

## HOST-PLANT EFFECTS ON THE KINETICS AND INHIBITION OF ACETYLCHOLINESTERASE IN THE COTTON WHITEFLY *BEMISIA TABACI* GENNADIUS.

Rawash, I.A.; El-Meniawi, Fatma, A.; El-Gayar, F.H. and Hussein, Hanaa, S.

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### ABSTRACT

The influence of two host-plants (tobacco and tomato) on the enzymatic kinetics and the inhibition of acetylcholinesterase (AChE) was investigated in the whole body homogenates of *Bemisia tabaci* male and female adults. The obtained results revealed that  $K_m$  values of AChE from both *B. tabaci* sexes reared on tobacco plants were significantly higher than those of tomato plants. It is noticed that,  $K_m$  values of tobacco culture increased to be 3.18 and 3.23 folds for females and males respectively when compared with the same sex from tomato culture. This may reflect a relative lower affinity of AChE from both sexes of tobacco culture to the substrate and in return lower activity than those of AChE from both sexes reared on tomato culture. On the contrary,  $V_{max}$  of females from tomato culture increased significantly to be about 1.3 folds of that of females from tobacco culture. The same trend was also recorded in the case of males but with no significant difference between  $V_{max}$  values of the two tested host-plants. In the same host-plant, males AChE were relatively lower in their affinity to the substrate than those of females. These latter findings may draw the attention to the criticism of not only segregated effect(s) of sex or host-plant alone but also to the combined sex: host-plant influence in this concern.

The in vitro inhibition of AChE by an organophosphorus insecticide, chlorpyrifos, and a carbamate insecticide, carbaryl, was investigated. The obtained results showed that, irrespective of carbaryl effect on males from tobacco culture, this insecticide showed to be the strongest inhibitor against AChE of both sexes of *B. tabaci* adults from tomato culture and only of females from tobacco culture. The lowest recorded  $I_{50}$  value for carbaryl was that against AChE from females reared on tobacco plants (0.13  $\mu$ M) followed by females from tomato culture (0.30  $\mu$ M) and then by males from tomato culture (0.71  $\mu$ M). The highest  $I_{50}$  value was that of chlorpyrifos against AChE from male adults of tomato culture (251  $\mu$ M) followed by that of females from tomato culture (56.2  $\mu$ M) by the same insecticide. Therefore, it could be concluded that, chlorpyrifos exerted a very weak inhibitory effect against AChE of both *B. tabaci* male and female adults reared on tomato plants. The obtained  $I_{50}$  values of chlorpyrifos, against *B. tabaci* adults from tobacco culture were very low (0.668 and 5.96  $\mu$ M) when compared to those of tomato culture. These findings revealed that chlorpyrifos caused its strong toxic effect on *B. tabaci* adults AChE reared on tobacco in comparison to AChE from *B. tabaci* tomato culture. This may reflect a host varying effect on AChE sensitivity to inhibition by chlorpyrifos. Moreover, within the same host-plant, AChE of *B. tabaci* female was more sensitive than that of males towards the two tested insecticides.

Generally, it could be concluded that the present study showed the existence of host-correlated variation phenomenon in *B. tabaci* AChE kinetics and inhibition by the two tested insecticides.

**Key words:** *Bemisia tabaci*, host-plant, Acetylcholinesterase, Kinetics, inhibition, insecticides.

### INTRODUCTION

The economic importance of whiteflies on the Egyptian agriculture has been recognized since 1930's (Priesner and Hosny, 1932), but viruliferous whiteflies have become very important pest during the last twenty five years. At least, twenty-one aleyrodid pests have been documented in Egyptian cropping system (Idriss *et al.* 1997). Three of them (*B. tabaci*, *B. argentifolii*, *Trialeurodis vaporariorum*) are able to transmit plant viruses (Bock *et al.*, 1974; Brown *et al.*, 1995a; McGrath and Harrison, 1995). Besides, *B. tabaci*, *B. argentifolii*, *Trialeurodis ricini* are vectors of plant geminiviruses (Idriss *et al.*, 1997). These geminiviruses are the largest and most economically significant group of plant viruses that cause devastating plant diseases to many crops in Egypt. In general, geminiviruses have become a significant group of plant diseases due to the modification of the ecology and other behavioural aspects of their natural vectors (Gerling, 1990).

