EFFECT OF SEWAGE SLUDGE AS AMENDMENT ON SOIL MICROBIAL POPULATIONS AND THEIR ACTIVITIES IN TWO DIFFERENT SOILS

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ABSTRACT: Sewage sludge may improve soil fertility but there is concern about effects of sludge metals on soil microorganisms and their activities. Laboratory studies were conducted to determine the effect of two levels of sewage sludge (2% and 4% w/w) on the microbial populations and their activities in two different soils (sandy clay loam and sandy soil) during 42 days periods. The application of sewage sludge with the rate of 2% increased the total counts of bacteria, fungi, actinomycetes, nitrifiers, aerobic and anaerobic nitrogen fixers significantly in both kinds of soil than the unamended and the amended soil with the rate of 4% sewage sludge. The treatment with the high rate of sewage sludge (4%) caused a reduction in the total counts of the tested microorganisms and the activities of some soil enzymes (dehydrogenase, phosphatase, and urease) than the treatment with 2% sewage sludge. However the values obtained with 4% sewage sludge was still higher than those found in the unamended soil (control). Sewage sludge with the rate of 4% caused an increase in total nitrogen reaching 634 and 378 mg kg dry soil after 7 days of incubation in sandy clay loam and sandy soils, respectively. Generally, the addition of sewage sludge to both kinds of soils with the rate of 2% increased the values of ammonia and nitrate nitrogen than the unamended and amended soil with the rate of 4% sewage sludge. No harmful effect to microbial population and their activities was observed when sandy clay loam and sandy soils were amended with sewage sludge with the rate of 2% consequently this rate can be used safely in agricultural soils.

Key words: Sewage sludge, microbial population, dehydrogenase, phosphatase, urease, ammonia, nitrate, total nitrogen

INTRODUCTION

Many problems are associated with environmental pollution

especially those concerning with heavy metals. Although metals such as Cu, Ni, and Zn are essential nutrients for living organisms (Alloway, 1995) they may be toxic at high concentration. Heavy metals are being added to the environment from a variety of sources including municipal, industrial, and agricultural wastes as well as dredge spoils (Huisingh, 1974, Austin, et al., 1977 and Timoney et al., 1978)

The addition of sewage sludge agricultural soil improves physical characteristics and acts as a valuable source of nutrients for growing crops (Smith, 1996). Also the addition of sewage sludge to soil, especially soils of low organic matter, significantly increased the population and the activity of soil microorganisms (Pichtel and Hayes, 1990, Barakah et al., 1995 and Banerjee et al., 1997) and also increased soil enzymes-activities especially dehydrogenase activity (Crecchio et al., 2004) and higher values promoted of phosphatase and urease activity (Qiling et al., 2002, Awad and Fawzy, 2004 and Crecchio et al., 2004).

However, there is concern over the use of sludge that has been contaminated with potentially toxic elements (PTEs) from both industrial and domestic sources. The concentration of PTEs in sludge is usually considered the main determinant in restricting its application to agricultural soils. Application of sewage sludge to agricultural land can lead to an accumulation of metals in the surface layers with estimated residence times of the metal contaminants in the order of thousands of years (Mc Grath, 1987). Moreover contamination with heavy metals arises also where sewage sludge containing pesticides or fertilizers are used extensively as amendments since it can affect the activities of the beneficial soil microorganisms, and consequently affect the cycling of plant nutrients in the soil (Giller et al., 1998). Heavy metal contamination can also inhibit soil functions, but it is often difficult to determine the degree of pollution. Enzyme assays offer potential indicators of biological functioning of soil (Belen, et al., 2004).

Several studies have shown metals influence that adversely microorganisms by affecting growth. their morphology, biochemical and activity, resulting in decreased dehydrogenase biomass. and urease activities by increasing rates of sewage sludge addition (Reddy et al., 1987, Fliessbach and Reber, 1990, Chander and Brookes, 1991, Fliessbach et al., 1994, Kandeler et al., 1996, Roane and Kellogg, 1996, Baath *et al.*, 1998, Rodriguez *et al.*, 2004 and Belyaeva, *et al.*, 2005).

The present work aimed to study the effect of amending two kinds of soils (sandy clay loam and sandy) with two levels of sewage sludge (2% and 4% w/w) on the microbial populations and the soilenzyme activities such (dehydrogenase, phosphatase and urease activities) as well as total nitrogen, ammonia, and nitrate nitrogen under controlled conditions in the laboratory.

MATERIALS AND METHODS

In this experiment, the effect of the treatment of the soil with different rates of sewage sludge on the enumeration and activities of different microbial groups was studied in two types of soil which have never received sewage sludge before. These types were sandy clay loam and sandy soils (sandy clay loam soil has been collected from El-Hosseinia district. Sharkia governorate, and sandy soil has been collected from Fakous district. Sharkia governorate).

The two soils were air dried, pulverized to pass through a 4- mm stainless steel sieve. The soils were analyzed for their physical and chemical properties (Table 1). Plastic pots 15cm in diameter were filled with 1.5 kg of sandy clay loam soil or sandy soil. The two soils were first treated by mixing thoroughly with sludge before putting them in the pots, as follows.

- 1- Soil without sludge.
- 2- Soil treated with sewage sludge (2% w/w).
- 3- Soil treated with sewage sludge (4% w/w).

The chemical properties and the heavy metals content of the sludge used in this experiment are shown in (Table 2) by plasma optical emission mass spectrometer (POEMS 3)

replicates for Three each were used. The treatment experiment was designed and the pots were placed in the incubator under controlled conditions (in the dark at $28\pm 2^{\circ}$ C. WHC 60%). The microbiological and chemical measurements have been carried out at intervals zero, 7, 14, 28, 42 days of incubation.

Microbiological Analysis

Determination of the total counts and some specific groups of microorganisms in soils during the time course of this study were carried out as follows:

An amount of 10 g of soil sample was accurately weighed

Table 1.Physical and chemical properties of the soils under investigation.

Type of soil	D	S	oil
analysis	Properties -	Hossenia	Fakous
Physical analysis	рН	7.6	8.7
	EC dsm ⁻¹	2.53	3.17
Mechanical	Sand %	55.89	93.16
analysis	Silt %	16.80	5.20
	Clay %	27.31	1.64
	Type of soil	Sandy clay loam	Sandy
Chemical analysis (meq / 100 g soil)	Cation Ca ++ Mg ++ Na K Anion HCO ₃ Cl SO ₄ CO ₃	2.5 1.5 0.8 0.06 0.63 0.46 1.32 Traces	1.8 0.9 2.85 0.12 2.8 1.2 0.81 Traces

 $\label{thm:content} Table~2. Heavy~metals~content~of~sewage~sludge~and~the~soils~under~investigation~(mg/L)$

	Sewa	ge sludge	Sand	y clay soil	Sai	ndy soil
parameter	Total	Available	Total	Available	Total	Available
Aluminum	2801	0.559	5013	1.960	1883	0.820
Boron	84.83	1.510	214.2	0.765	26.54	0.082
Calcium	14070	165.2	17410	113.7	2012	305.0
Cadmium	0.435	0.033	0.260	< 0.001	0.333	< 0.001
Cobalt	26.92	0.559	2.418	0.261	1.745	0.035
Chromium	49.46	0.292	19.80	0.166	9.011	< 0.007
Copper	52.99	26.28	38.88	6.629	15.38	0.330
Iron	38010	345.0	16910	84.67	2254	14.57
Magnesium	878.9	80.97	13580	916.0	569.3	55.39
Manganese	133.8	37.25	739.0	82.14	42.37	5.511
Molybdenum	139.8	0.700	9.798	0.391	5.975	0.064
Nickel	55.27	5.929	25.50	0.888	7.661	0.057
Lead	26.46	12.99	< 0.01	< 0.01	2.303	0.255
Strontium	171.5	3.193	71.41	2.011	18.93	2.648
Vanadium	143.1	7.305	12.40	0.375	7.546	0.258
Zinc	189.1	92.27	98.16	1.076	30.85	1.033

transferred to a bottle containing 90 ml of sterile phosphate saline solution (pH 7.3) according to Page (1982). The bottles were vigorously shaken for 10 minutes on a mechanical shaker. From each bottle, serial dilutions were made, and one ml from each desired dilution was used according to plate count technique and the dilution frequency method.

