BIOCHEMICAL BASES OF EGGPLANT SUSCEPTIBILITY TO SHOOT AND FRUIT BORER AND THEIR GENETIC CONTROL

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ABSTRACT

A series of experiments were conducted involving 70 genotypes to identify the source of resistance against shoot and fruit borer of eggplant and to understand the biochemical bases of resistance and with two distinct categories of genotypes: 3 least/moderately susceptible (lines) and 6 highly susceptible (testers) to determine their genetic control. The genotypes showed different susceptibility grades, 40 most susceptible, 13 highly susceptible, 9 susceptible, 7 moderately susceptible and 1 least susceptible. Highly significant positive correlations between sugar and protein contents and borer infestation and highly significant negative correlation between phenol content and borer infestation revealed the proposition of larval nonpreference and antibiosis mechanisms of resistance in the genotypes. Population means for shoot and fruit borer infestation, phenol and sugar contents in the fruits of the hybrids corresponded closely to the midparental values. It revealed much higher additive than dominance genetic variance for both shoot and fruit infestation. The inheritance was predominantly additive for phenol content and mainly dominance for protein content, while sugar content of the fruits was conditioned by additive and dominance variance, additive being greater in magnitude. Genotypes like HE 12. Pusa Purple Cluster having high phenois and lower sugar and protein contents in the fruits could effectively be utilized in breeding less susceptible eggplant varieties with good horticultural and cosmetic characters. The hybrids of such least/moderately susceptible x highly susceptible parents showed less susceptibility compared with the susceptible parent however, it will be essential to develop both the parental lines with higher phenol and lower protein and sugar contents in the fruits along with other good horticultural qualities to build up appreciable tolerance in the hybrids.

Key words: Eggplant, Shoot and Fruit borer

INTRODUCTION

The shoot and fruit borer (Leucinodes orbonalis) is the most serious insect-pest of eggplant (Solanum melongena) throughout India causing as high as 44% shoot and 100% fruit damage in severe infestation. The insecticide control method is relatively ineffective because of mode of damage, high operational cost and health hazard due to lasting pesticide residue in the fruits. Consequently, plant resistance would be useful either as a complete control measure or as a part of the integrated pest management programme with limited dependence on pesticides. Larval non-preference and antibiosis mechanisms operate in different crops for resisting the attack of many insect pests viz., shoot fly in sorghum (Dhawan et al 1993), tuber moth in potato (Das et al 1993), corn borer in maize (Williams et al 1997 and Kaur and Kanta, 2001), aphid in wheat (Havlickova 1996), brown plant hopper in rice (Soundararajan et al 2002), pod borer in cowpea (Oghiakhe et al 1993), aphid in mustard (Bhadauria et al 1996) and eggplant (Panda and Das 1975; Bajaj et al 1989 and Doshi 2004). Such resistance has reportedly been conferred on the host through over expression of phenolic compounds and down regulation of feeding stimulants for the insect pests like, free amino acids, protein, sugar contents..., etc. However, genetic control for the tolerance and the biochemical compounds conferring it in eggplant to shoot and fruit borer has not yet been reported. A series of experiments were thus conducted to identify the source of resistance against shoot and fruit borer of eggplant to understand the biochemical bases of resistance and to determine their genetic control towards framing strategies for resistance breeding.

MATERIALS AND METHODS

Seventy genotypes were grown in randomized block design with three replications for two consecutive years in autumn winter season (September-March) of 2001-2003 without any insecticide cover to allow natural infestation of shoot and fruit borer. Fortnightly observations were taken beginning 30 days after transplanting till the last fruit harvest was done. After recording the number of borer infested dead shoots, they were clipped off just above the point of insect burrow without destroying the larvae inside it. Single borer damage in the fruit was also considered as infested fruit. The percentage of borer infestation of both shoots and fruits were determined on number basis. Different biochemical compositions of fresh fruits of marketable maturity (15-

25 days after anthesis depending on the genotype) related to larval non-preference and antibiosis mechanism of host resistance were estimated from the sampled fruits of all the genotypes under second year screening (2001-2002) following standard methods: 1) total sugars by anthrone method (Dubois et al 1951) 2) crude protein through estimation of nitrogen by micro-kjeldahl method (Sadasivam and Manickam 1996), 3) total phenols by Folin-ciocalteau reagent method (Bray and Thrope 1954), and expressed on fresh weight basis. All these biochemical parameters were correlated with average shoot and fruit infestation over two years.

