MORPHOPHYSIOLOGICAL STUDIES ON EGYPTIAN COTTON PLANTS (Gossypium barbadense L.)

3. Effect of some growth regulators and some micronutrients on anatomical characteristics of cotton plants.

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ABSTRACT

The present investigation was carried out at Faculty of Agriculture Farm, during 1999 and 2000 growing seasons, Department of Agric. Botany, Faculty of Agriculture, Kafr El-Sheikh, Tanta University. The main objective of this investigation is: to study the effect of foliar spraying of the three PGRs *i.e.*, Pix, kinetin and Morphactin with two concentrations for each one (low and high) and three micronutrients *i.e.*, iron (Fe), zinc (Zn) and manganese (Mn) with one concentration of each one, and their combinations; on anatomical characteristics of cotton plants. The cotton cultivar was devoted in this study is Giza 86 an Egyptian long-staple.

The main obtained results may be summarized as follows

Pix, kinetin and morphactin applications significantly increased the stem diameter of cotton plants, thickness of epidermal cells, thickness of cortex layer (collenchyma and parenchyma). Also, significantly increased the diameter of vascular cylinder, thickness of pith, thickness of xylem tissue and No. of vascular bundles compared to the control. Kinetin applications significantly decreased the diameter of stem resulting in thinner stems compared to the control. Also, cortical layer thickness (collenchyma and parenchyma) significantly decreased compared to the control.. pix, kinetin and morphactin singly application significantly increased the thickness of blade and thickness of mesophyll compared to the control. Pix and kinetin singly applications significantly increased No. of vascular/midrib and diameter of xylem vessel, while decreased the thickness of xylem compared to the control. Morphactin applications significantly increased midrib zone thickness and thickness of xylem, while diameter of xylem vessel significantly decreased compared to the control.

Micronutrients applications had no significant effect on root and Leaf anatomy compared to the control. Micronutrients applications significantly increased the total thickness of cortex layer compared to the control.

combined treatments applications significantly increased diameter of the stem, thickness of cortex layer, thickness of pith, thickness of xylem tissue and diameter of xylem vessel and No. of vascular bundles compared to the control. Combined treatments applications especially included the iron (Fe) significantly increased thickness of blade, thickness of mesophyll, midrib zone thickness, thickness of xylem and diameter of xylem vessel compared to the control.

INTRODUCTION

Cotton in Egypt is considered the most important economical crop and represents the backbone of Agricultural income, having for many years played an outstanding role in the social economical and even the political life of the country. For this special situation all factors should be taken to improve its yield continuously among different factors affecting cotton yield micronutrients and plant growth regulators are considered most important factors influences cotton plant performance and determines its yield, micronutrients play an important role in the physiological and metabolic processes in cotton plants during different stages of growth.(Sharma et al 1988), (Azab et al 1992) and (Morgan 1980) emphasized that application of growth regulators to major field crops is relatively rare and that the beneficial dose not always involve direct increase in yields. The use of growth retardants such as Pix and Morphactin can cause large, mesophyll cells and elongatied palisade cell and increased the diameter of xylem vessels.number of bundles/midrib. (Gausman et al. 1980),(El-Nady 1994) and (Ali 1994). Kinetin at (10 mg/L) decreased thickness of the cortex then the stem was thinner of lupine plants but kinetin increased diameter of xylem vessels. (Ibrahim et al. Kinetin also decreased diameter of stem, however, the kinetin application also caused thickening of mesophyll tissue due to elongation of palisade cells and size of spongy tissue, on the other hand, kinetin also increased both diameter of xylem vessels in vascular bundles and number of vascular bundles/midrib of pepper plants(Shabaan 1996),. Therefore, the main objective of this investigation are: to study the foliar spraying effect with some plant growth regulators; PGRs (Pix, Kinetin and Morphactin), micronutrients (Fe, Zn, and Mn) and their combinations on anatomical characteristics (anatomical studies) for an Egyptian long-staple cotton cultivar, Giza 86 (Gossypium barbadense L.) during 1999 and 2000 growing seasons.

MATERIALS AND METHODS

The present investigation was conducted at the Faculty of Agriculture Farm, throughout the Department of Agricultural Botany, Faculty of Agriculture, Kafr El-Sheikh, Tanta University, during the two successive growing seasons 1999 and 2000. The study includes the effect of three plant growth regulators (PGRs) as well as, three micro-elements and all possible combinations of them with different concentrations and various times of application on anatomical characteristics of an Egyptian long-staple cotton cultivar, Giza. 86 (Gossypium barbadense L.). Cotton seeds were sown on April 18th and April 14th in 1999 and 2000seasons, respectively. Mechanical analysis and chemical composition of the soil of the experiment are presented in Table (1) and (2)

Table (1): Mechanical and chemical analysis of soil sample of the experiment during 1999 and 2000 growing seasons.