Plate count technique:

It was used to count bacteria, fungi and actinomycetes in soil samples using the following media:

- 1-Soil extract agar medium for counting the bacteria (Baruah and Barthakur, 1997).
- 2- Martin's rose bengal medium for counting the fungi (Baruah and Barthakur, 1997).
- 3-Dextrose- nitrate agar medium for counting the actinomycetes (Black, 1965).

The media were sterilized at 121°C for 20 minutes. In all cases, triplicate plates were prepared and inoculated with 1 ml of each desired soil solution under asceptic condition, and plates were incubated at 30 °C. The counting was carried out after 5, 7, and 14

days of incubation for plates of bacteria, fungi, and actinomycetes, respectively. The counts were then calculated to obtain a number of organisms per gram dry soil.

Most probable number (MPN) methods:

This technique was used according to Cochron Tables, (Alexander, 1982).to determine the densities of nitrifiers (Nitrosomonas and Nitrobacter), aerobic N₂- fixing Azotobacter and spore formers clostridia. In this connection, the following media were used:

- 1-Ammonium- calcium carbonate medium (Black, 1965).
- 2-Nitrite- calcium carbonate medium (Black, 1965).
- 3-Modified Ashby's medium for *Azotobacter* (Abdel- Malek & Ishac, 1968).
- 4-Modified Winogradsky's medium for anaerobic spore former nitrogen fixer *Clostridium* (Allen, 1959).

Ammonium oxidizers:

Ammonium oxidizers counting tubes were incubated at 30 °C for 30 days. After incubation period, spot test for nitrite was

carried out using Griess- Ilosvay reagent according to Black (1965).

Nitrite oxidizer:

Nitrite oxidizer counting tubes were incubated at 30 °C for 30 days. After this time, nitrate spot test was carried out using Griess-Ilosvay reagent according to Black (1965).

Azotobacter:

After 3 weeks of incubation at 30 °C, cultures were considered positive when turbidity and presence of characteristic pellicle were observed, and detection of the organism in the stained preparations.

Spore formers clostridià:

After subjecting the soil solution to pasteurization to get rid of the vegetative cells, the anaerobic spore former nitrogen fixer Clostridium, using the above mentioned selective liquid media, was determined. After inoculation, the tubes were covered with a sterile layer of vaspar and incubated at 30 °C for 3 weeks. The presence of clostridia was detected by the accumulation of gases and by observation of non mistakable plectridial sporangia microscopically.

Chemical Analysis of the Soil

Chemical analysis including ammonium, nitrate, total nitrogen, and the activities of some soil important enzymes, such as dehydrogenase, phosphatase and urease, were also determined periodically in the soil according to Page (1982).

RESULTS AND DISCUSSION

In this study two kinds of soil textures—were used for studying the effect of amending the soil with sludge as a source of heavy metals on the microbial densitics, soil enzyme activities and the changes in the chemical properties of the soil represented as total N, ammonia and nitrate nitrogen. Sewage sludge was used as an organic amendment (2% and 4%) besides an unamended soil as a control.

Effect of Application of Different Rates of Sewage Sludge to the Soil on the Microbial Counts.

Total bacterial counts:

Data in Table 3 show the effect of adding two different rates of sewage sludge to sandy clay loam and sandy soils on the total bacterial counts during 42 days of incubation. In both soils the treatment with 2% of sewage

Table 3. Total counts of bacteria, fungi, and actinomycetes in sandy clay loam and sandy soils as affected by applications of two rates of sewage sludge (2% and 4%)

		Sand	y clay loa	m soil	Sandy soil								
Days	0	7	14	28	42	0	7	14	28	42			
Treat.]	Bacteria (x 10° CF	U g-1 dr	y soil)		· · · · · · · · · · · · · · · · · · ·				
Control	82.02a	122.94b	110.67c	96.62b	75.27c	39.97b	51.95b	57.80a	43.90b	31.46b			
2%	82,59a	140.27a	176.35a	128.37a	116.62a	43.29a	56.24a	53.90b	51.46a	51.29a			
4%	83.78a	98.37c	153.51b	125.67a	85.81b	40.97b	49.39c	51.46c	49.63a	49.63a			
	LSD 3	.280		LSD 1.928									
			Fungi	$(x 10^2 \text{ fu})$	ingal proj	pagule g	g ⁻¹ dry so	oil)					
Control	119.31b	205.13c	208.64c	200.16b	198.41b	52.43b	71.09b	57.56b	49.75c	46.21c			
2%	120.67b	239.45b	244.59a	245.13a	229.72a	53.04b	82.07a	72.43a	89.63a	62.56a			
4%	123.64a	270.27a	215.13b	178.24c	167.97c	57.19a	60.24c	56.46b	72.19b	55.48b			
	LSD 1	.998				LS	D 1.621						
			Act	inomycet	es (x 10 ⁴	CFU g ^{-t}	dry soil))					
Control	83.41a	92.43c	95.43c	132.43c	128.10b	53.04a	53.29b	59.39c	62.80b	60,60c			
2%	84.45a	111.62a	128.78a	145.13a	147.16a	50.60b	52.19b	73.78a	77.32a	80.24a			
4%	81.48b	98.24b	115.81b	141.21b	120.81c	49.14b	58.90a	69.39b	77.07a	75.97b			
	LSD 1	.107				LS	SD 1.564	Į.					

sludge caused significant increase in the total counts of bacteria comparing with either of the unamended soil or treatment received 4% sewage sludge. These results were in agreement with those obtained by Mikanova et al., (2001) and Speir et al., (2004) who found that significantly higher numbers of total countable bacteria were found at the least-polluted sites. Numbers decreased with proximately to the pollution site (with increasing concentration of pollutants). Data also show that the highest bacterial count obtained after 14 days of incubation in sandy clay loam soil $(1.7 \times 10^7 \text{ CFU g}^{-1} \text{ dry soil})$. On the other hand, sandy soil gave the highest bacterial counts (5.6×10^6) CFU g⁻¹ dry soil) after incubation time of 7 days, when treated with 2% sewage sludge. Generally, the total counts of bacteria in sandy clay loam soil increased faster after 7 days of incubation reaching about 150%, 171% and 120% over total counts of bacteria obtained at zero time as influenced by the application of 0, 2 and 4% sewage sludge, respectively. The same trend was obtained in sandy soil after 7 days of incubation time reaching 130%, 141% and 124% over the control when 0, 2 and 4% of sewage sludge were used, respectively. Regardless the

incubation time, the data in Table 3 show also that the amended sandy clay loam and sandy soils with 2% of sewage sludge enhanced the rate of the bacterial counts. These results were in a harmony with those obtained by Barakah et al., (1995) and Stoven et al., (2005) who mentioned that amendment of loamy soil with sludge at the common rate (2%) enhanced the microbial flora with no harmful effects to plant growth or soil fertility and can be used agricultural safely in land. However, Kelly et al., (2003); Smejkalova et al., (2003) and Rajapaksha et al., (2004) found that heavy metals caused decrease in CFU in the case of oligotrophic bacteria.

Our results show also that the application rate of 4% sewage sludge to the soil caused higher counts of bacteria on both kinds of soils than the unamended soils (control), since the higher total counts obtained were 1.5×10^7 CFU g⁻¹ dry soil and 5.1×10^6 CFU g⁻¹ dry soil in the 14^{th} day of incubation time in the sandy clay loam and sandy soils, respectively. This indicates that there was no harmful effect on the total bacterial counts in both soils, under the use of sewage sludge (4%) to the soils.

