CHEMICAL, FUNCTIONAL AND BIOLOGICAL PROPERTIES OF MODIFIED CORN STARCH

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ABSTRACT

Chemical modified corn starch (CMS) such as; starch acetate (SA), starch succinate (SS) and starch phosphate (SP) were prepared, and their chemical, functional properties and biological effects were evaluated. Protein, fat and ash contents of corn starch were decreased by the modification process, except the ash content was increased in SP. Also, the amylose content was decreased in both SA and SP, while it was increased in SS. For functional properties, the obtained results showed that water absorption capacity (WAC) at 25°C and solubility at 90°C were increased in all CMS. Meanwhile, the viscosity was decreased in both SA and SP while, it was increased in SS. Swelling power at 90°C was decreased only in SP. Biological experiment was conducted on normal albino rats after feeding on diet containing 10% of modified starch, the result showed that feeding on both SA and SS decreased the final body weight, while feeding on diet containing SP caused slight increase in final body weight. The experiment on diabetic rats showed a significant decrease in body weight compared with normal rats. Also, feeding on SA significantly decreased serum glucose of healthy and diabetic rats . However , feeding on CMS had no significant differences on lipid fractions, kidney functions and liver functions on healthy rats compared with diabetic rats.

Keywords: starch acetate, starch succinate, starch phosphate, functional properties, biological properties, diabetic rats.

INTRODUCTION

Starch is the most important carbohydrate in all cereals and is a major source of carbohydrates in the human diet. The granules are used in foods because of their good thickening, gelling properties, and an excellent raw material for modifying food texture and consistency (Biliaderis,1991). However, use of native starches in industrial applications is limited because of several drawbacks , including low shear stress resistance, easy thermal decomposition , high retrogradation and syneresis .These shortcomings can be overcome by modifying the starch through chemical , physical or enzymatic methods (Agboola *et al.*, 1991) .

Chemically modified starches (CMS) are resistant to retrogradation, have higher viscosity and are more stable to acid and high temperatures than native starch. (Kishida *et al.*, 2000). Many studies approach the chemical modification of starch as an alternative way of improving its hygroscopic properties, therefore, they are increasingly used in food industry to improve the physical properties of various food items (Larotonda *et al.*,2005).

However, CMS may also have beneficial nutritional propperties. The puplished results of the joint FAO / WHO (1976) expert committee on food additives showed that modified starches were considered toxicologically safe

and may be used in foods without limitation or restrictions. Many investigators have observed a reduced susceptibility to enzymes in vitro and in vivo compared with the corresponding unmodified starch, therefore, CMS would be expected to exert physiological effects similar to those of dietary fiber. Some of the modified starches seem to be less digestible than the unmodified form (Ebihara et al., 1998). CMS as resistant starch may reduce the plasma glucose concentration and satiety after ingestion (Raben et al.,1994). The effect of modification and processing procedures of starch on human physiology (especially glycemic and insulinemic responses) have also been studied extensively (Bjorck,1996). So, many types of chemically modified starch, such as starches modified by acid hydrolysis, oxidation, etherification, esterification and cross-linking have attracted much attention (Adebowale and Lawal., 2003).

The aim of the present work is to modify corn starch chemically through acetylation, succinylation and phosphorylation processes then, study the effect of these process on the product properties and evaluate their biological effects on normal male albino rats compared with diabetic rats.

MATERIALS AND METHODS

Native starch: Food grade edible corn starch was obtained from the Egyption Starch and Glucose Co., Turra Factory, Cairo, A.R.E.

Commercial wheat flour: American wheat flour (72% extraction) was obtained from South Cairo Mills Company, Fysal, Giza, Egypt.

Animals: Male albino rats Winster strain weighing 138-143g were obtained from Holding Company of Biological Vaccines and Sera, Public Helwan farm, Cairo, Egypt.

Preparation of starch acetate (SA) and starch succinate (SS):

Starch acetate and succinate were prepared at laboratory scale as described by Phillips et al. (1998) following by the method of Walff et al. (1951) with minor modifications.

Prparation of starch phosphate (SP):

Starch was phosphorylated using the method of Inagaki and Seib (1992). Determination of degree of substitution (D.S.) of acetylated and succinylated starch:

The acetylation and succinylation level of the modified starch was determined using the titrimetric method of Wurzburg (1964).

Determination of phosphorus content in starch phosphate:

Phosphorus in both native corn starch and modified starches were determined by the phospho-molybdenum method which described by King (1932).

Chemical composition of native and modified corn starches:

Native and modified corn starches were determined for moisture, protein, fat, and ash contents according to A.O.A.C. (2000).

Determination of amylose and amylopectin content of native and modified corn starches:

The method described by McCready and Hassid (1943) was used for the estimation of linear fraction amylose in native and modified starches, and amylopectin was calculated by the difference as in the following equation %Amylopectin = 100- amylose (%).

Water absorption capacity (WAC):

WAC was measured for native and modified starches by the method described by Medcalf and Gilles (1965).

Solubility and swelling power:

Solubility and swelling power at 90°C were measured for native and modified starches using the methods of Leach et al. (1959).

Apparent viscosity:

Viscosity was determined for native and modified starches using the method of Wurzburg (1964).