For all the previously mentioned information about the increasing economic importance of the subject whitefly species in the Egyptian agriculture, and in the light of the previously recorded phenomena

of host-correlated variations in the subject whitefly (i.e. morphology, susceptibility, .....etc.), the present study was planned and achieved in order to gain some forward steps towards a successful integrated pest management (IPM) programme(s) of this pest. These phenomena clearly referred to the abruptive physiology of the whiteflies, and in return their promising, highly important role in enriching our knowledge of insect genetics and physiology.

The present study was conducted to evaluate the possible effects of host-plant on acetylcholinesterase (AChE) kinetics for both sexes of *B. tabaci* adults cultured on two different host-plants. Besides, experiments were carried out to investigate the host-correlated variations in relation with *B. tabaci* AChE inhibition by checking the potency of chlorpyrifos and carbaryl, as examples of OP- and carbamate- insecticides, respectively. Results of these investigations will surely add to the knowledge about insecticide susceptibility of the subject aleyrodid in consideration with host-correlated variations phenomenon. As AChE plays a key role in terminating excitatory neurotransmitter action in insect synapses (Gerschenfeld, 1973). This enzyme forms the primary site of action for the most widely used chemical

control agents in whitefly control, the systemic insecticides. Insects that used esterase to resist insecticides may alter and/or elevate AChE (Iwata and Hama, 1972; Devonshire and Moores, 1982; Moores *et al.*, 1988, French-Constant and Roush, 1990; Yu and Nguyen, 1992; Zhu *et al.*, 2000; Vontas *et al.*, 2001).

The present study along with all previous ones aimed to gain further knowledge and better understanding of the whitefly host-correlated variation phenomena in order to control this deleterious pest through suitable successful pest integrated management programmes (IPM) safe enough to man and environment.

## MATERIALS AND METHODS

### I- The whitefly *B. tabaci* Genn. Laboratory culture:

The subject insect has been taken from a laboratory culture of the whitefly *Bemisia tabaci* Gennadius, which has been first established by El-Helaly (1966) and which is still bred apart of any chemical treatments on tobacco plants *Nicotiana tabacum* in greenhouses at  $25\pm 7^\circ\text{C}$ ,  $65\pm 5\%$  RH and under natural light conditions. The identification of the mother culture was achieved by El-Helaly *et al.* (1971).

#### - The tomato culture of *B. tabaci* Genn.:

Adults of *B. tabaci* from the laboratory tobacco culture have been reared on tomato plants since 1999 in order to establish a new tomato culture to fulfil the requirements of present investigation about physiological host-correlated variations in AChE of the subject insect.

### II- Insect homogenate preparation:

Newly emerged *B. tabaci* adults [20 adults/0.3 ml buffer] of either sex (~14 and 20 ng body weight of males and females, respectively) were homogenized in ice cold 0.1M phosphate buffer, pH 7.0. Homogenization was achieved at low temperature by means of manually driven, glass minute homogenizer (1ml) especially developed for the whitefly adults.

### III- Determination of Acetylcholinesterase (AChE) kinetics:

Enzyme activity was measured using the spectrophotometric method of Ellman *et al.* (1961). The method is based on the hydrolysis of acetylthiocholine iodide (ATChI) as a substrate of AChE producing thiocholine iodide that reacts with 0.1M DTNB [5,5 dithiobis- (2-nitrobenzoic acid)] producing yellow color as function of the enzyme activity. The color intensity was measured spectrophotometrically at 412 nm as a rate of enzyme activity. The changes in absorbance were recorded at the beginning of the reaction and after 30 min. interval. The reaction mixture was kept at  $37^\circ\text{C}$ .

The affinity of AChE in whole body homogenates to the substrate were estimated using 0.1 to  $1 \times 10^{-3}$  M range of ATChI concentrations. The Lineweaver-Burk plot (L-B) was drawn by plotting  $1/v$  versus  $1/[S]$  where  $[S]$  is the molar concentration of (ATChI). Values of  $K_m$  (Micaeli's constant) and  $V_{max}$  (maximum velocity) were estimated from L-B regression lines.

Protein content in the whole body homogenates of the subject whitefly adults was assayed spectrophotometrically by the method of Lowry *et al.* (1951), at 750 nm wavelength, using bovine serum albumin (BSA) as a standard protein.

### Statistical analysis:

Values of  $K_m$  (the affinity) and  $V_{max}$  (the hydrolysing efficiency) were obtained by a least squares linear regression of double reciprocal plots of the points (Lineweaver and Burk, 1934). A *t*-test was adopted to compare the responses of AChE in both tested host-plants.