Total fungal counts

Data in Table 3 show the density of the total fungal counts in the sandy clay loam and sandy soils as affected by sewage sludge application. From this Table we can realize that sandy clay loam soil which was amended with 4% sewage sludge gave the highest value of fungal counts reaching 2.7 × 10⁴ fungal propagule g⁻¹ dry soil after 7 days of incubation time. Although the highest value of the total counts of fungi was observed after 7 days of incubation when the soil was amended with 4% sewage sludge. the counts were significantly decreased gradually from the second sampling time of incubation until the end of the experiment reaching 1.7×10^4 fungal propagule g^{-T} dry soil. On the other hand, the amended sandy clay loam soil with 2% sewage sludge gave a significant higher values of fungal counts through the 42nd day of incubation comparing with the unamended sandy clay loam soil.

Concerning the sandy soil, the data in Table 3 show that the fungal counts were significantly higher in the soil amended with 2% sewage sludge than either the unamended or the amended with 4% sewage sludge.

In conclusion, the application of sewage sludge with the rate of 2% to sandy clay loam and sandy soils was useful concerning the density of fungi. These results were in harmony with those Kacprzak obtained by and Stanczyk (2003), Awad and Fawzy (2004), Pisarek and Moliszewska, (2004) and Rajapaksha et al., (2004) who found that fungi activity initially increased with the level of metal contamination, being up to 3 and 7 times higher than that in the control samples during the first week at the highest level of Zn and Cu addition, respectively. The positive effect of metal addition on fungal activity was observed then decreased, but fungal activity was still higher in contaminated soil than in control soil after 35 days. Also Baneriee et al., (1997) found that single and repetitive applications of different amount of sludge significantly increased the amount of soil microbial biomass but reduce the functional diversity of microbial community at the highest rate of application. On the other hand, these results were in disagreement with those obtained by Ropek and Para (2002), Kelly et al., (2003), Gharieb et al., (2004) and Stoven et al., (2005) who found that heavy

metals ions inhibited fungal growth.

Total actinomycetal counts

Data in Table 3 show the effect of the addition of different rates of sewage sludge to sandy clay loam and sandy soils on the actinomycetal counts.

Generally, the amended soil with 2% sewage sludge increased of actinomycetes the counts significantly than both unamended soil (control) or the amended soil with 4% sewage sludge in sandy clay loam as well as in sandy soil. Data also show that the highest numbers of actinomycetes were obtained after the 42nd day of incubation time in both sandy clay loam and sandy soils treated with 2% sewage sludge reaching 1.47×10^6 and 8.02×10^5 CFU g⁻¹ dry soil, respectively. These results were in harmony with those obtained by Awad and Fawzy (2004) and Stoven et al., (2005) who found that the addition of sewage sludge with low heavy metal content as well as the addition of heavy metal polluted sludge showed increase counts of actinomycetes compared to the control. On the other hand, these results were disagreement with those obtained by Kelly et al., (2003) who stated that soils with higher levels of metals contamination showed decrease in actinomycetes.

Aerobic nitrogen fixers (Azotobacter spp)

Treatment of sandy clay loam and sandy soils with either 2% or 4% sewage sludge amendments increased the populations nitrogen aerobic fixers (Azotobacter spp.) significantly as compared with the control (unamended soil) through the experimental periods (Table 4). Data also show that in sandy clay Ioam soil, the highest counts of nitrogen aerobic fixer were obtained in the 14th day of incubation period with the two levels used from the sewage sludge $(2\% \text{ and } 4\%) \text{ reaching } 4.72 \times 10^3$ cells g⁻¹ dry soil. Comparing the counts of Azotobacter spp in sandy clay loam and sandy soils, it was clear that these counts were lower in the latter soil.

Concerning the sandy soil, the highest counts of Azotobacter spp was found at the 7th day of incubation period when 4% sewage sludge was applied reaching 1.45 x 10³ cells g⁻¹ dry soil. This count, decreased sharply in the 14th day since the counts reached approximately half of the value obtained at the 7th day.

The data show also that there is a some stimulation in *Azotobacer* spp counts after 14 days of

Table 4. Counts of aerobic and anaerobic N_2 fixers (Azotobacter spp and Clostridium spp) in sandy clay loam and sandy soils as affected by applications of two rates of sewage sludge (2% and 4%)

		Sandy	clay loa	ım soil	Sandy soil							
Days	0	7	14	28	42	0	7	14	28	42		
Treat. Azotobacter spp (x10 ³ cells g ⁻¹ dry soil)												
Control	2.78a	3.09b	3.09b	2.32c	1.45c	0.51a	0.56c	0.46c	0.46c	0.36b		
2%	2.32b	4.24a	4.72a	3.64a	2.47a	0.52a	0.71b	1.13a	1.08a	0.93a		
4%	2.63a	4.18a	4.72a	2.91b	1.96b	0.56a	1.45a	0.71b	0.50b	0.35b		
	LSD 0.	214			LSD 0.072							
			Cle	ostridiun	spp (x)	10 ³ cells	g ⁻¹ dry s	oil)				
Control	3.72a	4.18a	3.64b	3.25b	2.91b	0.78a	0.91c	0.89b	0.59b	0.58a		
2%	3.78a	4.24a	6.16a	3.64a	3.31a	0.80a	1.96a	2.08a	1.03a	0.52a		
4%	3.25b	2.91b	2.31c	2.16c	1.89c	0.63b	1.40b	0.71c	0.52b	0.46a		
	LSD 0.	125				LSD	0.137					

incubation period even though in control treatment. This increase in Azotobacter spp counts decreased sharply after 28 days of incubation time till the end of the experiment reaching 1.45 x 10³ cells g⁻¹ dry soil in sandy clay loam soil when no sludge was added. On the other hand, the counts of Azotobacter spp reaching 2.47 x 10³ and 1.96 x 10³ at the end of the experimental periods when the sludge was added with the levels of 2% and 4%, respectively.

Although the counts of Azotobacter spp were lower in sandy soil, the application of sewage sludge with the level of 2% gave a significantly higher values comparing with an unamended one. These results were agreement with those obtained by Badran, (1983) who reported that the sludge may also enhance growth of aerobic N2 fixers. And resuits these also were agreement with those obtained by Barakah et al., (1995) who found that the amended of the soil with sewage sludge (2%) significantly enhanced N₂ fixers.

By contrast the higher level of sewage sludge (4%) gave lower counts of *Azotobacter* spp reached about half of the counts obtained when the low level (2%) was used. These results were in harmony

with those obtained by Balzer and Ahrens (1991), Athar and Masood (2002) and Vasundhara *et al.*, (2002) who reported that numbers of *Azotobacter* in soils of treatments receiving 2.5 t ha⁻¹ of sewage sludge regularly at Giessen were tenfold greater than in soil receiving NPK; by contrast, they were only one-tenth of the numbers in the NPK control in a treatment receiving sludge at 5 t ha⁻¹.

Anaerobic nitrogen fixers (Clostridium spp)

Data in Table 4 show that the highest number of anacrobic spore formers bacteria was obtained at the 14th day of incubation period reaching 6.16×10^3 cells g^{-1} dry soil when sandy clay loam soil was treated with 2% of sewage sludge. Concerning the sandy soil, the highest number of anaerobic spore formers bacteria was obtained at the 14th day of incubation period reaching 2.08 x 10³ cells g⁻¹ dry soil. Data also show that at the end of the experiment, the counts of anaerobic spore formers bacteria in the treatment of 4% of sewage sludge were lower than those obtained with the control in both sandy clay loam and sandy soils, this decrease was significant in sandy clay loam and was non significant in sandy soil. Generally, data show that in sandy clay loam soil the counts of anaerobic spore formers bacteria with the treatment of 2% sewage sludge were increased significantly in the 14th day and then decreased until the end of the experiment. These results were in agreement with those obtained by Barakah et al., (1995) who found that the amended soil with sewage sludge (2%) significantly enhanced N₂ fixers. On the other hand, application of sewage sludge (4%) in sandy clay loam soil caused a reduction in the counts of the anaerobic spore formers bacteria reaching about 50% of the counts obtained in unamended one. This reduction was observed in the 7th day and continued until the end of the experiment. Our results were in agreement with those obtained by Smejkalova et al., (2003), Xu et al. (2003) and Heras et al., (2005) who found that supplementing of the Cu (II)-exchanged montmorillonite in the diet of broilers decreased the numbers of Clostridium. By contrast in sandy soil, the treatments with unamended as well as the addition of 4% sewage sludge caused an increase in the counts of anaerobic spore formers bacteria in the 7th day of incubation period and then decreased gradually until the end of the experimental periods.

