Effect Of Chemotherapy And Ultra-Violet Irradiation On Embryonation And Infectivity of *Toxocara canis* Eggs By Using Murine Model

By

Ramadan, M.Y. and Nagwa Eid Ahmed

Dept. of Parasitol. Fac. Vet. Med. Benha Univ.

Abstract

The present study aimed to investigate the in vivo effect of albendazole, ivermectin and UV irradiation on subsequent embryonation and infectivity of Toxocara canis eggs. For this propose, nine naturally infected puppies with Toxocara canis of two months old from the same environmental condition were allocated into three equal groups in separate cages. GroupI was received albendazole orally at a dose of 100 mg/kg b.w.; groupII was s/c injected with ivermectin at a dose of 0.3mg/kg. Group III was kept without treatment as control. Faecal samples were collected 3 times daily for 7 days post-treatment and the expelled T. canis worms were collected. At 7th day posttreatment, necropsy of all puppies was carried out and the adult T. canis worms were collected from their intestine, coecum and colon. The collected female worms from puppies groups I and II were dissected for preparation of eggs for embryonation. Eggs from worms of the third group (control group) were divided into 3 groups; one group (subgroup III a) was exposed to UV irradiation for 30 minutes before incubation, while the other two subgroups (subgroup IIIb & subgroup IIIc) were incubated till emberyonation. After embryonation, (subgroup III b) was exposed to irradiation for 30 x minutes while the other group (subgroup IIIc) was kept without irradiation as a control, non treated and non irradiated group. Embryonation was evaluated micro-scopically at 1 20th day. The infectivity of embryonated *T. canis* eggs was tested by mouse. The present data showed a rapid expulsion of worms in ivermectin treated puppies (48-120 hr. posttreatment) compared to (48-160 hr post- treatment) in puppies treated by albendazole. The lowest percentage of embryonation (35%) was recorded in albendazole treated group followed by (42%) in irradiated fertilized eggs (subgroup IIIa) as 65% and 58% of eggs cultures respectively were arrested at one cell stage, irregular cell division and a typical blastomeres and

gastrulae. Normal embryonation was recorded in (subgroups III b, c) and ivermectin treated groups. Comparatively lower mortalities were recorded in mice group inoculated with irradiated eggs containing second larval stage (subgroup III b). Lower number of larvae was detected from the liver of mice groups inoculated with irradiated larvae. Stained tissue section slides revealed that 20- 25 cross section of encapsulated *T. canis* larvae in mice belonged to group IV at 80th day post inoculation compared to 2-3 cross section of *T. canis* larvae in mice belonged to other mice groups. It was concluded that albendazole and UV irradiation showed an ovicidal effect against *T. canis* eggs and consequently reduce the occurrence of infective eggs in the environment. UV irradiation of embryonated eggs resulted in lower infectivity and mortality in inoculated mice compared to other groups.

Key words : Toxocara canis, emberyonation, infectivity, treatment, irradiation

Introduction

Toxocara canis is a parasite of canine carnivores and probably causes human toxocariasis. The bitches gradually accumulates T. canis larvae from the soil and tissues of prey and leak the larvae into its puppies both during the latter part of pregnancy (parentally) and during the nursing period through mammary gland (30). After birth, these larvae mature 25-40 days post partum and the puppies then contaminate the environment with eggs as every puppy can be expected to shad T.canis eggs (7).

(29) reported a syndrome of persistent eosinophilia in 2 children by administration of embryonated eggs of *T. canis*. (22) established the role of *T. canis* as an agent of human disease. (2) mentioned that, man being an aberrant host where *T. canis* can not complete its life cycle in man but cause a spectrum of diseases ranging from no symptoms to eosinophilia covert toxocariasis visceral and ocular larva migrans.

Most of the anthelmintic drugs are generally directed against larval and adult stages of helminthes (10). However (27) reported that, anthelmintics may also express ovicidal activity as in benzimidazoles. (25, 21) supposed other methods of treatments by using irradiated larvae of *Dictyocaulus viviparous* and *Anclystoma caninum* and found that, most larvae were died in the tissues of the host after few weeks. However little is known about the influence of anthelmintic treatment or effect of irradiation on the development and infectivity of *T. canis* eggs.

