Annals Of Agric. Sc., Moshtohor, Vol. 42 (3): 917-932, (2004).

# EFFECT OF DIFFERENT BIOFERTILIZERS ON PRODUCTIVITY OF SOME ACACIA SPECIES UNDER WADI SUDR CONDITIONS BY

# Saida O.M. Abd-Alla\*; Salwa A.M. El-Toukhy\*; Ibrahim, K.M.A. \* and Amal E.A. Abd El-Hamid\*\*

- \* Range Management Unit, Desert Research Center, Mataria, Cairo, Egypt.
- \*\* Soil Fertility and Microbiology Department, Desert Research Center, Mataria, Cairo, Egypt.

#### ABSTRACT

Response of three Acacia species i.e. Acacia viectoria, Acacia coriacea and Acacia saligna to different biofertilizer resources i.e. Bradyrhizobium, jabonicum, phosphate dissolving bacteria, (bacillus megatherium), Bradyrhizobium mixed with phosphate dissolving bacteria and control, were studied under Wadi Sudr conditions during the period extended from autumn 2001 to spring 2003. The obtained results revealed the following:

- 1. Superiority of biofertilizer treatments over all uninoculated ones.
- 2. The optimum conditions for both plant and soil microbial groups with mixed inoculation treated were mainly achieved for *Acacia saligna* at spring season.
- 3. The other high productive treatment gave sufficient options for using specific Acacia species engaged with appropriate biofertilizer.
- 4. There are significant relation between Bradyrhizobium fabonicum + Bacillus megatherium (Bio<sub>4</sub>) and crude fiber, total ash percentage, shrubs height, crown cover and crown volume of Acacia species particularly A. saligna.
- 5. The interaction between biofertilizer resources and Acacia species had a significant effect on fresh and dry forage yield which superior with Acacia coriacea.
- 6. The highest significant increase for total microbial counts, CO<sub>2</sub> evolution, PDB counts and aerobic cellulose decomposers recorded with (Bio<sub>4</sub>) and Acacia saligna at spring season.

Key words: Saline calcareous soil, Bradyrhizobium jabonicum, Bacillus megatherium, microbial parameters, Acacia viectoria, Acacia coriaceae, Acacia saligna, fresh and dry forage yield, chemical compositions.

#### INTRODUCTION

Using bioorganic agriculture has become a hobe for the Egyptian agriculture to minimize the non stop addition of the high doses of the chemical fertilizer especially when the economical and environmental point of view are considered.

Symbiotic N2 fixing bacteria inoculation is essential for increasing agricultural production through ability of fixing nitrogen, producing phytohormons biocontrol activity and stimulating other soil microorganisms (Abdel-Hamid, 2000). Phosphate dissolving bacteria as *Bacillus*, *Pseudomonas* and some fungi possess the ability to bring insoluble phosphate in soil into soluble forms were also used as biofertilizers (Taha et al., 1969, Ishac et al., 1985 and Mighaed et al., 2004).

However, the main problems under Wadi Sudr conditions are weak soil structure, light texture, salinity reached hazardeous levels poor fertility, wind erosion and arid climatic conditions (Abd El-Ghany et al. 1997). Using biofertilizers under desert stress conditions in new reclaimed desert soils and sand dune regions are very important for resistant wind erosion and increasing biological vegetation.

Acacia is one of the most successful leguminous shrubs used to overcome the lack of fodder in the desert conditions due to rapid growth and its drought resistance (Abou-Deya et al., 1990 and Topps, 1992). Also, Skerman, (1977) reported that leaves are palatable to livestock which used fresh or dried into hay. They are especially used as a supplementary feed for sheep and goats and it can be completely grazed off without harming the plants. Furthermore Ibrahem, 1981 pointed that Acacia includes several species which are introduced for developing the retrograded range lands, particularly under harsh environment, improvement soil and sand dune fixation in arid and semiarid regions.

Acacia saligna tolerates all desert environmental conditions and it is successfully grow in saline soil, (Sheha, 1984 and Zaght et al., 1993). Meanwhile, its leaves are high protein content which considered as forage source for sheep and goats, during dry seasons, (Gutteridge, 1991, Tiedman and Johnson, 1992 and Kandeel et al., 1994).

This work was carried out to investigate the effect of biofertilizer resources on soil microbial properties, resistant wind erosion by formation of difficult erodible grains and increasing the biological vegetation under desert stress conditions and sandune regions. Bioorganic agriculture without mineral fertilization was used for decreasing the environmental pollution and increasing the growth and forage yield as well as chemical content of different Acacia species under Wadi Sudr conditions.

#### **MATERIAL AND METHODS**

Two field experiments were carried out at Wadi Sudr experimental Station of Desert Research Center (DRC), South Sinai Governorate to study the effect of different resources of biofertilizer on productivity and quality of some Acacia species during the period extended from autumn 2001 (dry season) to spring 2003 (wet season). The soil at the experimental site is characterized as calcarous sandy clay texture (56% CaCO<sub>3</sub>, EC 8.3 mmhos/cm and pH value 8.0).

# Nursery site:

Seeds of the three Acacia species i.e. A. viectoria(S1), A. coriacea (S2)and A. saligna (S3) were planted in polyethylene bags filled with sand and clay soil (1:1)

on September 1999 in D.R.C. greenhouse under controlled conditions. After six months and before transplantation in the permanent site, seedling were treated with rhizobial suspention where CMC (0.5%) was used as an adhesive agent.

#### Permanent Site:

Seedlings of Acacia species, were transplanted in the permanent site at Wadi Sudr Exp. Station, after adding 30 m<sup>3</sup>/fed of sheep manure, which mixed with soil surface layer one month before cultivation without using chemical fertilization. Seedlings, were distributed at a distance of 2x2 meter (1050 shrubs/fed.). After six months biofertilizers were used as foliar application for plants and surrounding roots. After good established with shrubs two years old cutting was carried out during the period extended from autumn 2001 to spring 2003. The under ground saline water (8000 ppm) was used in irrigation.

