# EVALUATION OF SOME FEED ADDITIVES AT DIFFERENT LEVELS IN DIET OF NILE TILAPIA (OREOCHROMIS NILOTICUS)

#### **ADEL E. TOLAN**

Aquaculture Research Unit, Sakha, Central Laboratory of Aquaculture, Abassa, ARC, Egypt

(Manuscript received 1st October 2006)

#### **Abstract**

This study was carried out in order to estimate the influence of three levels of dried yeast, two kinds of probiotics (Bacillozyme and Bacillogene) and vitamin C on growth performance, feed utilization, blood parameters, hepatic and renal histopathology, chemical composition of the whole fish and economic feed efficiency of Nile Tilapia (O. niloticus) fingerlings. Total numbers of 390 of Nile Tilapia fingerlings with intial weight of 17.41 gram were divided into four treatment groups and control group. Each treatement group was divided into three sub-groups in three replicates each (39 aquaria). Each aquarium were contained 10 fish. Different types of additives were added to abasal diet (22% CP) at three levels each including dried yeast (10, 15 and 20 g/kg diet); probiotics (Bacillozym and Bacillogene), 1, 2 and 3 g/kg diet from each probiotic and vitamin C (500, 1500 and 2000 mg/kg diet. The best growth performance and feed utilization without any pathological signs in liver and kidney, normal blood parameters and chemical composition of meat and the best economic efficiency were obtained with bacillozyme at level three grams/kg diet. Dried yeast at level of 20 g/kg diet showed significantly better results than the control group. The lowest results were obtained with the highest level of vitamin c (2000 ppm/kg).

**Keywords:** Fish, growth performance, niltoicus, probiotics, vitamine C, growth performance, blood.

# INTRODUCTION

In Egypt, fish culture became widely supported to help in providing addition of animal protein resources. The most popular kind of fish raised in Egypt is *Oreochromis niloticus*. Intensive culturing of fish needs formulated ration of high quality, which represent 50% of production cost (Collins and Delmendo, 1979) which is the main target of nutritionist to minimize the cost of feed.

Feed additives including probiotics, antibiotics, vitamins and dried yeast were used to maximize utilization of ration ingredient by fish. Antibiotics had some seriously adverse effects as multiple drug resistant and residual in animal products (Wary and Davis, 2000), while vitamins such as vit. C is natural antioxidant nutrient that play an important role in animal health by inactivating harmful free radicals (McDowell, 1989).

Using natural food additives to substitute antibiotic became essential request (Kumar *et al.*, 2003). Therefore, the present study was carried out to determine the effects of different supplementation levels of some commercial probiotics (Bacillozyme and bacillogene zinc), vitamin C and dried yeast on growth performance, feed utilization, blood parameters and economic efficiency of *Oreochromis niloticus* fish.

# **MATERIALS AND METHODS**

The present study was carried out at Fish Laboratory, Animal Production Department, Faculty of Agriculture, Kafer El-Sheikh University during the period from July to August 2006.

### Fish and management:

A total number of 390 healthy *(O. niloticus)* fingerlings were purchased from Al-Manzalah Integrated Fish Farm, General Authority for Fisheries Resources Development, with an average initial body weight of about 17 g. Fish were randomly distributed into 13 treatments groups, each treatment was presented in three replicates. In each replicate 10 fingerlings were stocked in a glass aquarium (80 x 32 x 40 cm) containing about 102 liters of dechlorinated tap water and an air-stone concreted with an electric compressor. The aquaria water was partially replaced every day to renew the water. Electric light was used to complete the daylight to 14 hours.

Feeding system and feed additives:

The experimental fish were fed the tested diets twice daily at 9 a.m. and 3 p.m., six days/week for an experimental feeding period of 35 days. The daily feeding rate was 3% (on DM basis) of live body weight of the fish and the feed amount was adjusted biweekly on the basis of the actual average biomass of the fish within each replicate.

A ground basal diet was over fed to fish in all treatments at the same times. Proximate fed to satiation analysis of the basal diet is presented in Table (1).

Ingredient	%	Ingredient	%
Dry matter	93	Ash	8.11
Crude protein	22	NFE	5.22
Ether extract	16	Gross energy	5200
Crude fiber	3.0	Energy/Protein ratio	236,36

Table 1. Proximate analysis of the basal diet fed to fish in all treatments.

Dried yeast, probiotic bacillozym containing bacillus subtilus, 0.75x10<sup>10</sup>; cellulose enzyme, 15000; protease, 187500; alfa- amylaze, 300.000; beta-amylaze and saccaromyces services, 200x10<sup>10</sup> (IBEX pharmaceutical), the tested probiotic Bacillogene zinc contained 500 g bacillus subtilus garlic allicine, 0.247 micromil and hydrolytic enzyme plus zinc methionine (IBEX pharmaceutical) and vitamin C (ADWIA pharmaceutical) were used as feed additives as a rates .Beside the basal diet fed to the control treatment, four feed additives including dried yeast, Bacillozyme, Bacillogene zinc and vitamin C, three levels of each, were added to the basal diet as shown in Table (2).

Table 2. Experimental treatment groups with different levels of feed additives fed to *O. niloticus* fish.