Biological experiment:

Forty animals of adult male albino rats weighed 138 to 142g were used in present experiment. Animals were fed in the animal house, Crops Technology Department, Food Technology Research Institute (FTRI), Giza, Egypt, under normal conditions and fed on basal diet as shown in Table (1) for one week. After adaptation period, animals were weighed and divided into 8 groups each of 5 rats for one group as follow:

Group 1: Rats fed on basal diet.

Group 3, 4 and 5 : normal rats fed on basal diet containing 10 % of modified starch as shown in Table (1)

Groups 2,6,7 and 8 (diabetic group): Rats were injected by alloxan solution 150 mg/Kg body weight of recrystallized alloxan (Buko et al., 1996) to induce hyperglycemia, then the groups were fed on basal diet for 72 h. where hyperglycemia was developed. To ensure occurrence of diabetes in rats, blood sample was withdrawn after 72 h of alloxan injection and then fed on diets containing modified starch as shown in Table (1).

Weekly, rats were weighed and blood samples were collected by withdrawing from veva cava eye of both diabetic and normal rats and blood glucose level was determinated. After 5 weeks, animals were decapitated after fasting for 24h, and the blood was collected and centrifuged to obtain the serum which was kept at -18°C until analysis.

Determination of serum glucose:

Serum glucose was determined according to Trinder (1969).

Determination of serum lipid fractions:

Total lipids (T.L) were determined according to Knight *et al.* (1972), total cholesterol (T.C) was determined according to the method described by Allian *et al.* (1974), triglycerides (T.G) were determined according to the methods of Fossati and Prencipe (1982), HDL-cholesterol (HDL-C) was determined according to Lopez-virella *et al.* (1977) and LDL – cholesterol was calculated for serum samples using the formula of Friedewald *et al.* (1972) LDL-C= T.C-(T.G/5+HDL-C).

Determination of kidney function: Uric acid and Urea in serum were determined according to the methods of Kageyama(1971) and Tabacco *et al.* (1979) serum creatinine was determined according to the method described by Bartels and Bohmer (1971).

Determination of liver function: Serum transaminase enzymes serum aspartate transaminase (AST) and serum alanine transaminase (ALT) were

determined according to the method described by Retiman and Frankel (1957), serum total protein (TP) was determined according to the method described by Gornall et al. (1949)

Table (1) :Diet Composition (%)

Components	**G1	*G2	G3 SA	*G6 SA	G4 SS	*G7 SS	G5 SP	*G8 SP
Casein	15	15	15	15	15	15	15	15
Native starch	65	65	55	55	55	55	55	55
Starch acetate (SA)	-	Γ-	10	10	-	-	· -	-
Starch succinate (SS)	-	-	-	-	10	10	-	-
Starch phosphate (SP)	-	-	-	-	-	-	10	10
Corn oil	10	10	10	10	10	10	10	10
Cellulose	5	5	5	5	5	5	5	5
Salt mixture	4	4	4	4	4	4	4	4
Vitamin mixture	1	1	1	1	1	1	1	1

^{*} Diabetic rat groups

Statistical analysis:

Statistical analysis was carried out according to Fisher (1970). Least squares differences (LSD) at P < 0.05 test were used to compare the significant differences between means of treatment; where a,b,c values are means of five replicates \pm SE (Waller and Duncan, 1969).

RESULTS AND DISCUSSIONS

Chemical composition

Chemical composition of native, acetylated, succinylated and phosphorylated corn starch:

Proximate analysis was carried out in order to determinate the purity of the native starch as well as the effect of chemical modification on the composition of the starch. The results are presented on Table (2).

The results showed that the modified starches had the lowest content of fat and protein compared to native starch. The decrease in the contamination, is due to the washing of these component under the conditions used for the modification. Both acetylation and succinylation reduced the ash content due to the capacity of the chemical agent to solubilize minerals during the reaction. In this respect Betancur-Ancona et al. (2002) found a similar decrease in ash content when (Canavalia ensiformis) starch was modified using succinic anhydride. On the other hand, there was an increase in ash content of starch phosphate due to the formation of phosphate salts during the reaction. These results are agreed with those reported by Ola. S. Ibrahim. (1997).

At the same time amylose content in the acetylated and phosphorylated starches was decreased more than that found in the native

^{**}G1 fed on basal diet (normal control)

G3,G4and G5 healthy rats fed on modified starches

starch due to the process of substitution in these modified starch types that affect their degree of binding of iodine which gave lower value of the amylose content. Kasmsuwan and Jane (1994)reported that the cross-linked of maize starch or starch phosphate using phosphorous oxychloride (POCl₃) at 25°C at low level of cross-linking, the amylose molecules have been located in bundles between amylopectin clusters. It is randomly interspersed among amylopectin clusters in both amorphous and crystalline regions, so that the iodine blue colour was decreased. However, the presence of some succinyle groups interferes with the reassociation of amylose and amylopectin during the storage of starch molecules subjected to the gelatinization process. This cause the creation of more linear segments, which eases the absorption of higher amounts of iodine, so the value of amylose content in the succinylated starch was increased which is in consistence with our finding.