### Organisms:

Bradyrhizobium jabonicum was isolated from active nodules obtained from Acacia spp. grown in calcareous soil. Bacillus megatherium (active phosphate dissolver) was isolated from the rhizosphere of wheat plants grown in calcareous soil. Bacillus and Bradyrhizobia isolates were purified and identified according to Sneath, (1994) and using Bunt and Rovira media (Taha et al. 1969), respectively.

Fresh liquid culture of 7 days old pure local strain either of *Bacillus* or *rhizobia* were intended as biofertilized in the form of single or *mixture* (1:1) at the rate of  $\simeq 10^8$  cells/ml for each of the two bacterial strains. These bacterial cultures having the capability to with and stress desert soil conditions were provided from soil microbiology unit Desert Research Center, Cairo, (Abdel-Hamid, 2000).

The Biofertilizer treatments under study were: Control (BiO<sub>1</sub>); Bradyrhizobium jabonicum (BiO<sub>2</sub>); Bacillus megatherium

Control (BiO<sub>1</sub>); Bradyrhizobium jabonicum (BiO<sub>2</sub>); Bacillus megatherium (BiO<sub>3</sub>) and Bradyrhizobium jabonicum mixed with Bacillus megatherium (BiO<sub>4</sub>).

#### Plant Measurements:

The following measurements were determined at each season, shrub height (cm), crown cover (m<sup>2</sup>) and crown volume (m<sup>3</sup>) by using the following formula, respected by Thalen (1979).

crown cover =  $1/4 \pi \times D_1 \times D_2$ . crown volume =  $1/6 \pi \times D_1 \times D_2 \times H$ 

where  $\pi$ = 3.14,  $D_1$  and  $D_2$  = the shortest and the longest diameters of the shrub, respectively.

and H = plant height.

Fresh yield (ton/fed) was calculated by multiplying the average trender phytomass of the shrubs by the number of shrubs/fed. Dry yield (ton/fed) was determined by taking fresh samples to dry in oven 70°c. Each treatment was milled to fine powder and used for subsequent chemical analysis:

- Crude protein: total nitrogen was determined by modified micro Kjeldahl according to peach and Tracy (1956) and multiplied by 6.25 (Tripathi *et al.*, 1971).

- Crude fiber and total ash were determined by using the method outlined by A.O.A.C. (1970).
- Sodium and potassium were estimated by using a flamphotometer according to Brown and Lilleland (1946).

# Microbiological determinations:

Rhizosphere soil samples were taken during the period extended from autumn 2001 to spring 2003 and subjected to microbial determination. Total microbial counts on soil extract agar (Page et al., 1982), Phosphate dissolving bacterial count on Bunt and Rovira agar medium after modification by (Taha et al., 1970), CO<sub>2</sub> evolution was determined according to (Atef and Nannipien, 1995) as index to microbial activity in soil. Total nitrogen and organic matter content in rhizosphere soil was estimated as described by Black et al. (1982) as a creteion of microbial nitrogen fixation and fertility.

The twelve treatments were arranged in randomized complete block design with three replications.

Data obtained were statistically analyzed using computer statistical program COSTAT according to procedures otulined by Snedecor and Cochran (1980). Means were compared using Duncan's new multiple range test (Duncan, 1955).

#### RESULTS AND DISCUSSION

# 1. Shrub growth characters:

Significant differences in some growth character i.e. shrub height, crown cover and crown volume were observed between different biofertilizer resources during wet (spring) and dry (autumn) seasons in both years, except during spring in the second year (Table 1).

Bradyrhizobium jabonicum  $(BiO_2)$  + Bacillus megatherium  $(BiO_3)$  mixture increased the significantly shrub height as compared with no biofertilizer,  $(BiO_1)$  followed by bradyrhizobium  $(BiO_2)$  and phosphate dissolving bacteria (Bacillus megatherium)  $(BiO_3)$ .

Generally, the stimulating effect of bio mixed inoculation on growth (shrub height, crown cover &volume) may be attributed to provide the shrubs with continuous nitrogen supplying by nitrogen fixing bacteria (Abd El Hamid 2000) as well as providing them with available soluble phosphate produced by phosphate dissolving bacteria (Ishac, 1986). This may be produce phytohormones (Abd El-Hamid, 2000) and biological control elemination of the plant enemies including microbial pathogens (Lugtenberg et al., 1991), that play an important role in plant organ. In this respect, Basu and McKabi (1987) inoculated 7 species of acacia seeds with either Rhizobium sp. or Rhiz. + Azoto, after 150 days, they found that nodulation and the plant growth were enhanced in all treatments compared with control, and the most effective treatment varied with legume species.

Table (1): Growth parameters, fresh and dry productivity and chemical content of some *Acacia* species as affected by different resources of biofertilizer during the period extended from autumn 2001 to spring 2003 seasons.

