

EFFECT OF YEAST OR SELENIZED YEAST SUPPLEMENTATION TO RATIONS ON THE PRODUCTIVE PERFORMANCE OF LACTATING BUFFALOES.

S. M. Kholif¹ and M. M. Khorshed²

¹Dairy science Dept. National Research Centre, Dokki, Egypt.

²Animal production Dept., Faculty of Agriculture, Ain Shams Univ., Cairo, Egypt.

(Received 12/4/2006, accepted 3/9/2006)

SUMMARY

Two experiments were carried out in this study. In the first experiment, three Baladi castrated male goats were ranked in 3x3 latin square design experiment with 30 days interval periods to study the effect of adding yeast or selenized yeast to goat's diet on their rumen activity. The control group was given a basic diet consisting of (50% concentrate feed mixture (CFM) + 50 % berseem clover). The yeast or selenized yeast supplemented groups (2 and 3) were given the control diet with 2.5 g/h/d yeast or selenized yeast, respectively. Animals fed selenized yeast had a higher rumen total nitrogen ($P<0.01$), non protein nitrogen ($P>0.05$) and true protein nitrogen ($P<0.05$), while, ammonia nitrogen and total volatile fatty acids were not affected by any of the yeast supplementations.

In the second experiment, fifteen lactating buffaloes were divided randomly into three groups (of 5 animals each) using complete random block design experiment with 90 days period. The control group was given a diet consisting of (60% CFM: 20% berseem clover: 20% rice straw), the yeast group was fed the control ration plus 10 g/h/d yeast and the selenized yeast group was fed the control ration plus 10 g/h/d selenized yeast. Individual milk samples were collected every two weeks of the experimental period for chemical analysis. After 4 hrs of the morning meal, grab samples and blood serum samples were collected in the last day every month for chemical analysis. Yeasts supplementation were improved the nutrients digestibility coefficient compared with control. Supplementation of yeast or selenized yeast to buffaloes rations increased ($P<0.05$) milk yield, milk protein and lactose contents compared with control. 4% FCM yield, milk constituents yield and fat content were slightly higher with animals fed yeasts than those of the control group. Feed efficiency (Milk yield/DMI and FCM/DMI) was higher ($P<0.05$) for animals fed yeast or selenized yeast than animals fed control. Nutrients digestibility coefficient of organic matter, crude protein, crude fiber and nitrogen free extract were significantly higher with animals fed on yeasts than control. Blood serum glucose and albumin were higher ($P<0.05$) in animals fed on yeast or selenized yeast supplemented rations. It could be concluded that organic selenium supplementation to diets improved rumen activity and milk constituents, while the yeast supplementations were improved milk production and nutrients digestibility coefficients.

Keywords: *yeast, selenized yeast, rumen activity, digestibility, buffaloes, milk production, blood serum parameters.*

INTRODUCTION

Yeast and yeast cultures have been fed to dairy cattle for more than 60 yr with varied responses. In some studies, yeast cultures improved DMI (Williams et al., 1991; Wohlt et al., 1991; Dann et al., 2000) and milk production (Williams et al., 1991; Wohlt et al., 1991; Piva et al., 1993; Kung et al., 1997; Wang et al., 2001; Kholif et al., 2005), whereas other studies (Erdman and Sharma, 1989; Arambel and Kent, 1990; Soder and Holden, 1999) found no response to yeast cultures. Some of the benefits associated with *Saccharomyces cerevisiae* include increased DM and NDF digestion (Williams et al., 1991 and Carro et al., 1992; Kholif et al., 2005). Yeast cultures also have been shown to stimulate utilization of hydrogen by ruminal acetogenic bacteria (Chaucheyras et al., 1995). Wallace and Newbold (1992) reported that the responses of yeast culture are highly variable and apparently influenced by the composition of the diet. *Saccharomyces cerevisiae* (Yea-Sacc 1026) increased the number of ruminal total bacteria and cellulolytic bacteria (Newbold et al., 1995), increased the proportion of propionate (Mutsvangwa et al., 1992; Newbold et al., 1995), and decreased lactate concentration (Newbold et al., 1990). The Egyptian soil had a shortage in selenium which effect on plants, animals and human health. Selenium is an essential microelement for animal and human diets. It was identified as a part of cellular glutathione peroxidase, which provided evidence for selenium involvement in other metabolic processes (Heider and Bock, 1993). Its deficiency in nutrition may causes decreases the productivity of

domesticated animals (Foster and Sumer, 1997). Recently, researchers found evidence for selenium as a cancer-protective agent (Ip and Lisk, 1994). Therefore, selenium must be provided to human and animals as a part of nutritional intake. El-Batal and Fadel (2002) produced an edible food yeast (*S. cerevisiae*) having high levels of organically bound intracellular selenium in an assimilable non-toxic form, which is useful as a dietary supplement. Kholif (2005) used selenized yeast in lactating buffaloes diets to increase milk yield and its protein content.

