

Annals Agric. Sci., Ain Shams Univ., Cairo, 51(2), 551-558, 2006

[40]

CROSS RESISTANCE OF DELTAMETHRIN RESISTANT STRAIN OF PINK BOLLWORM PECTINOPHORA GOSSYPIELLA (SAUND.) TO SOME INSECTICIDES

Mona F. Rofail¹; M.S. El-Zemaity²; A.E. Bayoumi² and Hemat Z. Moustafa¹ 1- Plant Protection Research Institute, Agric. Research Center, Dokki, Giza, Egypt 2- Plant Protection Dept., Fac. Agric. Ain Shams Univ. Shoubra El-Kheima, Cairo, Egypt

Keywords: Pink bollworm, Deltamethrin, Crossresistance, Insecticides

ABSTRACT

Selection pressure under laboratory conditions of the field strain of pink bollworm Pectinophora gossypiella (Saund.) collected from Sharkeya Governorate, Egypt was carried out using the pyrethroid insecticide deltamethrin. The obtained results indicated that the resistance ratio attained a level of 215.11-fold comparing with the susceptible strain after 14 generations of selection. In addition, the response of the obtained deltamethrin resistant strain to some insecticides indicated the cross-resistance to esfenvalerate whereas no crossresistance occurred to thiodicarb (carbamate), chlorpyrifos (organophosphorus) and/or the bioinsecticides, Dipel 2X and Agerin (derived from Bacillus thuringiensis, subspecies kurstaki). These data may emphasize the possibility of rotation of deltamethrin with these insecticides in pest control program of pink bollworm.

INTRODUCTION

Pink bollworm *Pectinophora gossypiella* (Saund.) is one of the most serious pests attacking cotton crop in Egypt and the most cotton producing countries which cause a great damage in the quality and quantity of cotton yield (El-Naggar,

(Received June 3, 2006) (Accepted August 20, 2006) 2003). In the early of 1980's, pyrethroid insecticides were rapidly substituted for organophosphorus and organochlorine insecticides for control of the pink bollworm due to their wide spectrum, low dosage, high killing efficiency, low residue and low toxicity to humans and animals (Kranthi et al 2001). Unfortunately, resistance by the bollworm to such insecticides became more and serious because of indiscriminate applications (Wang, 1992). Since the resistance of such pests are expected, the aim of the present work is to investigate the development of resistance of *P. gossypiella* to a pyrethroid insecticide, deltamethrin. Also, cross-resistance to other insecticides was studied.

MATERIALS AND METHODS

1. Bioassay and selection pressure procedures for resistance

Newly hatched larvae of a susceptible strain of *P. gossypiella* (Saund.) were obtained from the Bollworm Research Division, Plant Protection Research Institute, ARC, Dokki, Giza, Egypt. Larvae were reared on semi-artificial diet under laboratory conditions for several generations away from exposure to any insecticidal pressure according to the method described by **Rashad and Ammar** (1984). Field strain (parent) was collected from Ebrahemia region, Sharkeya Governorate,

Egypt during 2002-2003 cotton season and reared under laboratory conditions as the same with susceptible strain.

Serial concentrations (0.1-500 ppm) of deltamethrin were sprayed into petri dishes (9 cm diameter) using a hand atomizer in a three replicates for each concentration. The treated surfaces were left to dry. Thirty newly hatched larvae were transferred with a clean brush to each treated dish. The dishes were covered with toilet paper then further covered with their covers and kept in an incubator adjusted at $27 \pm 2^{\circ}$ C and 70-85% R.H. After an hour of exposure, the treated and untreated (check) insects were transferred individually on semi artificial diet poured into glass tubes (2 x 7.5 cm) covered with cotton piece and kept under the previous constant conditions.