Year	2001/2002						
Treatments	<del> </del>	Autumn					
1		Biofertilizer resources Acacia species					
Triets		BiO <sub>1</sub> BiO <sub>2</sub> BiO <sub>3</sub> BiO <sub>4</sub>			SI	S2	S3
Shrub beight (cm)	87.17b		112.83ab		67.38b	95.00b	195.63a
Crown cover (m <sup>a</sup> )	0.423b		0.573ab	0.964a	0.328b	0.870a	0.892a
Crown volume (m³)	0.4236 0.254c		0.731ab	0.925a	0.328b	0.693a	0.923a
F. forage yield (ton/fed.)	0.132b	0.265b	0.164b	0.523a	0.104b	0.366a	0.349a
D. forage yield (ton/fed.)	0.050b	0.2030 0.111b	0.067b	0.323a	0.044b	0.300a	0.141a
Crude protein (%)	5.98a	5.86a	5.41a	5.06a	5.52a	5.10a	6.12a
Crude fiber (%)	23.91a	24.85a	26.89a	23,04a	24.76a	28.00a	21.26b
Total ash (%)	6.29a	5.02a	5.41a	6.39a	5.81a	6.08a	5.43a
Na <sup>+</sup> (%)	0.554ab	0.628a	0.424b	0.39a 0.443ab	0.571a	0.06a	0.500a
K (%)	0.371a	0.028a	0.4240 0.353a	0.344a	0.391a	0.400a	0.370a
K (70)	0.3/18	U.3988	0.333a		U.3918	0.3398	0.370a
Church beinha (mar)	102 025	149.17ab	120 17-1	Spring	00 00L	111 125	210.25a
Shrub height (cm)							
Crown cover (m²)	0.596b	0.805b	1,100b	1.89a	0.577b	1.500a	1.215a
Crown volume (m³)	0.715b	0.673b	1.224ab	1.975a	0.376b	1.747a	1.317a
F. forage yield (ton/fed.)	0.374b		0.467ab	0.818a	0.441b	0.346b	0.870a
D. forage yield (ton/fed.)	0.137a	0.165a	0.171a	0.270a	0.149b	0.137b	0.271a
Crude protein (%)	8.05a	9.14a	7.00a	9.06a	6.86a	9.00a	9.03a
Crude fiber (%)	16.37a	16.95a	18.13a	14.53a	18.65a	18.07a	12.77b
Total ash (%)	7.59ab	8.03a	7.59ab	6.56b	7.57a	7.06a	7.69a
Na (%)	0.593a	0.557a	0.533a	0.580a	0.623a	0.608a	0.468a
K (%)	0.465a	0.4360a	0.425a	0.518a	0.486a	0.505a	0.410a
1	1		- 7	2002/2003	,		
				Autumn	A = 2 = 4		
Shrub height (cm)		160.00ab			95.63b		217.75a
Crown cover (m <sup>3</sup> )	0.828b	1.390ab		2.230a	0.912b	1.824a	1.305ab
Crown volume (m³)	0.866b	1.389b	0,994b	2.561a	0.495Ъ	1.390a	1.932a
F. forage yield (ton/fed.)	0.700Ь	1.006ab		1.453a	0.840a	0.991a	1.221a
D. forage yield (ton/fed.)	0.318a	0.423a	0.413a	0.538a	0.357a	0.442a	0.470a
Crude protein (%)	6.04a	7.14a	6.17a	6.04a	6.41ab	4.88b	7.75a
Crude fiber (%)	26.49a	24.56a	24.09a	21.32a	21.43b	32.37a	18.54b
Total ash (%)	6.96a	4.76a	6.36a	6.66a	6.05a	5.51a	6.99a
Na (%)	0.617a	0.693a	0.593a	0.580a	0.680a	0.608a	0.575a
K*(%)	0.328a	0.383a	0.337a	0.332a	0.310a	0.331ab	0.394b
	175	T170 55	160.5	Spring	110.00	150 00	000.00
Shrub height (cm)	137.0a	160.33a		178.50		150.00b	
Crown cover (m²)	0.997a	0.802a	0.885a	1.357a	0.550b	1.434a	1.046ab
Crown volume (m)	1.072a	0.930a	0.968a	1.817a	0.419b	1.569a	1.603a
F. forage yield (ton/fed.)	1.234a	0.910a	1.811a	2.476a	0.834Ь	3.130a	0.860b
D. forage yield (ton/fed.)	0.555a	0.467a	0.837a	1.111a	0.420b	1.401a	0.406b
Crude protein (%)	3.67c	5.63b	5.62b	8.04a	5.13b	7.13a	4.96b
Crude fiber (%)	26.15a	21.85ab	20.77b	18.46b	21.25a	24.18a	19.99a
Total ash (%)	8.36a	4.49a	8.42a	8.35a	8.28b	6.52c	9.66a
Na (%)	0.240a	0.267a	0.238a	0.276a	0.291a	0.228b	0.247ab
K*(%)	0.262a	0.272a	0.253a	0.270a	0.257a	0.248a	0.288a
Views having the same letters are not different significantly at $P = 0.05$ level of significantly.							

Means having the same letters are not different significantly at P = 0.05 level of significantly.  $BiO_1 = Control$   $BiO_2 = Bradyrhizobium jabonicum$   $BiO_3 = Bacillus megatherium$  $BiO_4 = BiO_2 + BiO_3$  S1 = A viectoria S2 = A coriaceae. S3 = A saligna The growth characters (shrub height,crown cover &volume) differed significantly among acacia species, (Table 1). In general, it could be concluded that *Acacia saligna was* superior than other two species in most growth characters especially during autumn in both years followed by *Acacia coriacea*. This may be due to rapid growth and its drought resistance (Abou-Deya et al., 1990 and Topps, 1992). Furthermore, they tolerate all desert environmental conditions and they give a successful growth under saline soil, (Sheha, 1984 and Zaght et al., 1993).

The interaction between the two factors under study was not significant on the growth characters except crown cover and volume during spring in both years, (Table 2). The highest values were obtained when biomixed inoculation interacted with Acacia coriacea.

# II. Forage productivity:

Data in Table (1) reveal that the fresh forage yield of acacia species significantly affected by various biofertilizer resources. These trend was noticed during different seasons in both years. An exception was observed in spring of the second year. The highest fresh forage yield was recorded by application of *Bradyrhizobium* mixed with phosphate dissolving bacteria treatment, which superior than the control by 296%, 135%, 108% and 101% during autumn and spring of the two years, respectively. This may be attributed to the effect of this treatment on the promotion of growth parameters.

On the other hand, the insignificant increase was observed for dry forage yield with different biofertilizer treatments except during autumn in the first year. The maximum values were also obtained with *Bradyrhizobium* mixed with phosphate dissolving bacteria.

With respect to the different acacia species, data in Table (1) indicate that *Acacia saligna* gave significantly higher fresh forage yield in spring and autumn of the first and second year, respectively, while *Acacia coriacea* was superior in autumn and spring of the first and second year, respectively.

It is evident from (Table 1) that dry forage yield followed closely the same trend of the fresh forage yield. The maximum fresh and dry forage yields of *Acacia saligna* were recorded during both seasons in the two years except in the spring of the second year.