So, the present study aimed to investigate the in vivo effect of albendazole and ivermectin as two anthelmintics or UV irradiation on subsequent embryonation and infectivity of T. canis eggs by using murine model.

Materials and Methods

- **Experiment I**: Effect of albendazole and ivermectin on survival and expulsion of *T. canis*:
- *Animals:* Nine naturally infected puppies with *T. canis* of two months old were divided into three equal groups in separate cages and treated as follow:
- Group I: Three puppies were orally treated with albendazole 2.5% (Product of Pharma Swede Egypt) at a dose of 100 mg/kg b.w according to (19).
- **Group II:** Three puppies were injected s/c with ivermectin 1%(Nasromectin, product of El- Nasr Pharmaceutical Chemical Co.) at a dose of 0.3 mg/kg according to (24).
- **Group III:** Three puppies were kept without treatment as a control group. All puppy groups were kept on clean source of water and food.

Parasitological examination:

Faecal samples of all puppies were collected 3 times daily for 7 days post-treatment for collection of the expelled *T. canis* worms according to (5). At 7th day post- treatment, necropsy of all puppies was carried out and the adult *T. canis* worms were collected from the intestine, coecum and colon. The collected worms from each puppy were kept separately in saline and the female worms were dissected for preparation of eggs for emberyonation according to (12, 26).

Eggs from worms of group III (control group) were divided into 3 groups; one group (subgroup III a) was exposed to UV irradiation for 30 minutes before incubation and the other two groups (subgroup IIIb & subgroup IIIc) were incubated at $26C^{\circ}$ in 2.5% potassium-dichromate till emberyonation. After embryonation, (subgroup III b) was exposed to irradiation for 30 minutes according to (1) while (subgroup IIIc) was kept without irradiation as control non treated non irradiated group. The percentage of embryonation and stage of development of eggs were evaluated microscopically at 20^{th} day of incubation according to (5). The embryonated eggs from all female groups were used experiment II.

Experiment II : Infectivity of embryonated *T.canis* eggs after UV virradiation and the in vivo treatment:

1- Mice inoculation

' The infectivity of embryonated T. canis eggs was tested by mouse inoculation as described by (11, 6, 9). Forty -five mice were used and allocated into five equal groups as in table (1).

Group	Number of mice	Type of embryonated egg inoculum			
Group I	9	embryonated eggs from pups treated with albendazole			
Group II	9	embryonated eggs from pups treated with ivermectin			
Group III	9	Irradiated fertilized eggs			
Group IV	9	Irradiated embryonated . eggs			
Group V	9	Non treated Eggs			

Table (1): Mice groups inoculated by different types of treated eggs

The mice were inoculated with approximately 1500 embryonated eggs/ mouse by stomach tube according to (13). Mortalities among inoculated mice groups were recorded. For counting migratory larvae in different tissues, three mice from each group were scarified at 4 and 8th day post-inoculation and their intestine, liver and lung were minced separately for detection of larvae according to (18).

2-Histopathological examination:

The remaining mice were scarified at 80^{th} day post inoculation and specimens were taken from their intestine and immediately fixed in 10% neutral buffered formalin. After proper fixation a thin paraffin section of about 5-7 microns were routinely prepared and stained with haematoxylin and eosin stain for microscopical examination according to (8). Each stained tissue section was examined with light microscope (100X) and 15 random fields from each group were counted according to (11).

