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# PROPAGATION OF COTONEASTER HORIZONTALIS, DECNE THROUGH IN VITRO CULTURE BY

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#### ABSTRACT

This study has been executed on *Cotoneaster horizonta'is*, Decne at the Tissue Culture and Germplasm Conservation, Research Laboratory, Horticulture Research Institute, Agriculture Research Center during the years of 2006 and 2007. The goal of this research was to establish a protocol for micropropagation of Cotoneasters plants.

The data revealed that for sterilization explants, using (15% Clorox for 10 min) gave the best result for free contamination explants and the highest survival percentage.

For shooting behaviour, adding (3 mg/l BA) to MS-medium increased the shootlet number/explant; fresh and dry weights of shootlets. I owever shootlet length increased by adding (2 mg/l BA) to MS-medium, while the number of leaves increased by adding (1 mg/l 2iP) to MS-medium. For the type of media, culture the shootlet on MS-medium increased the number and length of shootlets, number of leaves, fresh and dry weight compared to culture the shootlets on WPM,  $B_5$  or LS media.

Concerning to chemical composition, the highest amoun of chlorophylla was recorded with MS-medium supplemented with 4 mg/l BA, while the highest amount of chlorophyll-b and carotenoids was recorded with MS-medium supplemented with 2 mg/l kin. Total indoles increased by adding 1 mg/l kin and total soluble phenols increased when using 3 mg/l 2iP.

Regarding to rooting behaviour, the highest percentage of rooted shootlet was recorded when culture the shootlets on full MS-strength with 1 mg/l IAA. For the number and the length of roots, no significant differences between all treatments on number of roots per shootlet and length of root (cm). For acclimatization, the survival percentage was 70%.

Key words: Cotoneaster horizntalis, in vitro culture, BA, Kin, 2iP, MS, WPM, B<sub>5</sub>, LS

#### INTRODUCTION

Cotoneaster horizontalis, Decne family Rosaceae are ornamental shrubs, many of them with decorative fruit remaining usually through the whole winter.

Cotoneaster horizontalis, Decne low shrub: branches almost horizontal and densely distinhously branched, Baily (1976).

Cotoneaster horizontalis, is very rare in Egypt, it is found only in Orman, Giza (Khalifa and Loutify, 2006).

There are many techniques available for the conservation of plant genetic resources of rare and endangered species. These include micropropagation, seed germination, regeneration from callus, micrografting and cryopreservation (Nitzsche, 1983; Rick, 1984; Stanilova et al., 1994). Micropropagation in Cotoneaster plants as an alternative to conventional methods for vegetative propagation attracts much attention, because of its advantages. It increases many times the multiplication level (Novak and Petra, 1981; Takayama and Misawa, 1982 and 1983; Van Aartijk and Blom-Barnhoorn, 1981; Van Aartijk et al., 1990 and Wickremesinhe et al., 1994) and enables material free virus and other diseases to be obtained (Blom-Barnhoorn and Van Aartijk, 1985 and Van Aartijk et al., 1990).

The success of tissue culture in propagation of ornamental plants is greatly influenced by the nature of culture media. The nutrient media has two major functions, the first is to supply the basic nutritional ingredients for continued growth of isolated explants and subsequent propagules; the second function is to direct growth and development through hormonal control (George and Sherrington, 1994).

The aim of this study was to investigate the effect of some sterilization treatments, some growth regulators, and different types of media on shooting behaviour and strength of salt MS-medium with or without auxin and with or without activated charcoal on rooting behaviour of Cotoneaster horizontalis,.

#### MATERIALS AND METHODS

This investigation was carried out in Tissue Culture and Germplasm Conservation Research Laboratory, Horticulture Research Institute, Agriculture Research Center during the years of 2006 and 2007 on Cotoneaster horizontalis, Decine.

The aim of this study was to establish a protocol for micropropagation of C. horizontalis, Decne. Therefore, clorox and period of disinfection were used for sterilization treatments. Multiplication of shootlet was studied by using different kinds of cytokinins (BA, kin or 2iP) with different concentrations adding to MS-medium. Moreover, studying the effect of different type of media (MS, WPM, B<sub>5</sub> or LS media). The rooted shootlets were studied by using strength of MS-medium with different kinds of auxine (free hormone, IAA, NAA and IBA) with or without activated charcoal.