Feed additive	Treatment	Level of supplementation /kg basal diet		
Without	Control	Basal diet without		
	DY1	10 g		
Dried yeast	DY2	15 g		
	DY3	20 g		
	BZ1	1 g		
Bacillozym	BZ2	2 g		
	BZ3	3 g		
	BG1	1 g		
Bacillogene zinc	BG2	2 g		
	BG3	3 g		
	VC1	500 mg		
Vitamin C	VC2	1500 mg		
	VC3	2000 mg		

# **Experimental procedures:**

Live body weight and feed intake of fish was weekly recorded, also initial and final live body weight as well as average total and daily gain, specific growth rate, feed conversion ratio and protein efficiency ratio were calculated. Survival rate was recorded, and all previous traits were calculated according to the following equations:

Total weight gain (g) =  $Wt_1 - Wt_0$ 

Where:

 $Wt_1$  is the final body weight (g) and  $Wt_0$  is the initial body weight (g).

Average daily gain (g) =  $Wt_1 - Wt_0 / T$ 

Where:

T is the experimental period (day),  $Wt_0$  is the initial body weight (g) and  $Wt_1$  is the final body weight (g) according to Castell and Tiews (1980).

Specific growth rate (%/d) =  $(Ln Wt_1 - Ln Wt_0/T) \times 100$ 

Where:

Ln is the natural logarithm of final and initial weight, respectively, and T is the experimental period (day) according to Pouomogne and Mbongblang (1993).

Feed conversion ratio=feed intake (g)/weight gain (g) (Tacon, 1987)

Protein efficiency ratio =  $(TWG (g)/TPI (g)) \times 100$ 

Where:

TWG is total weight gain and TPI is total protein intake according to (Davis and

Morries, 1997)

Survival rate = (No. of fish at end/ No. of fish at start)  $\times$  100

At the end of the experiment, 6 fish from each treatment, 2 fish from each replicate were randomly taken for chemical analysis of the whole body.

# Analytical procedures and blood samples:

The chemical analysis of the basal diet and the whole fish body at the end of experiment were carried out using the methods of A.O.A.C. (1990).

Blood samples were collected at the end of experiment from 6 fish of each treatment, two fish from each replicate. In blood serum of fish, concentrations of total protein (Merck, 1974) and albumin (Doumas *et al.*, 1971) were measured by colorimetric methods using commercial kits and spectrophotometer. However, concentration of globulin was determined by substracting concentration of total protein from albumin concentration.

Activity of liver enzymes, Aspartate amino transaminase (AST) and Alanine amino transaminase (ALT) was determined using commercial kits (Biomerieux) and spectrophotometer according to (Reitman and Frankel, 1957).

#### Histological study:

At the end of experiment, specimens from kidney and liver of three fish in each group were immediately fixed in buffered 10% neutral formalin. After a fixation period of 24-48 hours, all specimens were processed for routine paraffin technique. Paraffin sections (8-10  $\mu$ m) were stained by Harris Haematoxyline and Eosin according to Drury and Wallington (1980). Thereafter, slides were examined for histopathological signs.

#### Statistical analysis:

The obtained data were analyzed by F-test according to Sendecor and Cochran (1982) using SAS (1996) procedure for personal computer. Least significant difference according to Duncan (1955) was used for the comparison among the significant group means at level of P<0.05.

#### RESULTS AND DISCUSSION

#### **Growth performance:**

Data presented in Table (3) show significant (P<0.05) effects of dietary additives on final body weight, total weight gain, average daily gain and specific growth rate of fish in different treatment groups.

Among all dietary additives, fish in BG3 group showed significantly (P<0.05) the heaviest final weight (26.89 g), and the highest total gain (9.39 g), average daily gain (0.27 g) and specific growth rate (0.43 %/d). However, fish in VC3 group showed

significantly (P<0.05) the lowest values, being 20.93 g, 3.49 g, 0.10 g and 0.18 %/d, respectively (Table 3).

It is of interest to note that fish fed all levels of dried yeast (DY1, DY2 and DY3 groups) and those in BZ3 group showed significantly (P<0.05) higher values of final weight, total gain, average daily gain and specific growth rate than the control group and did not differ significantly than those in BG3. However, fish fed the low and medium levels in BZ1, BZ, BG1, BG2, VC1 and VC2 did not differ significantly than the control group in most growth traits studied (Table 3).

A participant observation could be noticed among the tested dietary additives that the all levels of dried yeast achieved significantly higher growth of tilapia fish, being the highest for a level of 15 g/kg diet. However, growth of fish increased by increasing level of Bacillogene zinc or Bacillozyme from 1 to 3 g /kg diet. Yet, levels of 500 and 1500-ppm vitamin C failed to achieve the expected growth; even growth was delayed when its level increased to 2000 mg in diet of tilapia fish.

On the basis of the obtained results, fish fed diet supplemented with Bacillozyme zinc at a level of 3 g/kg showed beneficial effects on their growth performance.

The beneficial effects of probiotics (Lacto-Sacc) on growth performance, gain and feed efficiency of rabbits have been reported by several; authors (El-Hindawy *et al.* 1993, 1994 & 1997 and Yamani *et al.*, 1992). The present results concerning growth performance traits are in agreement with those obtained by Abdelhamed *et al.* (2000) and Magouz *et al.* (2002) on Nile tilapia fingerlings fed on diets supplemented with dried yeast and Lacto-Sacc as probiotics. The beneficial effects of probiotics may be mediated by a direct antagonistic effect against specific groups of microorganisms, resulting in suppression of viable count, suppression of bacterial number, an alteration of microbial metabolism or by stimulation of immunity (Fuller, 1989 and Sissons, 1989). Also, the improvement in growth and gain of fish in BZ and BG groups may be related to a change in enteric flora and reduction of E. coli decreasing in the intestinal pH, production of antibiotic substances and/or reducing the toxic amines and ammonia level in the gut and blood of fish (Pollman, 1986).