The interaction between the main factors (biofertilizer resources and acacia species) had a significant effect on fresh and dry forage yields in autumn of the first year and during spring of the second year, Table (2). The highest fresh and dry forage yield were obtained when *Acacia coriacea* treated with *Bradyrhizobium* mixed with phosphate dissolving bacteria.

# **III. Chemical content:**

The influence of biofertilizer resources on chemical composition of different acacia species during the period extended from autumn 2001 to spring 2003 (Table, 1)showed that there was gradual and slight insignificant effects on crude protein, crude fiber, total ash and mineral content percent by using different biofertilizer resources. This trend was true during autumn in the two years, whereas, in spring there was significant effect for total ash in first year, crude fiber and crude protein in the second year.

Table (2): Effect of the interaction between different biofertilizer resources and some *Acacia* species on growth parameters, forage productivity and chemical content during the period extended from autumn 2001 to spring 2003 seasons.

	SUILS.							
Years	<u> </u>			200	01/2002			
Seasons	Autumn				<u>Spri</u>	ng		
Biofertilize					ļ			l .
season	BiO,	BiO <sub>2</sub>	BiO <sub>3</sub>	BiO <sub>4</sub>	BiO <sub>1</sub>	BiO <sub>2</sub>	BiO <sub>3</sub>	BiO₄
Acacia species	L	Ĺ	L	L	<u> </u>	L	<u> </u>	L
Crown Cover (m²)								
Acacia viectoria					0.189Aa	0.432Aa		1.031Ac
Acacia cariaceae	-	<u> </u>			0.551Da	0.709CDa		3.305Aa
Acacia saligna		-	<u> </u>	<del>-</del>	1.048Aa	1.276Aa	1.209Aa	.327Ab
		, <del></del> .	Crown	volume (		<del>,</del>		
Acacia viectoria	-				0.308Аа	0.122Aa	0.350Aa	
Acacia coriaceae				<u> </u>	1.494Aa	1.511Aa	1.975Aa	.008Aa
Acacia saligna		<u> </u>	<u> </u>		0.344Da	0.388CDa	1.347BCD	3.190Aa
			esh forag		on/fed.)			
Acacia viectoria		0.116Aa			-	-		
Acacia coriaceae	0.046Da	0.308BCD	.061CD	1.050Aa	-	<u> </u>		
Acacia saligna	0.309Aa	0.371Aa	0.329Aa	.387Ab				
		ŗ	ry forage	e yield (to	n/fed.)			
Acacia viectoria		0.055Aa			_	-		
Acacia coriaceae	.019cD	0.124BCD	0.019Da	0.396Aa				-
Acacia saligna	0.110Aa	0.154Aa	0.134Aa	.166Ab		•	-	•
			Tota	l asb (%	)			
Acacia viectoria	-	-	-	-	6.295DC	8.860Aa	8.440BCD	.680CD
Acacia coriaceae	-	•	-	-	6.310Abc	8.600Aa	6.545Aa	6.785Aa
Acacia saligna	-	-	-	-	10.175Aa	6.640CDa	7.710BCD	6.215Da
			Crud	e fiber (%	<del>(</del> 4)			
Acacia viectoria	-	-	-	-	20.050Aa	16.425Cb	20.940Aa	7.175BC
Acacia coriaceae	-	-	-	-	16.920Cb	21.510Aa	20.140Bb	3.695D
Acacia saligna	-	-	-	-	12.145Bc	12.905Ac	13.300Ac	2.730AB
			20	02/2003		<del></del>		*
			Crown	Cover (E	n')			
Acacia viectoria	-		-	- 1	0.494Aa	0.525Aa	1.093Aa	0.909Ac
Acacia comacene	-	-	-	-	.449BCD	0.82CDa	0.622Da	2.883Aa
Acacia saligna	-	-	-	-	1.049Aa	1.098Aa	0.940Aa	.097Ab
		·	Crown	volume (	m³)			
Acacia viectoria	-	-	_	-	0.340Aa	0.415Aa	0.845Aa	0.075Ac
Acacia coriaceae	-	-	-	-	1.425B-	0.659CDa	0.646Da	3.548Aa
					CDa			
Acacia saligna	<u>;</u>	-	-		1.453Aa	1.717Aa	1.413Aa	.830Ab
		Fr	esh forag	e yield (T	on/fed)			
Acacia viectoria	-		-	-	0.788Aa	1.155Aa	1.155Aa	0.237Ac
Acacia coriaceae	-	-	-		2.468CDa		3.255BCD	
Acacia saligna		-			0.447Aa	0.919Aa	1.024Aa	.050Ab
						<u> </u>		

Table	(2)	: Cont.
	· (-,	

Seasons		Aut	umn			Spri	ng		
Biofertilize season Acacia species	BiO <sub>1</sub>	BiO₂	BiO <sub>3</sub>	BiO <sub>4</sub>	BiO <sub>1</sub>	BiO₂	BiO <sub>3</sub>	BiO <sub>4</sub>	
•		I	ry forage	yield (T	on/fed.)				
Acacia viectoria	-	-	-	-	0.393Aa	0.565Aa	0.596Aa	0.125Ac	
Acacia cariaceae	-	-	-	-	0.064CDa	0.400Da	1.402BCD	2.738Aa	
Acacia saligna	-	-	-	-	0.209Aa	0.436Aa	0.511Aa	.470Ab	
			Tota	d ash (%	<del>)</del>				
Acacia viectoria	-	-	-	-	8.275Aab	7.185Abc	9.010Aa	8.635Aa	
Acacia coriaceae	-	-	-	-	6.280Ab	5.245Ac	7.180Aa	7.375Aa	
Acacia saligna	-	-	-		10.510Aa	10.350Aa	9.055Aa	9.040Aa	
-	Crude fiber (%)								
Acacia viectoria	-	-	-	-	21.710Ac		18.750Aa	2.820A	
Acacia coriaceae	-	J	•	-	34.665Aa	23.730BCD	9.875CD	8.450Da	
Acacia saligna	~		-	-	22.070Abc	20.100AB	23.695Aa	4.100B	

<sup>- :</sup> Not significant.