#### Plant material

The explant used for these experiments were taken from C. horizontalis, Decne shrub at Orman Garden, Giza

#### Culture media

Using MS-medium only or with different types of media (WPM,  $B_5$  and LS) enriched with sucrose 25 g/l and agar 7 g/l. The media were adjusted to pH 5.7  $\pm$  0.1, then poured at 25 ml in 200 ml capacity glass jars before autoclaving at 121°c and 1.2 kg/cm² for 15 min.

#### Culture conditions

The cultures were incubated in growth chamber at  $24 \pm 1^{\circ}$ c under 16 hr photoperiod (day light fluorescent tube) at 3 k lux.

### Effect of clorox concentrations and periods of disinfection on sterilization explant

The explants were surface sterilized with 70% (v/v) ethanol solution for one min and then they were treated with 5, 10, 15 and 20% (v/v) clorox with a few drops of tween 20 for 5, 10 and 15 min followed by rinsing three times with a sterile distilled water. After that, they were immersed in 0.1% mercuric chloride (MC) with a few drops of tween-20 for ten min. Finally there were rinsed three times with sterile distilled water and cultured on MS-medium free hormones for two weeks (twenty explants in five replicated). After that, free contamination and survival percentage of explant were calculated.

#### Effect of different concentrations of cytokinins (BA, kin or 2i.')

The free of contamination explants were cultured on MS-medium supplemented with different concentrations of benzyladenine (BA), kinetine (kin),  $6-\gamma-\gamma$ -dimethylallyl aminopurin (2iP) at the rate of (0, 1, 2, 3 or 4 mg/l). The explants were cultured incubated for one month (twenty explants were cultured in five replicate). The shootlets were subcultured three times, after that shootlets number/explant, shootlets length, number of leaves, fresh and dry weights were calculated and determine chemical composition.

For chemical composition, in three replicates, shootlet resulting from different concentrations of cytokinins were cut into small pieces and served for pigments, indoles and phenol analysis. For pigments determination, the ethanolic extraction was submitted to procedures of Saric *et al.* (1976) to qualitatively determine the endogenous chlorophyll-a,-b and carotenoids. For total indoles determination, the method of Selim *et al.* (1978) was applied. For total soluble phenols determination, the procedure of Daniel and George (1972) was used.

### Effect of different media types (MS, WPM, B<sub>5</sub> or LS)

The shootlets produced from culture on MS-medium with different kinds of cytokinins with different concentration (the best kind and concentration 2 mg/l BA) were added to different types of media. The media were used MS (Murashige and Skoog, 1962), WPM (Lloyd and McCown, 1980), B<sub>5</sub> (Gamborg et al., 1968) and LS (Linsmaier and Skoog, 1965). The shootlet (twenty shootlets

in five replicate) were culture on each medium used for one month and subcultured for three times. After this period shootlets number, shootlets length (cm), number of leaves, fresh and dry weight were calculated.

#### Effect of salt strength of MS-medium, types of auxin and activated charcoal

The resulted shootlets were cultured on different salt strength of MS-medium (full, half or quarter) free hormones or provided with 1 mg/l IAA, NAA or IBA without or with 1 g/l activated charcoal. The shootlets (twenty shootlet in five replicate) were incubated for 45 days. After this period rooting percentage, roots number per shootlet and root length (cm) was recorded.

#### Acclimatization rooted shootlets

The rooted shootlets resulting from rooting treatments were transferred to plastic pots containing peat moss irrigated with solution of 0.2 Topsin-M70 fungicides and covered by transparent polyethylene bags. The acclimatized vitroplants were kept in acclimatized galss house for four weeks before transplanting out-of-door after that survival capacity was calculated.

#### Experimental design and data analysis

The lay-out of the experiments was designed in completely randomized design and the test of LSD was used for comparison among means according to Steel and Torrie (1980).

#### RESULTS AND DISCUSSION

# Effect of clorox concentrations and periods of disinfection on sterilization explant

The data in Table (1) indicated that the increasing clorox concentrations increased percentage of free contaminated explants and decreased survival percentage of explants. The highest percentage of free contamination explants (73.33%) was recorded by using 20% clorox and decreased the survival percentage to the lowest percentage (46.67%).

On the other hand, the data indicated that increasing the time of immersed explants increased percentage of free contaminated but decreased the survival percentage of explant. The highest percentage of free contaminated explant (57.5%) was recorded by immersio the explant for 15 min, but gave the lowest percentage of survival explants (38.75%).

The data on the interaction between the concentration of clorox and immersion time indicated that the best percentage of free contaminated and survival percentage (70.0 and 60.0%) were observed when explants immersed in 10 min with 15% clorox.