It is of interest to note that higher increase in total gain of fish was observed with increasing level of DY, BZ and BG supplementation. However, the opposite was noticed with increasing VC level.

Table 3. Growth performance parameters of *O. niloticus* fish as affected by dietary additives in treatment groups.

Treatment group	Initial weight	Final weight	Total gain	Average daily gain (g/fish/day)	Specific growth rate (%/d)
Control	17.41	23.00±0.10 <sup>b</sup>	5.59±0.02d	0.16±0.001°	0.27 <sup>c</sup>
DY1	17.41.	25.90±0.15°	8.49±0.03 <sup>b</sup>	0.24±0.001°	0.40²
DY2	17.41	26.33±0.01ª	8.92±0.01 <sup>b</sup>	0.25±0.001°	0.41ª
DY3	17.41	26.42±0.02³	9.01±0.01³	0.26±0.010ª	0.43ª
BZ1	17.41	23.00±0.31b	5.59±0.04°	0.16±0.002°	0.35⁵
BZ2	17.41	23.40±0.30⁵	5.99±0.05 <sup>d</sup>	0.17±0.001°	0.29 <sup>c</sup>
BZ3	17.41	25.52±0.02ª	8.11±0.11 <sup>b</sup>	0.23±0.010b	0.39ª
BG1	17.41	23.80±0.13 <sup>b</sup>	6.39±0.03°	0.18±0.003°	0.31 <sup>b</sup>
BG2	17.41	24.50±1.10 <sup>b</sup>	7.09±0.07°	0.21±0.003b	0.34 <sup>b</sup>
BG3	17.41	26.80±1.30ª	9.39±0.11°	0.27±0.011°	0.43ª
VC1	17.41	24.80±0.17 <sup>b</sup>	7.39±0.05°	0.21±0.013 <sup>b</sup>	0.35 <sup>b</sup>
VC2	17.41	23.78±0.13 <sup>b</sup>	6.37±0.03°	0.18±0.014°	0.31 <sup>b</sup>
vc3	17.41	20.90±1.10°	3.49±0.01°	0.10±0.001d	0.18 <sup>c</sup>

Group with different superscripts within the same column are significantly different at P<0.05.

In this respect, Magouz *et al.* (2002) found similar trend by using Lacto-Sacc, Fermato and Bio-tonic as probiotics, whereas increasing level of supplementation from 1 up to 4 g of these probiotics//kg diet resulted in rather improvement in gain of Nile tilapia. Such trend reversed that reported on rabbit, being more benefit at 1 g/kg diet (Tawfeek and El-Hindawy, 1991). This was proved in the present study by increasing total gain of fish by increasing level of DY supplementation from 1 up to 3g/kg diet, which may indicated that fish require higher level to achieved the highest growth performance.

Concerning the negative trend of increasing level of VC on growth performance parameters in all VC groups may suggest a toxic effect of VC at high levels. It was stated that with dietary ascorbic acid 200 mg/kg diet, weight gain was improved in channel catfish (Duncan and Lovell (1994) and Nile tilapia (Abd Elaziz and Mahmoud, 2004).

# Feed utilization and survival rate:

Results shown in Table (4) revealed significant (P<0.05) effect of dietary additives on feed intake, feed conversion ratio, protein efficiency ratio and survival rate of fish in different treatment groups.

In comparison among treatment groups, fish fed DY3 and VC1 diets showed significantly (P<0.05) higher feed intake than the other treatment groups and the control group. However, fish in BG3 showed significantly (P<0.05) the lowest feed intake (Table 4).

Table 4. Feed intake and feed utilization of O. niloticus fish as affected by different dietary additives.

	· · · · · · · · · · · · · · · · · · ·				<del>,</del>
Treatment	Feed intake	Total gain	Feed	Protein	Survival rate
group	(g)	(g)	conversion ratio	efficiency ratio	(%)
Control	21.70 <sup>b</sup>	5.59±0.02d	3.88 <sup>b</sup>	1.37 <sup>b</sup>	100°
DY1	23.28 <sup>ab</sup>	8.49±0.03b	2.74 <sup>c</sup>	1,66ª	100ª
DY2	22.20 <sup>b</sup>	8.92±0.01b	2.48 <sup>c</sup>	1.80°	90⁵
DY3	26.01	9.01±0.01°	2.88∞	1,58ª	100a
BZ1	21.70 <sup>b</sup>	5.59±0.04 <sup>d</sup>	3.88 <sup>b</sup>	1.17 <sup>c</sup>	100°
BZ2	21.60 <sup>b</sup>	5.99±0.05°	3.61 <sup>b</sup>	1.26™	100ª
BZ3	22.40 <sup>b</sup>	8.11±0.11 <sup>b</sup>	2.76°	1.65ª	100°
BG1	23.45 <sup>b</sup>	6.39±0.03°	3.67 <sup>b</sup>	1. <b>2</b> 3°	90 <sup>p</sup>
BG2	22.58 <sup>b</sup>	7.09±0.07°	3.18 <sup>b</sup>	1.43 <sup>b</sup>	90⁵
BG3	21.50 <sup>b</sup>	9.39±0.11°	2.29°	1.74ª	100ª
VC1	24.54ª	7.39±0.05°	3.32⁵∞	1.37 <sup>b</sup>	90 <sup>b</sup>
VC2	22.02 <sup>b</sup>	6.37±0.03 <sup>c</sup>	3.46 <sup>b</sup>	1.32 <sup>b</sup>	100ª
VC3	22.38 <sup>b</sup>	3.49±0.01°	6.41ª	0.71 <sup>c</sup>	90 <sup>b</sup>

Group with different superscripts within the same column are significantly different at P<0.05.