Results

Experiment I:

Table (2) showed that ivermectin cause rapid expulsion of alive adult *T.canis* worms (48-120 hrs post treatment) in treated puppies, while albendazole led to delayed expulsion of died macerated worms from treated puppies after (48-160 hr p. t), The lowest percentage of embryonation (35%) was recorded in albendazole treated group as 65% of eggs cultures

	Mortality		Intestine		liver		lung	
	No	%	4 th day	8 th day	4 th day	8 th day	4 th day	8 th day
Group I	2	22.22	196.33	105.6	68.7	97.6	0.0	0.0
Group II	2	22.22	207.67	134.3	82.0	104.67	0.0	0.0
Group III	2 ,	22.22	192.3	123.0	78.5	88.75	0.0	0.0
Group IV	-	0.0	186.25	169.75	12.25	15.25	0.0	0.0
Group V	3	33.33	214.5	132.8	94.5	122.75	0.0	0.0

Table (3): Mortalities and mean number of detected *T. canis* larvae from internal organs of mice at 4th and 8th days post infection (pi)



Fig (1) :C.S in intestinal showed 20-25 cross section of encapsulated *T. canis* larvae in mice inoculated with irradiated embryonated eggs at 80^{th} day post inoculation (arrow). H&E (X100)



Fig (2): C.S in intestinal showed 2-3 cross section of encapsulated T. canis larvae in mice belonged to group I at 80th day post inoculation (arrow) H&E (X100)

Discussion

Although several studies were carried out on the biology, embryonation, and infectivity of *T. canis* in canine and paratenic hosts, the ovicidal effect of ivermectin or albendazole against its eggs have not been reported yet. Furthermore, most studies investigating ovicidal activity of anthelmintics are preformed the in vitro, while it had been suggested that in vitro effect of anthelmintics compounds may not necessarily reflect in the vivo activity (20). So the present study investigated the in vivo ovicidal effect of 2 different anthelmintic compounds ; ivermectin , albendazole as well as UV irradiation on subsequent embryonation and infectivity of *T. canis* eggs in mice

The present data showed that the two used drugs had good anthelmintic effect on alive adult *T.canis* worms. The alive worms were rapidly expelled alive from puppies treated with ivermectin. Such rapid expulsion may attributed to the paralytic effect of ivermectin on helminthes (5). Meanwhile albendazole led to delayed expulsion of died macerated worms from treated puppies.

The present study showed that albendazole had an ovicidal activity against T. canis eggs isolated from expelled females by inhibiting the subsequent egg development. Incubated eggs were largely arrested at one cell stage or irregular cell division followed by abnormal blastomeres and gastrulae and consequently low percentage of eggs reach complete development (eggs containing L2). In this respect, (15,16) reported that treatment with anthelmintic affect the eggs in the oviduct of the female worms that might impair subsequent egg development in vitro. (17) demonstrated the mode of action of albendazole and he reported that it inhibited tubulin polymerization in dividing cells. (5) recorded that albendazole had in vivo ovicidal activity against A. suum eggs that were isolated from deworming pigs including the arrestment of egg development at one cell stage followed by irregular cell division. On the other hand, the ovicidal effect of ivermectin was not clear as the incubated eggs were normally developed as the control group. These results were met with study results of (5) who declared that non benzimidazole compounds such as ivermectin have only shown larvicidal effect in vitro.

1

Exposure of *T canis* fertilized eggs to UV rays for 30 min. led to reduction in the number of embryonated eggs. Such reduction may be due to interference with the protein synthesis and the enzymes function of in UV irradiated eggs (23).

In the present study, the murine model had been used due to the similarity between the progression of T. canis in mouse and in human (28). The present study revealed that comparatively higher mortalities in all groups except in group IV (infected with irradiated embryonated eggs). This result was agreed with (4) that there was no mortality among mice infected by T.canis irradiated eggs. The low mortalities in mice belonged to group IV

may be attributed to the encapsulation of larvae in intestinal tissues and little number of larvae migrate from the intestine to liver and other tissues in such mice as mentioned by (3).

There were few differences between the infectivity of eggs of ivermectin, albendazole treated groups as well as between irradiated fertilized eggs and control groups. These results were agreed by (5) who mentioned that the small proportion of eggs that became embryonated followed albendazole treatment appears fully infective for mice.

The mean number of recovered larvae from intestine, liver and lung was comparatively low when compared to the inoculum. Similar results were obtained by (14). The mean numbers of recovered larvae from all mice groups after 8 days post infection were nearly similar, while there were reduction in its numbers specially in group IV. This may be due to the low ability of migration. This result was agreed with (3) who mentioned that irradiation of infective *T.canis* larvae reduced their migration.