Our results on sterilization of explants are in line with those of Hosni et al., (2000) on Limonium sinnuatum "Citron Mountion: and Hussein (2002) on Aglaeonema plants.

Table (1): Effect of clorox concentrations and periods of disinfection on free contamination and survival percentage of *Cotoneaster horizontalis*, explants.

|            | Free contamination |           |           |           | Survival percentage |           |           |           |
|------------|--------------------|-----------|-----------|-----------|---------------------|-----------|-----------|-----------|
| Treatments | 5<br>min           | 10<br>min | 15<br>min | Mean<br>B | 5<br>min            | 10<br>min | 15<br>min | Mean<br>B |
| Clorox 5%  | 5.0                | 15.0      | 25.0      | 15.00     | 100.0               | 85.0      | 60.0      | 81.67     |
| Clorox 10% | 25.0               | 40.0      | 40.0      | 35.00     | 95.0                | 75.0      | 45.0      | 70.00     |
| Clorox 15% | 65.0               | 70.0      | 75.0      | 70.00     | 85.0                | 60.0      | 35.0      | 60.00     |
| Clorox 20% | 60.0               | 85.0      | 90.0      | 78.33     | 65.0                | 60.0      | 15.0      | 46,67     |
| Mean A     | 38.75              | 52.50     | 57.50     |           | 86.25               | 68.75     | 38.75     |           |
| LSD 5% A   | 9.88               |           |           | 8,15      |                     |           |           |           |
| В          | 11.41              |           |           | 9.41      |                     |           |           |           |
| A×B        | 19.77              |           |           | 16.92     |                     |           |           |           |

# Effect of different concentrations of cytokinins (BA, kin or 2iP) on shooting behaviour of *Cotoneaster horizontals* explant

The data in Table (2) indicated that significant fluctuations in the behaviour of micropropagation of explants in vitro (i.e. number of the shootlets/explant, length of shootlet (cm), number of leaves/shootlet, fresh and dry weights) were attributed to the effect of cytockinins (BA, kin and 2iP) treatments.

For the number of shootlets/explant, the addition of BA at 3 or 4 mg/l BA in MS-medium produced the highest number of shootlets/explants (6.40 or 5.95). While the control (free hormone), 1 mg/l BA and all concentration of kin or 2iP gave the lowest number of shootlets/ explant ranging from (1.) to 2.95).

Regarding to shootlet length, the longest shootlet 2.13 cm) was recorded for explants grown on MS-medium supplemented with 1 mg/l BA. But culture the explants on MS-medium containing 3 mg/l BA reduced the shootlet length to the shortest length (0.84 cm).

Concerning to number of leaves/shootlet, the highest number of leaves/shootlet (7.63) was recorded when adding 1 mg/l 2iP. Whereas, the lowest number of leaves/shootlet (3.96) was recorded for explants grown on MS-medium free hormone.

As for fresh and dry weights, culturing the shootlets on MS-medium supplemented with 3 mg/l BA produced the highest fresh and dry weights of shootlets (1.03 and 0.252 g, respectively). On the other hand, culture the shootlets on MS-medium with 2 mg/l kin gave the lowest fresh weight (0.24 g). While all treatments except 3 mg/l BA had no significant effect on dry weight which ranging from (0.032 to 0.092 g).

From the above results, it could be generally suggested that adding 3 mg/l BA increased shootlets number/explant, fresh and dry weights of shootlets. Shootlet length increased by adding 2 mg/l BA, while number of leaves increased by adding 1 mg/l 2iP to MS-medium. Approving results were reported by Singh et al. (1994) on chrysanthemum morifolium; Monier (1995) on five Cotoneaster genotype; Sharma et al. (2003) on Crataeva adansonii and Parasharami et al. (2003) on Pinus roxburghii

Table (2): Effect of different concentrations of cytokinins (BA, kin or 2iP) on shooting behaviour of *Cotoneaster horizontals* explant in vitro.