The highest total gain along with the lowest feed intake of fish in BG3 was reflected in the best feed conversion ratio and protein efficiency ratio as compared to the other treatment groups and control group. Meanwhile, fish VC3 showed the lowest values, being in an opposite manner (Table 4).

Values of protein efficiency ratio were significantly (P<0.05) different, was higher in all DY groups, as well as the highest level of BZ and BG groups. While, it significantly (P<0.05) decreased with the low level of BZ and BG as well as the highest level of VC groups (Table 4).

Concerning the survival rate, no mortality cases were recorded in DY1, DY3, all BZ groups, BG3 and VC2. However, survival rate was 90% in the other groups.

Based on the present results, supplementation of 3 g from BG/kg to diets of Nile Tilapia had beneficial effects on feed utilization and survival rate of fish. Similar results were recorded on Nile tilapia using Lacto-Sacc, Fermacto and Bio-tonic as feed additives (Magouz *et al.*, 2002) or dried yeast and Lacto-Sacc as dietary supplementation (Abdelhamed *et al.*, 2000).

# **Blood parameters:**

Results shown in Table (5) revealed significant (P<0.05) effect of dietary additives on concentration of total protein, albumin (AL), globulin (GL), AL/GL ratio, urea and creatinin as well as activity of transaminases (AST &ALT) in blood serum of tilapia fish at the end of experiment.

As compared to the control group, albumen concentration significantly (P<0.05) decreased in all DY, BG and VC groups, being the lowest in VC3. Concentration of globulin significantly (P<0.05) increased in all BZ groups and DY2 and DY3 groups, and significantly (P<0.05) decreased in all VC groups and BG1 and BG2 groups. This is reflected in significant (P<0.05) decrease in concentration of total protein in BG1 and BG2 groups as well as all VC groups. Also, albumin/globulin ratio significantly (P<0.05) increased in BZ and BZ3, and significantly (P<0.05) decreased in DY2, DY3, BG2 and BG3 (Table 5).

Table 5. Concentration of total proteins and their fractions in blood serum of *O. niloticus* fish as affected by dietary additives in treatment groups.

	<del></del>	<del></del>	<del></del>	<del></del>
Treatment	Albumin (AL, g/dl)	Globulin	Total protein (g/dl)	AL/GL
group	rubumm (riz, gran)	(GL, g/dl)	, otal protein (g/ui)	rátio
Control	2,50	1.09 <sup>b</sup>	3.40 <sup>ab</sup>	2.30 <sup>b</sup>
DY1	2.13 <sup>b</sup>	1.02 <sup>b</sup>	3.15ab	2.10 <sup>b</sup>
DY2	2.15 <sup>b</sup>	1,25ª	3.40 <sup>ab</sup>	1.72°
DY3	2.17 <sup>b</sup>	1.24ª	3.31 <sup>ab</sup>	1.74 <sup>c</sup>
BZ1	3.17ª	1.28ª	4.45ª	2.48 <sup>b</sup>
BZ2	3.00ª	1.30ª	4.30ª	3.00ª
BZ3	2.90ª	1.50°	3.90°	2.90ª
BG1	1.00°	0.44 <sup>c</sup>	1.44 <sup>c</sup>	2.27 <sup>b</sup>
BG2	1.02°	0.64°	1.66°	1.60°
BG3	1.26 <sup>c</sup>	1.13 <sup>ab</sup> _	2.39 <sup>b</sup>	1.11 <sup>c</sup>
VC1	1.06°	0.47°	1.51 <sup>c</sup>	2.30b
VC2	1.04 <sup>c</sup>	0.45°	1.49 <sup>c</sup>	2.30°
VC3	1.00°	0.35⁴	1.35°	2.90ª

Group with different superscripts within the same column are significantly different at P<0.05.

Concentration of urea significantly (P<0.05) increased in all DY groups and BZ1 and BZ2. While, concentration of creatinin significantly (P<0.05) increased in all DY, BZ and BG groups, being the highest in all BZ groups. On the other hand, activity of AST significantly (P<0.05) increased in all DY and BZ groups, and activity of ALT significantly (P<0.05) increased in all BG and BZ groups. (Table 6). In general, fish of VC3 group showed significantly (P<0.05) the lowest albumin, globulin and total

protein concentrations, while those in BZ1 showed significantly (P<0.05) the highest concentration of urea and creatinin and the highest activity of AST and ALT (Table 6).

The information on the effect of probiotics used in this study on blood parameters of fish are scare. In agreement with the present results concerning the changes in ALT activity in fish fed DY, Ragheb *et al.* (2003) found that the effect of Lacto-Sacc on concentration of total protein was insignificant, while concentration of urea in blood plasma of growing calves significantly increased in supplemented than the control calves.

Table 6. Concentration of urea and creatinin and activity of transaminases (AST &ALT) in blood serum of *O. niloticus* fish as affected by dietary additives in treatment groups.