The same average in the same row is indicated by capital letters whereas in the same column in indicated by small letters.

 $BiO_1 = Control$ 

BiO<sub>2</sub> = Bradyrhizobium jabonicum BiO<sub>3</sub> = Bacillus megatherium

 $BiO_4 = BiO_2 + BiO_3$ 

No clear and insignificant effects were observed for the different chemical percentages concerning the different Acacia species in autumn and spring of the first year. An exception was noticed in crude fiber which, Acacia saligna had a lower value and there was no significant difference between the two other species. However, in the second year the contrary was true, there was significant differences between Acacia species in their chemical content. The maximum values of crude protein and total ash percentages were obtained from Acacia saligna in spring season, while, the lowest value of crude fiber percent was recorded.

Biofertilizer resources interacted with different Acacia shrubs species causing significant effect on crude fiber and total ash percentages in spring of the two years, Table (2).

# IV. Microbiological determinations:

#### A- Total microbial counts:

Regarding biofertilizer application either individuals or as a mixture could be significantly affected on microbial counts and inorder arranged as follows: (Bacillus megatherium (Bio<sub>3</sub>) + Bradyrhizobium japonicum (Bio<sub>2</sub>) > Bio<sub>2</sub> > Bio<sub>3</sub> > uninoculated (Table 3). Thus mixed inoculation increased counts as much as 3 folds if compared with uninoculated one.

The rhizosphere of A. saligna plant recorded the highest counts significantly followed in descending order A. coriaceae and the least significant by A. viectoria.

Thus, the highest total microbial counts recorded at spring season using mixed inoculation and A. saligna rhizosphere being 303x10<sup>5</sup> CFU/gm dry soil and increased counts N 3 folds if compared with uninoculated (control) treatment.

Table (3): Influence of Bradyrhizobium japonicum, Bacillus megatherium strains (individual or mixture) into rhizosphere of Acacia species. trial on microbial counts during autumn and spring seasons (2001/2003)

_\4	UU.	1/2	VV.	<u>"</u>

	(	Counts x 10 <sup>5</sup> C	FU g dry soil	l)
Treatment	Acacia viectoria	Acacia coriaceae	Acacia saligna	Mean
	Ţ	Autumn	season	
Cont. Bio1	55.50 c	66.67 c	85.33 d	69.17 d
Brad. Bio2	152.27 b	203.67 b	213.33 b	189.76 b
PDB Bio <sub>3</sub>	149.67 b	197.00 b	206.33 с	184.33 c
Brad + PDB Bio4	194.33 a	245.00 a	257.33 a	232.22 a
Mean	137.94 с	178.08 b	190.58 a	
		Spring	eason	
Cont. Bio,	78.28 c	99.74 c	118.68 c	98.90 c
Brad. Bio <sub>2</sub>	174.73 b	245.43 b	252.50 b	224.22 b
PDB Bio <sub>3</sub>	171.70 b	244.42 b	247,45 в	221.19 b
Brad + PDB Bio <sub>4</sub>	228.26 a	277.75 a	303.00 a	269.67 a
Mean	163.24 c	216.83 b	230.41 a	

Initial count =  $30 \times 10^3$  CFU g dry soil.

 $BiO_1 = Control.$ 

BiO<sub>2</sub> = Bradyrhizobium jabonicum

BiO<sub>3</sub> = Bacillus megatherium

 $BiO_4 = BiO_2 + BiO_3$ 

Mean values in the same box sharing an alphabet are not significantly different.

The results are in agreement with Subba Rao (1988). Abd-El Ghany (1996), Abdel-Hamid (2000) and Abd-El Rahman (2003) who stated that microbial inoculation increased the number and biological activity of desired microorganisms in the root environment.

#### B- CO<sub>2</sub> evolution:

Obtained results recorded in Table (4) apparently reveal that mixed inoculation gave the highest CO<sub>2</sub> evolution levels in comparison to individuals which gave slightly lower CO<sub>2</sub> evolution figures.

In all cases the rhizosphere of uninoculated Acacia species plants gave the lowest  $Co_2$  evolution values.

Results in Table (4) show that CO<sub>2</sub> evolution is positively correlated with total microbial counts under different inoculant either alone or as a mixture as concluded by Khalil *et al.* (1991), Faid (1994), Abd El-Hamied (1995), and Abo-Alaa (2002).

#### C-PDB

It is clear from the data presented in Table (5) that counts of PDB at spring season were higher than those at autumn season. Mixed inoculation significantly increased PDB  $\geq$  2 folds if compared with individuals and 3 folds in comparison to control.

Table (4): Influence of *Bradyrhizobium japonicum*, *Bacillus megatherium* strains (individual or mixture) into rhizosphere of *Acacia* species, on Co. evolution during autumn and spring seasons (2001-2003).

<u> </u>	Co <sub>2</sub> evolution (mg Co <sub>2</sub> /100g dry soil/24 hr)							
Treatment	Acacia viectoria	Acacia coriaceae	Acacia saligna	Mean				
		Autumn season						
Cont. Bio1	0.48 c	0.62 c	0.91 c	0.67 с				
Brad. Bio <sub>2</sub>	0.87 в	1.11 b	1.45 b	1.14 b				
PDB Bio <sub>3</sub>	0,86 b	1.11 b	1.41 b	1.13 b				
Brad + PDB Bio <sub>4</sub>	1.06 a	1.60 a	2.08 a	1.58 a				
Mean	0.82 c	1.11 b	1.46 a					
	Ţ. <u> </u>	Spring s	eason					
Cont. Bio1	0.56 c	0.86 с	1.24 c	0.89 с				
Brad. Bio <sub>2</sub>	1.11 b	1.43 b	1.74 b	1.43 b				
PDB Bio,	1.06 b	1.41 b	1.74 b	1.40 b				
Brad + PDB Bio <sub>4</sub>	2.34 a	2.57 a	2.79 a	2.57 a				
Mean	1.27 c	1.57 b	1.88 a					

Initial CO<sub>2</sub> evaluation 0.25

Mean values in the same box sharing an alphabet are not significantly different.