Table (2): Chemical composition of native corn starch and chemically modified starches (on dry weight basis).

Samples	Protein %	Ash %	Fat %	Amylose %	Amylopectin %
Native starch	0.54	0.126	0.060	26.00	74.00
Starch acetate (D.S.= 0.0910)	0.42	0.116	0.030	25.80	74.20
Starch Succinate (D.S.= 0.0263)	0.41	0.096	0.033	27.90	72.10
Starch phosphate (D.S= 0.0028)	0.36	0.165	0.031	24.60	75.40

Functional properties:

Results in Table (3) represent the functional properties of native and chemical modified starches. It could be observed that, the chemical modification increased the water absorption capacity (WAC) of the native starch. The higher increase in WAC was observed in succinylated starch (95%) followed by acetylated starch (74%) and phosphorylated starch (71%). This results are in good agreement with that obtained by Betancur-Ancona et al. (2002) and Nabeshima and Grossmann (2001). Waliszewski et al. (2003) showed that chemical modification improved starch water binding capacity because the hydrophilic groups were incorporated.

The chemically modified starches were more soluble than that of native starch. The low solubility value of native starch could be attributed to the increase of binding forces within starch molecules and resistance towards solubility. Under the same condition, there was an increase in solubility of modified starches, this increase may be due to hydrophilic groups or residual activity in starch (Aiyleye *et al.*,1993).

The swelling power was decreased at 90°C of starch phosphate compared to native starch which may due to two different possible interpretations, losing granular integrity, resulting in mixture of soluble starch and granules fragments, or larger granules could be preferentially broken down, leaving apopulation of smaller intact granules. These results agreed with Ola. S. Ibrahim. (1997).

However, succinylation and acetylation considerally increased granule swelling power. This observation is in agreement with the report of Betancur-Ancona et al. (2002) and Adebowale and Lawal (2003).

Also, results in Table (3) indicated that the value of apparent viscosity was decreased by acetylation or phosphorylation treatment. However succinlated starch had higer apparent viscosity. This results differs than that of Ola. S. Ibrahim. (1997) who reported that phosphorylation caused an increase in viscosity, but agreed with Betancur-Ancona *et al.* (2002) who reported that succinylation increased apparent viscosity. Derivatisation of starch with an ionic substituent group such as succinate, at low degree of succession (DS) converted it into a polyelectrolyte. Starch acquires typical rules of a polyelectrolyte such as increased water solubility and as a solution viscosity (Bhandari and Singhal, 2002).

(3): Functional properties of native and chemically modified starches.

Samples	Water absorption capacity (g)A	Apparent viscosity (CP)B	Solubility (%)C	Swelling power (g water/g starch)D
Native starch	1.20	13	6.20	9.80
Starch acetate	1.78	10	8.40	9.90
Starch succinate	2.03	20	8.6	10.10
Starch phosphate	1.73	10	7.09	6.04

A. Measured at 25°C

Biological effects of chemically modified starches (CMS): Effect on rat body weight :

Data in Table (4) represent the gain in body weight (g) in normal groups. It could be stated that, succinylated and acetylated starches significantly reduced the gain in body weight, while phosphorylated starch slightly decreased gain in body weight compared with normal control. The decrement of body weight gain may be due to the satiety ratings which are higher after meals with CMS than native starch (G1). These ratings are higher in WAC and viscosity and thus influencing the emptying and digestion of other compounds in the gut digestal (Raben et al.,1997 and Ebihara et al.,1998). These results are in agreement with Til et al. (1986) who reported that feeding on hydroxypropyl distarch phosphate and starch acetate caused slightly lower body weights. Also, feeding on five chemically modified starches (acetylated distarch phosphate, acetylated diamylopectin phosphate, starch acetate, hydroxypropyl distarch glycerol and phosphated distarch phosphate reduced body weights except that of the phosphated distarch phosphate (deGroot et al.,1974).

Table (5) represent gain in body weight in diabetic rats. It could be stated that, there was a significant decrease in body weight gain of diabetic rats compared to normal control and this due to break down of protein to obtain energy (Meyer et al., 1998). These results are in agreement with

B. Measured of 0.2 % starch suspension in Brookfield viscometer, spindle no.2, 25°C and 100 rpm.

C. and D. Measured at 90°C.

Yadav et al. (2004) who found that the body weight of alloxan-treated rats (diabetic rats) was significantly decreased after 21 days of diabetes compared to normal control.

Table (4): Initial body weight, final body weight and gain in body weight (g) of normal control and rats groups fed on 10% chemical modified starches.

Groups	Initial body weight (g)	Final body weight (g)	Gain in body weight (g)
G1	138.2 ± 7.06	185.4± 8.02	+47.00 ^a
G3	140.6 ± 4.83	136.4± 4.76	- 4.20°
G4	142.2 ± 5.88	134.2± 4.28	- 8.00°
G5	139.2 ± 7.20	145.4± 7.01	+ 6.20 ^b
L.S.D. at 5%			8.756

The column values followed by the same letters are not significant different.

The column values followed by different letters are significant different.

Table (5): Initial body weight, final body weight and gain in body weight (g) of normal control and diabetic rats groups fed on 10% chemical modified starches.