	Blood parameter		Enzyme activity	
Treatment	Urea	Creatinin	AST	ALT
group	(mg/dl)	(mg/dl)	(IU/dI)	(IU/dl)
Control	14 <sup>c</sup>	0.10 <sup>c</sup>	112 <sup>b</sup>	15 <sup>b</sup>
DY1	23 <sup>b</sup>	0.24 <sup>b</sup>	116°	16 <sup>b</sup>
DY2	19 <sup>b</sup>	0.20 <sup>b</sup>	114ª	15 <sup>b</sup>
DY3	15 <sup>b</sup>	2.19 <sup>b</sup>	114ª	15 <sup>b</sup>
BZ1	30ª	0.66ª	116ª	26ª
BZ2	22 <sup>b</sup>	0.51ª	115°	23ª
BZ3	10 <sup>c</sup>	0.45°	114ª	21 <sup>a</sup>
BG1	13°	0.32 <sup>ab</sup>	110 <sup>b</sup>	24ª
BG2	15 <sup>c</sup>	0.31 <sup>ab</sup>	110 <sup>b</sup>	25ª
BG3	14 <sup>c</sup>	0.19 <sup>ab</sup>	112 <sup>b</sup>	26ª
VC1	11 <sup>c</sup>	0.08 <sup>c</sup>	112 <sup>b</sup>	14 <sup>b</sup>
VC2	10°	0.08 <sup>c</sup>	111 <sup>b</sup>	13 <sup>b</sup>
VC3	12 <sup>c</sup>	0.09 <sup>c</sup>	112 <sup>b</sup>	1 <u>4</u> <sup>b</sup>

Group with different superscripts within the same column are significantly different at P<0.05.

#### Chemical composition and energy content:

Data in table (7) revealed that as compared to the control group, moisture content significantly (P<0.05) decreased in fish of all DY groups, the highest level of BZ and BG groups and the lowest level of VC group.

Content of CP significantly (P<0.05) decreased in all BZ and VC groups and the lower levels of DY and BG groups. However, contents of EE significantly (P<) increased in all treated groups, being the highest in DY1 group. Also, content of ash significantly (P<0.05) increased in all DY groups, BZ2 and BG1, and significantly (P<0.05) decreased, being the lowest in VC2 and VC3 groups (Table 7).

Table 7. Carcass chemical composition and energy content of *O. niloticus* fish as affected by additives in treatment groups.

Treat.	Moisture	Chemical con	Chemical composition (%)		
group	(%)	СР	EE	Ash	(kcol/kg)
Control	77.7±1.1ª	50.2±1.10 <sup>a</sup>	16.9±0.11 <sup>d</sup>	7.30±0.01 <sup>b</sup>	4822.2±0.7 <sup>d</sup>
DY1	74.1±1.20 <sup>b</sup>	49.8±1.30 <sup>b</sup>	30.22±0.15 <sup>a</sup>	8.22±0.07ª	5510.7±0.11 <sup>b</sup>
DY2	75.3±1.10 <sup>b</sup>	50.4±1.30 <sup>a</sup>	29.13±0.16 <sup>b</sup>	8.11±1.03 <sup>a</sup>	5543.8±1.24 <sup>b</sup>
DY3	71.1±0.01 <sup>c</sup>	53.3±0.92°	30.30±0.00ª	9.30±2.20°	5720.2±1.20ª
BZ1	76.0±0.13 <sup>a</sup>	45.8±1.01 <sup>b</sup>	29.41±0.40 <sup>b</sup>	7.70±0.11 <sup>b</sup>	5612.3±0.21°
BZ2	77.0±0.13ª	45.3±1.02 <sup>b</sup>	29.37±0.15 <sup>b</sup>	8.20±0.33ª	5677.0±0.30 <sup>a</sup>
BZ3	75.2±0.12 <sup>b</sup>	49.1±0.92 <sup>b</sup>	27.33±0.17 <sup>b</sup>	7.20±0.32 <sup>b</sup>	5522.3±1.05°
BG1	76.9±0.11 <sup>ab</sup>	46.8±0.88 <sup>b</sup>	28.21±0.11 <sup>b</sup>	8.66±0.31 <sup>a</sup>	5455.2±1.13 <sup>c</sup>
BG2	76.4±0.11 <sup>ab</sup>	47.9±0.01 <sup>b</sup>	25.33±0.11°	7.65±0.33 <sup>b</sup>	5511.3±1.70 <sup>c</sup>
BG3	73.2±0.13 <sup>c</sup>	54.7±0.09°	24.41±0.30 <sup>c</sup>	7.40±0.51 <sup>b</sup>	5422.7±1.20 <sup>d</sup>
VC1	73.9±0.18 <sup>c</sup>	48.1±1.10 <sup>b</sup>	23.47±0.33 <sup>c</sup>	7.50±0.50 <sup>b</sup>	4999.9±2.10 <sup>d</sup>
VC2	77.1±0.18 <sup>a</sup>	44.2±0.30 <sup>c</sup>	23.21±0.10 <sup>c</sup>	6.20±0.03 <sup>c</sup>	4822.8±1.01 <sup>d</sup>
VC3	78.9±0.00°	42.1±0.00 <sup>c</sup>	20.10±0.10 <sup>c</sup>	6.10±0.00 <sup>c</sup>	4800.3±1.30 <sup>d</sup>

Group with different superscripts within the same column are significantly different at P<0.05.

On the other hand, gross energy contents were significantly (P<0.05) increased in all DY and BZ groups, and significantly decreased in BG1 and BG2 groups (Table 7).