Three inbred genotypes viz. HE 12 (L₁), least susceptible, Pusa Purple Cluster (L₂) and Uttara (L₃), moderately susceptible containing low protein and sugar and high total phenol contents in the fruits (group 1) were crossed with 6 most susceptible inbred genotypes viz., BCB-1 (T₁), BCB-14 (T₂), BCB-15 (T₃), BCB-16 (T₄), BCB-72 (T₅) and BCB-87 (T₆) with high protein and sugar and very low total phenol contents in the fruits (group 2) in line x tester mating design. The 18 hybrids along with 9 parents were grown in autumn-winter season of 2003-04 to record natural infestation of shoot and fruit borer in them and to determine the protein, sugar and total phenol contents in the sampled fruits of marketable maturity with a view to elucidate their genetic control through data analysis following Kempthorne (1957).

RESULTS AND DISCUSSION

Field screening studies on fruit infestation suggested 40 genotypes as most susceptible showing infestation percentage to the tune and above the population mean (57.41%). By uniformly stepping down the population mean making four groups and subsequent clustering the genotypes in the respective groups, only one genotype HE12 (average 5.53 and 25.67% shoot and fruit infestation, respectively) could be grouped under tolerant/least susceptible category (Table 1). Of the 7 genotypes under moderately susceptible category, Pusa Purple Cluster (11.36 and 33.63% shoot and fruit infestation) and Uttara (12.18 and 35.17% shoot and fruit infestation) held promise for utilization in breeding programmes.

Table 1. Grouping of the 70 genotypes based on mean shoot and fruit infestation in two years.

	Sheet is	nfestation	Fruit infestation		
Group	No. of genotypes	Mean percent	No. of genotypes	Mean percent	
Least susceptible	1	5.53	1	25,67	
Moderately susceptible	4	12.02	7	36,98	
Susceptible	6	14.06	9	44.73	
Highly susceptible	18	15.84	13	52.13	
Most susceptible	41	17.65	40	64.04	

Marked differences for fruit compositions were recorded in the genotypes under different susceptibility grades (Table 2). Highly significant positive correlations between sugar and protein contents and borer infestation and highly significantly negative correlation between phenol content and borer infestation (Table 3) amply justified the proposition of larval non-preference and antibiosis mechanisms of resistance being operative in the genotypes. The least susceptible genotype, HE 12 contained 30.2 and 56.1% less protein and sugar, respectively and 67.13% higher phenol in the fruits than the most susceptible genotypes. Such association of biochemical characters with tolerance to insect-pest find support from Panda and Das (1975), Bajaj et al (1989) and Doshi (2004) for shoot and fruit borer in eggplant, Das et al (1993) for tuber moth in potato, Williams et al (1997) for corn borer in maize, Singh and Jotwani (1980) for shoot fly in sorghum.

Table 2. Fruit composition in the genotypes under different susceptibility category

-	Category of the genotypes					
Character mean	Least susceptible	Moderately susceptible	Susceptible	Highly susceptible	Most susceptible	
Fruit weight (g)	50.17	62.18	78,25	101.22	139.93	
Sugar content (g/100g fresh fruit)	1.48	2.10	2.51	2.79	3.62	
Crude protein (g/100g fresh fruit)	1.17	1.34	1.41	1.55	1,69	
Total phenol (mg/100g fresh fruit)	26.13	18.14	14,43	11.23	9.11	

Table 3. Correlations of different fruit compositions with fruit infestation

Characters	Correlation coefficients(r)			
Sugar content of fruit	0.86*			
Protein content of fruit	0.88*			
Phenol content of fruit	-0.59*			

The three lines (female parent) containing higher phenol and lower sugar and protein contents in the fruits and expressing less susceptibility to shoot borer satisfied their choice in such genetic analysis. High susceptibility of the 6 tester parents to the infestation of shoot and fruit borer did not vary much among themselves which indicated less likelihood of getting variable hybrids with respect to tolerance attributes.

Population means for shoot and fruit borer infestation, phenol and sugar contents in the fruits of the hybrids (Table 4) closely matched with the midparental values indicating the absence of dominance and suggesting that the number of genes involved for larval non-preference and antibiosis through expression of these biochemical factors was not many.