The mortality percent was determined and corrected after 24 hours using the Abbott's formula (1925). From the corrected mortality percent and the concentrations used, the toxicity regression lines of the tested compound was plotted and represented in Log/probit relation according to the method of Finney, (1972) using the computer program, Sigma Plot for Windows, Version 2.0. LC₃₀ and LC₅₀ values were estimated from the plotted toxicity lines. Selection for resistance was carried out on the newly-hatched larvae at the LC₃₀ levels. (0.078 ppm) of deltamethrin was applied on artificial diet in glass tubes of (2x7.5cm), each tube was infested by neonatal larvae and capped with cotton piece. Higher concentrations of deltamethrin were used in subsequent generations with the increase of resistance levels. LC50 values were estimated for 1st, 3rd, 5th, 6th, 7th, 10th and 14th generations. Development of resistance ratio as well as relative resistibility for each generation were calculated as follows:

Resistant ratio = LC_{50} of the selected strain / LC_{50} of susceptible strain.

Relative resistibility = LC_{50} of selected generation / LC_{50} of anterior generation.

2. Cross-resistance of deltamethrin resistant strain to the tested insecticides

Both of the obtained resistant strain to deltamethrin and the susceptible strain were exposed to various insecticides, i.e. esfenvalerate (a pyrethroid compound), chlorpyrifos (an organophosphorous), thiodicarb (a carbamate), Ecotech and Agerin (an bioinsecticides derived from *Bacillus thuringiensis*, subspecies *kurstaki*). The newly hatched larvae of each strain was exposed to serial dilutions of the tested insecticides as mentioned before. The corrected mortality percentages were estimated and the LC₅₀ values were evaluated according to Finney, (1972). The resistance ratio of the tested insecticides (LC₅₀ value of R-strain / LC₅₀ value of S-strain) was calculated and the differences of 5-fold or more were considered as indicating positive correlation while those between l : 4-fold were considered as indicating to no correlation. Differences less than 1-fold represented a probable negative correlation.

3. Statistical Analysis

All of the toxicity values (LC₅₀'s) which estimated from the plotted toxicity regression lines and their corresponding slope values were calculated using the Probit Analysis Program designed by Dr. Nabil AM. Abd EL-Salam, Plant Protection Institute, Dokki, Giza, Egypt. The significant differences between the mentioned values were statistically analyzed using the Computer program Statitica for Windows, version 4.5.

RESULTS AND DISCUSSION

1. Development of resistance of pink bollworm to deltamethrin

The estimated LC_{50} values of deltamethrin to the different strains of pink bollworm *P. gossypiella* (Saund.) during fourteen generations of selection are shown in **Table (1)**, while the plotted toxicity regression lines are illustrated in **Figure** (1). The results clearly indicate that LC_{50} values increased gradually during the first five generations. At G3 the resistance ratio attained a level of 11.49-folds then increased in generations 5 and 6 to be 20.82 and 69.69-folds, respectively. With further selection pressure, the resistance ratio increased again during generations 7 and 10 to be 99.80 and 165.84-folds and reached 215.11-folds in generation fourteen.

In addition, the toxicity regression lines were characterized by slight fluctuations in the slope values, which were comparatively low and were nearly close to each other. In this respect Hoskins and Gordon (1956) pointed out that the development of true resistance was characterized by regression line becoming shallower as it moves to the right direction, finally it becomes steeper again as resistance genotypes comes to characterize the

Selection generations	LC ₅₀ (ppm) (5%fiducial limits)	Slope	Resistance Ratio (Fold)	Relative Resistibility	
Susceptible	0.55 (0.28-1.06)	0.69	-	-	
Parent	1.03 (0.58-1.84)	0.63	1.87	-	
G1	2.93 (1.82-4.71)	1.31	5.32	2.84	
G3	6.32* (3.48-11.51)	0.73	11.49	2.16	
G5	11.45** (7.98-16.42)	1.30	20.82	1.81	
G6	38.33*** (23.46-62.62)	0.85	69.69	3.35	
G7	54.90*** (31.66-95.20)	0.85	99.80	1.43	
G10	91.21*** (61.60-135.06)	1.14	165.84	1.66	
G14	[18.3]*** (82.89-168.87)	1.31	215.11	1.30	

Table 1. Rate of development of resistance to deltamethrin in *P. gossypiella* (Saund.) during selection for 14 generations

Comparing to the parent generation, (***) highly significant $p \le 0.001$, (**) moderately significant $p \le 0.01$ and (*) significant $p \le 0.05$ (student *t*-test).