### IV- In vitro inhibition of AChE activity:

The inhibition of AChE activity was determined in both *B. tabaci* sexes from tobacco and tomato cultures using the organophosphorus insecticide, chlorpyrifos and the carbamate insecticide, carbaryl as inhibitors.

Estimation of  $I_{50}$  value (the concentration of the inhibitor which inhibit 50% of the enzyme activity) was carried out by preincubating the inhibitor with the enzyme (insect homogenate) at  $37^\circ\text{C}$  prior to the addition of Ellman's reagent (1961) (DTNB: at concentration of 0.1M & ATChI: at concentration of  $10^{-3}$  M).

Insect homogenate was preincubated with chlorpyrifos for 30 min and with carbaryl for 15 min. at  $37^\circ\text{C}$  before the substrate was added. The residual activity was then measured as described before in the determination of AChE kinetics. The inhibitors were used at  $10^{-7}$  to  $10^{-3}$  M concentrations. The percentage of the *in vitro* inhibition was calculated with respect to the activity in the absence of the inhibitor, using the equation of Ellman *et al.* 1961.

#### - Tested inhibitors:-

The organophosphorus insecticide, chlorpyrifos, (O,O-diethyl O-(3,5,6-trichloro-2-pyridinyl) phosphothioate) and the carbamate insecticide, carbaryl, (1-naphthalenyl methylcarbamate) were provided as technical grade insecticides from Frunol-chemie (W. Germany) and JinHung Fine Chem., Co. LTd Korea., respectively.

## RESULTS AND DISCUSSION

1- Host-plant effect on the AChE kinetics of *B. tabaci* adults.

The Kinetic studies were conducted to evaluate the possible effects of host-plant on AChE activity in both sexes of *B. tabaci* adults. This was conducted taking into consideration the previously observed host-correlated variation in the susceptibility of the subject insect to insecticides (El-Helaly, 1973; El-Meniawi, 1992; Anthony *et al.*, 1998).

The catalytic properties of AChE were estimated in whole body homogenates of male and female adults of two populations reared on tobacco and tomato plants. This was achieved by measuring reaction rates (V) over a range of ATChI concentrations, as a substrate (S), under optimum enzymatic activity conditions.

Fig.(1) shows the obtained Lineweaver-Burk (L-B) plots for both sexes of the subject insect adults from the two tested host plants. Table (1) summarizes the statistical analysis of the obtained values of  $K_m$  and  $V_{max}$  of AChE derived from all aforementioned L-B regression lines.

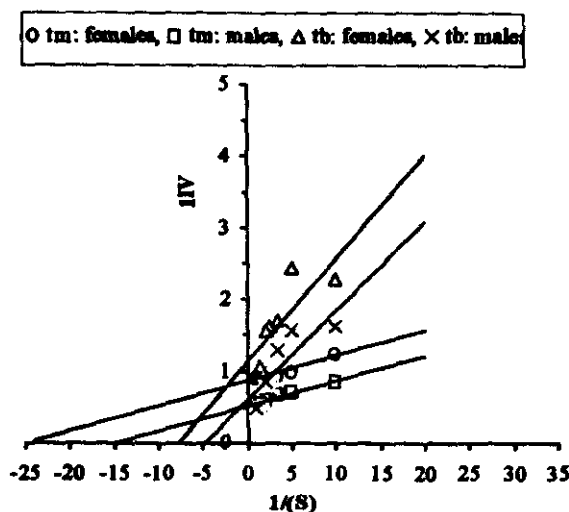


Fig.(1): Lineweaver-burk plots of AChE activities of *B. tabaci* female and male adults from the two tested cultures.

Tb-female: females from tobacco culture, Tb-male: males from tobacco culture, Tm-male: males from tomato culture, Tm-female: females from tomato culture,  $1/(S)$  = the inverse of ATChI concentrations in 0.1 to  $1 \times 10^{-3}$  M,  $1/v$  = the inverse of AChE activities in  $\mu$ M of ATChI.

Table (1): Host-plant effect on AChE kinetics of *Bemisia tabaci* adults from both tobacco and tomato cultures.

Host-plant	$K_m$		$V_{max}$	
	female	male	female	male
Tobacco	0.127	0.21	0.88	1.69
Tomato	0.04	0.065	1.15	1.89
t-calculated	16.82**	16.37**	6.44**	1.45 <sup>ns</sup>

$t_{0.05} = 2.776$ ,  $t_{0.01} = 4.604$

$K_m$ : Michaeli's constant= ATChI concentration  $\times 10^{-3}$  M

$V_{max}$ : Maximum velocity of the enzyme activity expressed as O.D.  $\lambda$  412  $\mu$ M/mg protein/min.