Concerning the infectivity of UV irradiated embryonated *T. canis* eggs, the present study revealed that there was no mortalities among mice and showed comparatively low mean number of recovered larvae. The majority of hatched larvae did not migrate from the intestine and became encapsulated as shown in stained tissue section slides. This result was in line with that of (4) who recorded no mortality among mice infected with irradiated *T. canis* eggs and also was met with that of (1) who recorded 27% reduction in the total larval recovery in mice infected by UV irradiated eggs compared to control. This was attributed by (3) who mentioned that irradiation of infective *T. canis* larvae reduced their pathogenesity, inhibited their migration from liver and such larvae died faster than non-irradiated larvae for the first 20 days of infection.

177

It was concluded that:

- 1- Albendazole showed an ovicidal effect against T. canis eggs and consequently reduced the occurrence of infective eggs in the environment after treatment with a single dose.
- 2- Ivermectin resulted in a rapid expulsion of alive *T. canis* female worms without an ovicidal effect against its eggs.
- 3- UV irradiation of fertilized *T. canis* eggs reduced its embryonation but not its infectivity
- 4- There were few differences between the infectivity of eggs of ivermectin, albendazole treated, irradiated fertilized and control groups.
- 5- UV irradiation of embryonated eggs decreased infectivity and subsequently mortality among inoculated mice.

Further studies are needed to determine the antigenisty and immunization of the paratenic hosts and puppies against *T. canis* infection.

Acknowledgment: Great thanks to Prof. Dr Abdel- baset El – mashad Professor of Pathology for his help in carrying out histopathological section.

References

- 1-Abo-Shehada, M.N., Al Zubaidy, B.A. and Herbert, S.V. (1991): Acquired immunity to *Toxocara canis* infection in mice. Vet. Parasitol. 38(4): 289-298.
- 2-Arambulo, P.V. and Steele, J.H. (1976): Urban dogs in houston Texas, Parasitic infection and environmental health impact. Int. J. of Zoonoses. 3:114-144.
- 3- Barriga, O.O. (1988): A critical look at the importance, prevalence and control of Toxocariasis and the possibility of immunological control. Vet. Parasitol. 29: 195-234.
- 4- Barriga, O.O. and Myser, W.C.(1987): Effect of irradiation on the biology of the infective larvae of *T. canis* in mouse. J. Parasitol. 73(1):89-94.
- 5-Boes, J.; Eriksen, L. and Nansen, P. (1998): Embryenation and infectivity of Ascaris suum eggs isolated from worms expelled by pigs treated with albendazole, pyrantel pamoate, ivermecten or piprazine dihydrochloride. Vet. Parasitol. (75): 181-190.