Fig. 1. Toxicity lines of deltamethrin to the tested generations of P. gossypiella

new population. Moreover, Osman et al (1991) reported that sixteen generations of larval selection of a field strain of pink bollworm P. gossypiella exposed to permethrin produced 9.7-folds resistance in adults. Higher slope values obtained in F12 through F16 generations indicated an increasing of homogeneity in later generations. On the other hand, the estimated slope values indicated that a population that was apparently heterogeneous at the beginning of the study tended to become relatively more homozygous as selection progressed. However, similar finding was obtained by Aldosari et al (1996) who found that selection of field populations of beet armyworm Spodoptera exigua for resistance to cyfluthrin for eight generations resulted in a 54.1-folds increase in the estimated LD₅₀ values. The regression lines of all strains, including the susceptible strain vielded relatively low slope values of 1.09 to 2.13 indicating that high degree of genetic heterogeneity exists among these strains.

Generally, various investigators reported that the selection pressure of tobacco budworm *Heliothis virescens* with pyrethroids produced an increase in levels of resistance compared to the susceptible strain. In such studies, it was found that the selection pressure of larvae of the diamondback moth *Plutella xylostella* with permethrin increased resistance. After twenty generations of continuous selection pressure resistance to permethrin reached to be over than 600-folds in this strain compared with the unselected parent strain. Strain differences as well as the degree of homozygosity of resistance may account for the discrepancy (Jensen *et al* 1984; Ibrahim *et al* 1996; Zhao *et al* 1996 & Yu and Nguyen 1996).

2. Cross-resistance to certain insecticides

The determined LC_{50} values of the tested insecticides on susceptible and deltamethrin resistant strains after the selection pressure of *P*. gossypiella for fourteen generations were listed in **Table (2)** and the corresponding plotted toxicity regression lines are illustrated in **Figures (2-6)**. The selected strain was apparently resistant to esfenvalerate, i.e. 25.57-folds, conferred low resistance to chlorpyrifos, i.e. 4.20-folds and low tolerant to thiodicarb, i.e. 3.18-folds. On the contrary, the impact of deltamethrin resistance as the potency of the biopesticides, Agerin and Ecotech, were negligible.

Tested	Susceptible strain		Deltamethrin resistance strain			
	LC ₅₀ (ppm) (Fiducial limits)	Slope	LC ₅₀ (ppm) (Fiducial limits)	Slope	Resistance ratio	
Esfenvalerate	0.14 (0.09-0.20)	1.19	3.58"(2.35-5.47)	1.19	25.57	
Chlorpyrifos	0.05 (0.04-0.07)	1.47	0.21 (0.14-0.31)	1.19	4.20	
Thiodicarb	70.80 (53.22-94.18)	1.88	225.02*** (159.34-317.77)	1.44	3.18	
Ecotech	380 (280-500)	1.41	350 (270-450)	1.59	0.92	
Agerin	350 (240-510)	1.12	340 (230-510)	1.02	0.97	

Comparing between deltamethrin resistant strain and susceptible strain, (***) highly significant $p \le 0.001$, (**) moderately significant $p \le 0.01$ and (*) significant $p \le 0.05$ (student *i*-test).



Fig. 2. Toxicity regression lines of esfenvalerate against susceptible and deltamethrin resistant strains of *P. gossypiella*



Fig. 3. Toxicity regression lines of chlorpyrifos against susceptible and deltamethrin resistant strains of *P. gossypiella*



Fig. 4. Toxicity regression lines of thiocarb against susceptible and deltamethrin resistant strains of *P. gossypiella*.



