.
- SAS (1989). Statistical Analysis System. SAS User's Guide : Statistics. SAS Institute Inc. Editors, Cary, NC.
- Tilley, J.M.A. and R.A. Terry (1963). A two stage technique for the digestion of forage crops. J. Br. Grassl. Soc. 18:104.

and the second second

a a state

and the second second

and the second second second

in a start and a

تأثير المعاملات الكيميانية والبيولوجية لبعض مخلفات الحقل على قيمتها الغذائية. ٢- تسأثير المعاملات البيولوجية على التركيب الكيماني ومعامل الهضم المعملي.

محمد على العشري ١ - حمدي محمد السيد ١ - محمد فاضل ٢ - حمدي موسى متولى ١ - محمد محمود خور شيد ١

١- قسم الانتاج الحيواني - كلية الزراعة - جامعة عين شمس - القاهرة - مصر
٢- قسم الميكروبيولوجي- المركز القومي للبحوث - الدقي - الجيزة - مصــــــر

تم استخدام سنة مخلفات محاصيل وهى تبن القمح – قش الأرز - حطب القطن - حطب الذرة – خسوص النخيل وجريد النخيل ومعاملتها بالمعاملات البيولوجية باستخدام التخمر الجاف لرفع قيمتها الغذائية للمساهمة بجـزه في علائق المجترات. تم رفع رطوبة المخلفات بنسبة ١ : ٢ للمخلف بواسطة البيئة السائلة والتى تتكون مـــن ٤% مولاس ، ٤. % يوريا ، ٢. % فوسفات بوتاسيوم أحادية الهيدروجين و ٢. . % كبريتات ماغنسيوم. تم تحضين المخلفات المضاف لها البيئة مع ١٠ % (حجم إلى وزن) فطر Trichoderma viride أو خميرة حــرارة الغرف. sec cerevisiae و % (حجم إلى وزن) بكل منهما وتم تحضين المخلفات المعاملة على درجة حــرارة الغرف. (٣٠ ± ٢) لمدة ٢١ يوم.

ارتفع محتوى البروتين الخام في كل من تبن القمح وقش الأرز وحطب القطن وخوص وجريد النخيل المعامل بالفطر والمعامل بالخميرة ومن جهة أخرى المعاملة بالفطر كانت الأفضل بالنسبة لحطب الذرة. كان الفقد في المادة الجافة الراجع للمعاملات البيولوجية ١١% لحطب الذرة يليه قش الأرز (٩%) ثـم تبسن القمـح (٨%). وكان الانخفاض في كل من خوص وجريد النخيل ٤.٣٧ و ٢.٥٥% على الترتيب.

المعاملات البيولوجية خفضت مكونات الألياف الخام ADF ، NDF معنوياً ، وارتفعت القيمة الغذائية للمخلفات المعاملة بيولوجياً في صورة معامل الهضم المعملي للمادة الجافة لكل من تين القمح وقش الأرز وحطب القطن ، وحطب الذرة وخوص وجريد النخيل من ٦٤,٣٩ ، ٦٢,٤٥ ، ٢٣,٠٤ ، ٢٣,٠٤ ، ٧٢,٣٥ ، ٣٢,٣٥ قبسل المعاملة إلى ١٩.٢١ ، ٢٠٢٤ ، ٨٠,٨٧ ، ٨٢,٥١ ، ٨٢,٨٩ ، ٢٦,٠٢ على لتو الى بعد المعالجة البيولوجية.