This study was conducted to evaluate the effects of organic selenium supplemented rations on the productive performance of lactating buffaloes.

MATERIALS AND METHODS

This study was conducted at the Experimental Farm in Shalakan, Faculty of Agriculture, Ain Shams University and Dairy Science Department, National Research Center, Dokki, Giza, Egypt, during 2004-2005.

Microorganisms:

Yeast (*Saccharomyces cerevisiae* F-25) and selenized yeast (*Saccharomyces cerevisiae* F-25 with organic selenium) were obtained from Microbial Chemistry Lab. National Research Center, Dokki, Cairo, Egypt. The cultures were maintained on Malt agar medium.

Preparation of high selenium yeast in shaking flasks:

Cane molasses medium composed of (g/L): molasses, 100 (42% sucrose 2.0; orthophosphoric acid, 2.0 and

sodium selenite, 0.05 (pH 5.5). The medium (50 ml in 250 ml Erlenmeyer flask) was autoclaved at 121°C for 15 min. The yeast culture was harvested by centrifugation at 3000 rpm for 15 minutes. The yeast yield was washed 5 times with distilled water. The yeast cells obtained was dried at air flow 18 °C till standard weight. Total count of yeast live cells was determined using agar plat count and selenium contents was determined according to (El-Batal and Fadel 2002). Fermentation was performed in 220 (rpm) shaking at 32°C for 72 h inoculation. The employed yeast cells have 4.2×10^{10} live cells/gas well as containing 1000 µg selenium/g dry cells.

Animals and diets:

Three Baladi castrated male goats (being 4 years old and weighting on average of 29 kg) were divided into three groups to study the effect of yeast or selenized yeast supplementation to goats ration on rumen activity using 3x3 Latin square design experiment for 30 days interval periods. The control diet used consisted of berseem clover (B) and concentrate feed mixture (CFM) (50 : 50 on dry matter basis). The experimental diets used were : control diet plus 2.5 gm/head/day yeast (*Saccharomyces cerevisiae*) (Yeast) and control diet plus 2.5 gm/head/day selenized yeast - contained 2.5 mg/h/d organic selenium - (Selenized yeast).

Fifteen lactating buffaloes after 7 days of parturition were divided to three groups - according to milk production and lactation season - to study the effect of yeast or selenized yeast supplemented buffalo's rations on blood serum parameters, nutrient digestibility coefficients and milk yield and composition using complete random block design experiment with 90 days

period. Experimental rations were control (60% CFM: 20% berseem : 20% rice straw), treatment I (control ration plus 10 gm/h/d yeast), treatment II (control ration plus 10 g/head/day selenized yeast - contained 10 mg/head/day organic selenium). The CFM was consisted of 35% yellow corn, 25 % wheat bran, 22% decorticated cotton seed meal, 15% rice bran, 1.5% ground limestone and 1.5% common salt. The chemical composition of ingredients is showed in Table (1). The offered feeds were assessed to cover the requirements for each animal (A.R.C. 1965). The CFM for each animal was offered individually once daily at 8.00 am, while berseem clover and rice straw were offered at 10.00 am and 4 pm. Dry matter intake was measured during the last refusals of the previous day. Clean water was available to animals at all times.

Analysis of feed samples:

Samples of ingredients and rations were analyzed for DM, ash, CF, organic matter (OM) and ether extract (EE) according to methods of A.O.A.C. (1995). While, nitrogen-free extract (NFE) was calculated.

Rumen liquor analysis:

At the last of each period, rumen liquor samples were collected from each goats at 4 hrs. post morning feeding by a stomach tube. The samples were strained through two layers of cheese cloth and then stored in glass bottles (10 ml) with 3 drops of toluene and a thin layer of paraffin oil just to cover the surface to stop microbial activity and to prevent volatilization and stored at - 18°C till they were analyzed. Ruminal pH was determined using a digital pH-meter, total nitrogen (TN), non-protein-nitrogen (NPN) and NH₃-N were determined according to A.O.A.C.

(1995). True protein nitrogen (True-PN) was calculated by difference (TN-NPN). Total volatile fatty acids (TVFA's) were determined by steam distillation as described by Warner (1964).

Digestibility trial:

Simultaneously three digestibility traits were carried out on all animals (buffaloes) of each treatment of feeding trail and repeated each 30 days of the experimental period. Grab sample method was used and silica as internal marker was applied for determining the digestibility. Feces grab samples were collected handily at 8.00 for three successive days from each animal. Solution of 10% H₂SO₄ were added to the representative samples then dried in oven at 70°C for 24 hours. The dried feces samples from each animal were mixed and stored at -18°C for chemical analysis. The digestibility coefficient was calculated according to the following formula according to (Gallup *et al.*, 1945 and Forbes and Garrigus 1948).