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Seasons	Soil	mechanical an	pН	Water table (cm)	
	Sand	Silt	Clay		(5)
1999	23.40	35.50	41.10	8.0	85
2000	24.10	35.20	40.70	7.8	87

Soil chemical analysis 1999

Depth		Anions	(meq/L)		Cations (meq/L)				EC	
(cm)	CO ₃	HCO ₃	Cľ	SO ₄ "	Ca ⁺⁺	Mg ⁺⁺	Na ⁺	K ⁺	dS m ⁻¹	
0-30		2.40	90.68	76.94	42.08	39.52	88.00	0.42	12.00	
30-60	-	2.60	23.25	40.05	12.38	14.02	39.25	0.25	5.20	
60-90	-	1.60	34.88	88.39	29.70	27.90	67.00	0.27	8.80	
90-120		1.80	32.55	47.12	14.75	16.35	50.00	0.27	6.40	

Soil chemical analysis 2000

Depth		Anions	(meq/L)		Cations (meq/L)				EC
(cm)	CO ₃	HCO ₃	Cl	SO ₄	Ca⁺⁺	Mg ⁺⁺	Na⁺	K ⁺	dS m ⁻¹
0-30	•	3.10	25.74	42.17	17.29	10.93	39.25	0.44	7.12
30-60	-	2.40	10.89	56.41	12.74	11.44	45.25	0.27	5.81
60-90	-	2.00	13.86	81.14	20.02	13.18	63.50	0.30	7.56
90-120	-	1.50	15.35	69.29	20.39	8.95	56.00	0.26	7.66

Table (2): Chemical analysis of experimental soil sample for microelements

during 1999 and 2000 growing seasons.

Season ·	Depth	Micronutreints of soil profiles (ppm)					
	ĺ	Fe	Zn	Mn			
1999	0-30	12.50	2.35	31.00			
1	30-60	14.00	2.85	32.30			
1	60-90	12.50	3.35	35.15			
}	90-120	14.00	1.10	33. 85			
2000	0-30	8.46	0.44	7.02			
ļ	30-60	7.27	0.29	7.97			
	60-90	5.83	0.32	7.71			
_ '	90-120	8.58	0.34	9.52			

Field Drainage Res. Dept., Sakha Agric. Res. Sta.

Table(3) Studies treatments and time of application

Treatment	Concentration	Time of Application
Control (Water)	0	
PGRs:-		
Pix1 (P1)	1000 ppm	PGRs were sprayed twice.
Pix2 (P2)	2000 ppm	The first spray was at 75 days from
Kinetin 1 (k1)	25 ppm	sowing.
Kinetin 2 (k2)	50 ppm	The second at 105 days from sowing.
Morphactin1(Mor.1)	10 ppm	
Morphactin 2(Mor. 2)	20 ppm	
Micronutrients:-		
Fe	2000 ppm	Micronutrients were sprayed three times:-
Zn	1500 ppm	The first spray was at 75 days from
Mn	1000 ppm	sowing.
PGRs+Micro.comb.		The second spray at 90 days from sowing.
P1+ Fe	.	The third spray at 105 days from sowing.
P1 + Zn	. 1	
P1 + Mn	İ	
P2 + Fe		*Sowing date in 1999 season:-April, 18.
P2 + Zn	1	*Sowing date in 2000 season:-April,14.
P2 + Mn		
kl + Fe		
kl + Zn	1	1
kl + Mn	ļ	
k2 + Fe		1
k2 + Zn		Ì
k2 + Mn		
Mor.1 + Fe		
Mor.1 + Zn		j
Mor.1 + Mn		· ·
Mor.2 + Fe Mor.2 + Zn		
Mor.2 + Zn Mor.2 + Mn	·	

The experiment included 28 treatments as follows in Table(3) The treatments were randomly distributed in 28 experimental plots in each replicate of the randomized complete block. Experiment with three replications. Each plot consisted of five rows 4.5 m in length and 0.6 m in width.

The effect of growth regulators and micronutrients on the anatomical structure of leaves, stem and root were studied after 80 days after sowing. Specimens 1 cm long were taken from the fourth upper internode and the fourth upper leaf including the midrib. The different samples were fixed 48 hours in FAA (10 ml formaline, 5 ml Glacial acid, 50 ml ethyl alcohol absolute and 35 ml distilled water). Dehydrated and cleared in a tertiary butyl series, and embedded in paraffin wax 56 c m.p. Sections (20 microns thick) were cut using a Rotary Microtom (AO Rotary Microtom Model 820 apparatus, stainedwith safranine-Light green and mounted in Canda Balsam(Ghamrawy and Zaher, 1953)

The Statistical analysis of variance for randomized complete block design was carried out for each character in each season as outlined by Snedecor and Cochran (1980). The differences between the different treatment combinations were tested using the Duncan's Multiple Range method outlined by (Leclerg et al. 1962).