In general, fish in DY3 group showed significantly (P<0.05) the lowest moisture content and the highest CP, EE, ash and energy contents. Similar trend of change in DM, CP EE and ash in body of fish fed DY were obtained by Magouz *et al.* (2002) in Nile tilapia fish fed diet supplemented with Lacto-Sacc. Generally, boy composition of fish in all groups is within the range reported by Abdelhamed *et al.* (2000)

#### Histopathological signs:

The histological examination of kidney and liver of fish in different treatment groups revealed normal histological structure of these organs, except signs of harmful in kidney of VC3 group showing abnormal architecture of the renal cortex and congestion of glomerulosa and leading to nephritis (Figs. 1 and 2). In liver of fish in VC3 group, only abnormal hepatic lobules and lymphatic nodules were seen between the hepatocytes (Fig. 3). Kidney of fish in DY1, DY2 and DY3 groups showed normal architecture of the renal cortex and intact structure of both glomerulosa and renal tubules (Figs. 4, 5 and 6).

However, liver of fish in all DY groups showed normal architecture of the hepatic lobules, central vein and hepatocytes, but small vacuoles were found within the portal lobules (Fig. 7). In liver of fish in BZ group, normal architecture of the hepatic lobules,

but the central vein was branched and wide sinusoid as well as lymphatic population within the hepatic lobules and the portal lobules (Figs. 8 & 9).

Liver of fish in DY3 showed normal architecture of the hepatic lobules, some vacuoles within the hepatic lobules (Fig. 10). On the other hand, kidney of fish in BG group showed normal architecture of the renal cortex and intact structure of both glomerulosa and renal tubules, but mild infiltration of fibroblast cells was observed between the renal tubules (Fig. 11).

# **Economic feed efficiency:**

In comparing price of each ton of feed used different treatment groups, BG3 group had the highest price and the control feed showed the lowest price. Such trend was associated with the highest price of each gram and high level of supplementation from Bacillozyme as compared to the other supplements (Table 8).

Based on feed cost per kg gain, fish in DY3, BZ3, BG3 and VC1 showed significantly (P<) lower feed price to produce one kg gain in weight as compared to the control group. This was mainly attributed to the higher total gain of fish in spite of the higher price of feed (Table 8).

Generally, fish in BG3 group showed the highest total gain and the lowest feed cost per kg gain, leading to the best economic feed efficiency (Table 7).

Table 8. Economic feed efficiency of *O. niloticus* fish as affected by dietary additives in treatment groups.

Treatment group	Feed intake (g)	Price of each ton feed (L.E.)	Total gain (g)	Feed cost/kg gain (L.E.)
Control	21.70	1600	5.59	6.21 <sup>b</sup>
DY1	23.28	1760	8.49	4.83 <sup>d</sup>
DY2	22.20	1840	8.92	4.58 <sup>d</sup>
DY3	26.01	1920	9.01	5.54 <sup>:</sup>
BZ1	21.70	1712	5.59	6.65 <sup>b</sup>
BZ2	21.60	1824	5.99	6.50 <sup>b</sup>
BZ3	22.40	1936	8.11	5.35°
BG1	23.45	1750	6.39	6.42 <sup>b</sup>
BG2	22.58	1900	7.09	6.05⁵
BG3	21.50	2050	9.39	5.35°
VC1	24.54	1615	7.39	5.36 <sup>c</sup>
VC2	22.02	1645	6.37	5.69 <sup>c</sup>
VC3	22.38	1660	3.49	10.6ª

Group with different superscripts within the same column are significantly different at P<0.05.

According to local market price, 2005, price was 1600 L.E. for each ton basal diet, 16 L.E. for each kg-dried yeast, 28 LE. For 250 g Bacillozyme, L.E. for 200 g Bacillogene and 15 L.E. for 500 g vit. C.

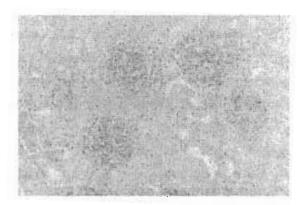


Fig. 1. Section in kidney of fish in VC3 group showing abnormal architecture of the renal cortex and congestion of glomerulosa and leading to nephritis (x 150, H&E stains).

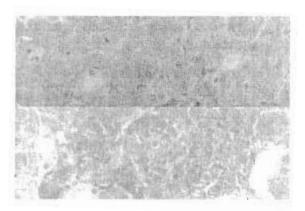


Fig. 2. Magnification of the previous section in kidney of fish in VC3 group showing sever congestion in glomerulosa. (x 250, H&E stains)

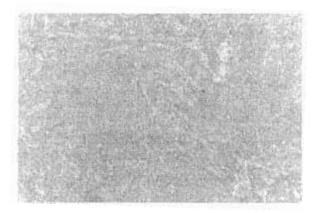


Fig. 3. Section in liver of fish in VC3 group showing abnormal hepatic lobules and lymphatic nodules were seen between the hepatocytes. (x 250, H&E stains).

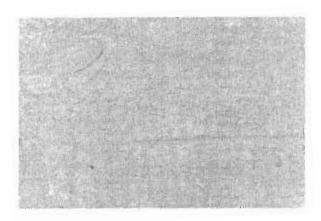


Fig. 4. Section in kidney of fish in DY1 group showing normal architecture of the renal cortex and intact structure of both glomerulosa and renal tubules. (x 120, H&E stains)



Fig. 5. Section in kidney of fish in DY2 group showing normal architecture of the renal cortex and intact structure of both glomerulosa and renal tubules. (x 120, H&E stains)

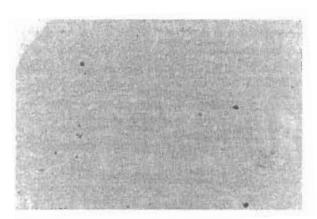


Fig. 6. Section in kidney of fish in DY3 group showing normal architecture of the renal cortex and intact structure of both glomerulosa and renal tubules. (x 120, H&E stains)