Groups	Initial body weight (g)	Final body weight (g)	gain in Body weight (g)
G1	138.2 ± 7.06	185.4 ± 8.02	+47.2 ^a
G2	143.0 ± 5.47	107.6 ± 4.34	-35.4 ⁵
G6	140.4 ± 6.93	116.2 ± 6.11	-24.2 ^b
G7	142.4 ± 6.46	108.4 ± 5.48	-34.0 ^b
G8	139.0 ± 7.77	111.0 ± 6.42	-28.0°
L.S.D. at 5%			12.251

The column values followed by the same letters are not significant different. The column values followed by different letters are significant different.

Effect on serum blood glucose:

Tables (6 and 7) represents the effect of chemically modified starches on blood glucose in normal and diabetic rats. The data revealed that starch acetate significantly decreased blood glucose level of group 3 and group 7. These results agree with Raben et al. (1997) who found that chemical modification of potato starch (acetylation or β – cyclodextrin) improved the stateing, glycemic index and insulinemic properties of the meal. The glycemic index and insulinemic index of foods have been found to correlate with hunger and satiety after a meal. Thus foods with low indexes were more satiating than foods with high indexes (Holt et al., 1992). Also , CMS are resistant to small intestinal amylosis, thus there capacity to lower the glycemic response may be more predictable than the other types of resistant starches (Brid and Topping., 2002). Acetylated , propionylated or butyrylated starches raise large bowel of short chain fatty acids (SCFAs) when fed to rats and these reduced plasma glucose levels. (Ferguson and Jones .,2000 and Annison et al., 2003).

Table (6): Blood glucose level (mg/dl) in normal rats fed on diets containing 10% chemical modified starches

Groups	Zero time	1week	2weeks	3weeks	4weeks	5weeks
G1	94.42 ^a	96.14ª	97.17 ^a	95.64 ^a	94.8°	95.92ª
G	±7.06	±6.67	±5.81	±4.75	±5.61	±3.81
62	97.21ª	96.71ª	94.41 ^a	90.4ª	84.99ª	79.26 ^b
G3	±6.82	±5.87	±5.47	±4.98	±3.76	±3.49
C4	91.03ª	90.43 ^a	89.72 ^a	87.8ª	88.6ª	90.17 ^{ab}
G4	±5.61	±4.87	±5.44	±4.33	±4.8	±4.14
CE	98.01 ^a	96.79ª	93.86 ^a	95.2°	97.4ª	96.73 ^a
G5	±7.37	± 5.49	±6.19	±6.53	±5.73	±5.34
L.S.D. at 5%	20.243	16.775	17.210	15.645	15.110	12.7538

The column values followed by the same letters are not significant different. The column values followed by different letters are significant different.

Table (7): Blood glucose level (mg/dl) in diabetic rats fed on diets containing 10% chemical modified starches

Groups	Zero time	1week	2weeks	3weeks	4weeks	5weeks
G1	94.42 ^b	96.14 ^b	97.17 ⁵	95.64 ^b	94.8 ^c	95.92°
	±7.06	±6.67	±5.81	±4.75	±5.61	±3.81
	273.4ª	271.2ª	267.7ª	265.9ª	264.8 ^a	262.46ª
G2	±12.08	±10.88	±9.82	±8.54	±8.21	±7.31
	265.6ª	262.8ª	256.4ª	245.5ª	228.6 ^b	212.6 ^b
G6	±9.56	±8.56	±8.37	±7.75	±7.47	±6.58
	270.4 ^a	269.8ª	265.6ª	262.5ª	259.2ª	257.2 ^a
G7	±11.108	±9.77	±9.29	±10.07	±10.10	±9.47
	259.9 ^a	257.6ª	254.62 ^a	250.8ª	248.4 ^{ab}	245.8 ^a
G8	±9.016	±9.24	±8.69	±9.14	±7.10	±7.47
L.S.D. at 5%	29.266	26.947	25.132	24.266	23.125	21.149

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Effect on serum lipid fractions:

Tables (8 and 9) represents the level of total lipids (T.L), total cholesterol (T.C), triglycerides (T.G), high density lipoprotein - cholesterol (HDL-C) and low density lipoprotein - cholesterol (LDL-C) in normal and diabetic rats. It could be observed that in normal group (G3,G4, G5) their were no significant differences in lipid fractions compared to normal control (G 1). In contrast, the levels of T.C., T.G., T.L. and LDL-C were higher in diabetic groups. These results are in the line with those of Anderson *et al.* (1973) who found that Pitman-Moore miniature pigs fed on formulated diets containing 5.4% unmodified starch and 5.6% phosphated distarch phosphate had non significant difference in cholesterol and triglycerides levels. Also, Raben *et al.* (1997) reported that, serum triacylglycerol concentrations were not significantly different in the fasted state and after meal ingestion of rats fed on chemical modified potato starch (acetylation β – cyclodextrin). Our results are disagreed with those found by Ebihara *et al.* (1998) who showed

that two different types of CMS (hydroxypropyl starch and hydroxypropyl distarch phosphate) decreased the plasma cholesterol level of rats but their was no effect on T.G level.