|            | Should be chartout of Coton-case, northwest Capitale we view. |                         |                  |                     |                |  |
|------------|---|-------------------------|------------------|---------------------|----------------|--|
| Treatments | Number of shootlet  | Length of shootlet (cm) | Number of leaves | Fresh<br>weight (g) | Dry weight (g) |  |
|            | <del></del>   | <del></del>             |                  |                     |                |  |
| Control    | 1.23  | 0.87                    | 3.96             | 0.27                | 0.032          |  |
| 1 mg/l BA  | 1.68  | 2.13                    | 5.89             | 0.71                | 0.092          |  |
| 2 mg/l BA  | 2.95  | 1.46                    | 5.24             | 0.85                | 0.078          |  |
| 3 mg/l BA  | 6.40  | 0.84                    | 4.90             | 1.03                | 0.252          |  |
| 4 mg/l BA  | 5.95  | 0.99                    | 5.82             | 0.95                | 0.082          |  |
| 1 mg/l kin | 1.20  | 1.03                    | 5.63             | 0.42                | 0.052          |  |
| 2 mg/l kin | 1.08  | 1.52                    | 5.19             | 0.24                | 0.036          |  |
| 3 mg/l kin | 1.20  | 1.50                    | 5.19             | 0.49                | 0.058          |  |
| 4 mg/l kin | 1.00  | 1.59                    | 5.85             | 0.41                | 0.058          |  |
| 1 mg/l 2iP | 1.00  | 1.71                    | 7.63             | 0.56                | 0.046          |  |
| 2 mg/l 2iP | 1.38  | 1,30                    | 4.86             | 0.55                | 0.066          |  |
| 3 mg/l 2iP | 1.50  | 1.56                    | 5.56             | 0.37                | 0.048          |  |
| 4 mg/l 2ip | 1.30  | 1.82                    | 6.33             | 0.54                | 0.052          |  |
| LSD (5%)   | 0.882   | 0.433                   | 1.025            | 0.267               | 0.1216         |  |

## Effect of different concentrations of cytokinins (BA, kin or 2iP) on chemical composition

The results in Table (3) indicated that the amounts of chlorophyll-a, -b, carotenoids, total indoles and total soluble phenols which were mined in the shootlets tissues were significantly affected by different concentrations of cytokinins (BA, kin or 2iP).

For chlorophyll-a, the data showed that using MS-medium with 4 mg/l BA produced the highest amount (287.2 mg/100g fw). While adding 4 mg/l kin, 1, 2, 3 or 4 mg/l 2iP to MS-medium reduced the amount of chlorophyll-a to the lowest amounts (85.56, 85.52, 83.90, 101.9 or 89.50 mg/100g fw, respectively).

As for chlorophyll-b, the highest amounts (45.96 mg/100g fw) was recorded by using MS-medium supplemented with 2 mg/l kin, while the lowest values (18.22 or 15.16 mg/100g fw) was recorded by using MS-medium supplemented with 1 or 4 mg/l 2iP.

Concerning carotenoids content, adding kin at concentration 2 mg/l increased the amount of carotenoids to the highest value (45.35 mg/100g fw). But using 3 or 4 mg/l kin or 1, 2, 3 or 4 mg/l 2iP decreased the amounts of

carotenoids to the lowest values (12.56, 10.72, 10.88, 8.28, 11.88 or 11.86 mg/100g fw, respectively).

Regarding the total indole, using MS-medium containing 1 mg/l kin increased the total indole to a maximum amount (187.9 mg/100g fw). While adding 3 mg/l 2iP decreased the content of total indole to minimum amount (18.32 mg/100g fw)

Relating to the total soluble phenols, MS-medium contain 3 mg/l 2iP produced the highest amount of total soluble phenols (195.0 mg/l00g fw). But culture shootlet on MS-medium free hormones (control) decreased the content of total soluble phenols to the minimum amount (112.5 mg/l00g fw).

In conclusion, the highest content of chlorophyll-a was recorded with MS-medium supplemented with 4 mg/l BA, while the highest amount of chlorophyll-b and carotenoids was recorded with MS-medium supplemented with 2 mg/l kin. Total indoles increased by adding 1 mg/l kin and total soluble phenols increased when using 3 mg/l 2iP. In this respect Tung (1997) on Cucumber seedling studied the effect of different concentrations of cytokinins BA, BA-ribosid (BAR), BA-3G, BA-7G, BA-9G, Zeatin (Z), Z-riboside, dihydrozeatin and 2iP on the formation capacity of chlorophyll-a,-b and carotenoids. He declared that the production of such pigments was negative related with the concentrations of the other cytokinins examined.