Table 4. Shoot and fruit borer infestation and biochemical composition in the fruits (per

100 g fresh) of the parents and hybrids.						
Parent/ hybrid	Protein (g)	Sugar (g)	Phenol (mg)	Shoot infestation (%)	Fruit infestation (%)	
Li	1.21	1.43	25.86	7.38	29,77	
\tilde{L}_2	1.26	1.72	21.19	11.36	33.63	
L_3	1.32	2.13	17.57	12.18	35.17	
$\overline{\mathbf{T_1}}$	1.77	3.89	8.16	18.16	62,43	
T ₂	1.72	3.75	9.29	18.29	64.35	
T ₃	1.81	3.92	8.14	17.93	60.18	
T ₄	1.79	3.83	9.28	18.64	73,10	
T ₅	1.66	3.69	9.22	18.38	65.94	
\tilde{T}_6	1.73	3.76	8.26	18.21	67.27	
$L_1 \times T_1$	1.47	2.17	17.49	9.59	43.54	
x T ₂	1.47	2.30	17.73	11.89	48.41	
x T ₃	1.52	2.21	17.17	11.87	51.76	
x T ₄	1.49	2.25	17.24	11.60	45.96	
x T ₅	1.45	2.25	17.59	12.06	46.19	
x T ₆	1.46	2.34	17.19	11.25	49.08	
$\mathbf{L}_{2} \times \mathbf{T}_{1}$	1.49	2.24	14.59	13.57	47.51	
x T ₂	1.50	2.13	15.15	12.17	46.45	
x T ₃	1.56	2.29	14.71	12,28	48.53	
x T ₄	1.55	2.15	14.78	13.18	51,70	
x T ₅	1.48	2.33	15.12	13.72	51.38	
x T ₆	1.49	2.21	14.55	12,96	45,28	
$L_3 \times T_1$	1.57	2.27	13.16	13.58	51.40	
x T ₂	1.54	2.21	14.22	14.47	52.17	
x T ₃	1.49	2.31	12.97	13.13	48.61	
x T ₄	1.47	2.37	13.22	13.14	47.68	
x T ₅	1.52	2.34	13.17	14.19	44.97	
x T ₆	1.55	2,36	12.88	13.32	53.21	

Additive $(\sigma^2 A)$ and dominance $(\sigma^2 D)$ variances calculated from the general combining ability (gca) and specific combining ability (sca) variances at F=1 (eggplant being self pollinated crop) revealed much higher additive than dominance genetic variance for both shoot and fruit infestation (Table 5) suggesting the importance of additive effect of the genes conditioning resistance through larval non-preference and antibiosis mechanisms. The inheritance was predominantly additive for phenol content and mainly dominance for protein content, while sugar content of the fruits was conditioned by additive and dominance variance, additive being greater in magnitude. Earlier reports of greater dominance effect for protein content in wheat (Dong et al 1995), higher additive variance for the control of protein content in lucerne (Guines et al 2002), predominant additive genetic control for tannin content in sorghum (Rodrigues et al 1998) and importance of additive gene effects for sugar content in sorghum stems (Cheng et al 1986) acceded to our findings. However, some other reports suggesting higher additive variance for the control of protein content in lucerne (Guines et al 2002) and cowpea (Hazra et al 1996) did not support the present findings.

Table 5. Estimates of combining ability and genetic component of variance

Character	Combining a	bility variances	Genetic component		Predictability
	gca	sca	$\sigma^2 A$	$\sigma^2 \mathbf{D}$	ratio
Shoot infestation	0.83	0.45	1.66	0.45	0.79
Fruit infestation	1.41	0.58	2.83	0.58	0.83
Protein content	0.15	1.57	0.29	1.57	0.16
Sugar content	0.77	1.16	1.53	1,16	0.57
Phenol content	2.92	0.81	5.85	0.81	0.88

The genotypes like HE 12, Pusa Purple Cluster having high phenols and low sugar and protein contents in the fruits could effectively be utilized in breeding less susceptible eggplant varieties with good horticultural and cosmetic characters. However, striking proper balance for these biochemical compounds is necessary because both bitterness and discolouration in the fruits increase with increasing total phenol content (Chadha et al 1990), although high phenol level of 26.13mg/100 g fresh did not impart bitternesses in the fruits of HE 12 indicating higher threshold level of phenol for bitterness.