Table (8): Triglycerides, total cholesterol, HDL- cholesterol, LDL-cholesterol and total lipids, levels (mg/dl) in normal rats fed on diets containing 10% chemical modified starches.

Groups	Triglycerides	Total cholesterol	HDL- cholesterol	LDL- cholesterol	Total lipids
G1	112.6°±	95.2°±	49.02 ^a ±	23.660 ^a ±	326.0 ^a ±
	3.97	7.74	3.29	10.41	13.89
G3	109.6°±	88.0° ±	47.87°±	18.212 ^a ±	321.2 °±
	5.76	8.07	2.80	6.74	13.01
G4	105.8°±	92.0°±	50.02°±	20.816°±	330.6 ^a ±
	4.30	6.77	2.95	4.78	10.99
G5	116.2°± 4.61	87.4 ^a ± 7.44	45.22°± 1.94	18.940 ^a ± 6.57	318.6 ^a ± 9.87
L.S.D. at 5%	14.12	22.5647	8.336	22.234	36.1247

The column values followed by the same letters are not significant different.

The column values followed by different letters are significant different.

Table (9): Triglycerides, total cholesterol, HDL- cholesterol, LDL-cholesterol and total lipids, levels (mg/dl) in diabetic rats fed on diets containing 10% chemical modified starches.

Groups	Triglycerides	Total cholesterol	HDL- cholesterol	LDL- cholesterol	Total lipids
G1	112.6 ^c ± 3.97	95.2°± 7.74	49.020 ^a ± 3.29	23.66°± 10.41	326.0 b ±13.89
G2	217.2°± 10.93	178.4° ± 4.22	33.082 b± 1.41	101.87°± 4.03	516.0 a ±17.05
G6	185.6 b ± 8.87	149.0 ^b ± 9.93	40.092 ^b ± 1.64	71.79 ^b ± 8.66	412.2° ±12.55
G 7	210.0 ^{ab} ± 9.86	172.0° ± 6.01	36.574 ^b ± 2.39	93.42 ^{ab} ± 8.50	507.8° ±14.91
G8	202.4 ^{ab} ± 8.72	169 ^{ab} ± 6.80	35.796 ⁵ ± 1.78	92.72 ^{ab} ± 6.97	499.6° ±17.56
L.S.D. at 5%	25.9707	21.224	6.5292	23.622	47.00315

The column values followed by the same letters are not significant different.

The column values followed by different letters are significant different.

Effect on kidney function:

The results of kidney function, i.e., urea, uric acid and creatinine were reported in Tables (10 and 11). The data showed that there were no significant differences on serum blood urea, uric acid and creatinine between normal group(G3, G4 and G5) and normal control (G1). However, data in Table (11) showed that there are significant increments in serum urea ,uric acid and creatinine in case of diabetic groups compared with normal control.

These results are in agreement with Anderson et al. (1973) who found no significant differences on biochemical analysis of blood (urea nitrogen, alkaline phosphatase and total protein) of pig fed on diets containing 5.6% phosphated distarch phosphate. In this respect, Cervenka and Kay (1963) reported that no adverse effects were observed on blood chemistry such as, urine parameters and liver functions in rats groups after given gelatine capsules containing 50, 250 and 1250 mg modified starch/kg b.wt. This results differs from the results of Til et al. (1986) who found slight urinary changes in rats fed on hydroxypropyl distarch phosphate and starch acetate.

Table (10): Urea, uric acid, and creatinine levels (mg/dl) in normal rats fed on diet containing 10% chemical modified starches.

Groups	Urea	Uric acid	Creatinine
G1	25.00° ± 2.91	3.76°±.341	0.802°±.081
G3	22.40° ± 2.60	3.36° ±.266	0.768 ^a ±.09
G4	20.56 ^a ± 2.07	3.59 ^a ±.281	0.726 ^a ±.074
G5	27.00°± 3.20	3.28 ^a ±.402	0.702 ^a ±078
L.S.D. at 5%	8.1930	0.2421	0.9825

The column values followed by the same letters are not significant different. The column values followed by different letters are significant different.

Table (11): Urea, uric acid, and creatinine levels (mg/dl) in diabetic rats fed on diet containing 10% chemical modified starches

Groups	Urea	Uric acid	Creatinine
G1	25.0 b ± 2.91	3.76 ^c ±.341	0.802 b ±.081
G2	54.0 a ±4.89	8.56 a ±.423	1.76 a ±.121
G6	42.4° ±3.77	6.86 b ±.577	1.42 a ±.185
G7	50.4 a ±3.97	7.64 ^{ab} ±.465	1.56 a ±.131
G8	47.8 a ±3.70	8.03 ab ±.528	1.62 a ±.156
L.S.D. at 5%	11.5206	1.3983	0.4101

The column values followed by the same letters are not significant different. The column values followed by different letters are significant different.