Table (5): Influence of Bradyrhizobium japonicum, Bacillus megatherium strains (individual or mixture) into rhizosphere of Acacia species, on phosphate dissolving bacteria during autumn and spring seasons (2001-2003)

	Densities of P-dissolvers 10 <sup>2</sup> CFU/g dry soil						
Treatment	Acacia viectoria	Acacia coriaceae	Acacia saligna	Mean			
	Autumn season						
Cont. Bio1	49.24 c	63.13 c	76.26 c	62.87 c			
Brad. Bio2	152.76 в	206.04 b	216.14 b	191.65 b			
PDB Bio <sub>3</sub>	152.01 b	202.00 b	214.12 в	189.38 b			
Brad + PDB Bio <sub>4</sub>	189.38 a	252.50 a	277.75 a	239.88 a			
Mean	135.85 с	180.92 b	196.07 a				
	1	Spring season					
Cont. Bio	66.66 c	92.42 c	102.01 c	87.03 c			
Brad. Bio2	188.87 b	228,51 b	245.43 Ь	220.94 b			
PDB Bio,	186.85 b	224.22 b	245.43 b	218.83 b			
Brad + PDB Bio <sub>4</sub>	205.03 a	287.85 a	328.25 a	273.71 a			
Mean	161.85 c	208.25 b	230.28 a				

Initial count =  $30 \times 10^2$  CFU/  $g^{-1}$  dry soil.

Mean values in the same box sharing an alphabet are not significantly different.

Also, the counts of PDB was significantly increased by acacia species in ascending order 161.85, 208.25, 230.28 x  $10^2$  CFU/gm dry soil as means average in the rhizosphere of A. viectoria, A. coriaceae and A. saligna at spring season, respectively.

Thus, mixed inoculation (PDB + Brad) stimulated the counts of PDB in the rhizosphere of A. saligna which significantly doubled if compared with control.

# D. Aerobic cellulose decomposers:

Cellulose decomposers are the first group of bacteria attacking organic matter and breaking its complexity that encourage the proliferation of other group of microorganisms.

Table (6) show that densities tended to increase by using different types of biofertilizers rather than the control. The highest density was recorded in the treatment inoculated with (PDB + Bradyrhizosphere) followed in descending order by individuals.

The highest cellulose decomposers densities were significantly recorded with A. saligna followed in descending order by A. coriaceae and A. viectoria at both seasons.

The highest significant value was recorded with rhizosphere of A. saligna plant using (PDB + Bradyrhizobium) as mixed inoculation, this increased densities as much as 4 folds if compared with uninoculated treatment at spring season.

# E- Organic Matter(O.M):

The mixed inoculation significantly increased O.M.% followed in descending order by individual treatments and uninoculated one under A. saligna > A. coriaceae > A. viectoria at spring and autumn seasons, (Table 7).

# F. Total Nitrogen(T.N).:

Data presented in table (8) show that the highest total nitrogen content was observed by using biofertilized treatment for A. saligna at spring season. The high pH value, high calcium carbonate may cause N volatization in the from of ammonia resulting in decrease of the total nitrogen available for the growing plants.

Finally, the study maintains a lot of options which could be used up in any farm conditions.

It could be concluded from the above results that application of biofertilizers activated soil microorganisms particularly in plant rhizosphere region and was more efficient for root colonization and tolerant desert stress conditions. (Subba Rao, 1988, Abd El-Hamid 2000, Migahaed et al., 2004) using biofertilizer essential for wind erosion control under Wadi Sudr conditions because biofertilization enhancing the formation of difficult erodible grains (Abd El Ghany et al., 1997) and also the interaction between biofertilizer resources and acacia species had a significant effect on shrubs height, crown cover, crown volume, crude fiber and total ash. Finally it is very important for resistant wind erosion by formation of difficult erodible grains and increasing the biological

vegetation under desert stress conditions and sandune regions. Using bioorganic agriculture with might mineral fertilization for decreasing the environmental pollution and increasing the growth and forage yield as well as chemical content of different Acacia species under Wadi Sudr conditions.

Table (6): Influence of Bradyrhizobium japonicum, Bacillus megatherium strains (individual or mixture) into rhizosphere of Acacia species. plants, on aerobic cellulose decomposers densities during autumn

and spring seasons (2001-2003).

	MPN of cellulose decomposers 10 <sup>5</sup> cells/g d					
Treatment	Acacia viectoria	Acacia coriaceae	Acacia saligna	Mean		
		Autumn	season			
Cont. Bio1	0.61 c	0.63 d	0.91 d	0.71 d		
Brad. Bio <sub>2</sub>	0.94 b	1.21 ь	1.82 b	1.32 b		
PDB Bio <sub>3</sub>	0.93 b	1.11 c	1.72 c	1.25 c		
Brad + PDB Bio <sub>4</sub>	1.41 a	1.82 a	2.22 a	1.82 a		
Mean	0.97 c	1.19 b	1.67 a			
<u> </u>		Spring	scason			
Cont. Bio1	1.11 d	1.41 c	1.72 c	1.41 d		
Brad. Bio <sub>2</sub>	2.22 c	2.83 b	2.83 в	2.63 c		
PDB Bio <sub>3</sub>	2.42 b	2.83 b	2.85 в	2.70 b		
Brad + PDB Bio.	2.83 a	3.54 a	5.45 a	3.94 a		
Mean	2.15 c	2.65 b	3.21 a			

Initial count =  $0.3 \times 10^3$  cells/g dry soil

Mean values in the same box sharing an alphabet are not significantly different.

Table (7): Influence of Bradyrhizobium japonicum, Bacillus megatherium strains (individual or mixture) into rhizosphere of Acacia species.

plantson organic matter percent in soil.