Statistical analysis of the obtained results revealed that  $K_m$  values of AChE from both *B. tabaci* sexes reared on tobacco plants were significantly higher than those from tomato plants. It is noticed that  $K_m$  values in tobacco culture increased to 3.18 and 3.23 folds for females and males respectively when compared with the same sex from tomato culture. This may reflect a relative lower affinity of AChE from both sexes of tobacco culture to the substrate and in return lower activity than those of AChE from both sexes reared on tomato culture.

On the contrary,  $V_{max}$  of females from tomato plants increased significantly to about 1.3 folds of that of females from tobacco culture (Table 1). The same trend was also recorded in the case of males but with no significant difference between  $V_{max}$  values of the two tested host-plants.

The shift of the Michaeli's Mentin constants was sufficient to explain the changes which occurred due to host-plant. The observed reduction in  $K_m$  values of either sex from tomato culture indicated that the enzyme acquired more affinity to ATChI. The  $V_{max}$  values of AChE in both sexes from tomato culture were raised referring to a probable increase in the number of active sites due to the effect of host-plant, but this needs further investigations. In other words, these findings are almost host than sex correlated.

Present results suggest that the two tested host-plants induced some alteration in the AChE nature in both sexes of the subject whitefly adults probably by increasing the active centers of the enzyme molecules. This alteration might lead to meet its defined physiological functions. Taking into consideration the probability of the presence of certain allelochemicals in each tested host-plant.

Moreover, the present results show that in the same host-plant, males AChE are relatively lower in their affinity to the substrate than those of females as  $K_m$  values in males exceeded those of females within the same culture. Therefore, it could be concluded that the kinetics of *B. tabaci* AChE were found to be different in either males and females of both tobacco or tomato cultures. This may refer to some probable

difference in the reproductive biotic potentials of either sex. In addition, *B. tabaci* AChE were found to have higher  $V_{max}$  and lower  $K_m$  values in tomato than tobacco host-plants. These latter findings may draw the attention to the criticism of not only segregated effect(s) of sex or host-plant alone but also to the combined sex: host-plant influence in this concern. In other words, the already noticed increasing effect of tobacco host-plant on female AChE  $K_m$  value compared with that of tomato females has been also noticed here for male AChE. Therefore, it could be concluded that irrespective of sex, host-correlated variation in AChE is pronounced in the subject aleoideid *B. tabaci* adults, and in return worth further investigation.

At this point of discussion, it could be clearly concluded that the obtained results proved the existence of both studied phenomena of sex- and host-correlated variations in the AChE kinetics of the subject whitefly *B. tabaci*. In addition, the recorded similar kinetical attitude in both sexes which may reflect probable effects for such kinetical attitude in relation to the subject whitefly toxicity should not be overlooked.

As far as the writer is aware, no information is available in the literature about the influence of host-plants on the AChE kinetics in Homopteran insects. However, kinetics of general esterase by using  $\alpha$ -naphthyl acetate as a substrate were found to be affected by host-plants in *Bemisia tabaci* (El-Meniawi, 1992); *Spodoptera littoralis* (El-Aw and Hashem, 2001); *Aonidiella aurantii* (Grafton-Cardwell et al., 2004). In addition, changes in the electrophoretic profiles of general esterase due to host-plant have been reported (Costa and Brown, 1991; Perring et al., 1992; El-Meniawi, 1992; Wool et al., 1993; Brown et al., 1995b).

## 2-AChE inhibition by organophosphate (OP) and carbamate (CR)- insecticides of *B. tabaci* tobacco and tomato cultures.

From the obtained  $I_{50}$  values (Table 2), remarkable inhibition of the enzyme activity by the two tested insecticides was observed with relatively varying degrees between the tested host-plants.

Table (2): The  $I_{50}$  ( $\mu$ M) values of the tested pesticides (chlorpyrifos and carbaryl) against AChE activity of *Bemisia tabaci* adults from both tobacco and tomato cultures.