Effect on liver functions:

Tables (12 and 13) represents the amount of total protein (TP), the activities of serum aspartate amino transaminase (AST)and serum alanine amino transaminase (ALT). The data showed that there was no significant change in normal groups compard to normal control in serum TP. However, a significant decrease of serum total protein in rats from 7.12 g/dl in normal control to 5.34 g/dl in diabetic control. In this study, the decrease in serum protein may be due to the effect of alloxan on RNA and protein synthesis. Also, the data show that AST and ALT activities were significantly increased in serum after injecting with alloxan (150mg/Kg body weight), while there was no significant different in AST and ALT values between rats fed chemical modified starches and normal control. These results agreed with Anderson et al. (1973) and Cervenka and Kay (1963).

Table (12): Total protein (g/dl), activities of ALT, AST (IU/I) in normal rats fed on diets containing 10% chemical modified starches.

Groups	TP	ALT	AST
G1	7.12 ^a ±0.227	23.20°±1.854	33.8°±2.417
G4	7.08 ^a ±0.0149	22.10 ^a ±2.216	36.2°±2.922
G5	6.80°±0.330	19.76 ^a ±1.636	29.8°±4.271
G6	7.08°±0.159	25.16 ^a ±1.745	32.6°±4.045
L.S.D. at 5%	0.68349	5.06237	10.4908

The column values followed by the same letters are not significant different. The column values followed by different letters are significant different.

Table (13): Total protein (g/dl), activities of ALT, AST (IU/I) in diabetic rats fed on diets containing 10% chemical modified starches.

Groups	TP	ALT	AST
G1	7.12 ^a ± 0.227	23.2°±.1.854	33.8°±2.417
G2	5.34 ^{bc} ±0.150	50°±2.829	59 ^a ±.4.159
G6	6.014 ^b ±0.185	38.6°±.3.572	46.4 ^{ab} ±.5.420
G7	5.46 ^{bc} ±0.276	46.8 ^{ab} ±.3.184	57.2°±.4.397
G8	5.26°±0.242	48.2 ^{ab} ±.3.929	54.2°±.4.694
L.S.D. at 5%	0.6502	9.3081	12.6527

The column values followed by the same letters are not significant different. The column values followed by different letters are significant different.

Conclusion

Chemical and functional properties of native and chemical modified starches (CMS) were investigated. All chemical modification process reduced fat, protein and ash content except phosphorylation process increased ash content. Water absorption capacity and solubility were increased with chemical modification process, the succinylated starch gave a higher value. Acetylation and phosphorylation reduced apparent viscosity, while succinylation process gave a higher value. These properties give pastes and gels a better thickening capacity, clarity and stability at low temperatures.

The biological effects demonstrated that the succinylated and acetylated starches reduced the final body weight, while phosphorylated starches caused slight increase in final body weight. Also, acetylated starches reduced serum glucose in healthy and diabetic rats. In general the CMS had no affect on serum lipid fractions ,kidney and liver functions in healthy rats but improved serum lipid fractions, kidney and liver functions in diabetic rats in case of using starch acetate.

REFERENCES

- A.O.A.C. (2000). Association of Official Analytical Chemists. Official Methods of Analysis. 17th ed., Washington .DC, USA.
- Adebowale, K. O. and Lawal, O. S. (2003). Functional properties and retrogradation behaviour of native and chemically modified starch of mucuna bean (*Mucuna pruriens*). J. Sci. Food Agric., 83:1541.
- Agboola, S. O.; Akingbala, J. O. And Oguntimein, G. B. (1991). Physicochemical and unctional properties of low DS cassava starch acetates and citrates. Starch, 43:62.
- Aiyleye, F. B.; Akingbala, J. O. and Oguntimein, G. B. (1993). Chemical factors affecting actylation of cassava starch. Starch, 45: 443.
- Allian, C. C.; Poon, L. S.; Chan, C. S.; Richamand, W. and Fu, P. C. (1974). Enzymatic determination of total serum cholesterol. Clin. Chem., 20:470.
- Anderson, T. A.; Filer, L. J.; Fomon, S. J.; Andersen, D. W.; Jensen, R. L. and Rogers, R. R. (1973). Effects of waxy corn starch modification on growth, serum biochemical values and body composition of Pitman-Moore miniature pigs. Food Cosmet. Toxicol., 11:474.
- Annison, G.; Illman, R. J. and Topping, D. L. (2003). Acetylated,propionylated or butyrylated starches raise large bowel short chain fatty acids.preferentially when fed to rats. J. Nutr., 133:3523.
- Bartels, H. and Bohmer, M. (1971). Creatinine standard and measurement of serum creatinine with picric acid. Clin. Chem., 17:81.
- Betancur-Ancona, A. D.; Garcia, C. E.; Cañizares, H. E. and Chel, G. L. (2002). Chemical modification of jack bean (*canavalia ensiformis*) starch by sucinylation. Starch, 54:540.
- Bhandari, P. N. and Singhal, R. S. (2002). Studies on the optimisation off preparation of succinate derivatives from corn and amaranth starches. Carbohydrate Polymers, 47: 277.
- Biliaderis, C. G. (1991). The structure interactions of starch with food constituents. Can. J. Phys. Pharm., 69:877.
- Bjorck, I. (1996). Starch nutritional aspects. In: Carbohydrates in Food. Ed., Eliasson, A.pp. 505-553. Academic Press: New York.
- Brid, A. R. and Topping, D. L.. (2002) Short-chain fatty acids and human colonic function: roles of resistant starch and nonstarch polysaccharides. Physiol. Rev., 82:1031.
- Buko, V.; Lukivskaya, O.; Niktin, V.; Tarasov, Y.; Zavodnik, L.; Borodassky, A.; Goren, S. B.; Janz, B. and Gundermann, K. J. (1996): Hepatic and pancreatic effects of polyenoylphatidyl choline in rats with alloxan-induced diabetes. Cell Biochem. Funct., 14(2): 131.
- Cervenka, H. and Kay, J. H. (1963). Subacute oral toxicity of phosphate starch. Clin. Chem., 8:682.
- de Groot, A. P.; Til, H. P.; Feron, V. J.; Van der Meullen, H. C. and Willems, M. I. (1974). Two-year feeding and multigeneration studies in rats on five chemically modified starches. Food Cosmet. Toxicol., 12:651.