Nitrifiers bacteria

Data in Table 5 show the effect of application of sewage sludge on Nitrosomonas spp counts in sandy clay loam and sandy soils. The data show that the highest counts of Nitrosomonas spp was observed in the 14th day in the amended sandy clay loam soil with 2% sewage sludge reaching 2.35×10^3 cells g^{-1} dry soil. Data also show that the counts at the rate of 2% sewage sludge were higher than those unamended or 4% sewage sludge. Also, in sandy soil higher counts of Nitrosomonas spp was found with the treatment of 2% sewage sludge compared with unamended or 4% sewage sludge.

Data also show that highest number of *Nitrosomonas* spp in sandy soil was obtained at the 7th day of incubation period reaching 1.40 x 10³ cells g⁻¹ dry soil when the soil was amended with 2% sewage sludge. Generally in sandy soil, the counts of spp increased Nitrosomonas 7^{th} significantly in the comparing with the control and then decreased until the end of the experimental periods.

However, a great increase was observed in sandy soil at the 7th day of incubation time when it was treated with 2% sewage sludge

Table 5. Counts of nitrifying bacteria (Nitrosomonas spp and Nitrobacter spp) in sandy clay loam and sandy soils as affected by applications of two rates of sewage sludge (2% and 4%)

		Sand	y clay lo	am soil		Sandy soil					
Days	0	7	14	28	42	0	7	14	28	42	
Treat.	<u></u>	· 	Niti	rosomon	as spp (2	x10 ³ cells	s g ⁻¹ dry	soil)			
Control	0.89a	1.21b	1.32b	0.94c	0.72b	0.40a	0.52c	0.48b	0.42b	0.35b	
2%	0.94a	1.41a	2.35a	1.85a	1.08a	0.42a	1.40a	0.94a	0.60a	0.50a	
4%	0.94a	1.32ab	1.17c	1.08b	1.06a	0.39a	0.59b	0.42c	0.39c	0.31b	
	LSD 0	.136				LSD 0.065					
			Ni	trobacte	r spp (x1	10 ³ cells	g ⁻¹ dry s	oil)			
Control	0.54ab	0.84b	0.93c	0.66c	0.66c	0.17a	0.31b	0.37b	0.29a	0.26a	
2%	0.64a	1.63a	1.85a	1.41a	1.13a	0.20a	0.87a	0,60a	0.35a	0.31a	
4%	0.52b	1.56b	1.32b	0.93b	0.84b	0.23a	0.35b	0.32b	0.32a	0.24a	
	LSD 0	.109				LSD 0.076					

reaching 3.5 folds of the counts obtained at the beginning of the experiment.

Also data in Table 5 show the influence of two levels of sewage sludge applied to sandy clav loam and sandy soils on Nitrobacter spp counts. Data show that the highest value of Nitrobacter spp counts was found at the 14th day in sandy clay loam soil which was amended with 2% of sewage sludge reaching 1.85 x 10³ cells g⁻¹ dry soil. On the other hand, in sandy soil, the highest number was found at the 7th day with the application of 2% scwage sludge reaching 8.7 x 10² cells g⁻¹ dry soil. Data also show that, the amended soil with 2% caused a significant increase in the number of Nitrobacter spp comparing with either the control or the application of 4% of sewage sludge in both sandy clay loam and sandy soils. These results and the results obtained with the spp Nitrosomonas were agreement with those obtained by Yang et al., (2005) who found that the addition of cadmium at low levels stimulated the nitrifying activity in soil, while inhibitory influence were shown at high levels. Nitrifying bacteria was proved to be the most sensitive one.

In conclusion, the application of sewage sludge with the rate of 2% caused a significant increase in Nitrobacter spp counts in sandy clay loam soil as well as in sandy soil reaching 1.85×10^3 and 8.7×10^3 10² cells g⁻¹ dry soil, respectively. Our results concerning nitrifiers bacteria were in agreement with those obtained by Barakah et al., (1995), Furczak and Joniec (2002) and Jezierska and Frac (2005) who found that the amended soil with sewage sludge (2%) significantly enhanced nitrifying bacteria. On the other hand, the measurement of the potential ammonia oxidation indicated that ammonia oxidizing bacteria were completely inhibited in the contaminated soil with heavy metals.

Assay of the Soil-Enzyme Activities

Dehydrogenase activity:

Data in Table 6 show the effect of addition of sewage sludge to the soil on the dehydrogenase activity in sandy clay loam and sandy soils. The data show that dehydrogenase activity was higher in the soil amended with 2% of sewage sludge than those of the unamended and amended with 4%

Table 6. Dehydrogenase and urease activities in sandy clay loam and sandy soils as affected by applications of two rates of sewage sludge (2% and 4%)

			_			~~	-					
		Sand	y clay lo	am soil		Sandy soil						
Days	0	7	14	28	42	0	7	14	28	42		
Treat.		Dehydrogenase activity (micro mole H ₂ / g dry soil / day)										
Control	3.83	4.29	5.63	4.46	3.70	1.39	1.63	1.86	0.90	0.77		
2%	4.07	5.57	5.86	5.06	4.72	1.41	1.99	2.60	1.91	1.62		
4%	3.83	4.46	5.35	4.59	3.76	1.40	1.94	1.97	1.09	0.94		
		\mathbf{U}_{1}	rease act	tivities (ppm uti	lized ure	a / g dry	soil / da	ay)			
Control	60.05	62.32	57.76	47.94	43.66	14.47	18.69	20.10	20.91	19.63		
2%	61.11	65.51	66.51	57.32	47.47	16.02	32.84	26.98	24.91	23.72		
4%	59.06	65.40	61.98	51.36	43.78	16.80	27.40	22.15	17.83	15.62		

of sewage sludge. These results might be due to the lower concentrations of heavy metals which could be reflected on the total counts of bacteria as well as dehydrogenase activity. In addition

the data show that the highest values of dehydrogenase activity were observed in the 14th day in amendment with 2% of sewage both kind of soils sludge in reaching 5.86 and 2.60 micro mole H₂ g⁻¹ dry soil/day in sandy clay loam and sandy soils, respectively. It was obvious that dehydrogenase activity increased sharply in the 14th day then decreased gradually. These phenomena appeared in both sandy clay loam and sandy soils in all the treatments used. These results were in harmony with those obtained by Furczak and Joniec (2002), Crecchio et al., (2004) and Stoven et al., (2005) who found that the addition of municipal solid waste to cropped plots increased dehydrogenase activity by 9.6%.

Concerning the application of sewage sludge with the level of 2%, it was obvious that this treatment increased the dehydrogenase activity in both kinds of soils. On the other hand, the application of sewage sludge with 4% to sandy clay loam and sandy soils caused a reduction in the dehydrogenase activity

comparing with the unamended soil (with no sludge). This reduction in dehydrogenase activity with the application of 4% sewage sludge might be due to the higher concentrations of heavy metals (Table 2). Similar results were obtained by Chander and Brookes, (1991), Mikanova et al., (2001), Lee et al., (2002) and Ying et al., (2002) who mentioned that activity the lowest dehydrogenase was found in contaminated soil with heavy metals, Also Reddy et al., (1987) and Fliessbach and Reber, (1990) found that dehydrogenase was more sensitive to sewage sludge.

As a conclusion, dehydrogenase activity at 42nd day of incubation time was still higher when the soil was amended with 2% sewage sludge than either of the treatment with 4% sewage sludge or unamended one.

Urease activity:

Data in Table 6 show the effect of addition of sewage sludge to the soil on the urease activity in sandy clay loam and sandy soils. Data also show that the highest value of urease activity was observed in sandy clay loam soil at the 14th day in the treated soil with 2% of sewage sludge reaching 66.51 ppm utilized urea g⁻¹ dry soil/day. On the other hand,

in sandy soil, the highest value of urcase activity was observed at the 7th day in amendment with 2% of sewage sludge reaching 32.84 ppm utilized urea g⁻¹ dry soil/day. These results were in harmony with those obtained by Oiling et al., (2002) who found that the activity of urease in soil increased with increasing rates of sewage sludge. At the same time, Stuczynski et al. (2003) demonstrated that Cd has stimulatory effect on urease activity in soil.