Host-plant	Sex	Tested pesticides	
		Chlorpyrifos	Carbaryl
Tobacco	Female	0.668	0.13
	Male	5.96	31.6
Tomato	Female	56.2	0.30
	Male	251	0.71

In comparing the inhibition potency of the two tested insecticides, it is clear that, irrespective of carbaryl effect on males from tobacco culture, this insecticide showed to be the strongest inhibitor against AChE of both sexes of *B. tabaci* adults from tomato culture and only of females from tobacco culture. The lowest recorded  $I_{50}$  value (the highest inhibition potency) for carbaryl was that against AChE from *B. tabaci* tobacco females (0.13  $\mu$ M) followed in ascending order by females from tomato culture (0.30  $\mu$ M) and then by males from tomato culture (0.71  $\mu$ M). These results reflect that carbaryl had the highest inhibition potency against AChE from females of tobacco culture.

On the other hand, the highest  $I_{50}$  value was that of chlorpyrifos against AChE from male adults reared on tomato plants (251  $\mu$ M) followed by that of females from tomato culture (56.2  $\mu$ M) by the same insecticide. Therefore, it could be concluded that, chlorpyrifos exerted a very weak inhibitory effect against AChE of both *B. tabaci* male and female adults from tomato culture. Concerning the inhibitory potency of the OPs insecticide, chlorpyrifos, against *B. tabaci* adults from tobacco culture, it is clear that the obtained  $I_{50}$  values were very low (0.668 and 5.96  $\mu$ M) when compared to those of *B. tabaci* tomato culture (56.2 and 251  $\mu$ M). This may reflect a host varying effect on AChE sensitivity to inhibition by chlorpyrifos. These findings revealed that chlorpyrifos causing its strong toxic effect on AChE *B. tabaci* adults reared on tobacco in comparison to AChE from *B. tabaci* tomato culture. Such varying effect of the tested insecticides on *B. tabaci* AChE in respect to host-plant could be understood in the light of the results of El-Helaly, (1971 and 1973); El-Meniawi (1992); Anthony et al. (1998); Byrne and Devonshire (1997) about the host-correlated variations in the insecticide susceptibility.

Moreover, it was also noticed that, within the same host-plant, AChE of *B. tabaci* female was more sensitive than males towards the two tested insecticides (Table 2). As  $I_{50}$  values in the case of chlorpyrifos were 0.668, 56.2  $\mu$ M for females and 5.96, 251  $\mu$ M for males from tobacco and tomato cultures, respectively. Also, in the case of carbaryl,  $I_{50}$  values were 0.31, 0.30  $\mu$ M for females and 31.6, 0.71  $\mu$ M for males from tobacco and tomato cultures, respectively. These findings could be understood in the light of the highest total protein content in *B. tabaci* females when compared to those of males (Table 3).

Table (3): Means of protein content mg/ml as affected by the interaction between sex, and host-plant.

Host-plant	Female	Male
Tobacco	2.51 b $\pm$ 0.110	1.43 c $\pm$ 0.160
Tomato	3.57 a $\pm$ 0.104	1.49 c $\pm$ 0.191

L.S.D<sub>0.05</sub> = 0.19

Means followed by the same letter (s) are not significantly different according to L.S.D at 0.05 level of probability.

In discussion of the present findings on AChE inhibition with respect to changes in the kinetic parameters in the former item, it could be concluded that, the changes in  $K_m$  values between the two tested host-plants indicate change in the affinity of such esterase to ATChI. However, the tobacco culture had generally higher  $K_m$  values, or, in other words, had lower affinities to ATChI within the same sex. This indicates that the AChE affinity to ATChI may vary in response to changing the sort of host-plant under which *B. tabaci* adults were reared.

The greater affinity of tomato culture (or males than females within the same culture) enzyme for the substrate, as indicated by its lowest  $K_m$  values would have a greater protective influence on its interaction with tested insecticides than that expected for tobacco culture enzyme. If this reflects the situation *in vivo*, this greater affinity for substrate means that AChE should be partially protected from inhibitors by any substrate present in the synapse. This suggestion may be explained with the highest  $I_{50}$  values in the case of *B. tabaci* tomato culture when compared with tobacco culture. Acceptable or rejection of such latter comment surely needs further investigation. However, such latter comment looks acceptable in the light of the studies of Devonshire and Moores, 1984 and Byrne and Devonshire, 1997, who concluded that the low  $K_m$  value could be afforded some degree of protection from the weaker inhibitors such as monocrotophos. Taking into consideration the conclusion in the study of Byrne and Devonshire (1997), that increased release of Acetylcholine would also be expected in a hyperactive nervous system following OP- or carbamate- poisoning, and this might partially alleviate the inhibition even more. The study of Grafton-Cordwell *et al.* (2004) about the effect of host-plant tissue on the activity of inhibition of AChE in *Aonidiella aurantii* suggested that this insect is using increased amounts of esterase enzymes, including AChE, to sequester OP and carbamate insecticides, rather than modified AChE.