- Ebihara, K.; Shiraishi, R. and Okuma, K. (1998). Hydroxypropyl-modified potato starch increases fecal bile acid excretion in rats. J. Nutr., 128:848.
- FAO/WHO.(1976). On wholesomeness of irradiated foods. Joint Expert Committee Meeting Report.Food and Agriculture Organization of the United Nations: Geneva.
- Ferguson, M. J. and Jones, G. P. (2000). Production of short-chain fatty acids following *in vitro* fermentation of saccharides, saccharide esters, fructo-oligosaccharides, starches, modified starches and non-starch polysacharides. J. Sci. Food Agric., 80:166.
- Fisher, R. A. (1970). Statistical Method for Research Workers, pp. 140. Edinburg - Oliver &Boyd-Germany.
- Fossati, P. and Prencipe, L. (1982). The determination of triglyceride using enzymatic methods. Clin. Chem., 28:2077.
- Friedewald, W.I. stewart, S.W. and Arnold, T.F.(1972): Estimated calculation of low density lipoprotein. Clin. Chem., 18: 499.
- Gornall, A. C.; Bardawill, C. J. and David, M. M. (1949). Deterination of serum proteins by means of the biuret reaction. J. Bio. Chem., 177: 751.
- Holt, S.; Brand, J.; Soveny, C. and Hansky, J. (1992). Relationship of satiety to post prandial glycemic, insulin and cholecystokinin responses. Appetite., 18:129.
- Inagaki, T. and Seib, P. A. (1992). Firming of bread crumb with cross-linked waxy barley starch substituted for wheat starch. Cereal Chem., 69:321.
- Kageyama, N. (1971). A direct colorimetric determination of uric acid in serum and urine with uricase-catalase system. Clin. Chim.Acta, 31:421.
- Kasmsuwan, T. and Jane, J. (1994). Location of amylose in normal starch granules. I. Locations of phosphodiester cross-linking revealed by phosphorus 31nuclear magnetic resonance. Cereal Chem., 71:282.
- King, E. J. (1932). The colorimetric determination of phosphorus. Biochem. J., 26:292.
- Kinght, J. A.; Anderson, S. and Rowle, J. M. (1972). Chemical bases of the sulfo-phospho vanillin reaction for estimating total serum lipids. Clin. Chem., 18:199.
- Kishida, T.; Nakai, Y. and Ebihara, K. (2000). Hydroxypropyl-distarch phosphate from tapioca starch reduces zinc and iron absorption, but not calcium and magnesium absorption, in rats. J. Nutr., 131:294.
- Larotonda, F. D.; Matsui, K. N.; Sobral, P. J. and Laurindo, J. B. (2005). Hygroscopicity and water vapor permeability of Kraft paper impregenated with starch acetate. J. Food Engineering., 71:394.
- Leach, H. W.; McCowen, L. D. and Schoch, T. J. (1959). Structure of the starch granule. 1. Swelling and solubility patterns of various starches. Cereal Chem., 36:534.
- Lopez-virella, M. F.; Stone, S.; Ellis, S. and Collwel, J. A. (1977). Cholesterol determination in high density lipoproteins separated by three different methods. Clin. Chem., 23: 882.
- McCready, R. M. and Hassid, W. Z. (1943). The separation and quantitative estimation of amylose and amylopectin in potato starch. J. Am. Chem. Oil Soc., 65:1154.