Sampling and analysis of milk:

Individually milk samples were collected every two weeks of the experimental period (90 days). The buffaloes were handily milked (twice/day), milk yield was recorded and pH of milk was determined (Ling, 1963). Milk samples were also, analyzed for fat, total solids (TS), total protein (TP) and ash (Ling, 1963) lactose (Barnett and Abd El-Tawab, 1957). solids-not-fat (SNF) was calculated by difference.

Blood serum analysis:

Blood samples were collected from the jugular vein of each animals (buffaloes) at the last day of each period (4 hr. post morning feeding). The collected blood samples were

centrifuged at 4000 r.p.m./20 min. to separate the serum. The obtained serum was stored at -18°C till it was analyzed. Serum total protein was determined as described by Armstrong and Carr (1964), albumin (Doumas *et al.* 1971), urea (Patton and Crouch,1977), glucose (Siest *et al.*,1981), serum GOT and GPT (Reitman and Frankel, 1957) and cholesterol (Raltiff and Hall 1973). Globulin and albumin/globulin ratio were calculated.

Statistical analysis:

Data obtained from this study were statistically analyzed according to procedures outlined by Snedecor and Cochran (1982). The procedure was;

- 1- Latin square design for rumen liquor data using the general linear model procedure:

$$Y_{ijk} = \mu + R_i + C_j + T_k + e_{ijk}$$

Where Y_{ijk} is the parameter under analysis of the *ijkl* goat, μ is the overall mean, R_i is the effect due to the period on the parameter under analysis, C_j is the effect due to the animals on the parameter under analysis, T_k is the effect due to treatment on the parameter under analysis, e_{ijk} is the experimental error for *ijk* on the observation.

- 2- Complete random block design for milk, blood and digestibility data using the general linear model procedure:

$$Y_{ijk} = \mu + R_i + T_k + e_{ijk}$$

Where Y_{ijk} is the parameter under analysis of the *ijkl* goat, μ is the overall mean, R_i is the effect due to the lactation period on the parameter under analysis, T_k is the effect due to treatment on the parameter under

analysis, e_{ijk} is the experimental error for ijk on the observation.

The Duncan's multiple range test was used to test the significance between means (Duncan, 1955).

RESULTS AND DISCUSSION

Dry matter intake:

Dry matter intake did not differ among treatments, being 15.56, 15.35 and 15.65 kg/h/d in control, yeast and selenized yeast, respectively. The animals of selenized yeast treated group consumed higher ($P>0.05$) amount of DM following by control and then yeast treated group. Values of grams consumed/kg metabolic body size (MBS) were similar among treatments being 136.40, 136.10 and 136.86 g/MBS. These results are in agreement with El-Ashry et al., (2001)

Rumen liquor parameters:

Effects of treatments on characteristics of ruminal fermentation are shown in Table (2). Ruminal pH values were significantly higher ($P<0.05$) in yeasts treated groups than control group. All values were above pH 6.0 which indicated a better digestion of cellulolytic materials (Mertens, 1978). Also, TVFA's value was slightly increased with animals fed yeast supplemented groups compared with control group. The major effect of yeast on Ruminal fermentation included increased concentrations of VFA and propionate (Lila et al., 2004).

Rumen total nitrogen ($P<0.01$) and true protein nitrogen ($P<0.05$) were significantly increased with animals fed on selenized yeast compared with other treatments. Its interested to note that, the highest improvement of nutrients

digestibility (Table, 3), and highest values of rumen total nitrogen and true protein nitrogen and TVFA's which observed with animals fed on selenized yeast followed by yeast led to concluded the highest improvements of rumen environment and microflora activity which produced more microbial protein. Hieder and Bock (1993) suggested that selenium was identified as a part of cellular glutathione peroxidase, which provided evidence for selenium involvement in other metabolic process. It has been suggested that increased bacterial flora in animals fed *S. cerevisiae* is central to the action of yeast in the rumen and increased bacterial population leads to an increase in both the degradation of fiber in the rumen and the flow of microbial protein from the rumen (Wallace and Newbold, 1992).

Ammonia-N concentration was not modified by the addition of yeasts which was similar to the results of Mutsvangwa et al., (1992) in lactating dairy cows fed yeast (10 g/d). Although no significant difference ($P>0.05$) were detected among different treatments in ammonia-N and non protein nitrogen concentrations, values of ammonia-N seemed to be sufficient to cover the microbial demands for microbial protein synthesis as obtained from the reports of previous investigators (Mehrez et al., 1977, Wallace, 1979 and Erdmann et al., 1986).