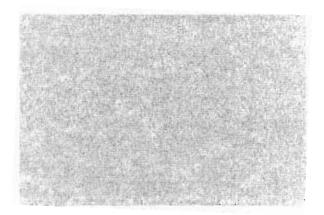


Fig. 7. Section in liver of fish in DY showing normal architecture of the hepatic lobules, central vein and hepatocytes, but small vacuoles were found within the portal lobules. (x 120, H&E stains)

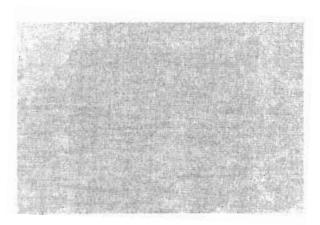


Fig. 8. Section in liver of fish in BZ showing normal architecture of the hepatic lobules, but the central vein was branched and wide sinusoid as well as lymphatic population were observed within the hepatic lobules and the portal lobules. (x 120, H&E stains)

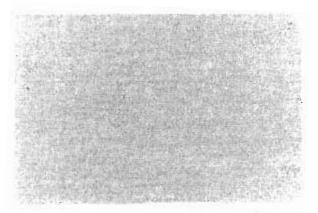


Fig. 9. Section in liver of fish in BZ showing normal architecture of the hepatic lobules, but the central vein was branched and wide sinusoid as well as lymphatic population were observed within the hepatic lobules and the portal lobules. (x 120, H&E stains)

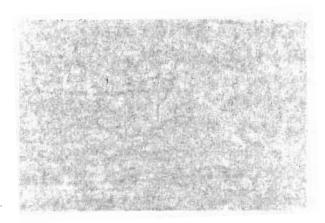


Fig. 10. Section in liver of fish in DY3 showing normal architecture of the hepatic lobules, some vacuoles within the hepatic lobules. (x 150, H&E stains)

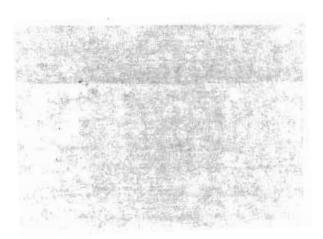


Fig. 11. Section in kidney of fish in BG group showing normal architecture of the renal cortex and intact structure of both glomerulosa and renal tubules, but mild infiltration of fibroblast cells was observed between the renal tubules (x 150, H&E stains).

# REFERENCES

- Abed El-Aziz, M. A. and M. A. Mahmoud. 2004. Study on the effect of vitamin C deficiency on *Oreochromis Niloticus* under intensive culture. The first Sci. Conf. Fac. Vet. Med Moshtohor. Banha-Ras-Sedr,527-548.
- Abdelhamed, A. M., F. F. M. Kalil and M. A. A. Seden. 2000. Possibility of using dried yeast and lactosacc in Nile tilapia fingerlings diets. J. Agri. Sci. Mansora Univ. 25:4905-4911.
- A. O. A. C. 1990. Association of Official agricultural chemists Official methods of analysis 15 <sup>th</sup> ed. Published by the A.O.A.C Benjamin Franklin station. Washington. D.C.
- 4. Castell, J. D. and K. Tiews. 1980. Report of the EIFAC. IUNS and ices Woring group on the standardization of methodology in fish nutrition research Hamburg fed. Rep. Germany, 21-23. March 1979. EIFAC Teeh.pap.. No.36, 24 pp.

- Collins, R. A. and M. N. Delmando. 1979. Comparative economics of aquaculture in cages ,raceways and enclosures. In: Advance in aquaculture, PP.427-437. England, Fishing News Books.
- 6. Davies, S. J. and P. C. Morris. 1997. Influence of multiple amino acid supplementation on the performance of rainbow trout, *Oncorhynchus mykiss* (walbaum), fed soya based dists .Aqua Res., 28:65-74.
- 7. Doumas, B. T., W. A. Watson and H.G. Biggs. 1971. Albumin standards and the Measurements of serum Albumin with Bromocresol Green. Clinical chemistry Acta, 31, 87-96.
- 8. Duncan, D. B. 1955. Multiple rang and multiple F tests. Biometrics, 11: 1-42.
- 9. Duncan P.L. and Lovell, R. T. 1994. Growth, Survival and Blood comparison of channel catfish (*Ictalurus punctatus*).
- 10. Drury, R. A. B. and E. B. Wallington. 1980. Carlton's Histology Techniques. 5<sup>th</sup> ed., pp. 139- 307, Oxford Univ. Press, England.
- 11. El-Hindawy, M. M, M. I. Tawfeek and A. M. Omar. 1997. Growth performance of broiler chicks as affected by dietary energy levels and biological feed additives supplementation. International Conference on animal production and health The Eg6yptian international center for Agriculture, Cairo, Egypt PP401-414.
- El-Hindawy, M. M., K. A. Yamani and M. I. Tawfeek. 1993. Effect of probiotic (lacto sacc) in diets with different protein levels on growth performance, digestibility and some carcass aspects of growing rabbits. Egypt J. Rabbits. Sci.3:13-28.
- 13. El-Hindawy, M. M., K. A. Yamani, M. I. Tawfeek and B. M. Khashaba. 1994. Performance and weaning rabbitsas affected by energy level and inclusion of probiotics in the diet. Proceeding of the first international Conference on rabbits production in Hot climates, Egypt, 157-163.
- 14. Fuller, R. 1989. A review probiotics in man and animals. J. Appl. Bacteriol, 66:365-378.
- 15. Kumar, B. S., S. K. Vijaysarathi and S. Raq. 2003. Effect of feeding probiotics on the performance of broilers in experimental fowl typhoid. Indian Vet.J., 80:52-55.
- 16. Magous, F., M. K. Mohsen, and A. H. Gooda. 2002. Effect of including some biological feed additives in the diet on growth performance and feed efficiency of Nile Tilapia (*Oreochromis niloticus*). Proc.2<sup>nd</sup> Conf. Food borne Contamination and Egyptian's Health El-Mansoura Egypt. 329-339.
- 17. McDowell, H. R. 1989. Vitamins in Animal Nutrition. Acad. Press, San Diego, CA.
- 18. Merck, E. 1974. Clinical laboratory. 11<sup>th</sup> Ed. Of Micro-chimical investigation Methods. Darmstadt, Federal Republic Germany.