- 6- Daniela S. Moreira and Gutemberg M. Rocha (2005): Toxocara canis: Impact of preweaning nutritional deprivation on the pathogenesis of pneumonia in the mouse Experimental Parasitology, Volume110, Issue 4, August 2005, Pages 349-352
- 7-Douglas, J.R and Baker, N. F. (1959) : The choronology of experimental intrauterine infection with *Toxocara canis* in dogs. J. Prarasitol.45:43-44.
- 8- Drury ,R. and Wallington, E. (1986) : Carleton ,s histological technique . 4th ed. Oxford Univ. press . NewYork , Toranto.
- 9- Elena, P., Sietze, B., Jan D., Manoj, F., Clare M. Hamilton and Joke van der Giessen (2007): Toxocara canis: Effect of inoculum size on pulmonary pathology and cytokine expression in BALB/c mice. Experimental Parasitology, Volume 115, Issue 1, January 2007, Pages 76-82
- 10- Ergerton, J.R. (1969): The ovicidal and larvicidal effect of thiabendazole on various helminth species. Tex. Rep. Biol. Med. 27:561.
- 11- Fan,C.K .; Lin,Y.H. and Du,W.Y. (2003) : infectivity and pathogenisity of 14 month cultured embryonated eggs of Toxocara canis in mice. Vet. Parasitol. April 18, 113 (2) : 154-155
- 12-Fayek, A,S. Nada, M.S. El-Gawady H.M.(1992): Toxocara canis eggs embryonation in vitro. Zag. Vet. J. (EGVMC) vol. 20 (2): 239-246.
- 13- Fok, E. and Kassai, T. (1998): Toxocara canis infection in the paratenic host. A study on the chemo susceptibility of the somatic larvae in the mice. Vet. Parasit. 74(2-4): 243-259.
- 14- Ghoneim, N.H., Reda, W.W. and Sabry, M.A.(2001): Toxocarsis and paratenic hosts. Vet. Med. J. Giza 49(2) 249-260.
- 15- Kirsch, R. (1978): In vitro and in vivo studies on the ovicidal activity of fenbendazole. Res. Vet. Sci. (25): 263-265.
- 16- Kirsch, R. And Schleich, H. (1982): Morphological changes in trichostrongyloid eggs ofter treatment with fenbendazole. Vet. Parasital. (11): 375-380.
- 17- Lacy, E. (1988): The role of the cytoskeletal protein tubulin in the mode of action and mechanism of drug resistance to benzimidazole carbamates. Int. J. Parasital. (18):885-936.
- 18- Lescano, C.Z.; queiroz, M.L. nd Chieffi, P.P. (2004): Larval recovery Toxocara canis in tissues of experimentally infected Rattus norvegicus. Mem. Inst oswaldo Cruzi Oct. 99 (6): 627-628.

١

ł

- 19- Lioyd, S. and Soulsby, E.J.L. (1983): prenatal and transmammary infection of *Toxocara* canis in dogs: effect of benzimidazole carbamate anthelmintic on various developmental stages of the parasite. J. of small animal practice. 24 (12): 763-768.
- 20- Lubega, G.W. and Prichard, R.K. (1991): Interaction of benzimidazole anthelmintic efficacy. Exp. Parasitol. (73): 203-213.
- 21- Miller, T.A. (1971): Vaccination against canine hook worm disease. Adv. Parasitol. 9: 153-183
- 22- Molk, R. (1983): Ocular Toxocariasis A review of the literature. Ann. Ophthalmal. 15: 216-231.
- 23- Okada, S. (1970): In "Radiation Biochemistry" vol. J. Academic press, New York.
- 24- Payne, P.A. and Ridley, R.K. (1999): Stratgic use of ivermectin during pregnancy to control *Toxocara canis* in greyhound puppies. Vet. Parasit. 85(4): 305-312.
- 25- Poynter, D. (1964): Commercial production of an irradiated vaccine against *Dictyocaulus vivipara*. IAEA.Tech. Rep. Ser. 30. Vienna, 17-23.
- 26- Ramadan, M. Y. (2004) : A trial to control *Toxocara canis* infecting puppies in Kalubyia . Governorate Egypt. Zag. Vet. J. (ISSN III0- 1458) Vol.32 No.1 : 120- 128.
- 27- Sinniah, B.; Singh, M.; Subramaniam, K. and Ranakrishnan, K. (1995): Deformity of *Trichuris trichura* eggs following treatment with albendazole. Inf. Dis. Clin. Prod. (4): 15-18. suum. Amer. J. Vet. Res. 47:869-873.
- 28- Smith, H.V. (1991): Immune evasion and immunopathology of *T. canis* infection. In parasitic Nematodes- antigens membranes and genes (ed. Kennedy, M.W) Taylor and Francis. Ltd London. Pp. 116-139.
- 29- Smith, M.H.D. and Beaver, P.C. (1953): Persistance and distribution of *Toxocara* larvae in the tissue of children and mice. Pediatrics. 12:491-497.
- 30- Sprant, J. F.A. (1952): On migratory behavior of larvae of various Ascaris in white mice.J. inf. Dis. 90: 165 176.