RESULTS AND DISCUSSION

1. Anatomical studies:

1.1Effect on the stem structure:

The micrographs in Fig. (1) and the data in Table (4) showed that foliar spraying treatments with PGRs, micronutrients and their combinations had a significant effect on stem anatomy of cotton plant and there were significant differences between these treatments and the control.

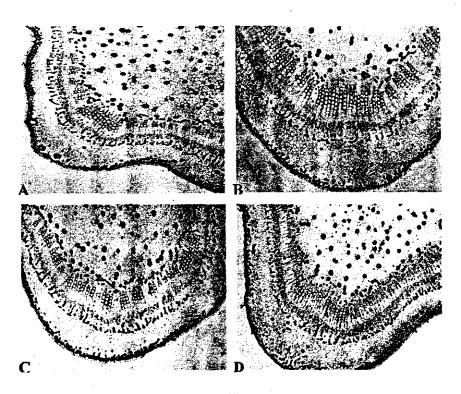
1.1.1Effect of Pix:

In cross sections all pix treatments (1000 and 2000 ppm) significantly increased the stem diameter of cotton plants as compared to the control. Thickness of epidermal cells show a tendency to slight increase at the low concentration of pix (1000 ppm) as compared to the control. It is also clear from the data (Table 4) and the micrographs in Fig. (1) that increasing pix concentration significantly increased diameter of stem and thickness of epidermal cell.

Table (5): Stem structure (x 100) of cotton plants as influenced by foliar spraying treatments with PGRs, micronutrients and their combinations.