The data in Table 6 show also that the treatment of sandy soil with 4% sewage sludge stimulates the activity of urease at the beginning of the experiment at the 7th day then caused an inhibition effect reaching 15.6 ppm utilized urea g⁻¹ dry soil / day at the end of the experimental period. This value was lower than those obtained with the unamended or the amended soil with 2% sewage sludge which reaching 19.63 and 23.72 ppm utilized urea g⁻¹ dry soil/day, respectively. From this data we can conclude that higher percentage of sewage sludge application caused an inhibitory effect than the lower percentage of sewage sludge. The latter caused a stimulatory effect. These results were in agreement with those obtained by Giridhara and Siddaramappa, (2002), Wen et al., (2002), Liu-Chung et al., (2002), Liu-Xia, et al., (2003) and Belyaeva, et al., (2005) who found that urease activity in the soil treated with heavy metals was decreased compared with the control. On the other hand, Renella et al., (2005) found that urease activity was not affected by sludge amendment.

Phosphatase activity:

Data in Table 7 show the effect of the addition of different concentrations of sewage sludge to sandy clay loam and sandy soils on the phosphatase activity. Generally, it could be observed that alkaline phosphatase gave lower values than those of acidic phosphatase activity in sandy clay loam soil. By contrast, no clear different values were obtained between the two kinds phosphatase activity in sandy soil. On the other hand, alkaline phosphatase measurements gave the higher values in sandy soil comparing with those obtained in sandy clay loam soil. Data also show that, in sandy clay loam soil the highest values of acidic phosphatase were observed at the 14th day in either of unamended or an amended soil with 4% of

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Table 7. Phosphatase activity (μg P / g dry soil / day) in sandy clay loam and sandy soils as affected by applications of two rates of sewage sludge (2% and 4%)

	0		7		14	4	28	42				
Acidic	alkaline	e Acidic	alkaline	Acidic	alkaline	Acidic	alkaline	Acidic	alkaline			
Sandy clay loam soil												
40.21	2.39	41.94	3.36	43.28	3.51	40.31	3.11	33.36	2.13			
40.03	2.68	41.34	3.21	42.25	3.26	40.09	3.27	34.95	2.58			
39.32	2.31	42.24	3.52	43.26	3.91	41.21	3.68	33.67	2.35			
				sand	ly soil							
6.66	8.36	7.03	10.20	6.83	10.33	6.72	10.36	5.93	9.63			
7.41	8.21	6.49	10.51	6.95	10.50	6.34	10.41	6.41	10.11			
6.35	8.96	5.44	11.31	8.08	10.42	7.03	10.32	6.52	10.23			
	40.21 40.03 39.32 6.66 7.41	40.21 2.39 40.03 2.68 39.32 2.31 6.66 8.36 7.41 8.21	Acidic alkaline Acidic 40.21 2.39 41.94 40.03 2.68 41.34 39.32 2.31 42.24 6.66 8.36 7.03 7.41 8.21 6.49	Acidic alkaline Sa 40.21 2.39 41.94 3.36 40.03 2.68 41.34 3.21 39.32 2.31 42.24 3.52 6.66 8.36 7.03 10.20 7.41 8.21 6.49 10.51	Acidic alkaline Acidic Sandy cla 40.21 2.39 41.94 3.36 43.28 40.03 2.68 41.34 3.21 42.25 39.32 2.31 42.24 3.52 43.26 sand 6.66 8.36 7.03 10.20 6.83 7.41 8.21 6.49 10.51 6.95	Sandy clay loams Sandy clay loams 40.21 2.39 41.94 3.36 43.28 3.51 40.03 2.68 41.34 3.21 42.25 3.26 39.32 2.31 42.24 3.52 43.26 3.91 sandy soil 6.66 8.36 7.03 10.20 6.83 10.33 7.41 8.21 6.49 10.51 6.95 10.50	Sandy clay loam soil 40.21 2.39 41.94 3.36 43.28 3.51 40.31 40.03 2.68 41.34 3.21 42.25 3.26 40.09 39.32 2.31 42.24 3.52 43.26 3.91 41.21 sandy soil 6.66 8.36 7.03 10.20 6.83 10.33 6.72 7.41 8.21 6.49 10.51 6.95 10.50 6.34	Sandy clay loam soil 40.21 2.39 41.94 3.36 43.28 3.51 40.31 3.11 40.03 2.68 41.34 3.21 42.25 3.26 40.09 3.27 39.32 2.31 42.24 3.52 43.26 3.91 41.21 3.68 sandy soil 6.66 8.36 7.03 10.20 6.83 10.33 6.72 10.36 7.41 8.21 6.49 10.51 6.95 10.50 6.34 10.41	Sandy clay loam soil 40.21 2.39 41.94 3.36 43.28 3.51 40.31 3.11 33.36 40.03 2.68 41.34 3.21 42.25 3.26 40.09 3.27 34.95 39.32 2.31 42.24 3.52 43.26 3.91 41.21 3.68 33.67 sandy soil 6.66 8.36 7.03 10.20 6.83 10.33 6.72 10.36 5.93 7.41 8.21 6.49 10.51 6.95 10.50 6.34 10.41 6.41			

sewage sludge reaching 43.28 and 43.26 µg P g⁻¹ dry soil/day, respectively. Also, in sandy soil the highest value of the acidic phosphatase was obtained at 14th day when 4% of sewage sludge

was applied reaching 8.08 µg P g⁻¹ dry soil/day. From these results it could be concluded that in both soils no harmful effect was observed when sewage sludge was applied even though at the high level of application.

Data also show that the highest value of alkaline phosphatase was observed at the 7th day in an amended soil with 4% of sewage sludge in sandy soil reaching 11.31 µg P g⁻¹ dry soil /day. On the other hand, in sandy clay loam soil the highest value was found in the 14th day in an amended soil with 4% of sewage sludge reaching 3.91 µg P g⁻¹ dry soil /day. These results were in agreement with those obtained by Awad and Fawzy, (2004),Crecchio et al., (2004) and Speir et al., (2004) who found that the increasing rates of sewage sludge higher values promoted of phosphatase activity. By contrast, Lee et al., (2002); Wang et al., (2002) and Stuczynski et al., (2003) found that the pollution with heavy metals (Cu and Zn) caused an inhibitory influence on phosphatase activity. However Brookes et al., (1984), Reddy et al., (1987) and Gzik et al., (2003) stated that phosphatase activity was not affected with increasing metals.

Effect of Different Concentrations of Sewage Sludge on Total Nitrogen, Ammonia and Nitrate of the Soil.

Total nitrogen

Data in Table 8 show the influence of the addition of two rates of sewage sludge to sandy clay loam and sandy soils on total soil nitrogen content. Total nitrogen in two kinds of soils increased with the addition of sewage sludge to both soils.

Concerning the application of 2% sewage sludge, it was clear that this treatment caused an increase in the total nitrogen with comparing the control treatment. These values reaching 131, 136, 137, and 135% over the values obtained with the control after 7, 14, 28, and 42 days of incubation time. Data also show that in sandy clay loam soil, the highest value of total nitrogen was obtained after 7 days of incubation time in the soil which was treated with 4% sewage sludge reaching 634 mg kg⁻¹ dry soil. On the other hand, the values of the total

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Table 8 . Total nitrogen (mg kg $^{-1}$) in sandy clay loam and sandy soils as affected by applications of two rates of sewage sludge (2% and 4%)

Treatments	Days									
	0	7	14	28	42					
		Sandy	clay loar	n soil						
Control	435	438	448	420	407					
2%	510	572	609	577	550					
4%	547	634	630	612	580					
		9	Sandy soil							
Control	286	249	261	236	199					
2%	320	332	340	323	273					
4%	361	378	368	336	311					

nitrogen decreased gradually with increasing the incubation time, which reaching 580 mg kg⁻¹ dry soil at the end of the experimental period In sandy soil, total nitrogen was increased with the addition of 2% as well as 4% sewage sludge. Although the highest value was obtained when sewage sludge was used at the rate of 4% reaching 378 mg kg⁻¹ dry soil after 7 days only, but when 2% sewage sludge was added, the highest value was obtained after 14 days reaching 340 mg kg⁻¹ dry soil. These values with the two rates were decreased gradually until the end of the experimental time reaching 273 and 311 mg kg⁻¹ dry soil when 2% and 4% sewage sludge were used, respectively.