Development of hybrids emerged as the best breeding strategy in eggplant because of the preponderance of allelic and non-allelic interaction for

fruit yield and yield components as revealed in several studies (Kathiria et al 1998; Vaghasiya et al 2000; Singh et al 2002 and Hazra and Roy, 2004). The hybrids of least/moderately susceptible x most susceptible parents in the present investigation showed less susceptibility to the borer compared to the most susceptible parent. However, it will be essential to develop both the parental lines with higher phenol and lower protein and sugar contents in the fruits along with other good horticultural qualities to build up appreciable tolerance in the hybrids.

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REFERENCES

- Bajaj K.L., D. Singh and G. Kaur (1989). Biochemical basis of relative field resistance of eggplant (Solanum melongena) to shoot and fruit borer. Veg. Sci., 16: 145-149.
- Bhadauria, N.S., S.S. Jakhmola, N.K.S. Bhadauria, and V.S. Bhadauria (1996). Reaction of mustard genotypes against aphid, Lipaphis erysimi (Kalt.) Adv. Plant Sci. 9: 139-142.
- Bray H.G. and W.V. Thrope (1954). Analysis of phenolic compound of interest in metabolism. Methods of Biochemistry Analysis, 1: 27-52.
- Chadha M.L., C.M. Sharma and K.L. Bajaj (1990). Inheritance of bitterness in brinjal (Solanum melongena L.) Indian J.Hort., 47: 244-249.
- Cheng, B.C., Q.Y.Liu, H. Jiang, and L.R. Liu (1986). Analysis of genetic effects for sugar content in sorghum stems. Acta Agronomica Sinica. 12: 39-42.
- Das, G.P., E.D.Magattona, , V.K. Raman, and C.B. Adalla (1993). Growth and development of the potato tuber moth, *Phthorimaea operculella* (Zeller), on resistant and susceptible potato genotypes in storage. Philipp. Entomol. 9: 15-27.
- Dhawan, P.K., S.P. Singh, A.N. Verma, and D.R. Arya (1993). Antibiosis mechanism of resistance to shootfly, *Antherigona soccata* (Rondani) in sorghum. Crop Res. (Hisar) 6: 306-310.
- Dong, J.Y., Z.D. Zhao, J.J. Lu, K.J. Gong (1995). Genetic analysis of protein content and sedimentation value in winter wheat grains. Acta Agronomica Sinica. 21: 330-333.

- Doshi K.M. (2004). Influence of biochemical factors on the incidence of shoot and fruit borer infestation in eggplant. Capsicum and Eggplant Newsl., 23: 145-148.
- Dubois, M., K.A. Gilles, J.K. Hamilton, Robers P.A. and F. Smith (1951). A calorimetric method for the determination of sugar. Nature, 168-167.
- Guines F., B. Julier, C. Ecalla, and C. Uygha (2002). Genetic control of quality traits of Lucerne (*Medicago sativa* L.) Aust. J.Agril. Res. 53: 401-407.
- Havlickova (1996). Differences in winter wheat cultivars in aphid infestation in relationship to biochemical characteristics. Rostlinna-Vyroba 42: 41.45.
- Hazra, P. and U. Roy (2004). Inheritance of fruit yield and its components in brinjal (Solanum melongena) through analysis of generation means. Bangladesh J.Genet. Biotechnol 5: 15-18.
- Hazra, P., P.K. Das, and M.G. Som (1996). Combining ability for a pod yield and seed protein in cowpea (Vigna unguiculata (L.) Walp.). Indian J. Genet. 56: 553-555.
- Kathiria K.B, M.H Vaghasiya, M.K Bhalala and K.M. Doshi (1998). A note on genetics of fruit yield components in two crosses of brinjal. Veg. Sci. 25: 199-200.
- Kaur, R. and U. Kanta (2001). Antibiosis mechanism of stem borer resistance in maize germplasm. Insect Environ. 7: 19-20.
- Kempthorne, O. (1957). An introduction to genetic statistics. XVII, John Wiley & Sons Inc., New York, 545 pp.
- Oghiakhe, S., W.A. Makanjuola, and L.E.N. Jackai (1993). Antibiosis mechanism of resistance to legume pod borer, Maruca testulalis Geyer Crop Res (Hisar) 6: 306-310.
- Panda N. and R.C. Das (1975). Antibiosis factor of resistance in brinjal varieties to shoot and fruit borer. South Indian Hort., 23: 43-48.
- Rodrigues, W.A., P.C. Magalhaes, F.G. Santos, G.A. Tosello, and F.G. Dos Santos (1998). Analysis of partial diallel crosses for tannin content in sorghum. Pesquisa Agropecuaria Brasileira. 33: 1079-1083.
- Sadasivam S. and A. Manickam (1996). Biochemical Methods. New Age International (P). Ltd. Published, (2nd Ed.) New Delhi, India.
- Singh, Major, G. Kalloo, M. Banerjee, and S.N. Singh (2002). Genetics of yield and its component characters in brinjal. Veg. Sci. 29(1): 24-26.
- Singh, S.P. and M.G. Jotwani (1980). Mechanism of resistance in sorghum to shortfly II. Antibiosis. Indian J.Entomol. 42: 353-360.
- Soundarajan, R.P., N. Chitra, and K. Gunathilagaraj (2002). Evaluation of antibiosis resistance to brown plant hopper, *Nilaparvata lugens* stal. in rice. J. Appl. Zool. Res. 13: 14-18.