In general, the present study proved the existence of host-correlated variation phenomenon in AChE kinetic parameters and inhibition by the tested insecticides. In addition, the present investigation revealed the interaction between the effects of the differences in both sexes and host-plants.

#### REFERENCES

- Anthony, N.M.; Brown, J.K.; Feyereisen, R. and Ffrench-Constant, R.H. (1998). Diagnosis and characterization of insecticide insensitive acetylcholinesterase in three populations of the sweetpotato whitefly *Bemisia tabaci*. *Pesticide Science*. 52 (1): 39-46.
- Brown, J.K.; Frohlich, D.R. and Rosell, R.C. (1995a). The sweetpotato or silverleaf whiteflies: Biotypes of *Bemisia tabaci* or a species complex? *Annu. Rev. Entomol.* 40: 511-534.
- Brown, J.K.; S.A. Coats; I.D. Bedford; P.G. Markham; J. Bird; and D.R. Frohlich. (1995b). Characterization and distribution of esterase electromorphs in the whitefly, *Bemisia tabaci* (Genn.) (Homoptera: Aleyrodidae). *Biochemical Genetics*, 33(7/8): 205-214.
- Bock, K.R.; Guthrie, E.J. and Woods, R.D. (1974). Purification of maize virus and its relation to viruses associated with streak diseases of sugarcane and *Panicum maximum*. *Ann. Appl. Biol.* 77:289-296 (c.f. Gerling, D,1990).
- Byrne, Frank J. and Alan L. Devonshire. (1997). Kinetics of insensitive acetylcholinesterases in organophosphate- Resistant tobacco whitefly, *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae). *Pesticide Biochemistry and physiology*. 58, 119-124.
- Costa, H. S. and J. K. Brown.(1991). Variation in biological characteristics and esterase patterns among populations of *Bemisia tabaci*, and the association of one population with silverleaf symptom induction. *Entomol. Exp. Appl.* 61: 211-219.
- Devonshire, A.L. and Moores, G.D. (1982). A carboxylesterase with broad substrate specificity causes organophosphorous, carbamate, and pyrethroid resistance in potato aphids (*Myzus persicae*). *Pestc. Biochem. Physiol.* 18: 235-246.
- Devonshire, A.L. and Moores, G.D. (1984). Different forms of acetylcholinesterase in insecticide-resistant house flies (*Musca domestica*). *Pestc. Biochem. Physiol.* 21, 336-340.
- El-Aw, M. A. and M. Hashem. (2001). Effect of host plant and day time on the susceptibility of the cotton leafworm, *Spodoptera littoralis* (Boisd.), larvae to profenofos and on the kinetics of head esterases. *Alex. J. Agric. Res.* 46 (1): 115-129.
- El-Helaly, M.S. (1966). Studies on whiteflies. M.Sc. Thesis, Faculty of Agriculture, University of Alexandria, Egypt.
- El-Helaly, M.S.; El-Shazli, A.Y. and El-Gayar, F.H. (1971). Morphological studies on immature stages of *Bemisia tabaci* Gennadius (Homoptera, Aleyrodidae). *Z. ang. Ent.*, Bd. 68 (4): 403-408.
- El-Helaly, M.S. (1973). Further studies on the whiteflies. Ph.D. Thesis, Faculty of Agriculture, University of Alexandria, Egypt.
- Ellman, George L.; K. Diane Courtney; Valentino Andres, J. and Robert M. Featherstone. (1961). A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochemical Pharmacology*, 7, 88-95.