The increase in total nitrogen as influence of application of 2% and 4% sewage sludge might be due to the increasing numbers of aerobic and anaerobic nitrogen (Azotobacter fixers spp and Clostridium spp) (Table 4). Our results were in agreement with those obtained by Balzer and Ahrens (1991) who found that nitrogen increased with increasing application of sewage sludge. Also Barakah et al., (1995) found that total nitrogen in rhizosphere and nonrhizosphere of alfalfa and wheat significantly increased with application of organic amendment. Also they added that amendment of the soil with sewage sludge (2%) significantly enhanced nitrogen fixers.

Our results also were in harmony with those obtained by Brofas et al., (2000) who found that the addition of sewage sludge at seven rates (0, 10, 20, 40, 60, 80, and 120 tonnes ha⁻¹) in a field experiment increased nitrogen. Also, they found that the nitrogen total concentration decreased with the time after application. sewage sludge Generally, our results show that the total counts of bacteria, fungi and actinomycetes in sandy clay loam soil (Table 3) were higher when sewage sludge was applied comparing with the control. This increase might be due to the higher production in sugar and organic acid which were necessary for growing Azotobacter spp and hence stimulate the nitrogen fixation in the soil (Table 8). These results were in agreement with those obtained by Barakah (1995); Crecchio et al., (2004), Speir et al., (2004), Stoven et al., (2005) and Heras et al., (2005) who mentioned that the amendment of the soil with sewage sludge 2% significantly enhanced nitrogen fixers.

Ammonia and nitrate nitrogen

Data in Table 9 show the effect of the addition of two levels

Table 9. Ammonia and nitrate nitrogen (mg kg $^{-1}$) in sandy clay loam and sandy soils as affected by applications of two rates of sewage sludge (2% and 4%)

Days	0		7		1	4	2	8	42	
-	NH ₄	$\overline{NO_3}$	NH ₄	NO ₃	NH ₄	$\overline{NO_3}$	NH ₄	NO ₃	NH ₄	NO_3
Treat.										
				Sa	ndy clay	loam so	il			
Control	18.7	30.7	18.8	32.7	17.0	30.1	14.9	25.6	12.1	20.0
2%	18.6	30.5	19.2	42.0	21.6	44.3	16.8	40.3	15.3	36.8
4%	19.1	30.5	20.5	39.2	16.8	36.4	15.1	35.6	14.2	34.2
					sandy	soil				
Control	10.0	15.3	10.6	13.2	11.2	11.2	10.2	9.6	8.4	8.6
2%	10.2	16.6	14.9	20.0	14.2	28.0	13.1	22.3	10.2	18.6
4%	10.3	16.9	14.0	26.1	13.1	22.6	11.2	20.1	9.3	15.2

of sewage sludge (2% and 4%) on ammonia and nitrate nitrogen in sandy clay loam and sandy soils. The data show also that in sandy clay loam soil the highest values of ammonia nitrogen were found at the 14th day of incubation periods when the rate of sewage sludge application was 2% reaching 21.6 mg kg⁻¹. This value decreased gradually until the end of the experiment reaching 15.3 mg kg⁻¹. Concerning sandy soil, the peak of ammonia was obtained during 7-14 days being 14.9 and 14.2 mg kg⁻¹, respectively when 2% sewage sludge was applied. Generally the addition of sewage sludge to the two soils with the rate of 2% increased the values of ammonia nitrogen than the unamended and the amended soil with the rate of 4% sewage sludge. Although, the application with 4% sewage sludge increased the values of ammonia nitrogen than the control, but these values were lower than those obtained with the application of 2% sewage sludge in both kinds of soils. The highest values of ammonia nitrogen were observed at the 7th day in sandy clay loam and sandy soils when the soils were amended with 4% sewage sludge reaching 20.5 and 14 mg kg⁻¹, respectively. These values decreased gradually until the end experimental the periods of

reaching 14.2 and 9.3 mg kg⁻¹ in sandy clay loam and sandy soils, respectively. These results were in agreement with those obtained by Furczak and Joniec (2002) and Yang et al., (2005) who found that the addition of cadmium at low levels stimulated the ammonifying activity in soil, while inhibitory influence were shown at high levels. Also Jezierska and Frac (2005)found that fertilizer application to the soil with diary sewage sludge resulted in the stimulation of the number of ammonifying bacteria and ammonium nitrogen concentration.

The data also show that the highest value of nitrate nitrogen was found at the 14th day of incubation period with the rate of 2% sewage sludge in sandy clay loam soil reaching 44.3 mg kg⁻¹. The same trend was obtained in sandy soil which gave the highest value reaching 28 mg kg⁻¹. It's obvious that the incubation more than 14 days caused almost plateau line until the end of the experiment in both kinds of soils. The data also show that the addition of sewage sludge to the sandy clay loam and sandy soils with the rate of 2% increased the values of nitrate nitrogen. These results were true in both kinds of soil. These results were in agreement with

those obtained by Furczak and Joniec (2002) who stated that strongly crumbled sludge stimulated most biochemical activities of the soil (ammonification and nitrification). By contrast, these results were in disagreement with those obtained by Kelly et al., (2004), Gremion et al., (2004) and Wilke et al., (2005) who found that ammonia oxidizing bacteria were completely inhibited in the contaminated soil with heavy metals.

The results of this investigation clearly showed that, no harmful effect to microbial population and their activities was observed when sandy clay loam and sandy soils were amended with sewage sludge with the rate of 2% consequently this rate can be used safely in agricultural soils.

REFERENCES

- Abd-EL-Malek, Y. and Y. Z. Ishac. 1968. Evaluation of methods used in counting *Azotobacter*. J. Appl. Bact .31:267.
- Alexander, M. 1982. Most probable number method for microbial population. In: Methods of soil analysis part 2 chemical and microbiological properties pp.815-820 (A.L. page Eds.) Number 9 (part 2)

- in the series Agronomy. Madison, Wismosin USA.
- Allen, O. N. 1959. Experiments in soil bacteriology. Burgess publ.Co., Minneapoles 15, Minnesota, USA.
- Alloway, B. J. 1995. Soil processes, and the behavior of metals. In: Alloway, B.J.(Ed), Heavy metals in soils. Blackie Academic and professional, Glasgow, pp.3-10
- Athar, R. and A. Masood 2002. Heavy metal toxicity: effect on plant growth and metal uptake by wheat, and on free living *Azotobacter*. Water Air Soil Pollution. 138 (1/4): 165-180.
- Austin, B., D. A. Allen, A. L. Mills and R. R. Colwell 1977. Numerical taxonomy of heavy metal-tolerant bacteria isolated from an estuary. Can. J. Microbial. 23:1433-1477.
- Awad, N. M. and K. S. M. Fawzy 2004. Assessment of sewage sludge application on microbial diversity, soil properties, and quality of wheat plants grown in a sandy soil. Annals. Agric. Sci. Cairo. 49 (2): 485-499.
- Baath, E., M. Diaz-Ravina, A. Frostegard and C. Cacpbell 1998. Effect of metal-rich sludge amendments on the soil

- microbial community. Appl. Environ. Microbiol. 64: 238-245.
- Badran, N. M. 1983. Organic manuring and its effect on some chemical and microbiological properties of some Egyptian soils. M.Sc. Thesis, Faculty of Agriculture, Moshtohor, Zagazig University, Egypt.
- Balzer, W. and E. Ahrens 1991.

 Mikrobioloische bewertung von boden aus dauerversuchen mit fouls chlamm. In: Sauer beck, D, and Liibben, S. (Eds.)