- Vaghasiya, M.N., K.B. Kathiria, M.K. Bhalala and K.M. Doshi (2000). Gene action for yield and its components in two crosses of brinjal. Indian J.Genet. 60: (1): 127-130.
- Williams, W.P., F.M. Davis and J.A. Mihm (1997). Mechanisms and bases of resistance in maize to southwestern corn borer and fall armyworm. *Proc. Intern. Symp. Insect resistant maize: recent advances and utilization*. International maize and wheat Improvement Center, 27 Nov-3 Dec., 1994, 29-36.

الأسس الكيموحيوية لوراثة صفة الحساسية لحفار ساق وثمار الباذنجان ب.هازرا ، ر. دونا ، ت. روى ، أ.ك. ماندال و يو روى تسم محاصيل الخضر – ولاية البنغال الغربية – الهند

أجريت سلسلة من التجارب على سبعين تركيبا وراثيا من الباذنجان بفرض معرفة الأسس الكيموحيوية التي تتحكم في وراثة صفة المقاومة لحفار ساق وثمار الباذنجان. وقد أظهرت هذه الستراكيب الوراثية درجات مختلفة من الحساسية ، فقد وجد ٤٠ تركيبا وراثيا شديد الحساسية ، ١٣ تركيبا وراثيا علية الحساسية ، وتركيبا واحد أقل علية الحساسية ، ٩ تراكيب وراثية معتدلة الحساسية وتركيبا واحد أقل حساسية. ووجد ارتباط معنوى وموجب بين محتوى الثمار من السكر والبروتين ودرجة الإصابة ، كذلك وجد ارتباط سالب ومعنوى بين محتوى الثمار من الغينولات ودرجة الإصابة.

ولفهم الأسس الكيموحيوية لوراثة صفة المقاومة لحفار ساقى وثمار الباننجان تم التهجين بيسن مجموعتين من التراكيب الوراثية للباننجان ، المجموعة الأولى: استعملت كأمهات (Lines) وشسملت تراكيب وراثية معتدلة في درجة إصابتها والمجموعة الثانية إستعملت كآباء (Testers) وشملت ٦ تراكيب وراثية شديدة الحساسية في درجة إصابتها. ويدراسة صفة المقاومة للأبساء (٩) والسهجن (١٨) وعلاقة المقاومة بمحتوى السكر ، والبروتين والفينولات في الثمار ، وجد أن الجينات المضيفة كانت أكثر أهمية من الجينات السائدة في التحكم في صفة المقاومة ويتحكم الفعل المضيف للجينات فسي محتسوى الثمسار مسن الفينولات ، بينما يتحكم الفعل السيادي للجين في محتوى الثمار من البروتين ، بينما يتحكم كل مسن الفعل المضيف والسيادي في وراثة صفة محتوى الثمار من السكر.

أظهر التركيب الوراثى Pusa Purple Cluster, HE12 درجة معتدلة من المقاومسة ، وذلك راجع إلى محتوى الثمار العالى من الفينولات والمحتوى المنخفض من السكر والبروتين ، كما أن ثمار هذين التركيبين الوراثيين ذات مواصفات بستانية جيدة ، لذلك يمكن تحسين صفة المقاومة لحفار سلساق وثمار الباذنجان عن طريق استخدام هذين التركيبين الوراثيين في برامج مستقبلية للتربية.

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