Fig. 5. Toxicity regression lines of Ecotech against susceptible and deltamethrin resistant strains of *P. gossypiella*



Fig. 6. Toxicity regression lines of Agerin against susceptible and deltamethrin resistant strains of *P. gossypiella*

Similar finding was reported by Riskallah et al (1983) who showed that the selection pressure of a field strain of Egyptian cotton leafworm Spodoptera littoralis with fenvalerate or deltamethrin resulted in the development of resistance to all pyrethroids, DDT and aminocarb but showed no cross resistance to the orparathion. methyl ganophosphate, Several investigators reported that there is no cross pyrethroids resistance and between organophosphorus, carbamate and microbial insecticides, (Yu and Nguyen 1996; Liu and Jiang 1995; Wu et al 1994 and Ayad and Phillabaum 1990). Also, Jensen et al (1984) found that a permethrin resistant strain of tobacco budworm H. virescens had developed cross resistance to cy-

permethrin which was evaluated by 7.9-folds. Tabashnik et al 1987 suggests that cross-resistance between conventional insecticides and B.thuringiensis is unlikely in diamondback moth. In contrary, Elzen (1995) and Zhao et al (1996) found that the tobacco budworm H. virescens resistance strain to cypermethrin proved cross resistance to organophosphates, i.e. profenofos and azinophos methyl and carbamates, i.e. thiodicarb and methomyl but the data did not indicate any resistance to B. thuringiensis. Also, Aldosari et al (1996) reported that cyfluthrin selected strain of beet armyworm S. exigua (Hubner) showed cross resistance to methomyl whereas it was only weakly cross-resistant to profenofos.

REFERENCES

Abbott, W.S. (1925). A method of computing the effectiveness of an insecticide. J. Econ. Entomol. 18: 265-267.

Aldosari, S.A.; T.F. Watson; S. Sivasupramaniam and A.A. Osman (1996). Susceptibility of field populations of beet armyworm (Lepidoptera: Noctuidae) to cyfluthrin, methomyl, profenofos, and selection for resistance to cyfluthrin. J. Econ. Enotmol. 89 (6): 1359-1363.

Ayad, H.M. and C.R. Phillabaum (1990). Toxicity of thiodicarb to susceptible and resistant strains of tobacco budworm and full armyworm larvae. Proceedings Beltwide Cotton Conference. 1: 239-241.

El-Naggar, A.Z.A. (2003). Evaluation of Certain New Approaches of Control Measures in an Integrated Pest Management Program of Cotton Bollworms. p. 1. Ph. D. Thesis. Fac. Agric. Alexandria University, Egypt.

Elzen, G.W. (1995). Trends in *Heliothis* resistance. Proceedings Beltwide Cotton Conference 2: 964-965.

Finney, D.J. (1972). Probit Analysis: a Statistical Treatment of the Sigmoid Response Curve. p. 33.. Cambridge Univ. Press, London.

Hoskins, W.M. and H.T. Gordon (1956). Arthropod resistance to chemicals. Ann. Rev. Entomol. 1: 89-122.

Ibrahim, S.A.; A.M. Younis and J.A. Ottea, (1996). Biochemical mechanisms of pyrethroid resistance in cypermethrin selected *Heliothis virescens*. Proceedings Beltwide Cotton Conference. 2: 1054-1059.

Jensen, M.P.; L.A.Crowder and T.F. Watson (1984). Selection for permethrin resistance in the tobacco budworm (Lepidoptera: Noctuidae). J. Econ. Entomol. 77: 1409-1411.

Kranthi, R.K.; D. Jadhaw; R. Wanjani; S. Kranthi and D. Russel (2001). Pyrethroid resistance and mechanisms of resistance in field strain of *Helicoverpa armigera* (Lepidoptera: Noctuidae). J. Econ. Entomol. 94 (1): 253-263.

Liu, Y.X. and Y.C. Jiang (1995). Cross resistant patterns of insecticide selected strains of cotton bollworm (*Helicoverpa armigera*) Hubner. Resistance Pest Management 7 (2): 13-14.