n:	Thickens of	Thickness of cortex layer (µ)			Diameter of	No. of	Xylem tissue	
Diameter of Stem (µ)	epidermal cell (µ)	Collenchyma tissue	Parenchyma tissue	Total	vascular cylinder (μ)	vascular bundles	Thickness	Diameter of vessel (μ)
								25.00
4936.67 cd	20.0 fgh	225.0 gh					1	
5106.67 bc		256.67 efg			l i		1	
3240.00 m	20.00 fgh	250.00 fgh	100.00 k			-		
3256.67 m	20.00 fgh	225.00 gh	133.33 hij	· •		•	,	35.00 a-
4862.67 d	23.00 bc	258.33 efg	150.00 e-h				•	. 23.33 f
5068.33 bc	25.00 a	271.67 b-f	175.00 a-d	446.67 g	4125.0 bc	22.00 bcd	[276.67 a	25.00 f
							l	21.67.4
3781.33 jk	19.00 gh	275.00 b-f						
3818.00 jk	19.00 gh	281.67 a-f	150.00 e-h					
3745.00 k	18.33 h	291.67 a-e	150.00 e-h	441.67 h	2825.0 lm	19.00 fgh	186.67 gh	31.67 de
1	•			1				
4331.67 gh	20.00 fgh						1	33.33 b-
4375.00 fgh	22.33 bcd	255.00 ef	160.00 de				1 1	36.67at
4212.33 h	22.00 cde	278.33 b-f				_	ł I	
5208.33 b	25.00 a	308.33 ab	175.0 a-d					
5405.00 a	25.00 a	300.00 a-d	190.0 a					38.33 a
4228.00 h	24.00 b	275.00 b-f	156.67 d-g					
3490.001	20.00 fgh	250.00 fgh	104.33 k	354.33 q	2750.0 mn	-		
3782.33 jk	22.00 cde	216,67 h	131.67 hij	348.33 г	3041.67 jk	-	1	38.33 a
		225.0 gh	130.0 ij	355.00 q			, ,	31.67 d
		241.67 fgh	141.67 f-i	383.33 p	3125.0 ej			33.33 b
4045.00 i	20.00 fgh	256.67 efg	141.67 f-i	398.33 m	3208.33 hi			36.00 al
3853.00 ik	19.00 gh	265.00 def	121.67 j	386.67 no	3041.67 jk			33.33 b-
	-	306.67 abc	158.33 def	465.00 d	3541.67 e		1 :	30.00 e
		300.0 a-d	140.0 g-j	440.00 hi	3750.0 d			30.00 e
4456.67 fe			170.0 bcd	461.67 e	3500.0 ef		4 1	30.00 e
	1)		445.00 g	4041.67 bc	17.33 hij	246.67 b	30.00 e
1				_	4041.67 bc	21.33 cde	285.00 a	30.67 e
1 1						23.33 ab	250.00 b	30.00 6
		l .	l i					30.00 e
	_						222.02	32.54
	Stem (µ) 4936.67 cd 5106.67 bc 3240.00 m 3256.67 m 4862.67 d 5068.33 bc 3781.33 jk 3818.00 jk 3745.00 fgh 4212.33 h 5208.33 b 5405.00 a 4228.00 h 3490.00 I 3782.33 jk 3208.67 m 3931.67 jj 4045.00 i 3853.00 jk 4517.00 ef 4640.00 e 4456.67 fe 4971.67 cd 5038.33 bc 4987.67 cd 3724.67 k	Stem (µ) 4936.67 cd cell (µ) 4936.67 cd 20.0 fgh 5106.67 bc 25.00 a 3240.00 m 3256.67 m 4862.67 d 20.00 fgh 20.00 fgh 20.00 fgh 3256.67 m 4862.67 d 20.00 fgh 4862.67 d 25.00 a 3781.33 jk 19.00 gh 19.00 gh 3745.00 k 18.33 h 4331.67 gh 4212.33 h 22.00 cde 5208.33 b 5405.00 a 25.00 a 22.00 cde 5208.33 b 22.00 cde 5208.33 jk 22.00 cde 328.67 m 20.00 fgh 3931.67 ij 4045.00 i 20.00 fgh 3931.67 ij 4045.00 i 20.00 fgh 3853.00 jk 4517.00 ef 4640.00 e 4456.67 fe 420.00 cde 4300 bc 4971.67 cd 20.33 efg 5038.33 bc 25.00 a 3724.67 k 19.00 gh 4295 44 21.55	Stem (μ) epidermal cell (μ) Collenchyma tissue 4936.67 cd (μ) 20.0 fgh 25.00 a 256.67 efg 3240.00 m 20.00 fgh 225.00 gh 4862.67 d 23.00 bc 5068.33 bc 25.00 a 271.67 b-f 25.00 a 256.67 efg 25.00 gh 225.00 gh 225.00 gh 225.00 gh 271.67 b-f 3781.33 jk 19.00 gh 3745.00 k 19.00 gh 3745.00 k 18.33 h 291.67 a-e 273.33 b-f 25.00 a 271.67 b-f 4331.67 gh 4375.00 fgh 4212.33 h 22.00 cde 4212.33 h 22.00 cde 5208.33 b 25.00 a 308.33 ab 5405.00 a 25.00 a 300.00 a-d 4228.00 h 24.00 b 275.00 b-f 3490.00 l 20.00 fgh 250.00 fgh 3782.33 jk 22.00 cde 216.67 h 20.00 fgh 250.00 fgh 250.00 fgh 3931.67 ij 20.00 fgh 250.00 fgh 256.67 efg 3853.00 jk 19.00 gh 265.00 def 4456.67 fe 23.00 bc 4640.00 e 21.00 def 300.0 a-d 4250.00 e 21.00 def 4456.67 fe 23.00 bc 4456.67 fe 23.00 bc 291.67 a-e 4971.67 cd 20.33 efg 281.67 a-f 5038.33 bc 25.00 a 320.0 a 4987.67 cd 23.33 a 275.00 b-f 3724.67 k 19.00 gh 266.67 c-f 4295.44 21.55 268.63	Stem (μ) epidermal cell (μ) Collenchyma tissue Parenchyma tissue 4936.67 cd (μ) 20.0 fgh 25.00 gh 256.67 efg 180.0 abc 180.0 abc 180.0 abc 25.00 gh 250.00 fgh 100.00 k 13256.67 m 20.00 fgh 225.00 gh 133.33 hij 1862.67 d 23.00 bc 258.33 efg 150.00 e-h 175.00 a-d 271.67 b-f 175.00 a-d 175.00 gh 271.67 b-f 175.00 a-d 175.00 gh 271.67 b-f 175.00 a-d 175.00 fgh 271.67 a-e 150.00 e-h 150.00 e-h 150.00 e-h 150.00 e-h 150.00 e-h 150.00 ab 160.00 de 160.00 de 160.00 de 175.00 ab 175.0 a-d 175.0 a-d 175.0 ab 17	Stem (μ) epidermal cell (μ) Collenchyma tissue Parenchyma tissue Total 4936.67 cd 5106.67 bc 5106.67 bc 3240.00 m 20.00 fgh 3250.00 fgh 3256.67 m 20.00 fgh 4862.67 d 23.00 bc 5068.33 bc 25.00 a 271.67 b-f 180.0 abc 133.33 hij 385.33 op 4862.67 d 23.00 bc 258.33 efg 150.00 e-h 408.33 m 446.67 g 3781.33 jk 19.00 gh 19.00 gh 3745.00 k 18.33 h 291.67 a-e 175.00 a-d 150.00 e-h 41.67 h 4331.67 gh 4375.00 fgh 421.33 h 22.00 cde 4212.33 h 25.00 a 25.00 a 25.00 a 25.00 a 308.33 ab 175.0 a-d 4228.00 h 24.00 b 275.00 b-f 160.00 de 438.33 ij 360.00 a 4228.00 h 24.00 b 275.00 b-f 160.00 de 438.33 ij 3782.33 jk 22.00 cde 278.33 b-f 160.00 de 438.33 ij 3782.33 jk 22.00 cde 278.00 b-f 156.67 d-g 431.67 k 3490.00 l 20.00 fgh 250.00 fgh 104.33 k 354.33 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Means designed by the same letter at each cell are not significantly different at the 5% level according to duncan's multiple rang test