- El-Meniawi, Fatma, A. (1992). Physiological studies on the cotton whitefly *Bemisia tabaci* Genn. Ph.D. Thesis, Faculty of Agriculture, University of Alexandria, Egypt.
- Ffrench-Constant, R.H.; and Roush, R.T. (1990). Resistance detection and documentation: the relative roles of pesticidal and biochemical assays, pp. 4-38. In R.T.Roush and B.E.Tabashnik [ed], pesticide resistance in arthropods. Chapman and Hall, New York. (c.f. Grafton-Cardwell et al. 2004).
- Gerling, D., Ed. (1990). Whiteflies: their bionomics, pest status and management. Wimborne, UK. Intercept. 348pp.
- Gershenfeld, H.M., (1973). Chemical transmission in invertebrate central nervous system and neuromuscular junctions. *Physiol. Rev.*, 53:1-119. (c.f. Anthony 1998).
- Grafton-Cardwell, E.E.; Yuling O.; Rebecca A. T.; Julie, A.; Black, C.S. (2004). Role of esterase enzymes in monitoring for resistance of California red scale, *Aonidiella aurantii* (Homoptera: Diaspididae), to organophosphate and carbamate insecticides. *J. Econ. Entomol.* 97 (2): 606-613.
- Idriss, M.; Abdallah, N.; Aref, N.; Haridy, G.; Madkour, M. (1997). Biotypes of the castor bean whitefly *Trialeurodes ricini* (Mitra) (Hom., Aleyrodidae) in Egypt: biochemical characterization and efficiency of geminivirus transmission. *J. Appl. Ent.* 121, 501-509.
- Iwata, T.; and Hama, H. (1972). Insensitivity of cholinesterase in *Nephotettix cincticeps* resistant to carbamate and organophosphate insecticides. *J. Econ. Entomol.* 65: 643-644. (c.f. Grafton-Cardwell et al. 2004).
- Lineweaver, H. and Burk, D. (1934). The determination of enzyme dissociation constants. *J. Am. Chem. Soc.* 56: 658-666.
- Lowry, H.O.; Rosebrough, N.J.; Farr, A.L. and Ranball, R.J. (1951). Protein measurement with the folin phenol reagent. *J. Biol. Chem.* 193: 265-275.
- McGrath, P.F. and Harrison, B.D. (1995). Transmission of tomato leaf curl geminiviruses by *Bemisia tabaci* effects of virus isolate and vector biotype. *Ann. App. Boil.* 126: 307-316.
- Moore, G.D.; Denholm, L.; Byrne, F.J.; Kennedy, A.L. and Devonshire, A.L. (1988). Characterizing acetylcholinesterase genotypes in resistant insect populations. Proceedings of the Brighton Crop Protection Conference-Pests and Diseases, 451-456.
- Perring, T.M.; Cooper, A. and Kazmer, D.J. (1992). Identification of the poinsettia strain of *Bemisia tabaci* (Homoptera: Aleyrodidae) on Broccoli by electrophoresis. *J. Econ. Entomol.* 85(4): 1278-1284.
- Priesner, H. and Hosny, M. (1932). Contributions to a knowledge of the whiteflies (Aleyrodidae) of Egypt. (*Bull. Minist. Agric. Egypt tec.scient. serv.* 121: 1-8.
- Vontas, J.G.; Cosmidis, N.; Lukas, M.; Tsakas, S.; Hejazi, M.J.; Ayoutanti, A.; and Hemingway, J. (2001). Altered acetylcholinesterase confers organophosphate resistance in the olive fruit fly *Bactrocera oleae*. *Pestic. Biochem. Physiol.* 71: 124-132.
- Wool, D.; Gerling, D.; Bellotti A.C. and Morales F.J. (1993). Esterase electrophoretic variation in *Bemisia tabaci* (Genn.) (Hom., Aleyrodidae) among host plants and localities in Israel. *J. Appl. Ent.* 115:185-196.
- Yu, S.J.; and Nguyen, S.N. (1992). Detection and biochemical characterization of insecticide resistance in the diamond-back moth. *Pestic. Biochem. Physiol.* 44: 74-81. (c.f. Grafton-Cardwell et al. 2004).
- Zhu, K.Y.; Gao, J.; and Strarkey, S.R. (2000). Organophosphate resistance mediated by alterations of acetylcholinesterase in a resistant clone of the greenbug, *Schizaphis graminum* (Homoptera: Aphididae). *Pestic. Biochem. Physiol.* 68: 128-147. (c.f. Grafton-Cardwell et al. 2004).