	Organic matter content %					
Treatment	Acacia viectoria	Acacia Coriaceae	Acacia saligna	Mean		
		Autumn	SCASOR			
Cont. Bio1	0.21 c	0.30 с	0.39 c	0.30 c		
Brad. Bio2	0.49 b	0.68 b	0.81 b	0.66 b		
PDB Bio,	0.54 Ь	0.66 b	0.81 b	0.67 b		
Brad + PDB Bio4	0.73 a	0.87 a	1.17 a	0,92 a		
Mean	0.49 с	0.63 b	0.80 a			
	Spring season					
Cont. Bio,	0.41 c	0.46 d	0.57 d	0.48 d		
Brad. Bio <sub>2</sub>	0.57 b	0.78 Ъ	0.94 в	0.76 b		
PDB Bio <sub>3</sub>	0.56 b	0.74 с	0.89 c	0.73 с		
Brad + PDB Bio4	0.83 a	1.05 a	1.39 a	1.09 a		
Mean	0.59 с	0.76 b	0.95 a			

Initial soil organic matter content 0.12.

Mean values in the same box sharing an alphabet are not significantly different.

Table (8): Influence of Bradyrhizobium japonicum, Bacillus megatherium strains (individual or mixture) into rhizosphere of Acacia species. plants on total nitrogen percent in soil during autumn and spring seasons (2001-2003).

	Total nitrogen %					
Treatment	Acacia viectoria	Acacia coriaceae	Acacia saligna	Mean		
		Autumn	season			
Cont. Bio1	0.028 d	0.028 c	0.051 c	0.036 d		
Brad. Bio <sub>2</sub>	0.034 b	0.030 в	0.064 b	0.034 b		
PDB Bio <sub>3</sub>	0.030 c	0.030 b	0.063 b	0.041 c		
Brad + PDB Bio <sub>4</sub>	0.057 a	0.071 a	0.081 a	0.069 a		
Mean	0.037 a	0.040 b	0.064 a	,		
	Spring season					
Cont. Bio1	0.0323 d	0.0343 d	0.0566 d	0.0404 d		
Brad. Bio <sub>2</sub>	0.0444 b	0.0626 b	0.0758 Ь	0.0609 ხ		
PDB Bio <sub>3</sub>	0.0424 c	0.0606 с	0.0737 c	0.0589 с		
Brad + PDB Bio <sub>4</sub>	0.0667 a	0.0838 a	0.0949 a	0.0818 a		
Mean	0.0460 c	0.0603 b	0.0752 a			

Initial soil total nitrogen 0.016 %

Mean values in the same box sharing an alphabet are not significantly different.

#### REFERENCES

- Abd El-Ghany F. Bouthaina (1996): Influence of different bacterial strains as biofertilizers on wheat crop production in new cultivated land. Desert Inst. Bull., Egypt 46, No. 2 (1996).
- Abdel-Ghany F. Bouthania,; Khalil, K.W.; El-Sersawy; M.M. and Awadalla, S.Y. (1997): Improvement of Wadi Sudr soil properties using modern Bioorganic techniques and their effects on desertification combat and barley production. Desert Inst. Bull., Egypt. 47, No. 1, pp. 69-100.
- Abd El-Hamid E.A. Amal, (2000): Effect of bacterization using rhizobia and phosphate dissolvers on the growth of some leguminous crops cultivated in desert soils. Ph.D. Thesis, Fac. of Agric., Ain Shams Univ.
- Abd El-Hamid, E.A. Amal (1995): Studies on the role of micro-organisms in utilizing phosphate in desert soils. M.Sc. Thesis, Fac. of Agric., Ain Shams Univ.
- Abdel-Rahman M. Mona (2003): Study on some factors affecting activity and productivity of growth regulators by azotobacters desert soil. M.Sc. Thesis, Fac. Sci., Al-Azhar Univ., Cairo, Egypt.
- Abo-Alaa, K.A. (2002): Biofertilization techniques used for improving production of some medicinal plants in desert soil. Ph.D. Thesis. Institute of environmental studies and research, Ain Shams University, Cairo, Egypt.
- Abou-Deya, I.B.; Nassar, Z.M. and Salem, M.O. (1990): Performance of *Acacia* saligna under rainfed conditions in the North-West Coast. Proc. 4<sup>th</sup> Conf. Agron. Cairo, 2: 671-678.

- Association of Official Agriculture Chemists (1970): Official Methods of Analysis, A.O.A.C. Washington, 11th Ed. D.C. 832 pp.
- Atef, K. and Nannipieri, P. (1995), Methods in Applied soil Microbiology and Biochemistry, Academic Press, Harcout Brace and Company, Publishers, London, PP 214-217.
- Basu, P.K. and McKabi, (1987): Effect of application of biofertilizers on the growth and nodulation of seven forest legumes. Indian-Forester. 113 (4): 249-257.
- Black, C.A.; Evans, D.O.; Ensminger, L.E.; White, J.L.; Clark, F.E. and Dinauer, R.C. (1982): Methods of soil Analysis. Part 2. Chemical and Microbiological properties. 2nd edition. Soil Science Society of America Inc. Publ., Madison, Wisconsin, U.S.A.
- Brown, J.D.; and Lilleland, O. (1946): Rapid determination of potassium and sodium in plant material and soil extracts by flamphotometry. Proc. Amer. Soc. Hort. Sci., 48: 341.
- Duncan, D.D. (1955): Multiple range and multiple F-tests. Biometrics II: 1'-42.
- Faid, E.Y. (1994): Studies on biofertilizer in desert soils. M.Sc. Thesis, Fac. of Agric., Ain Shams Univ.
- Gutteridge, R.C. (1991): Agronomic evaluation of tree and shrub species in South East Queensland. Fer. Abst., 52(3): 1366.
- Ibrahem, M.K. (1981): Shrubs for fodder production. Advances in food producing systems for arid and semi arid lands, National Agricultural Research Station, Kenya 601-642p.
- Ishac, Y.Z; El-Haddad, M.; El-Borollosy, M.A.; Racal, A.G. and Mostafa, M.I. (1985): Effect of organic acids and carbon dioxide on a symbiotic nitrogen fixation. Egypt. J. Microbial; Special Issue.,:1-11.
- Ishac, Y.Z.; El-Haddad, M.E.; Kherbawy; M.E.; Saleh; E.A.; El-Borollosy; M.A. and El-Demerdash; M.E. (1986): Effect of seed bacterization and phosphate uspplementation on wheat yield and mycorrhizal development. Proc. 2<sup>nd</sup> AABNF Conf. Cairo, Egypt, 597-610.
- Kandeel, S.A.; Kherallah, A.E. and Bourtham, B.E. (1994): Biomass utilization of juvenile *Acacia cyanophylla* Lindl. grown in Egypt. First International Symposumon Silviculture of Protection of Forestry in Arid Regions and the Agroforestry potential. ARC, NARP and USAID, Alexandria, 126-142pp.
- Khalil, K.W., El-Sersawy, M.M., Abd El-Ghani, B.F. and Hashem, F.A. (1991): Profitability of using some organic wastes with P fertilization on wheat production under saline irrigation water and Wadi Sudr conditions. Egypt. J. Appl. Sci., 6 (7), 267-284.
- Lugtenberg, B.J.J.; De-Wager, L.A. and Bennett, J.W. (1991): Microbial stimulation on plant growth and protection from disease. current Opinion in Biotechnlogy, 2:3, 457-464.
- Mighaed, H.A.; Amal, E.A. and Bouthaina, F.A. (2004): Effect of different bacterial strains as biofertilizer agents on growth production and oil of *Apium Graveolens* under calcareous soil. Arab Univ. J. Agric. Sci., Ain Shams Univ., Cairo, 12 (2), 2004.
- Page, A.L.; Miller, R.H. and Keeny, D.R. (1982): Methods of soil analysis, Part 2; 2<sup>nd</sup> Am. Spec. Agronomy, Inc., Mad Wisconsin, U.S.A.