Table (3): Effect of different concentrations of cytokinins (BA kin or 2iP) on chemical composition (mg/100g fw) of Cotoneas er horizontals shootlets in vitro.

| Treatments | Chloro-<br>phyll –a | Chloro-<br>phyll -b | Carote-<br>noids | Total<br>indole | Total<br>soluble<br>phenols |
|------------|---------------------|---------------------|------------------|-----------------|-----------------------------|
| Control    | 199.80              | 33.64               | 29.77            | 132.80          | 112.50                      |
| 1 mg/l BA  | 220.10              | 37.93               | 26.67            | 181.50          | 147.60                      |
| 2 mg/l BA  | 167.40              | 27.20               | 20.82            | 175.30          | 182.90                      |
| 3 mg/l BA  | 227.80              | 37.13               | 30.69            | 165.00          | 119.80                      |
| 4 mg/l BA  | 287.20              | 33.47               | 27.23            | 181.50          | 140.40                      |
| 1 mg/l kin | 212.90              | 38.16               | 28.33            | 187.90          | 141.90                      |
| 2 mg/l kin | 245.50              | 45.96               | 45.35            | 172.00          | 177.40                      |
| 3 mg/l kin | 114.30              | 25.59               | 12.56            | 117.90          | 163.9                       |
| 4 mg/l kin | 85.56               | 23.31               | 10.72            | 26.07           | 120.40                      |
| 1 mg/l 2iP | 85.52               | 18.22               | 10.88            | 88.42           | 180.50                      |
| 2 mg/l 2iP | 83.90               | 23.43               | 8.28             | 58.80           | 181.50                      |
| 3 mg/l 2iP | 101.90              | 21.60               | 11.88            | 18.32           | 195.00                      |
| 4 mg/l 2ip | 89.50               | 15.16               | 11.86            | 52.71           | 158.40                      |
| LSD (5%)   | 53.19               | 12.36               | 13.29            | 53.07           | 49.23                       |

Effect of different media types (MS, WPM,  $B_5$  or LS) on shooting behaviour of Cotoneaster horizontals explants

The behaviour of shooting due to media type including the number of shootlets/explant, length of shootlet (cm), number of leaves/shootlet, fresh and dry weights (g) were illustrated in Table (4).

For number of shootlets/explant and number of leaves/shootlet the data indicated that all types of media (MS, WPM, B<sub>5</sub> or LS) had no significant effect on these characters.

Relating to the length of the formed shootlet, the data declared that the tallest shootlets was found by using MS or LS media, which gave (1.28 or 1.25 cm). But the shortest length (0.93 cm) was observed when culture the shootlet on WPM medium.

Concerning fresh weight of shootlets, culture the shootlet on MS-medium increased fresh weight of shootlet (1.36 g) compared to culture the shootlet on WPM, B<sub>5</sub> or LS, which decreased the fresh weight (0.63, 0.62 or 0.68 g, respectively).

Regarding to dry weight of shootlet, the shootlet culture on MS-medium gave the highest dry weight (0.31 g), while shootlet culture on LS-medium decreased dry weight (0.062 g) of shootlet.

A wide survey of the obtained data indicated that culture the shootlet on MS-medium increased the number and length of shootlets, the number of leaves, fresh and dry weights compared to culture the shootlet on WPM, B<sub>5</sub> or LS media. Approving results were reported by Monier (1995) on five Cotoneaster genotypes, Youssef (1996) on Melaleuca armillars, Ipekci et al. (2001) on Paulownia elongata and Brum et al. (2003) on Ficus carica.

Table (4): Effect of different media types (MS, WPM, B<sub>5</sub> or LS) on shooting behaviour of *Cotoneaster horizontals* explant in vitro.

| Treatments     | Number of shootlet | Length of shootlet (cm) | Number of leaves | Fresh<br>weight (g) | Dry<br>weight (g) |
|----------------|--------------------|-------------------------|------------------|---------------------|-------------------|
| MS             | 5.90               | 1.28                    | 6.20             | 1.36                | 0.310             |
| WPM            | 4.81               | 0.93                    | 5.66             | 0.63                | 0.088             |
| $\mathbf{B_5}$ | 5.43               | 1.07                    | 6.27             | 0.62                | 0.070             |
| LS             | 4.13               | 1.25                    | 6.13             | 0.68                | 0.062             |
| LSD (5%)       | NS                 | 0.279                   | NS               | 0.304               | 0.242             |

Effect of salt strength of MS-medium, types of auxins and activated charcoal on rooting behaviour of Cotoneaster horizontals shootlets in vitro.

It is quite clear from the data in Table (5) that the various treatments caused a significant influence on rooting percentage, while they had no significant effect on the number and the length of root.

For rooting percentage, full strength of MS-medium with 1 mg/l IAA produced the highest percentage of rooting shootlets (30.0%). On the other hand, the different strength of MS-medium (full, half or quarter) without auxine and without or with 1 g/l activated charcoal failed to form roots (0.0%).