## الملخص العربي

### تأثير النبات المائل على تنشيط وتثبيط إنزيم الأستيل كولين استيريز في حشرة ذبابة القطن البيضاء .*Bemisia tabaci* Genn.

إبراهيم عبده رواشن، فاطمة أحمد المنيلوي، فاروق حلمي الجبار، هناء صالح حسين  
قسم علم الحشرات الاقتصادية، كلية الزراعة، جامعة الإسكندرية

تم دراسة تأثير نوعان مختلفان من النباتات المائل (نخان وطماطم) على كل من تنشيط وتثبيط إنزيم الأستيل كولين استيريز، في متجانس أجسام الحشرات الكاملة لحشرة ذبابة القطن البيضاء وذلك لكل من الذكور والإناث. وقد أوضحت النتائج المتحصل عليها أن قيم  $K_m$  لهذا الإنزيم المستخلص من كل من ذكور وإناث الحشرة موضع الدراسة والمرباه على نبات النخان كانت تفوق معنوياً مثلثتها في الحشرات المرباه على نبات الطماطم. فقد لوحظ أن قيم  $K_m$  لكل من الإناث والذكور المرباه على النخان كانت تعادل ٣,١٨، ٣,٢٣ مرة قدر مثلثتها من نفس الجنس في حالة مزرعة الطماطم. وهذا ربما يمكن إنخفاض ميل الإنزيم للإرتباط بمادة التفاعل في كلا جنسي الحشرة المرباه على النخان بالمقارنة بتلك المرباه على الطماطم. وعلى العكس من ذلك، فقد كانت قيم  $V_{max}$  (المسرعة القصوى) في حالة الإناث المرباه على نبات الطماطم تفوق معنوياً بمقدار ١,٢ مرة قدر الإناث المرباه على النخان. وقد سجلت نفس الإستجابة في حالة الذكور ولكن كانت الفروق غير معنوية في قيم  $V_{max}$  بين مزرعتي النخان والطماطم. وبالنسبة لكل عائل نباتي على حدة، فقد أظهر إنزيم AChE في حالة الذكور ميلاً منخفضاً للإرتباط بمادة التفاعل بالمقارنة بالإناث من نفس المائل.

كذلك تم دراسة تثبيط إنزيم الأستيل كولين استيريز *in vitro* باستخدام مييد فوسفوري وهو الكلوربيريفوس Chlorpyrifos وآخر كاربماتي وهو الكارباميل Carbaryl. وقد أوضحت النتائج المتحصل عليها أنه بإستثناء تأثير الكارباميل على ذكور الحشرة من مزرعة النخان، فإن هذا المييد كان أكثر كفاءة في تثبيط AChE في كلا جنسي الحشرة من مزرعة الطماطم وكذلك للإناث في مزرعة النخان. وقد سجلت أقل قيمة للـ  $I_{50}$  (تركيز المييد اللازم لتثبيط ٥٠% من النشاط الإنزيمي) لمييد الكارباميل في حالة إناث مزرعة النخان (٠,١٣ ميكرو مول/لتر) بينما تصاعداً الإناث من مزرعة الطماطم (٠,٣٠ ميكرو مول/لتر) ثم الذكور من مزرعة الطماطم (٠,٧١ ميكرو مول/لتر). أما بالنسبة لمييد الكلوربيريفوس Chlorpyrifos فقد سجلت أعلى قيمة للـ  $I_{50}$  في حالة AChE المستخلص من ذكور الحشرات المرباه على الطماطم (٢٥١ ميكرو مول/لتر) بينما تنزلاً الإناث من مزرعة الطماطم (٥٦,٢ ميكرو مول/لتر). ومن هذه النتائج يمكن إستنتاج أن، الكلوربيريفوس أظهر تأثير تثبيطي ضعيف جداً على AChE لحشرات مزرعة الطماطم في كل من الذكور والإناث.

أما قيمة للـ  $I_{50}$  لمييد الكلوربيريفوس ضد الحشرات الكاملة المرباه على نبات النخان فقد كانت منخفضة جداً (٠,٩٦، ٠,١٦٨ ميكرو مول/لتر) وذلك بمقارنتها بالـ AChE من حشرات مزرعة الطماطم. ومن هنا فإن النتائج المتحصل عليها أثبتت أن الكلوربيريفوس له تأثير تثبيطي قوي على AChE للحشرات الكاملة المرباه على النخان بمقارنتها بتلك المرباه على الطماطم. وهذا يمكن إختلافاً مرتبطاً بالنبات المائل بالنسبة لحساسية هذا الإنزيم للتثبيط بالكلوربيريفوس.

كذلك أثبتت النتائج أنه على مستوى المائل الواحد فقد كانت الإناث أكثر حساسية من الذكور بالنسبة لكلا المبيدين المختبرين. هذه الدراسة أثبتت وجود ظاهرة الإختلافات المرتبطة بالمائل في القيم الحركية لإنزيم AChE في حشرة ذبابة القطن البيضاء وكذلك في تثبيط هذا الإنزيم بالمبيدين المختبرين.