Fig(1)Cross sections of cotton stem illustrating the effect of pix, kintineAnd morphactin(x100)

A-control C-kinetin(50ppm) B-pix(2000ppm) D-morphactin(20ppm)

Concerning the cortical layers thickness, it is apparent that the Collenchyma tissue is represented by 55.24% of the cortex layer at the low concentration of pix (1000 ppm), while it is represented by 58.78% of the cortex layer at the higher concentration of pix (2000 ppm), compared to 65.31% for Collenchyma layer in the control. This indicates that pix applications (1000 and 2000 ppm) compressed the outer Collenchyma region by about 10.07% and 6.53% less than the control, respectively, but increased the size of cortex layer cells in high pix application compared to the (control). Similar results were obtained by Gausman (1986) on cotton and El-Nady (1994) on eggplants.

Also, pix applications (1000 and 2000 ppm) significantly increased the diameter of vascular cylinder, No. of vascular bundles, thickness of xylem tissue and diameter of vessel as compared to the control.

1.1.2Effect of kinetin:

The micrographs in Fig. (1) and the data in Table (4) showed that kinetin applications (25 and 50 ppm) significantly decreased the diameter of stem resulting in thinner stems compared to the nonkinetin-treated cotton plants (control). Thickness of epidermal cells show a tendency to slight increase at the two concentrations of kinetin as compared to the control. It is also clear that cortical layer thickness including both Collenchyma and Parenchyma tissues significantly decreased compared to the control. Also, kinetin applications significantly reduced the diameter of vascular cylinder, No. of vascular bundles in low concentration only and thickness of xylem tissue. On the other hand, xylem vessel significantly increased compared to the control, these results are in agreement with those obtained by Ibrahim et al. (1990) on lupine and Ateya (2001) on soybean. The decrease of stem diameter may be due to that kinetin may increase elongation of cell wall modification (increased plasticity) and decrease diameter of cell.

1.1.3Effect of morphactin:

As shown from the micrographs in Fig. (1) and the data in Table (4) morphactin treatments significantly increased stem diameter, thickness of epidermal cells, thickness of cortex layer including both Collenchyma and Parenchyma tissues, diameter of vascular cylinder, No. of vascular bundles and thickness of xylem tissue as compared to the non-morphactin treated plants (control). Visually, the morphactin and non-morphactin treatments were easily distinguishable by the swollen stems in the first case.

Concerning the diameter of xylem vessel, it appeared more narrower than the control. These results were in harmony with those obtained by Dierig and Backhans (1990) on Guayule (Rubber), Ali et al. (1994) on vicia faba, El-Nady (1994) on eggplant and Shabaan (1996) on sweet pepper.

1.1.4Effect of micronutrients:

Table (4) showed that the micronutrients applications did not significantly differ from the control with regard to stem diameter, thickness of epidermal cells, diameter of vascular cylinder, No. of vascular bundles, thickness of xylem tissues and diameter of xylem vessels.

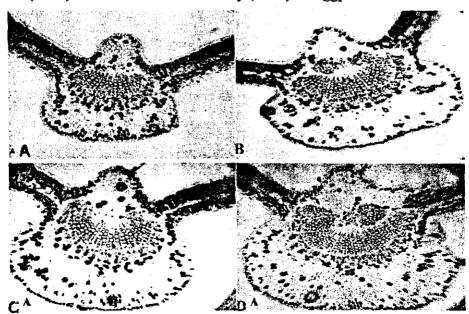
1.1.5Effect of PGRs + micronutrients combinations:

As shown from the data in Table (4) all combined treatments including pix or morphactin or kinetin at high concentration (some cases) plus one microelements (Fe, Zn, or Mn) significantly increased diameter of stem, thickness of cortex layer including both Collenchyma and Parenchyma, diameter of vascular cylinder, No. of vascular bundles, thickness of xylem tissue and diameter of xylem vessels. The highest values of stem anatomy were in favour of the combined treatment any PGR plus Zn.