Nutrients digestibility coefficient:

Table (3) showed nutrients digestibility coefficient as affected by experimental treatments. Organic matter and crude fiber digestibilities were significantly increased ($P<0.01$) with animals fed ration supplemented with yeasts compared with control. This improvements of crude fiber digestibility may be due to the increase

Table (1): Chemical composition of concentrate feed mixture (CFM), rice straw (RS) and berseem clover (B) (DM basis).

<i>Item</i>	<i>CFM</i>	<i>RS</i>	<i>B</i>
Dry matter	91.29	93.39	18.0
Ash	10.11	13.23	12.1
Organic matter	89.89	86.77	87.9
Crude protein	14.15	3.96	12.8
Ether extract	4.05	4.55	2.5
Crude fiber	15.33	35.5	28.2
Nitrogen free extract	56.36	42.76	44.4

Table (2): Rumen liquor contents as affected by supplementing goat's ration with yeast or selenized yeast (the first experiment).

<i>Item</i>	<i>Control</i>	<i>Yeast</i>	<i>Selenized yeast</i>	\pm <i>SE</i>
Total nitrogen (mg/100ml)	148.7 ^B	150.45 ^B	195.10 ^A	0.278
Non protein nitrogen (mg/100ml)	65.34	63.62	74.10	0.685
True protein nitrogen (mg/100ml)	83.36 ^b	86.83 ^b	120.07 ^a	0.472
Ammonia nitrogen (mg/100ml)	30.77	27.37	30.58	1.191
TVFA's (m.eq/100ml)	6.28	6.77	6.60	0.157
pH	6.25 ^B	6.28 ^A	6.30 ^A	0.020

Table (3): Nutrients digestibility coefficient as affected by supplementing lactating buffalo's rations with yeast or selenized yeast (the second experiment).

<i>Item</i>	<i>Control</i>	<i>Yeast</i>	<i>Selenized yeast</i>	\pm <i>SE</i>
DM	58.96	61.59	60.29	1.721
OM	59.88 ^B	63.25 ^A	62.91 ^A	2.130
CP	59.69 ^b	65.93 ^a	63.11 ^a	1.115
EE	58.31	64.31	64.11	0.932
CF	57.44 ^B	68.55 ^A	67.31 ^A	1.583
NFE	56.99 ^b	65.39 ^a	63.21 ^a	2.343

A,B,C means with different superscripts are significant ($P < 0.01$) difference.

a,b,c, means with different superscripts are significant ($P < 0.05$) difference.

of the numbers of rumen total viable bacteria and cellulolytic bacteria with animals fed yeast (Lila et al., 2004 and Newbold et al., 1995). The digestion coefficients of crude protein and NFE were also significantly improved ($P < 0.05$) by the yeasts supplementation compared with control. Digestibility of dry matter and ether extract were followed the same trend, however, the effect of yeasts failed to be significant. There was no differences effect between two yeasts on nutrients digestibility coefficient. In total, there is no evidence for an improvement on nutrients digestibility coefficient through additional selenium to yeast. Similar results obtained by El-Waziry et al., (2000) and El-Ashry et al., (2001). Williams and Newbold (1990) suggested that yeast culture alter the site of digestion and that total tract digestibility.

Milk yield and composition:

Data of milk and milk constituent yields of the experimental buffaloes are summarized in Table (4). Milk yield was significantly ($P < 0.05$) higher in yeasts supplemented groups representing an increase of about 9.35 % than buffaloes receiving control. Also, 4% FCM was insignificantly ($P > 0.05$) higher in yeasts supplemented groups representing an increase of about 11.95 and 12.23% for yeast and selenized yeast, respectively, than buffaloes receiving control. As an impact of the increased milk yield, daily fat, TS, SNF, TP, lactose and ash yields were insignificantly higher ($P > 0.05$) in groups received yeast or selenized yeast than that received control ration.

Data of milk composition of the experimental buffaloes are also summarized in Table (4). Milk protein and lactose contents were higher ($P < 0.05$) in T₃ (selenized yeast)

followed by T₂ and then control. From these data we can concluded the positive effect of selenium on the metabolic process in the mammary gland which led to the increase of milk protein and lactose synthesis. The higher milk yield with the animals fed yeasts supplemented ration might be attributed to the positive effect of yeasts on the digestibility of organic matter and its nutrients as shown in Table (3). These results also probably attributed to the higher of blood serum glucose and albumin concentration of animals fed yeasts supplemented ration as shown in Table (5). It led to an increase in milk lactose synthesis and consequently milk production being increase.

Milk fat content was insignificantly increased with yeasts treated groups compared with control. However, milk total solids, solids not fat and ash contents. Milk pH values were not significantly differ among treatments. Generally, feed efficiency calculated as milk yield / DMI and 4% FCM / DMI were significantly improved ($P < 0.05$) with animals fed yeast supplemented ration followed by selenized yeast supplemented ration and then control.