- Medcalf, D. G. and Gilles, K. A. (1965). Wheat starches.1.comparison of physicochemical properties. Cereal chem, 42:558.
- Meyer, C.; Stumvoll, M.; Nadkarni, V.; Dostou, J.; Mitrakou, A. and Gerich, J. (1998). Abnormal renal and hepatic glucose metabolism in type 2 diabetes mellitus. J. Clin. Invest., 102: 619.
- Nabeshima, E. H. and Grossmann, M. V.(2001). Functional properties of pregelatinized and cross-linked cassava starch obtained by extrusion with sodium trimetaphosphate. Carbohydrate Polymers.,47: 365.
- Ola. S. Ibrahim. (1997). Evaluation of modified starch properties in respect to their application in food processing. Ph.D. Thesis Fac. Agric., Cairo Univ.
- Phillips , D. L.; Pan, D. H.; Liu, H. J. and Croke, H. (1998). Raman spectrscopic determination of the level of acetylation in modified wheat starch. Anal. Lett., 31:2105.
- Raben, A.; Tagliabue, A.; Christensen, N. J.; Madsen, J.; Holst, J. J. and Astrup, A. (1994). Resistant starch: The effect on postprandial glycemic, hormonal response, and satiety. Am. J. Clin. Nutr., 60:544.
- Raben, A.; Anderson, K.; Karberg, M. A.; Holst, J. J. and Astrup, A. (1997). Acetylation of or β cyclodextrin addition to potato starch: benefical effect on glucose metabolism and appetite sensations. Am. J. Clin. Nutr., 66:304.
- Retiman, A. and Frankel, S. (1957). A colorimetric method for the determination of serum glutamic oxaloacetic and glutamic pyruvic transaminases. Am. J. Clin. Path., 28:56
- Tabacco, A.; Searcy, R. L. and Reardon, J. E. (1979). Standard Methods of Clinical Chemistry, Edited Seligon, D. Clin. Chem., 25:336.
- Til, H. P.; Feron, V. J. and Immel, H. R. (1986). Chronic (89-week) Feeding study with hydroxypropyl distarch phosphate, starch acetate, lactose and sodium alginate in mice. Food.Chem.Toxicol.,24:25.
- Trinder, P. (1969). Determination of blood glucose using an oxidation peroxidase system with anon carcinogenic chromogene. Ann. Clin. Biochem., 6:24.
- Walff, I. A.; Olds, D. W. and Hilbert, G. E. (1951). Acetylation of starch, amylose and amylopectin. J. Am. Chem. Soc., 73:346.
- Waliszewski, K.N.; Aparicio, M.A.;Bello, L.A.and Monroy, J.A.(2003). Changes of banana starch by chemical and physical modification. Carbohydrate Polymers., 47:85.
- Waller, W. M. and Duncan, D. B. (1969). A boys role for symmetric multiple problem. Am. State Assoc. J., 65:1485.
- Wurzburg, O. B. (1964). Starch derivatives and modification. In: Methods in Carbohydrate Chemistry. Eds., Whistler, R. L.; Smith, J. R. and Wolfrom, M. L.pp.240-241. Academic Press: New York.
- Yadav, U. C.; Moorthy, K. and Baquer, N. Z. (2004). Effects of sodium-orthovanadate and *Trigonella foenum-graecum* seeds on hepatic and renal lipogenic enzymes and lipid profile during alloxan diabetes. J. Biosci., 29:81.

الخواص الكيماوية والوظيفية والبيولوجية لنشا الذرة المحور محمد ابراهيم قبيصى*، سميحة محمد عبد السلام**، ابتسام عبد المسنعم محمسود* و زهرة العلا محمود محمد**

- قسم الكيمياء الحيوية كلية الزراعة جامعة القاهرة الجيزة
- ** قسم بحوث تكنولوجيا المحاصيل معهد بحوث تكنولوجيا الأغذية مركز البحوث الزراعية -الجيزة

تم اجراء عمليات تحوير كيميائية لنشا الذرة و ذلك بتحضير ثلاث مــ شنقات منــه و هــم خلات النشا و فرسفات النشا و سكسينات النشا و تم تقيــيم الخــواص الكيماويــة و الوظيفيــة و التأثيرات البيولوجية لهم وقد وجد ان محتواهم من البروتين و الدهون والرماد يقل بعمليات التحوير فيما عدا محتوي الرماد يزداد في حالة فوسفات النشا.ايضا حدث انخفاض في محتوي الاميلوز في كلا من خلات و فوسفات النشا بينما زاد محتواه في سكسينات النشا.

اظهرت نتائج الخواص الوظيفية حدوث زيادة في القدرة على امتصاص الماء عند ٢٠°م و خاصية الذوبان في الماء عند ٩٠°م في جميع انواع النشا المحور و ايضا حدث انخفاض للزوجة في كلا من خلات و فوسفات النشا بينما زادت في سكسينات النشا كما حدث انخفاض في قوة الانتفاخ عند ٩٠°م في حالة فوسفات النشا فقط.

ولتقييم المشتقات الناتجة تم اجراء تجربة بيولوجية على ذكور الفئران البيضاء بعد تغنيتها على وجية محتوية على ١٠% من النشا المحور و اظهرت النتائج ان التغذية على كلا مسن سكسينات و خلات النشا احدثت انخفاض في وزن الجسم بينما التغذية على وجبة محتوية على فوسفات النشا اظهرت زيادة طفيفة في وزن الجسم بينما حدث انخفاض ملحوظ في وزن الجسم للفئران المريضة بالسكر مقارنة بالفئران الطبيعية و ايضا وجد ان التغذية على خلات النسشا ادت لحدوث انخفاض معنوي لمستوي جلكوز الدم الفئران الطبيعية والمريضة بالسكر بينما التغذية على مشتقات النشا لم يحدث اختلافات معنوية على مكونات دهون الدم ، وظائف الكلسي والكبد في الفئران الطبيعية مقارنة بالفئران المريضة بالسكر.

ولذلك توصى بادخال هذه المشتقات في منتجات المخابز لما لها من فاندة تصنيعية و علاجية هامة.