1.2 Effect on the leaf structure:

1.2.1 Effect of pix:

As shown from the micrographs in Fig. (2) and the data in Table (5), it is apparent that pix applications (1000 and 2000 ppm) significantly increased the thickness of blade, thickness of mesophyll tissues compared to the control. Cross section and the data also showed that pix applications significantly increased No. of vascular/midrib and diameter of xylem vessel, while decreased the thickness of xylem. Similar results were reported by Gausman et al. (1980) on vicia faba and El-Nady (1994) on eggplant.



Fig(2)Cross sections of cotton leaf illustrating the effect of pix, kintine And morphactin(x100)

A-control C-kinetin(50ppm) B-pix(2000ppm) D-morphactin(20ppm) Table (6):Leaf structure (x 100) of cotton plants as influenced by foliar spraying treatments with PGRs, micronutrients and their combinations

spraying treatments with PORS, interonutrients and their combinations										
1	Thickness	Thickness of		Vascular bundle						
Treatments	of blade	mesophyl	tissue µ							
ļ	μ	Palisade	Spongy	No. of	Thickness	Diameter/				
i i				vascular/	of xylem	Vessel				
				midrib	μ	μ]				
PGRs		,								
Pix 1	271.67 ab	110.00 ab	118.33 abc	21.67 b-e	170.00 gh	30.00 bcd				
Pix 2	271.67 ab	100.00 c-f	121.67 a	22.67 bc	163.33 hui	36.67 a				
Kinetin I	230.00 gkl	90.00 g-j	108.33 de	16.33 klm	171.67 gh	30.00 bcd				
Kinetin 2	243.33 g-j	90.00 g-j	111.67 bcd		155.00 lj	33.33 ab				
Morphactin 1	251.67 e-h	95.00 d-h	100.0 ef	19.00 f-i	203.33 def					
Morphactin 2	251.67 e-h	93.33 f-i	123.33 a	25.00 a	196.67 ef	26.67 de				
Micro-nutrients:										
Fe	215.00 mn	83.33 jkl	96.67 f	16.00 lm	176.67 g	30.00 bcd				
Zn	213.33 mn	76.67 1	100.00 ef	15.33 m	165.00 hi	30.00 bcd				
Mn	213.33 mn	80.00 kl	98,33 f	16.67 j-m	17167 gh	30.00 bcd				
PGRs + Micr. comb.	'		1		- , <u> </u>	00.00				
P ₁ + Fe	270.00 abc	113.33 a	125.00 a	16.00 lm	203.33 def	31.67 bc				
$P_1 + Z_n$	240.00 h-k	90.00 g-j	110.00 cd	17.67 h-1	198.33 ef	33.33 ab				
P _i + Mn	256.67 c-g	98.33 d-g	118.33 abc	16.33 klm	200.0 def	30.00 bcd				
P ₂ + Fe	276.67 a	115.00 a	121.67 a	19.67 e-h	203.33 def	31.67 bc				
P ₂ + Zn	246.67 f-i	. 90.00 g-j	116.67 ad	16.00 lm	195.00 f	31.67 bc				
$P_2 + Mn$	270.0 abc	100.00 c-f	121.67 a	18.33 g-k	206.67 cde	31.67 bc				
K ₁ + Fe	223.33 lm	86.67 h-k	110.00 cd	20.00 d-g	150.00 j	30.00 bcd				
$K_1 + Zn$	203.33 n	76.671	96.67 f	17.00 i-m	125.00 k	33.33 ab				
K ₁ + Mn	223.33 lm	93.33 f-i	108.33 de	20.33 d-g	180.00 g	33.33 ab				
K ₂ + Fe	256.67 с-д	98.33 d-g	116.67 a-d	23.00 Б	161.67 hi	31.67 bc				
K ₂ + Zn	228.33 ki	85.00 i-1	108.33 de	19.00 f-i	128.33 k	33.33 ab				
K ₂ + Mn	253.33 d-h	98.33 d-g	116.67 a-d	22.00 bcd	200.00 def	33.33 ab				
Mor ₁ + Fe	263.33 a	96.67 efg	120.00 ab	23.00 ъ	220.00 ab	25.00 с				
Mor ₁ + Zn	236.67 i-l	85.0 i-l	108.33 de	17.67 h-1	203.33 def	28.33 cde				
Mor ₁ + Mn	258.33 b-f	95.00 e-h	118.3 abc	20.67 c-f	225.00 a	28.33 cde				
Mor- + Fe	266.00 a-d	108.33 abc	121.67 a	22.00 bcd	210.00 bcd	26.67 de				
Mor ₂ + Zn	265.00 a-e	106.67 a-d	121.67 a	18.67 f-j	200.00 def	26.67 de				
Mor ₂ + Mn	266.67 a-d	103.33 be	125.00 a	23.00 б	215.00 abc	28.33 cde				
Water (control)	210.0 m	80.00 kl	95.00 f	15.00 m	176.67 g	30.00 bcd				
Mean Mean	245.57	94.23	112.80	19.54	183.04	30.36				