Concentrations of selenium in colostrum and milk were about 1.8 times greater when cows were fed selenized yeast (Weiss and Hogan, 2005). From this finding we can produce selenium supported milk for humans. Selenium deficiency in nutrition may causes cardiological and oncological human diseases (Foster and Sumer, 1997). Researchers found evidence for selenium as a cancer-protective agents (Ip and Lisk, 1994). Therefore, selenium must be provided to human and animals as a part of nutritional intake.

Blood serum metabolites:

Table (4): Milk yield and composition as affected by supplementing lactating buffaloes rations with yeast or selenized yeast (the second experiment).

Item	Control	Yeast	Selenized yeast	±SE
No. of animals	5	5	5	
Live body weight	553	545	555	2.124
Metabolic body size (W ^{0.75})	114.4	112.80	114.35	0.524
Dry matter intake (kg/head/day):	15.56	15.35	15.65	1.023
kg DM/kg W ^{0.75}	136.4	136.1	136.86	0.221
Milk yield (kg/d)	8.98 ^b	9.82 ^a	9.82 ^a	0.216
4% FCM (kg/d)	12.13	13.58	13.62	0.558
Yields (g/d)				
Fat	569	643.1	645.9	4.16
Protein	380.3	437.8	424.2	2.985
Lactose	369.6	408.5	433.6	2.411
TS	1486.1	1645.1	1618.1	5.909
SNF	903.8	1002	972.1	4.37
Ash	70.36	79.88	79.12	1.212
Milk composition %				
Fat	6.28	6.36	6.40	0.134
Protein	4.14 ^b	4.25 ^{ab}	4.36 ^a	0.045
Lactose	4.15 ^b	4.30 ^{ab}	4.46 ^a	0.073
TS	16.42	16.48	16.48	0.109
SNF	10.25	10.13	10.07	0.053
Ash	0.777	0.812	0.795	0.030
PH	7.01	7.06	7.05	0.013
Feed efficiency:				
Milk yield/DMI	0.577 ^b	0.640 ^a	0.627 ^a	0.896
FCM yield/DMI	0.780 ^b	0.885 ^a	0.870 ^a	0.685

^{a,b,c} means with different superscripts are significant (P<0.05) difference.

Table (5): Blood serum contents as affected by supplementing lactating buffalo's rations with yeast or selenized yeast (the second experiment).

Item	Control	Yeast	Selenized yeast	±SE
Total protein (mg/100ml)	7.40	7.01	7.49	0.122
Albumin (mg/100ml)	4.11 ^b	4.50 ^{ab}	4.83 ^a	0.081
Globulin (mg/100ml)	3.29	2.52	2.65	0.225
A/G ratio	1.52	1.95	2.03	0.193
AST (U/100ml)	29.66	30.11	28.30	0.536
ALT (U/100ml)	13.2	12.90	12.98	0.235
Glucose (mg/100ml)	55.37 ^b	61.57 ^a	63.06 ^a	0.559
Urea (mg/100ml)	38.34	40.08	37.84	0.273
Cholesterol (mg/100ml)	109.39	100.54	105.23	0.823

^{a,b,c} means with different superscripts are significant (P<0.05) difference.

Data in Table (5) showed no significant differences ($P < 0.05$) among different treatments on some blood serum parameters, except glucose and albumin values ($P < 0.05$), which were increased with animals fed selenized yeast followed by yeast and then control. These results maybe due to the improvements occurred in metabolic process as a selenium and yeast response.

Diets containing yeast selenium tend to increase total protein and A/G ratio however, globulin was slightly decreased with animals fed yeasts. Serum glucose had the same trend of milk yield (Table, 4) which was in accordance with the results of Clark et al., (1977) who claimed a positive correlation between blood glucose and milk yield. Yeasts supplementation to animals ration were not affected serum cholesterol. Blood serum glutamic-oxaloacetate-transaminase (GOT) and glutamic-pyruvate-transaminase (GPT) values were not significantly affected by treatments. These results indicated that supplementing yeast or selenized yeast to lactating buffaloes' rations were not affecting on liver activity or animals health.

The concentration of Se in serum at calving and until 28 day in milk was about 1.4 times greater for cows fed selenized yeast (0.3 mg Se/kg DM) than for those fed sodium selenate (0.3 mg Se/kg DM) (Weiss and Hogan, 2005).

CONCLUSION

It could be concluded that supplementing buffaloes' ration with the proposed of yeast or selenized yeast improved rumen fermentation and nutrients digestibility coefficients. Also, yeasts supplementation improved feed efficiency, milk production and

composition with no deleterious effect on general health of the treated animals as compared to animals fed the control ration. Milk supported with selenium can protect humans against cancer. Although inorganic selenium in the diets was enough, lactating buffaloes have a positive response to organic selenium supplementation.