Regarding to the root number/shootlet and root length (cm), the different strengths of MS-medium (full, half or quarter) without auxine or with 1 g/l IAA, NAA or IBA without or with 1 g/l activated charcoal had no significant effect on the roots number which ranged from (0.25 to 1.25 root/shootlet) and length of roots which ranged from (0.63 to 1.88 cm).

These findings go in line with those reviewed by Rajendra et al. (1998) on Artocarpus heterophyllus; Sakr et al. (1999) on Yucca elephantips; Hussein (2002) Aglaeonema plants and Sayed and Abou-Dahab. (2006) on Faucaria tuberculosa.

Table (5): Effect of strength of salt MS-medium, types of auxine and activated charcoal on rooting behaviour of Cotoneaster horizontals shootlets in vitro.

| norizoniais shootiets in vitro. |             |             |                |  |  |  |
|---------------------------------|-------------|-------------|----------------|--|--|--|
|                                 | Rooting (%) | Number of   | Length of root |  |  |  |
| <u>Treatments</u>               |             | root        | (cm)           |  |  |  |
| F. MS                           | 0.0         |             | Į <b></b> ]    |  |  |  |
| H. MS                           | 0.0         | <del></del> |                |  |  |  |
| Q. MS                           | 0.0         | <b></b>     | <b></b> ,      |  |  |  |
| F. MS+IAA                       | 30.0        | 1.25        | 1.88           |  |  |  |
| H. MS+IAA                       | 15.0        | 0.25        | 0.75           |  |  |  |
| Q. MS+IAA                       | 15.0        | 0.75        | 1.63           |  |  |  |
| F. MS+NAA                       | 20.0        | 0.25        | 0.63           |  |  |  |
| H. MS+NAA                       | 5.0         | 0,75        | 1.38           |  |  |  |
| Q. MS+NAA                       | 10.0        | 0.75        | 1.50           |  |  |  |
| F. MS+IBA                       | 20.0        | 0.75        | 0.63           |  |  |  |
| H. MS+IBA                       | 15.0        | 0.75        | 1.25           |  |  |  |
| Q. MS+IBA                       | 10.0        | 1.00        | 1.38           |  |  |  |
| F. MS+AC                        | 0.0         |             |                |  |  |  |
| H. MS+AC                        | 0.0         |             |                |  |  |  |
| Q. MS+AC                        | 0.0         |             | <b></b>        |  |  |  |
| F. MS+IAA+AC                    | 10.0        | 0.50        | 0.63           |  |  |  |
| H, MS+IAA+AC                    | 10.0        | 0.50        | 0.75           |  |  |  |
| Q. MS+IAA+AC                    | 10.0        | 1.25        | 1.50           |  |  |  |
| F. MS+NAA+AC                    | 15.0        | 0.50        | 1.13           |  |  |  |
| H. MS+NAA+AC                    | 5.0         | 0.25        | 0.75           |  |  |  |
| Q. MS+NAA+AC                    | 5.0         | 1.50        | 0.88           |  |  |  |
| F. MS+IBA+AC                    | 10.0        | 1.00        | 1.38           |  |  |  |
| H. MS+IBA+AC                    | 15.0        | 0.50        | 0.75           |  |  |  |
| Q. MS+IBA+AC                    | 10.0        | 1.00        | 1.38           |  |  |  |
| LSD (5%)                        | 18.49       | NS          | NS             |  |  |  |

F.: Full

H.: Half

Q.: Quarter

#### Acclimatization rooted shootlets

All rooted shootlets were acclimatized on peat moss. The survival of plantlet after one mother were calculated and gained 70% percentage