١

تأثير العلاج بالأدوية والأشعة فوق البنفسجية على النمو الجنيني والغدرة على العدوى لبويضات إسكارس الكلاب باستخدام نماذج من الفنران

قسم الطفيليات - كلية الطب البيطرى - جامعة بنها

استهدفت الدراسة معرفة تأثير عقارى البندازول و ايفرمكنين في داخل الجسم الحي وكذلك الأشعة فوق البنفسجية على النمو الجنيني والقدرة على العدوي لبويضات إسكارس الكلاب باستخدام نماذج من الفنران. تم اختيار ٩ كلاب صغيرة عمر شهرين من نفس الظروف البينية ومصابة إصابة طبيعية بالإسكارس وقسمت الى ثلاث مجموعات متساوية عولجت كالاتى: المجموعة الاولى تم تجريعها بعقار البندازول٥و٢% بمعدل ١٠٠ ملج/كجم مرة واحدة. و المجموعة الثانية تم حقنها بعقار أيفرمكتين تحت الجلد بمعدل ٣ملج/ كجم , المجموعة الثالثة تركت بدون علاج كمجموعة ضابطة وتم تجميع البراز ثلاث مرات يوميا ولمدةُ أسبوع بعد العلاج بهدف تجميع ديدان الاسكارس التي تم طردها من الجسم نتيجة تأثير العلاجات وفى اليوم السَّابع تم تشريح الكلاب وتجميع ديدان الاسكارس من امعانها وتم تحديد إناث ديدان الإسكارس وأخذ البويضات المخصبة من الأ جزاء العلوية من الأرحام للمجموعتين الاولى والثانية وتحضينها في ثاني كرومات البوتاسيوم°و٢% عند درجة حرار ٢٦٤ م لمدة ٢٠ يوم وتم حساب نسبة التكوين الجنيني ومراحله المختلفة أما بويضات المجموعة الثالثة فقسمت الى ثلاث مجموعات عرضت إحداهن(تحت المجموعة الثالثة أ) الى الأشعة فوق البنفسجية لمدة نصف ساعة وتحضينها مع المجموعتين(تحت المجموعة الثالثة ب وتحت المجموعة الثالثة ج) في ثاني كرومات البوتاسيوم°و ٢ عند درجة حرارة٢٦ م لمدة ٢٠ يوم وتم حساب نسبة التكوين الجنيني ومراحله المختلفة وبعد اكتمال النمو الجنيني تم تعريض (تحت المجموعة الثالثة ب) للاشعة فوق البنفسجية لمدة نصف ساعة أما (تحت المجموعة الثالثةج) فتركت بدون تعرض للأشعة وقد تم تجريع الخمس مجموعات المختلفة من البويضات لخمس لمجموعات متساوية من الفنران عن طريق أنبوبة اللي المعدي. أثبتت الدراسة أن عقار أيفرمكتين أدى إنى طرد سريع لديدان حية من إسكارس الكلاب وعند تحضينها كانت نسبة التكوين الجنيني بها مرتفعة (٧٦%) أما عقار البندازول٥و٢% فقد أدى إلى طرد ديدان ميئة من إسكارس الكلاب وعند تحضينها كانت نسبة التكوين الجنيني بها منخفضة(٣٥%) أما بالنسبة لتعريض البويضات المخصبة (تحت المجموعة الثالثة أ) للأشعة فوق البنفسجية لمدة نصف ساعة أظهرت النتائج ان نسبة التكوين الجنيني بها كانت منخفضة أيضا (٤٢%) .وقد أوضحت الدراسة أن القدرة على العدوى كانت مرتفعة كما كانت نسبة الوفيات مرتفعة أيضا في كل المجموعات ماعدا (تحت المجموعة الثالثة ب) إلتي تم تعريضها للأشعة فوق البنفسجية لمدة نصف ساعة بعد اكتمال نموها الجنيني. وقد بينت الدراسات الهستوباتُولوجية التي أجريت بعد ٨٠ يوما على الفنران بعد تجريعها بالأنواع المختلفة من البويضات ان البويضات الناضجة والتي تعرضت للأشعة فوق البنفسجية لمدة نصف ساعة لم يهاجر الكثير من يرقاتها الى الكبد ولكن تحوصلت في أنسجة الأمعاء .

الباحثّان محمد یوسف رمضان و نجوی عید احمد