REFERENCES

- A.O.A.C. (1995). Official Methods of Analysis. 16th Ed. Vol. 1, "Agricultural, Chemicals, Contaminants, Drugs". Washington, D. C., USA.
- Arambel, M. J., and B. A. Kent. (1990). Effect of yeast culture on nutrient digestibility and milk yield response in early to midlactation dairy cows. *J. Dairy Sci.* 73:1560–1563.
- A.R.C. (1965). The Nutrient Requirement of Livestock. no.2 Ruminants Tech. Rev. and summaries Agric. Res. Council, London, UK.
- Armstrong W. D. and C. W. Carr (1964). *Physiological Chemistry: Laboratory Directions*, 3rd ed. PP. 75, Bures Publishing Co. Minneapolis, Minnesota, USA.
- Barnett A. J. G. and G. Abd El-Tawab (1957). Determination of lactose in milk and cheese. *J. Sci. Food Agric.*, 8: 437.
- Carro, M. D., P. Lebzien and K. Rohr. (1992). Effects of yeast culture on rumen fermentation, digestibility and duodenal flow in dairy cows fed a silage based diet. *Livest. Prod. Sci.* 32:219–229.
- Chaucheyras, F., G. Fonty, G. Bertin and P. Gouet. (1995). In vitro H₂

- utilization by a ruminal acetogenic bacterium cultivated alone or in association with an archaea methanogen is stimulated by a probiotic strain of *Saccharomyces cerevisiae*. Appl. Environ. Microbiol. 61:3466–3469.
- Clark, J. H.; H. R. Derring and M. R. Bennink (1977). Milk production, nitrogen utilization and glucose synthesis in lactating cows infused postpartum with sodium caseinate and glucose. J. Nutrition, 107: 631.
- Dann, H. M., J. K. Drackley, G. M. McCoy, M. F. Hutjens and J. E. Garrett. (2000). Effects of yeast culture (*Saccharomyces cerevisiae*) on prepartum and postpartum intake and milk production of Jersey cows. J. Dairy Sci. 83:123–127
- Doumas, B., W. Wabson and H. Biggs (1971). Albumin standards and measurement of serum with bromocresol green. Clin. Chem. Acta, 31: 87.
- Duncan, D.B. (1955). Multiple range and multiple F tests. Biometrics, 11: 1.
- El-Ashry M. A., A. M. Kholif, H. A. El-Alamy, H. M. El-Sayed and T. A. El-Hamamsy (2001). Effect of different yeast cultures supplementation to diet on the productive performance of lactating buffaloes. Egypt. J. Nutr. & Feeds, 4: 21-34.
- El-Batal, A. L. and M. Fadel (2002). Production of high selenium yeast in pilot scale batch fermentation. Egypt. J. Biomed. Vol. 9, April, :87-98.
- El-Waziry A. M.; H. E. M. Kamel and M. H. M. Yacout (2000). Effect of bakers yeast (*Saccharomyces cerevisiae*) supplementation to berseem (*Trifolium alexandrinum*) hay diet on protein digestion and rumen fermentation of sheep. Egypt. J. Nutrition and Feeds, 3 71-82.
- Erdmann R. A.; G. H. S. Proctor and J. H. Vandersall (1986). Rumen ammonia concentration on In situ rate and extent of digestion of feed stuffs. J. Dairy Sci., 69: 2312.
- Erdmann R. A. and B. K. Sharma. (1989). Effect of yeast culture and sodium bicarbonate on milk yield and composition in dairy cows. J. Dairy Sci. 72:1929–1932.
- Forbes R. M. and W. P. Garrigus (1948). Application of a lignin ratio technique to the determination of the nutrient intake of grazing animals. J. Anim. Sci., 7: 373-382.
- Foster, L. H. and S. Sumar (1997). Selenium in health and disease. A review. Crit. Rev. Food Sci. Nut. 37:211-228.
- Gallup, W. D.; C. S. Hobbs and H. M. Briggs (1945). The use of silica as a reference substance in digestion trials with ruminants. J. Anim. Sci., 4: 68-71.
- Heider, J. and A. Bock (1993). Selenium metabolism in microorganisms. Adv. Microb. Physiol. 25: 71-109.
- Ip, C. and D. J. Lisk (1994). Characterization of tissue selenium profiles and anticarcinogenic response in rat fed natural source of selenium rich product. Carcinogenesis. 15:573-576.
- Kholif, A.M. (2005). "Production, Development and Evaluation of Healthy food products": Evaluation of some feed additives for dairy animals. 1st Annual Report, for Project No. 704106 supported by