- 19. Pollman, D. S., D. M. Danielson and E. R. Peo. 1980. Effects of microbial feed additives on performance of starter and growing finishing pigs. J. Animal Sci..51:577-581.
- 20. Pouomogne, V. and J. Mongblang. 1993. Effect of feeding rate on the growth of tilapla (*Oreochromis nillotitcus*) in earthen ponds. Bamidgh, 45:147-153.
- 21. Ragheb E. E., A. F. Mehrez and A. E. Abdel-Khalek. 2003. Digestibility coefficients, blood parameters, feed efficiency and growth performance of weaned friesian calves fed diet supplemented with Lacto-Sacc. The 9<sup>th</sup> Conference on Animal Nutrition, Hurghada, 14-17 October, Egyptian J. Nutr. Feeds, 6: (Spicial Issue).
- 22. Reitman S. and S. Frankel. 1957. Determination of AST and ALT in serum. Am. J. clinic. Path., 28: 56-68.Sisson, J. W.1989. Potential of probiotic organisms of prevent diarrhoea and promote digestion in farm animals A review. J. Sci. Food Agri.,49:1-13.
- 23. Snedecor, G. W. and W. G. Cochran. 1982. Statistical Methods, 7<sup>th</sup> ed. Iowa State Univ. Press, Ames, Iowa, USA.
- 24. Statistical Analysis System. SAS. 1992. SAS/STAT User's Guide Release 6.03 edn. SAS institute. Cary, NC. 1028 pp.
- 25. Tacon, A. 1987. The nutrition and feeding of farmed fish and shrimp a training manual.V61. 1. The essential nutrients FAO.PP. 117-130.
- 26. Tawfeek, M. I. and M. M. El-Hindawy. 1991. Reproduction and growth performance of NZW and CAL rabbits as affected by supplementation p of Lactosaccduring summer. Egypt. J. Rabbit. Sci 1:124-135.
- 27. Wary, C. D. and R. H. Davies. 2000. Comparative exclusion an alternative to antibiotics, Vet. J.,59:107-108.
- 28. Yamani, K. A., H. Ibrahim A. A. Rashowan and K. M. El-Gendy. 1992. Effect of a pelleted diet supplementation with probiotic Lacto-Sacc and Water supplemented with a combination of probiotic and acidifier (Acid-Pak4-Way) on digestibility, growth, carcass and physiology aspects of weaniling rabbits. J.Appl. Rabbit. Res. 15:1087-1100.

# بعض الإضافات الغذائية في مستويات مختلفة في عليقه البلطي النيلي

# عادل عزت طولان

المعمل المركزي لبحوث الثروة السمكية بالعباسة مركز البحوث الزراعية- جيزة - مصر

تم القيام بهذا العمل لتقيم تأثير بعض الإضافات الغذائية في عليقه بعض البلطي النيلي، وقد استخدمت الأنواع الانتها الخميرة ونوعين من البروبيوتيك (الإضافات الحيوية) باسبلوزيم و باسبلوجين وفيتامين ج في ثلاث مستويات على الحاله الانتاجيه وكفاءة التمثيل الغذائي والهستوباثولوجي للكبد والكلي التركيب الكيماوي لجسم السمك والتحليل الاقتصادي وقد تم استخدام عدد ٣٩٠ اصبعية من نوع بلطي نيلي في ٣٩ حوض زجاجي مقسمة آلي أربع معاملات بالإضافة ألي المعاملة الضابطة وكل معامله مقسمة الى ثلاث مستويات وكل مستوى له ثلاث مكررات وتم تخزين ١٠ اصعيات في كل حوض وكان العلف المصنع يحتوي علي نسبة ٢٢ % بروتين وقد تم إضافة الخميرة بالنسب آلاتية ( ٢٠٠٠،١٥٠٠ جرام/كيلوجرام علف) باسيلوجين وياسلوزيم (٢٠٠٠ جرام كيلو جرام علف جرام /كيلو جرام علف) وفيتامين ج بالنسب آلاتية ( ٢٠٠٠،١٥٠٠ ملج / كيلوجرام علف دون اي ظهور لاي صورة مرضية في الدم او للفحص الهستوباثولوجي وكذلك بالنسبة للخميرة ٢٠٠٠ ملج /كيلوجرام علف الكل كيلوجرام علف وكانت أسوء النتائج التي تم تسجيلها في حالة فيتامين ج النسبة مدين حد ٢٠٠٠ ملج /كيلوجرام علف كيلوجرام علف وكانت أسوء النتائج التي تم تسجيلها في حالة فيتامين ج ٢٠٠٠ ملج /كيلوجرام علف .