 Auswirkungen von Siedlungsabfallen auf Boden.

 Bodenorganismen und Pflanzen, pp.359-389. Berichte aus de Okologische Forschung Band 6, Forschungszentrum, Julich.
- Banerjee, M. R., D. L. Burton and S. Depoe 1997. Impact of sewage sludge application on soil biological characteristics. Agric. Ecosys. Environ. 66: 241-249.
- Barakah, F. N. 1995. Effect of irrigation with sewage water on the nodulation and nitrogen fixation of alfalfa grown on calcareous soils .Annals Agric. Sci., Ain Shams Univ., Cairo, 40 (2): 579-595.

- Barakah, F. N., S. H. Salem, A. H. Heggo and M. A. Bin-Shiha 1995. Activities of Rhizosphere Microorganisms as affected by Application of organic amendments in a calcareous Carbon loamy soil [-Assimilation. Arid Soil Research and Rehabilitation. 9:189-200.
- Baruah, T. C. and H. P. Barthakur 1997. A textbook of Soil Analysis. Vikas Publishing House PVT LTD.
- Belen, Hinojosa, M., J. A. Carreira, R. Garcia, Ruiz and R. P. Dick 2004. Soil moisture pretreatment effects on enzyme activities as indicators of heavy metal contaminated and reclaimed soils. Soil Biol. Biochem. 36 (10): 1559-1568.
- Belyaeva, O. N., R. J. Haynes and O. A. Birukova 2005. Barley yield and soil microbial and enzyme activities as affected by contamination of two soils with lead, zinc or copper. Biol. Fertil. Soils 41 (2): 85 94.
- Black, C. A. 1965. Methods of Soil Analysis part 2 Chemical and Microbiological Properties. American Society of Agronomy, Inc., Publisher Madison, Wisconsin, USA.

- Brandt, K. K., P. H. Krogh and J. Sorensen 2003. Activity and population dynamics of heterotrophic and ammonia-oxidizing microorganisms in soil surrounding sludge bands spiked with linear alkylbenzene sulfonate: a field study. Environ. Toxicol. Chem. 22 (4): 821-829.
- Brofas, G., P. Michopoulos and D. Alifragis 2000. Sewage sludge as an amendment for calcareous bawxite mine spoils reclamation. J. Environ. Quality 29 (3): 811-816.
- Brookes, P. C., S. P. Mc Grath, D. A. Klein and E. T. Elliot 1984. Effects of heavy metals on microbial activity and biomass in field soils treated with sewage sludge. In: Environmental Contamination, CEP, Edinburgh, pp. 574-585.
- Chander, K. and P. C. Brookes 1991. Is the dehydrogenase assay invalid as a method to estimate microbial activity in copper-contaminated soils? Soil Biol. Biochem. 23: 909-915.
- Crecchio C., M. Curci, M. D. R. Pizzigallo, P. Ricciuti and P. Ruggiero 2004. Effects of municipal solid waste compost amendments on soil enzyme

- activities and bacterial genetic diversity. Soil Biol. Biochem. 36: 1595-1605.
- Fliessbach, A. and H. Reber 1990.

 Effects of long term sewage sludge applications on soil microbial parameters. Joint Seminar of the Commission of the European Community and German Federal Res. Centre of Agriculture (FAL), 6-8 June 1990, Braunschweig.
- Fliessbach, A., R. Martens and H. H. Reber 1994. Soil microbial biomass and activity in soils treated with heavy metal contaminated sewage sludge. Soil Biol. Biochem. 26:1201-1205.
- Furczak, J. and J. Joniec 2002. Studies of the effects of the level of sewage sludge crumbling on microbial and biochemical activities of soils. Pol. J. Soil Sci. 35 (1): 59-67.
- Gharieb, M. M., M. A. Hefnawy and A. M. Soliman 2004. Alteration of the fungicidal effect of copper oxychloride against Fusarium oxysporum f. sp. lycopersici and Alternaria solani by heavy metals and salinity. Afric. J. Mycol. Biotech. 12 (1): 11-33
- Giller, K. E., E. Witter and S. P. McGrath 1998. Toxicity of

- heavy metals to microorganisms and microbial processes in agricultural soils. A review. Soil Boil. Biochem. 30: 1389-1414.
- Giridhara, M. and R. Siddaramappa 2002. Effect of heavy metals on urease activity on soil. Current Research University, Agric. Sci. Bangalore. 31 (1/2): 4-5.
- Gremion, F., A. Chatzinotas, K. Kaufmann, W. Sigler and H. Harms 2004. Impacts of heavy metal contamination and phytoremediation on a microbial community during a twelve-month microcosm experiment. FEMS- Microbiol. Ecol. 48 (2): 273-283.
- Gzik, A., M. Kuchling, I. Schneider and B. Tschochner 2003. Heavy metal contamination of soils in a mining area in South Africa and its impact on some biotic systems. J. Soil Sediments 3 (1): 29-34.
- Heras, J. de-las, P. Manas and J. Labrador 2005. Effects of several applications of digested sewage sludge on soil and plants. J. Environ. Sci. Health. Part A, Toxic/Hazardous Substances Environ. Eng. 40 (2): 437-451.

- Huisingh, D. 1974. Heavy metals: implications for agriculture. Annu. Rev. phytopathol. 12:375-388.
- Jezierska T. S. and M. Frac 2005. Changes in enzymatic activity and the number of proteolytic microorganisms in brown soil under the influence of organic fertilization and cultivation of spring wheat. Pol. J. Soil Sci. 38 (1): 61-68.
- Kacprzak, M. and E. Stanczyk, Mazanek 2003. Changes in the structure of fungal communities of soil treated with sewage sludge. Biol. Fertil. Soils. 38 (2): 89-95.
- Kandeler, E., C. Kampichler and O. Horan 1996. Influence of heavy metals on the functional diversity of soil microbial communities. Biol. Fertil. Soils 23: 299-306.
- Kelly, J. J., M. M. Haggblom and R. L. Tate 2003. Effects of heavy metal contamination and remediation on soil microbial communities in the vicinity of a zinc smelter as indicated by analysis of microbial community phospholipid fatty acid profiles. Biol. Fertil. Soils. 38(2): 65-71
- Kelly, R. T., I. D. S. Henriques and N.G. Love 2004. Chemical

- inhibition of nitrification in activated sludge. Biotech. Bioeng. 85 (6): 683-694.
- Lee, Insook; Kim, Okkyung, Y. Chang, Yoon, H. Bae, Bum, Kim, H. Hyun and H. Baek, Kyung 2002. Heavy metal concentrations and enzyme activities in soil from contaminated Korean shooting range. J. Biosci. Bioeng. 94 (5): 406-411.
- Liu-Chung, S., Y. Chang-Hong, Y. Sun-Bai, H. Sun-Yu, L. Ye-You and S. Zhang-Fu 2002. Effects of external copper on enzyme activity in soil and apple tree system. Acta-Pedologica Sinica. 39 (1): 37-44.
- Liu-Xia, Q. Liu-Shu and H. Tan-Zhao 2003. The relationship between heavy metal forms and soil enzymatic activities in alluvial meadow soils and meadow cinnamon soils. Acta-Pedologica-Sinica 40 (4): 581-587.
- Mc Grath, S. P. 1987. Long-term studies of metal transfer following application of sewage sludge. In: Pollutant transport and fate in ecosystems. British Ecological Society Special Publication No. 6 (P. J. Coughtrey, M. H. Martin and

- M. H. Unsworth Eds.), pp. 301-317- Blackwell, Oxford.
- Mikanova, O., J. kubat, N. Mikhailovskaya, I. Voros and B. Biro 2001. Influence of heavy metal pollution on some soil-biological parameters in the alluvium of the Litavka River. Rostlina Vyroba. 47 (3): 117-122.
- Page, A. L. 1982. Method of soil analysis part 2 American Soc. of Agron. Soil Sci. Am. Madison, Wisconsin. USA.
- Pichtel, I. R. and Y. M. Hayes 1990. Influence of fly ash on soil microbial activity and populations. J. Environ. Quality 19: 593-597.
- Pisarek, I. and E. B. Moliszewska 2004. Effect of the addition of sewage sludge on the composition of humic substances, content of heavy metals and the presence of *Trichoderma* spp. in the soil. Polish J. Soil Sci. 37 (2): 171-179.
- Qiling, T., H. X. Cheng, Z. J. Hou, R. M. Mclaren, C. Li and W. S. Li 2002. Forms of heavy metal in sewage sludge and its effects on soil enzyme activities of Aquent soil. J. Huazhong, Agric. Univ. 21 (1): 36-39.