- National Research Center (2004-2007).
- Kholif A.M., M. A. El-Ashry; H. A. El-Alamy; H. M. El-Sayed; M. Fadel and S. M. Kholif (2005). Biological treatments of banana wastes for feeding lactating goats. *Egyptian J. Nutr. & Feeds*, 8 (2):149.
- Kung, Jr., L. E., M. Kreck and R. S. Tung. (1997). Effects of a live yeast culture and enzymes on in-vitro ruminal fermentation and milk production of dairy cows. *J. Dairy Sci.* 80:2045–2051.
- Lila Z. A., N. Mohammed, T. Yasui, Y. Kurokawa, S. Kanda and H. Itabashi (2004). Effects of a twin strain of *Saccharomyces cerevisiae* live cells on mixed Ruminal microorganism fermentation in vitro. *J. Anim. Sci.* 82:1847-1854.
- Ling E. R. (1963). "Text Book of Dairy Chemistry" Vol. II. Practical, 3rd ed Chapman and Hall, L.T.D., London, UK.
- Mehrez, A. Z.; E. R. Orskov and I. McDonald (1977). Rates of rumen fermentation in relation to ammonia concentration. *Br. J. Nutr.*, 38: 447.
- Mertens, D. R. (1978). Effect of buffers upon fiber digestion. Invited paper at Regulation of Acid-Base Balance Symposium, 8:9, PP. 65, Arizona Inn, Tucson, Arizona.
- Mutsvangwa, T., I. E. Edwards, J. H. Topps and G. F. M. Paterson. (1992). The effect of dietary inclusion of yeast culture (Yea-Sacc) on patterns of rumen fermentation, food intake and growth of intensively fed bulls. *Anim. Prod.* 55:35–41.
- Newbold, C. J., P. E. V. Williams, N. McKain, A. Walker and R. J. Wallace. (1990). The effects of yeast culture on yeast numbers and fermentation in the rumen of sheep. *Proc. Nutr. Soc.* 49:47A. (Abstr.) .
- Newbold, C. J., R. J. Wallace and F. M. McIntosh. (1995). Different strains of *Saccharomyces cerevisiae* differ in their effects on ruminal bacterial numbers in-vitro and in sheep. *J. Anim. Sci.* 73:1811–1818.
- Patton C.J. and S. R. Crouch (1977). Spectrophotometric and kinetics investigation of the berthelot reaction for the determination of ammonia. *Anal. Chem.*, 49: 469.
- Piva, G., S. Belladonna, G. Fusconi and F. Sicaldi (1993). Effects of yeast on dairy cow performance, ruminal fermentation, blood components and milk manufacturing properties. *J. Dairy Sci.* 76:2717–2722.
- Raltiff C. R. and F. Hall (1973). Laboratory manual of clinical biochemistry. Scott and Memorial Hospital Publication Office, Temple, TX.
- Reitman, S. and S. Frankel (1957). Colorimetric method for the determination of serum glutamic-oxaloacetic and glutamic – pyrovate transaminase. *An. J. clin. Path.*, 28: 56.
- Siest G., J. Henny and F. Schiele (1981). Interpretation des examens de laboratoire. Karger ed. 206.
- Snedecor, G. W. and W. G. Cochran (1982). Statistical Methods. 7th ed. Iowa State Unvi. Press, Ames, Iowa, USA.
- Soder, K. J. and L. A. Holden (1999). Dry matter intake and milk yield and composition of cows fed yeast prepartum and postpartum. *J. Dairy Sci.* 82:605–610.

Egyptian J. Nutrition and Feeds (2006)

- Wallace R. J. (1979). Effect of ammonia concentration on the composition, hydrolytic activity and nitrogen metabolism of the microflora of the rumen. *J. Appl. Bacteriol.*, 47: 443.
- Wallace R. J., and C. J. Newbold (1992). Probiotics for ruminants. Page 317 in *Probiotics: The Scientific Basis*. R. Fuller, ed. Chapman and Hall, London, U.K.
- Wang, Z., M. L. Eastridge and X. Qiu. (2001). Effects of forage neutral detergent fiber and yeast culture on performance of cows during early lactation. *J. Dairy Sci.* 84:204–212.
- Warner A. C. J. (1964). Production of volatile fatty acids in the rumen. *Methods of Measurements. Nutr. Abst. & Rev.*, 34:339.
- Weiss W. P. and J. S. Hogan (2005). Effect of selenium source on selenium status, neutrophil function, and response to intramammary endotoxin challenge of dairy cows. *J. Dairy Sci.* 88:4366-4374.
- Williams, P. E. V. and C. J. Newbold (1990). Rumen probiosis. The effects of novel microorganisms on rumen fermentation and ruminant productivity. In: W. Hresign and D. J. A. Cole (Ed.) *Recent Advances in Animal Nutrition* pp 211-227. Butterworths, London, UK.
- Williams, P. E. V., C. A. G. Tait, G. M. Innes and C. J. Newbold (1991). Effects of the inclusion of yeast culture (*Saccharomyces cerevisiae* plus growth medium) in the diet of dairy cows on milk yield and forage degradation and fermentation patterns in the rumen of steers. *J. Anim. Sci.* 69:3016–3026.
- Wohlt, J. E., A. D. Finkelstein and C. H. Chung (1991). Yeast culture to improve intake, nutrient digestibility and performance by dairy cattle during early lactation. *J. Dairy Sci.* 74:1395–1400.