Means designed by the same letter at each cell are not significantly different at the 5% level according to duncan's multiple rang test

1.2.2 Effect of kinetin:

The micrographs in Fig. (2) and the data in Table (5) showed that kinetin applications had a significant effect on the cotton leaf anatomy. It is clear that kinetin significantly increased the thickness of blade as a result of increasing the thickness of mesophyll tissue, No. of vascular/midrib and diameter of xylem vessel only in case of

high concentration of kinetin (50 ppm) as compared to the control. Thickness of xylem significantly decreased by using the two kinetin concentration compared to the control. These results are in opposite with those obtained by Gassman et al. (1980) and Ateya (2001).

1.2.3 Effect of morphactin:

As shown from the micrographs in Fig. (2) and the data in Table (5) it is obvious that morphactin applications significantly increased the thickness of blade, palisade tissue thickness, spongy tissue thickness, midrib zone thickness, thickness of xylem. While diameter of xylem vessel significantly decreased compared to the control. Cross section and the data revealed that palisade tissue thickness at low pix concentration was significantly higher (95 μ) than high concentration (93.33 μ), while spongy tissue thickness behaved an opposite trend where, it was thicker at higher morphactin concentration (123.33 μ) than low one (100.0 μ). The increases on midrib zone thickness compared to the control, this may be attributed to shrinked and contract sample of mesophyll tissue on the lower surface of the main vein. These results are in parallel with those obtained by Ali et al. (1994) on vicia faba and Abd El-Aziz (2003) on vicia faba.

1.2.4 Effect of micronutrients:

Table(5) showed that micronutrients applications insignificantly affected the leaf anatomy. All leaf anatomy characteristics did not significantly differ from the control.

1.2.5 Effect of PGRs + micronutrients combinations:

As shown from data in Table (5), it is clear that all combined treatments including one PGR + one microelement significantly increased thickness of blade, thickness of mesophyll (palisade and spongy tissue), midrib zone thickness, thickness of xylem and diameter of xylem vessel, with an exception of low kinetin application (25 ppm) + zinc. However, this treatment gave higher value of xylem diameter vessel compared to the control.

REFERENCES

Abd El-Aziz, Kh.A. (2003). Morphophysilogical studies on *Orobanche* spp. plants. M.Sc. Thesis, Fac. Agric., Tanta Univ Ali, S.A.; E.R. El-Desoki; R.R. El-Masry and S.I. Omara (1994). Effect of Morphactin CF 125 on the anatomical structure of *Vicia faba* L. Annals Agricultural Science, Moshtohor, 32(2): 875-888.

Ateya, A.G.E. (2001). Morphophysiological studies on soybean (Glycine max Merill) plants. M.Sc. Thesis, Fac. Agric., Tanta Univ..

Azab, A.S.M.; M.A. Eweida and A.W. Shalaby (1992a). Response of two long staple cultivars of Egyptian cotton to foliar application with some micronutrients. Zagazig. J. Agric. Res. Vol. 19(1); 49-60

Dierig, D.A. and R.A. Backhaus (1990). Effects of morphactin and DCPTA on stem growth and Bioinduction of rubber in Guayule. HortScience. 25(5): 531-533..

El-Nady, M.F. (1994). Effect of some growth regulators on eggplant *Solanum melongena* L. growth. M.Sc. Thesis, Fac. Agric., Kafr El-Sheikh, Tanta Univ.

Gausman, H.W. (1986). Onion bioregulators including Pix and cycocel and their biorelevancy. West Printing. Lubbock, Tx.

Gausman, H.W.; J. Stabenow; F.R. Rittig; D.E. Escobar; M.V. Garz and M. Abdel-Rahman (1980). Mepiquat chloride effects on cotton leaf anatomy. Proceeding of the plant growth regulator working group 1980, 8-14, 17 ref.

Ghamrawy, A.K. and A. Zaher (1953). Anatomical studies on Berseem (*Trifolium alexandrinum* L.). 1- The seedling. Cairo Univ., Fac. Agric. bull., 30: 1-14.

Ibrahim, D.N.; M.A. Khafagy and A.M. Abo El-Khaeer (1990). Some growth substances affecting the growth, chemical composition and alkaloidal content of *Lupinus termis* L. Egypt. J. Appl. Sci., 5(7): 367-381.