Osman, A.A.; T.F. Watson and S. Sivasupramaniam (1991). Reversion of permethrin resistance in field strains and selection for azinphosmethyl and permethrin resistance in pink bollworm (Lepidoptera: Gelechiidae). J. Econ. Entomol. 84(2): 353-357.

Rashad, A. and E.D. Ammar (1984). Mass rearing of the spiny bollworm *Earias insulana* (Boisd.) on semi artificial diet. Bull. Entomol. Soc. Egypt. 65: 239-244.

Riskallah, M.R.; S.F. Abd-Elghar; M.R. Abo-Elghar and M.E. Nassar (1983). Development of resistance and cross resistance in fenvalerate and deltamethrin selected strain of *Spodoptera littoralis* (Boisd.). Pestic. Sci. 14: 508-512.

Tabashnik, B.E.; N.L. Cushing and M.W. Jonson (1987). Diamondback moth (Lepidoptera: Plutellidae) resistance to insecticides in Hawaii: intra island variation and cross resistance. J. Econ. Entomol. 80 (6): 1091-1099.

Wang, S. (1992). Pyrethroid resistance of cotton bollworm and its management in the north China cotton region. Proceedings Beltwide Cotton Conferences, p. 900.

Wu, Y.D.; J.L. Shen and Z.P. You (1994). Laboratory selection for fenvalerate resistance and susceptible strains in cotton bollworm *Heliothis armigera* (Hubner). Acta Entomologica Sinica. 37 (2): 129-136.

Yu, S.J. and S.N. Nguyen, (1996). Insecticide susceptibility and detoxication enzyme activities in permethrin selected diamondback moths. Pestic. Biochem. Physiol. 56: 69-77.

Zhao, G.; R.L. Rose; E. Hodgson and R.M. Roe (1996). Biochemical mechanism and diagnostic microassays for pyrethroid, carbamate, and organophosphate insecticide resistance/crossresistance in the tobacco budworm, *Heliothis virescens.* Pestic. Biochem. Physiol. 56: 183-195.

حوليات العلوم الزراعية جامعة عين شمس، القاهرة مجلد(٥١)، عدد (٢)، ٥٥١–٥٥٨، ٢٠٠٦



عبور صفة مقاومة سلالة دودة اللوز القرنفلية المقاومة للدلتامثرين لبعض المبيدات الحشرية

[٤.]

مونا فكرى روفائيل' - محمد السعيد صالح الزميتي' - علاء الدين بيومي' -همت زكريا محمد مصطفى' ١- معهد بحوث وقاية النباتات - مركز البحوث الزراعية - الدقسي - الجيسزة - مصر ٢- قسم وقاية النبات - كلية الزراعة - جامعة عين شمس - شبرا الخيمة - القاهرة - مصر

للدلتامثرين أنه يوجد عبور لصفة المقاومة بين الدلتامثرين و اس فنفاليرات بينما لا يوجد عبور لصفة المقاومة بين المبيدات الحشرية المعتادة كلوربيريفوس، ثيوديكارب أو المبيدات الحيوية إكونيك و أجرين في هذه السلالة. و تؤكد هذه النتائج علي إمكانية مناوبة الدلتامثرين لهذه المبيدات في برنامج مكافحة دودة اللوز القرنفلية.

تم إجراء الضغط الإنتخابى تحت ظروف معملية لسلالة حقلية من دودة اللوز القرنفلية جمعت من محافظة الشرقية باستخدام المبيد البيرثرويدى دلتامثرين. ودلت النتائج المتحصل عليها على أن مستوى المقاومة قد وصل إلى ٢١٥,٧٥ ضعفا وذلك مقارنة بالسلالة الحساسة بعد ١٤ جيلا. وأظهرت دراسة إستجابة سلالة دودة اللوز القرنفلية المقاومة

> تحکیم: أ.د عبد الحمید عبد الحمید زیدان أ.د العیـــداروس أحمــد جمعــه