#### REFERENCES

- Baily, L. H. (1976): Hotus third. Aconcis Dictionary Plants Cultivated in the United States and Canada Mechmilan Publishing Co., Inc. New York.
- Blom-Barnhoorn, G. J. and Van Aartijk, J. (1985): The regeneration of plants free of LSV and TBV from infected *Lilium* bulb-scale explants in the presence of virazole. Acta Hort, 164: 163-168.
- Brum, G. R.; Pasqual, M.; Silva, A. B.; Chalfum, N. N. J.; Lopez, C. M. and Bernalte, M. J. (2003): Sucrose, culture media and their interactions during in vitro proliferation of "Roxa de Valinhos" (Ficus carica L.). Acta Hort., 605: 131-135.
- Daniel, H. D. and George, C. M. (1972): Peach seed dormancy in relation to endogenous inhibitors and applied growth substances. J. Amer. Soc. Hort. Sci., 97: 651-654.
- Gamborg, O. L.; Miller, R. A. and Ojima, K. (1986): Nutrient requirement of suspension cultures of soybean root cells. Exp. Cell Res., 50, 151.
- George, E. F. and Sherrington, P. D. (1994): Plant propagation by tissue culture. Handbook and Directory of Commercial Laboratories. Edington. Westbury Wiis, England.
- Hosni, A. M.; Hosni, Y. A. and Ibrahim, M. A. (2000): In vitro micropropagation of Limonium sinnuatum "Citron mountion" a hybrid static newly introduced in Egypt. Annals. Sci. Ain Shams Univ., Cairo, 45 (1) 327-339.
- Hussein, M. M. (2002): In vitro propagation of three species of Agaeolnema plants. Bull. Fac. Agric. Cairo Univ., 53: 465-488.
- Ipekci, Z.; Altinkut, A.; Kazan, K.; Bajrovic, K. and Gozukirmizi, N. (2001): High frequency plant regeneration from nodal explants of *Paulowina elongata*. Plant Biol., 3 (2): 113-115.
- Khalifa, S. F. and Loutify, M. H. (2006): Ornamental cultured plant collection. In the Occasion of the First International Conference on "Strategy of Botanic Gardens".
- Linsmaier, E. M. and Skoog, F. (1965): Organic growth factor requirements of tobacco tissue cultures. Physiol. Plant, 18: 100-127.
- Lloyd, G. and McCown, B. (1980): Commercially feasible micropropagation of mountain lourel, *Kalmia latifolia*, by using of shoot tip culture. Com. Proceed. Int. Plant Prop. Soc., 30: 421-427.
- Monier, C. (1995): Establishing micropropagation conditions for five Cotoneaster genotype. Plant Cell, Tissue and Organ Culture, 42 (3): 275-281.
- Murashige, T. and Skoog, F. (1962): A revised medium for rapid growth and bioassays with tobacco tissue culture. Physiol. Plant, 15: 473-497.
- Nitzsche, W. (1983): Germplasm preservation. In D.A. Evans, W.R. Sharp, P.V. Ammirato and Y.Yamada (eds), Handbook of Plant Cell Culture. VI.Collier Macmillan Publishers, London, pp. 782-805.
- Novak, F. J. and Petra, E. (1981): Tissue culture propagation of *Lilium* hybrids. Sci. Hot. 14: 191-199.

- Parasharami, V. A.; Poonauala, L. S. and Nadgauda, R. S. (2003): Bud break and plantlet regeneration in vitro from mature trees of *Pinus roxburghii* Sorg. Culture Sci., 84 (2): 203-208.
- Rajendra, S.; Tiwari, J. P. and Singh, R. (1998): *In vitro* clonal propagation of jackfruit (*Artocarpus heterophyllus* Lamk). Ind. J. Hort., 55 (3): 213-217.
- Rick, C. M. (1984): Plant germplasm resources. In P.A. Ammirato, D.A. Evans, W.R. Sharp and Y.Ymada (eds), Handbook of Plant Cell Culture V2. Collier Macmillan Publishers, London, pp. 9-37.
- Sakr, S. S.; El-Khateeb, M. A and Abd-El-Kareim, A. H. (1999): In vitro production of Yucca elephantips. Bull of Fac. of Agric. Univ. of Cairo, 50 (2): 265-282.
- Saric, M.R.K.; Curic, R.; Cupina, T.C. and Gerir, I. (1967): Chlorophyll determination. Univ. U. Novon. Sadu Praktikum Iz Fiziologiz Biljaka, Beograd. Haucua, Anjiga, pp.215.
- Sayed, S. Sawsan and Abou-Dahab, T. A. (2006): Propagation of *Faucaria tuberculosa* by *in vitro* culture. Arab. Biotech., 9 (2): 351-362.
- Selim, H. H.; Fayek, M. A. and Sweidan, A. M. (1978): Reproduction of Bircher cultivars by layering. Ann. Agric. Sci., Moshtohor, 9: 159-166.
- Sharma, P. K.; Purnima, T.; Sharma, K. C.; Kothari, S. L. and Tyagi, P. (2003): Clonal propagation of *Crataeva adansonii* (DC.) Prode.: a multipurpose tree. *In vitro* Cellular and Developmental Biology Plant, 39 (2): 156-160.
- Singh, K.; Arora, J. S. and Singh, K. (1994): In vitro multiplication of Chrysanthemum morifolium Ramata cv. Riot. J. Orn. Horti., 2 (1-2): 63-68.
- Stanilova, M. I.; Ilcheva, V. P. and Zagorska, N. A. (1994): Morphogentic potential and *in vitro* micropropagation of endangered plant species *Leucojum aestivum* L. and *Lilium rhodopaeum* Delip, Plant Cell Re J. 13: 451-453.
- Steel, R. G. D. and Torrie, J. H. (1980): Principle of statistics. A biometrical approach. Second Ed., Mc Graw-Hill Kogakusha, L. T. D.
- Takayama, S. and Misawa, M. (1982): Regulation of organ formation by cytokinins and auxin in *Lilium* bulbscales grown in vitro. Plant Cell Physiol., 23: 67-74.
- Takayama, S. and Misawa, M. (1983): A scheme for mass propaga ion of *Lilium in vitro*. Sci. Hot., 18: 353-362.
- Tung, M. S. (1997): Cytokinins efficiently enhanced pigment production in detached cotyledons of dark grown cucumber seedlings. J. Agric. And Fore., 46 (2): 85-91.
- Van Aartijk, J. and Blom-Barnhoorn, G. J. (1981): Growth regulator requirements for adventitious regeneration from *Lilium* bulb-scale in viti 2, in relation to duration of bulb storage and cultivar. Sci. Hort., 14: 261-168.
- Van Aartijk, J.; Blom-Barnhoorn, G. J. and Van der Linde, P. C. G. (1990): Lilies, In P.V. Ammirato, D.A. Evans, W.R. Sharp and Y.P.S. Bajaj (eds), Handbook of Plant Cell Culture V5. Collier Macmillan Publishers, London, pp. 553-576.
- Wickremesinhe, E. R. M., Holcomb, E. J. and Arteca, R. N. (1994): A practical methods for the production of flowering Easter lilies from callus cultures. Sci. Hort., 60: 143-152.