LeClerg, E.L.; W.H. Leonard and A.G. Clark (1962). Field plot technique. Burges Publishing Company.

Mahmoud, W.Sh. (1987). Effect of growth retardants on tomato structure. Al-Azhar J. Agric. Res., Vol. 7.

Morgan, P. (1980). Synthetic growth regulators: potential for development Botanical Garjette, 141: 337-346

Shabaan, F. M.R. (1996). Morphological and physiological responses of sweet pepper to some growth regulators. M.Sc. Thesis, Fac. Agric., Kafr El-Sheikh, Tanta Univ.

Sharma, J.C.; N.K. Tomar and V.K. Gupta (1988). Effect of levels and methods of zinc application and growth, yield attributes and yield of cotton (*Gossypium hirsutum* L.). Agricultural-Science-Digest-Karnal. 1988, 8: 3, 165-169; 6 ref.

Snedecor, G.W. and W.G. Cochran (1982). Statistical methods. The Iowa State University Press. 7th Edit. 2nd Printing. 507 pp.

دراسات مورفوفسيولوجية على نباتات القطن المصرى"

دراسات مورقوقسيونوجيه على ببانات الفطن المصرى ثالثا: تأثير بعض منظمات النمو والعناصر الصغرى على الصفات الثير بعض التشريحية لنبات القطن

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تم إجراء هذا البحث في مزرعة كلية الزراعة خلال مواسم ٩٩٩م، ٢٠٠٠م ــ قسم النبات الزراعي ــ كلية الزراعة الزراعة كفرالشيخ ــ جامعة طنطا. وكان الهدف الأساسي لهذا البحث هو دراسة تأثير ثلاثة منظمات نمو هي البيكس والكينتين والمورفاكتين بتركيزين لكل منهما (منخفض وعال) وثلاثة عناصر صغرى هي الحديد والزنك والمنجنيز بتركيز واحد لكل منهما والتوافيق الممكنة لكل من منظمات النمو مع العناصر الصغرى على الصفات التشريحية لنبات القطن وكان صنف القطن المستخدم في هذه الدراسة هو جيزه ٨٦ وهو صنف مستنبط حديثا من طبقة الأقطان الطويلة.

ويمكن تلخيص النتائج المتحصل عليها فيما يلى:

ادت المعاملة بكل من البيكس أو المورفاكنين منفردا إلى زيادة معنوية في قطر الساق لنباتات القطن وسمك خلايا البشرة وسمك طبقة القشرة (النسيجين الكولنشيمي والبارانشيمي) أيضا ادت نفس المعاملة إلى زيادة معنوية في قطر الأسطوانة الوعائية وسمك نسيج الخشب وعدد الحزم الوعائية أما من حيث المعاملة بالكينتين فقد أدت إلى نقص معنوى في قطر الساق معطية سيقان نباتات رفيعة مقارنة بالنباتات الغير معاملة كما أدت الكولنشيمي والبارانشيمي) وقللت معنويا من قطر الأوعية الأسطوانية الكينتين أو المورفاكتين منفردا إلى زيادة معنوية لكل من البيكس أو الكينتين أو المورفاكتين منفردا إلى زيادة معنوية لكل من البيكس أو الورقة وسمك نسيج الميزوفيل مقارنة بالنباتات الغير معاملة في حين أدت معاملات البيكس أو الكينتين إلى زيادة معنوية في عدد أوعية العرق الوسطى وسمك أوعية الخشب في حين قالت من سمك الخشب مقارنة بالكنترول، أما المعاملة بالمورفاكتين فقد أدت إلى زيادة معنوية في سمك

منطقة عرق الورقة وسمك الخشب. بينما قطر أوعية الخشب نقص معنويا مقارنة بالنباتات الغير معاملة (الكنترول).

لم نتأثر الصفات التشريحية للورقة بالمعاملة بالعناصر الصغرى ولم تختلف كثيرا عن الكنترول و أدت المعاملة بالعناصر الصغرى إلى زيادة معنوية في السمك الكلي لطبقة القشرة مقارنة بالكنترول. كذلك أعطت المعاملات المشتركة زيادة معنوية في قطر الساق وسمك طبقة القشرة وسمك أنسجة الخشب وقطر الأوعية الخشبية وعدد الحزم الوعائية مقارنة بالكنترول.و أدت المعاملات المشتركة إلى زيادة معنوية في سمك نصل الورقة وسمك الميزوفيل ومنطقة عرق الورقة وسمك الخشب وقطر الأوعية الخشبية مقارنة بالنباتات الغير معاملة (الكنترول).