- Peach, K. and Tracy, M.R. (1956): Modern Methods of Plant Analysis, Vol. 1 Springer Verlage, Berlin, 643 pp.
- Sheha, M.Y. (1984): Adaptation of plants in Kuait. Kuait foundation for Sci. Development, First Eddition.
- Skerman, P.L. (1977): Tropical range legumes. FAO Plant Production and Protection Publication, No. 2, 524 pp.
- Sneath, P.H.A. (1994): Gram-Negative aerobic microaerophilic rods and cocci. "Bergy's Manual of systematic Bacteriology, Group. 4: 78-125.
- Snedecor, G.W. and Cochran, W. (1980): Statistical Methods. 7<sup>th</sup> Ed., p. 507. Iowa Stat. Univ., Press, Ames, Iowa, USA.
- Subba Rao, N.S. (1988): Biofertilizers in Agriculture Oxford and TBH Pub. Co. Ltd., New Delhi, Bombay and Calcutta, 134-141.
- Taha, S.M.; Mahmoud, S.A.Z. and Mohbarek, M.S. (1970): Effect of reclamation of sandy soil on some chemical and microbiological properties. Plant and Soil, 32(2): 282-292.
- Taha, S.M.; Mahmoud, S.A.Z.; El-Damaty, A.H. and Abdel-Hafez, A.M. (1969): Activity of phosphate-dissolving bacteria in Egyptian soils. Plant and soil, 31: 149-160.
- Thalen, D.C.P. (1979): Ecology and utilization of desert shrub range lands in Iraq. Dr. W. Junk by. Publishers, The Hague.
- Tiedman, J.A. and Johnson, D.E. (1992): Acacia cyanophylla for forage and fuel wood in North africa. Agroforestry Systems, 17: 169-180.
- Topps, J.H. (1992): Potntial, composition and use of legume shrubs and trees as fodder for livestock in tropics. J. Agric. Sci. Cambridge. 118: 1-8.
- Tripathi, R.D.; Srivastava, G.P.; Misra, M.S. and Pandey, S.C. (1971): Protein content in some varieties of legumes. The Allah Abad Farmer, 16: 291-294.
- Zaght, M.F.; Tag El Din, S.S. and Elsh. Osama (1993): Guide for the important plants of the desert garden. Center of Research Studies Station, King Saud University Kingdom of Saudi Arabia, Riyadh.

دراسات على إنتاجية ونوعية بعض أنواع الاكاسيا تحت تأثير مصادر مختلفة من التسميد الحيوى بمنطقة وادى سدر

سيده عثمان محمد عبد الله "، سلوى على محمد الطوخي"، كرم محمود أحمد ابراهيم "، آمال السيد أحمد عبد الحميد " "

وحدة المراعى - مركز بحوث الصحراء - المطرية - القاهرة .

\* قسم خصوبة ميكروبيولوجيا الأراضى - مركز بحوث الصحراء - المطرية - القاهرة

أجرى هذا البحث لدراسة تأثير مصادر مختلفة من التسميد الحيوى ( مثبتات الروت تكميلية - بكستريا مديبة للفوسفات - مخلوط مثبتات الأروت + بكتريا مديبة للفوسفات - والمقارنة) وذلك على إنتاجية ونوعية بعض أنواع الآكاسيا تحت ظروف رأس سدر - محافظة جنوب سيناء خلال الفترة من خريف ٢٠٠١ حتى ربيع ٢٠٠٣ . وكانت أهم النتائج المتحصل عليها:

- ۱- أدى استخدام السماد الحيوى بكل أشكاله الى زيادة النشاط الميكروبي مقارنة بمعاملة الكنترول.
- ٢ أدى استخدام مخلوط مثبتات الأزوت التكاملية مع البكتريا المذيبة للفوسفات الى زيادة معنوية في صفات النمو الخضرية وبعض المكونات الكيميائية .
  - ٣ تفوقت الأكاسيا سالجنا على الأنواع الأخرى في معظم الصفات.
- ٤ تم الحصول على أعلى قيمة في معظم القياسات عند استخدام المخلوط الحيوى مع الاكاسيا سالجنا.
- آزیادة المدد الکلی للمیکروبات ومعدل تصاحد ثانی اکسید الکربون و احداد المیکروبات المذیبة للفوسفات و کذلك اعداد المیکروبات المحللة للسیلیلوز هوائیا و ذلك عند استخدام المخلوط الحیوی مع الاکاسیا سالجنا فی موسم الربیع .