Youssef, E. M. A. (1996): *In vitro* microporopagation of tee tree (*Melaleuca armillars* Smith). Proceed. 4<sup>th</sup> Arab. Conf. Hort. Crops, Fac. Agric. Minia Univ., 66: 981-902.

### إكثار الكوتنيستر هوريزنتالس من خلال زراعة الأنسجة

### سوسن سامى سيد ، سهام جاد الكريم معهد بحوث البساتين – مركز البحوث الزراعية

أجرى هذا البحث بمعمل زراعة الانسجة وحفظ الاصول الوراثيــة بمعهــد بحوث البساتين– مركز البحوث الزراعية خـــلال الاعـــوام ٢٠٠٦و ٢٠٠٧ ويهــدف البحث الى عمل بروتوكول للكثار الدقيق لنبات الكوتنيستر وقد اظهرت النتائج:

بالنسبة لمرحلة التعقيم: فقد ادى استخدام تركيز ١٥% كلــوركس لمـــدة ١٠ دقيقة للحصول على احسن نسبة من الاجزاء النباتية الحية و الخالية من التلوث.

بالنسبة لمرحلة تكوين اللافرع فقد ادى اضافة ٣ ملليجرام/لتر BA الى زيادة عدد الفريعات والوزن الطازج والجاف للفريعات. وقد ادى اضافة ٢ ملليجرام/لتر BA الى زيادة طول الفريعات. بينما عدد الاوراق زاد بإضافة ١ ملليجرام/لتسر 2iP. أما بالنسبة لانواع البيئات المختلفة فان زراعة الفريعات على بيئة MS زاد من عدد وطول الفريعات كما زاد عدد الاوراق و الوزن الطازج والجاف للنبيتات.

بالنسبة للمحتوى الكيماوى فأن اعلى محتوى من الكلوروفيل  $\dashv$  تم تقديره فى بيئة بها ٤ ملليجرام/لتر BA بينما اعلى كمية من كلوروفيل - ب والكاروتين تسم تقديرها فى النبيتات المزروعة على بيئة MS بها ٢ ملليجرام/لتر أما الاندولات فقد زادت بأضافة ١ ملليجرام/لتر kin والفينولات الكليسة الذائبسة زادت عند اضسافة ٣ ملليجر ام/لتر 2iP.

بالنسبة لمرحلة التحذير فان أعلى نسبة لتكوين الجذور كان للفريعات المزروعة على بيئة MS كاملة التركيز مضاف إليها ١ ملليجرام/لتر IAA أما طول وعدد الجذور فلم يوجد بين المعاملات فروق معنوية. وقد وصلت نسبة نجاح الأقلمة إلى ٧٠% في النبيتات المتكون عليها جذور