- Rajapaksha, R. M. C. P., M. A. Tobor, Kaplon and E. Baath 2004. Metal toxicity affects fungal; and bacterial activities in soil differently. Appl. Environ. Microbiol. 70: 2966-2973.
- Reddy, G. B., A. Faza and R. Bennett 1987. Activity of enzymes in rhizosphere and non-rhizosphere soil amended with sludge. Soil Biol. Biochem. 19: 203-205.
- Renella, G., M. Mench, A. Gelsomino, L. Landi and P. Nannipieri 2005. Functional activity and microbial community structure in soils amended with bimetallic sludge; Soil Biol. Biochem. 37 (8): 1498-1506.
- Roane, T. M. and S. T. Kellogg 1996. Characterization of bacterial communities in heavy metal contaminated soil. Candian J. Microbiol. 42: 593-603.
- Rodriguez, B. B., J. A. Bolbot and I. E. Tothill 2004. Urease-glutamic dehydrogenase biosensor for screening heavy metals in water and soil sample. Analyt. Bioanalyt. Chem. 380 (2): 284-292.

- Ropek, D. and A. Para 2002. The effect of heavy metal ions and their complexons upon sporulation growth, and pathogenicity of the entomopathogenic fungus Verticillium lecanii. ... Invertebrate Pathology. 79 (2): 123-125.
- Smejkalova, M., O. Mikanova and L. Boruvka 2003. Effects of heavy metal concentrations on biological activity of soil micro-organisms. Plant, Soil Environ. 49(7): 321-326.
- Smith, S. R. 1996. Agricultural recycling of sewage sludge and the environment. CAB International, Walling Ford.
- Speir, T. W., J. Horswell, A. Pvan Schaik, R. G. McLaren and G. Fietje 2004. Composted biosolids enhance fertility of a sandy loam soil under dairy pasture. Biol. Fertil. Soils. 40 (5): 349-358.
- Stoven, K., A. Al-Issa, J. Rogasik, S. Kratz and E. Schnug 2005. Effect of long term sewage sludge applications on microorganisms in an arable soil. Landbauforschung-Volkenrode. 55 (4): 219-226.

- Stuczynski, T. I., G. W. McCarty and G. Siebielec 2003. Response of soil microbiological activities to cadmium, lead, and zinc salt amendments. J. Environ. Quality 32(4): 1346-1355.
- Timoney, T. F., J. Port, J. Gites and J. Spanier 1978. Heavy metal and antibiotic resistance in the bacterial flora of sediments of New York Bight. Appl. Environ. Microbiol. 36:465-472.
- Vasundhara, G., G. M. Kurup, V. B. Jacob, M. R. Sethuraj and R. Kothandaraman 2002. Nitrogen fixation by *Azotobacter chroococcum* under cadmium stress. Indian J. Microbiol. 42 (1): 15-17.
- Wang, X. L., J. M. Xu, Z. M. Xie, H.Y. Yao and W. Y. Shi 2002. Effects of Cu and Zn soil contamination on biological indicator of environmental J. quality. Zhejiang University Agric. Life Sci. 28 (2): 190-194.
- Wen, H. X., H. F. Ying, Z. E. Ming and Z. Yiping 2002. Effect of Hg and Cd on soil urease activity. Acta-

- Pedologica Sinica 39 (3): 412-420.
- Wilke, B. M., M. MaiKe, A. Gattinger, M. Schloter and P. Gong 2005. Effects of fresh and aged copper contaminations on soil microorganisms. J. Plant Nutrition Soil Sci. 168 (5): 668-675.
- Xu, Z. R., Y. L. Ma, C. H. Hu, M. S. Xia, T. Guo and H. L. Jin 2003. Effects of Cu (II)-exchanged montmorillanite on growth performance, intestinal microflora, bacterial enzyme activities and morphology of broilers. Asian Australasian. J. Animal Sci. 16 (11): 1673-1679.
- Yang, Y., C. X. Ying, T. M. Guang and Z. J. Zhang 2005. Microbial activity related to N cycling in the rhizosphere of maize stressed by heavy metals. J. Environ. Sci. 17 (3): 448-451.
- Ying, T., H. Y. Chang, J. Long and Y.Y. Huai 2002. Studies on soil enzymatic activities in areas contaminated by tailings from Pb, Zn, Ag mine. China Environ. Sci. 22 (6): 551-555.

تأثير حمأة المجارى على المجاميع الميكروبية وأنشطتها في نوعين مختلفين من التربة

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استخدمت من قبل حمأة المجاري كبديل السماد العضوي لتحسين خصوبة التربة ولكن هناك تخوف من تأثير المعادن الموجودة في الحمأة على ميكروبات التربة وكذلك على نشاط هذه المبكروبات. وفي هذا البحث تم استخدام مستويين من حمأة المجاري مقدرة وزن / وزن بنسية ٧% و ٤% من خلال تجربة معملية لدراسة تأثير هذه الحمأة على كل من أعداد ونشاط الميكروبات وذلك من خلال استخدام نوعين من التربة إحداهما كانت رملية طميية طينية والأخرى كانت تربة رملية حيث أن كلاهما لم يتعرضا لأى إضافة من حمأة المجارى . تم أخذ عينات من هذه التجربة على فترات بعد التحضين على درجة حرارة ٢٨ ± ٢م واستمر ذلك لمدة ٤٢ يوم . واستخلص من هذه التجربة ما يلي : وجود زيادة في الأعداد الكلية أكل من البكتريا والغطر والاكتينوميسيتات وكذلك أعداد كل من بكتريا التأزت والبكتريا المثيتة للنيتروجين اللاتكافلية سواء كانت هوائية أو لا هوائية عند استخدام معدل إضافة بمقدار ٣٠% من الحمأة في كلا النوعين من التربة المستخدمة مقارنة بالتربة غير المعاملة بالحمأة أو تلك التي عومات بمعدل ٤% , كما لوحظ أن المعاملة بالمعدل الأعلى (٤% من الحمأة) تسبب في انخفاض أعداد الميكروبات التي تم تقديرها في هذه التجربة وكذلك انخفضت الأنشطة الخاصة ببعض إنزيمات التربة مثل (الديهيدروجينيز ، الفوسفاتيز واليورييز) عن تلك التي عومات بمعدل ٢% من الحمأة وعلى الرغم من انخفاض أعداد وأنشطة الميكروبات في النربة المعاملة بالتركيز الأعلى إلا أن تلك الأعداد لهذه المجاميع الميكروبية وكذلك أنشطتها كانت في مستوى أعلى مما وجد في التربة غير المعاملة. وقد أدِى استخدام الحمأة بالمعدل الأعلى الى زيادة فى النيتروجين الكلى وصلت الى ١٣٤ و ٣٧٨ مللجم / كجم تربة جافة بعد ٧ أيام فقط من التحضين فى كل من التربة الرملية الطميية الطينية والرملية على التوالى.

ويصفة عامة فإن إضافة حمأة المجارى الى كلا النوعين من التربة المستخدمة فى هذه التجربة بمعدل ٢% حمأة قد أدت الى زيادة القيم المتحصل عليها من الامونيا والنترات عن التربة الاخرى غير المعاملة أو المعاملة بالمعدل الأعلى ولم يلاحظ من هذه التجربة أى تأثير ضار على أعداد الميكروبات أو نشاطها عند استخدام المعدل المتوسط من الحمأة . ونخلص من هذه التجربة أن هذا المعدل (٢%) من الحمأة يمكن استخدامه بأمان في التربة الزراعية .