تأثير إضافة الخميره أو الخميره المدعمة بالسيلينيوم على الأداء الإنتاجي للجاموس الحلاب

صبحى محمود محمد خليف^١ و محمود محمد خورشيد^٢

^١المركز القومى للبحوث ، قسم الألبان، شارع التحرير - الدقى - مصر.

^٢كلية الزراعة ، جامعة عين شمس، قسمالاتج الحيوانى، شبرا الخيمة - القاهرة - مصر.

أجريت تجربتين فى هذه الدراسة: فى التجربة الأولى تم استخدام ثلاثة ذكور ماعز بلدى مخصصة لدراسة تأثير إضافة الخميره أو خميره السيلينيوم على نشاط الكرش باستخدام نظام المربع اللاتينى 3x3 حيث تغذت الحيوانات على العلائق التالية:

١- المجموعة الأولى (المقارنة): علف مركز ٥٠% + برسيم أخضر ٥٠%.

٢- المجموعة الثانية: علفه المقارنة + ٥ جم خميره/ رأس/يوم.

٣- المجموعة الثالثة: علفه المقارنة + ٥ جم خميره مدعمة بالسيلينيوم/ رأس/يوم.

وكانت أهم النتائج المتحصل عليها ارتفاع تركيز كل من النيتروجين الكلى والبروتين الحقيقى معنويا والنيتروجين الغير بروتينى بإضافة الخمائر بينما لم يتأثر تركيز الأمونيا و الأحماض العضوية الطيارة فى الكرش بالمعاملات.

وفى التجربة الثانية تم استخدام ١٥ جاموسة حلابة لدراسة تأثير إضافة الخميره أو خميره السيلينيوم على الأداء الإنتاجي للجاموس الحلاب. حيث قسمت الحيوانات ألى ثلاثة مجموعات باستخدام نظام القطاعات العشوائية الكاملة حيث تغذت الحيوانات على العلائق التالية:

٤- المجموعة الأولى (المقارنة): علف مركز ٦٠% + برسيم أخضر ٢٠% + قش أرز ٢٠%.

٥- المجموعة الثانية: علفه المقارنة + ١٠ جم خميره/ رأس/يوم.

٦- المجموعة الثالثة: علفه المقارنة + ١٠ جم خميره مدعمة بالسيلينيوم/ رأس/يوم.

وكانت أهم النتائج المتحصل عليها كما يلى:

١- ارتفع معامل الهضم معنويا (٥%) لكل من المادة العضوية والبروتين الخام والألياف الخام والمستخلص الخالى من النيتروجين كما ارتفع كل من المادة الجافة الدهن الخام للمجاميع التى تغذت على كلا نوعى الخمائر مقارنة بالمجموعة المقارنة.

٢- ارتفع إنتاج اللبن معنويا (٥%) بإضافة نوعى الخمائر ونتيجة لذلك ارتفع محصول كل مكونات اللبن بإضافة نوعى الخمائر مقارنة بالمجموعة المقارنة. كما ارتفعت نسبة بروتين ولاكتوز اللبن معنويا (٥%) بإضافة خميره السيلينيوم تبعثها إضافة الخميره مقارنة بالمجموعة المقارنة. كما ارتفع دهن اللبن بإضافة الخمائر بينما ل(٥%) بإضافة مكونات اللبن معنويا بإضافة الخمائر. كما ارتفع معدل الاستفاداة من الغذاء معنويا (٥%) بإضافة نوعى الخمائر مقارنة بالمجموعة المقارنة.

٣- ارتفع جلوكوز الدم والبيومين الدم معنويا (٥%) بإضافة الخمائر لعلائق الجاموس الحلاب بينما لم يتأثر تركيز باقى مكونات اللبن معنويا بالمعاملات.

ومن هذه النتائج يتضح أن إضافة الخميره عامة قد حسن من إنتاج اللبن وتركيبه إلا أن إضافة السيلينيوم العضوى إلى علائق الجاموس الحلاب قد أثر ايجابيا على محتوى اللبن الناتج من السيلينيوم مما يودى إلى حماية المستهلك من بعض الأمراض السرطانية. ولذلك يوصى بإضافة خميره السيلينيوم لتدعيم نقص الاستفادة من السيلينيوم فى أغذية الحيوانات الحلابة دون التأثير على صحة الحيوان.