GENETICAL AND ANATOMICAL ANALYSIS OF NORMAL AND ABNORMAL FLOWERS OF DATE PALM CV. 'BARHY' FROM OFFSHOOT AND TISSUE CULTURE DERIVED PLANTS

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Abstract

Random Amplified DNA Polymorphism (RAPD) analysis, between 6 normal flower producing offshoot derived and 6 abnormal, multiple carpel, flower producing tissue culture (TC) derived trees of cv. 'Barhy', was performed with the objective to resolve genetic variation if any at DNA level. DNA samples were extracted from pollinated and un-pollinated flowers from both sets of plants. Amplified RAPD products were clearly detected with 30 primers used in this experiment but only 3 gave a few polymorphic bands which shows low level of genetic variation among the offshoot and TC derived plants. Cluster analysis by the unweighted paired group method of arithmetic means (UPGMA) showed close genomic similarity among the 12 DNA samples with the range of 0.486-0.904 Nei and Li's coefficient in the similarity matrix. The average similarity among the 12 DNA samples was more than 50%. Floral abnormalities in TC derived plants were also studied microscopically. Abnormalities like more than three carpel development, abnormal ovule development and deformities of style and stigma were observed. The results show that the composition and the abnormalities of flowers in TC derived plants of cultivar 'Barhy' may be attributed to epigenetic changes that takes place at different stages of tissue culture and not due to major changes at DNA level.

Keywords: *Phoenix dactylifera*, RAPD, epigenetic, embryogenesis, fruit abnormality, Flower abnormality

INTRODUCTION

Date palm (*Phoenix dactylifera* L.) is the major fruit tree crop in the Middle East and North of Africa since the beginning of the history of mankind. Saudi Arabia is among the top most countries of the Middle East that produces and exports the best quality dates of the world (Zaid and *de We*t, 1999). Here date palm is of great socioeconomic importance and at least 450 different cultivars are alone found in this country (Bashah, 1996). Traditionally date palm cultivars are propagated through offshoots but limited number bars their distribution. With the advent of tissue culture techniques in date palm massive expansion of elite cultivars was made possible but commercially TC propagated plants were introduced only in the last decade.

Propagation either through offshoots or TC techniques like somatic embryogenesis (Al-Khalifah et al., 2004; Al-Khayri, 2003; Tisserat, 1982, 1979; Zaid and *de Wet*, 1999; Bhansali et al., 1988) must result in true-to-type plants. However, off-types that are not identical to their progenitor are also produced due to somaclonal variation caused by genetic and or epigenetic changes during TC process (Kunert et al., 2003, 2002; Kaeppler et al., 2000; Sala et al., 1999; Cullis et al., 1999). In TC derived date palm many off-type phenotypes are detected that include stunted or severely retarded growth, leaves with wide leaflets, variegated leaf, malformation of inflorescence, abnormal flowers and low levels of fruit setting (Gurevich et al., 2005; Cohen, et al., 2004; Djerbi, 2000; McCubbin et al., 2000). Some of these abnormalities in many trees become normal after 8-10 years of plantation and are therefore believed to be due to epigenetic changes (Cohen et al., 2004). Epigenetic changes are expressed under stress conditions possibly due to DNA methylation, DNA amplification and or activation of transposable elements (Kaeppler et al., 2000; Brar and Jain, 1998; Hirochika et al., 1996; Brettell and Dennis, 1991).

Abnormal flowers and low levels of normal fruit setting, especially in TC plants of cultivar Barhy, are common in Saudi Arabia. Large numbers of tissue cultured plants introduced in the country in and around 1995 are producing an average of more than 60% abnormal flowers and fruitlets per plant causing low levels of fruit set which is of a great economic loss to the farmers (Kunert et al., 2003). A plausible reason of low levels of fruit setting may be inadequate pollination due to abnormalities in the florwers (carpels, style and stigma). As a consequence all the three unfertilized carpels develop into abnormal and small fruitlets (local name *Shees*). In normal date flowers, after pollination, only a single carpel develops into a fruit and the other two carpels degenerate. More than three carpelled florets and fruitlets (4-7 carpels) are also observed (Gurevich, et al., 2005; Cohen et al., 2004; Al-Wasel, 2000, 2001; Djerbi, 2000; Reuveni, 1986). It is possible that the supernumerary carpels other than the main three carpels are staminodial primordia transformed into carpel-like structures (Cohen et al., 2004). Almost the same abnormalities have been observed in flowers of TC derived oil palms (Corley et al., 1986)

Molecular genetical analysis of TC plants showing abnormal phenotypes was performed by using RAPD (Saker et al., 2000) and isozymes (Saker et al., 2000; Azeqour et al., 2002). Both techniques detected plantlets that were showing difference in the traits at TC stage but were unable to resolve differences in the abnormal traits that appeared later at mature stage of the plant growth. Gurevich, et al., (2005) have recently performed AFLP analysis of both normal and off type (low fruit setting) TC

derived plants of cv. Barhy but were unable to detect any AFLP marker linked to abnormal flowering and low level of fruit setting traits.

In the present study RAPD analysis, between normal flower producing offshoot derived and abnormal, multiple carpel, flower producing TC derived trees of cv. 'Barhy', was performed with the objective to resolve genetic variation if any at DNA level. Floral abnormalities in TC derived plants were also studied microscopically.

MATERIALS AND METHODS

Plant material

Fresh flowers from 12 different plants; 6 normal flower producing offshoot plants and 6 multiple carpel flower (*shees*) producing TC derived plants, were collected from an orchard near Al-Kharj area of Saudi Arabia (Table 1). Flower samples were taken ten days before and after the pollination from both true-to-type offshoots and off-type TC plants.

Total genomic DNA extraction

Total genomic DNA was extracted from 10 fresh flowers of each of the 12 samples. The whole flowers were first ground into a fine powder in liquid nitrogen by usinf pestle and mortar and then by following the steps of the protocol (Dellaporta et al., 1983) pure and highly intact DNA was extracted. The quality and quantity of the DNA was determined by using a fluorometer (Hoefer DyNA Quant 200; Pharmacia Biotech). The integrity of the DNA was also determined by agarose minigel electrophoresis.

Primers and PCR amplification

A total of 30 RAPD primers of A, B, C, and D series (OPERO Tech., CA, USA) were used for polymerase chain reaction (PCR) amplification of the 12 different DNA templates to construct DNA fingerprinting profiles. PCR amplification was performed as described by Al-Khalifah and Askari, 2003. The RAPD products of each primer was separated by electrophoresis according to their molecular weight in 1.4% agarose gels submerged in 1 \times TBE buffer and then stained with ethidium bromide (10 μ g/ml) solution for 20 minutes. The DNA were visualized on UV-transilluminator and documented with Gel Documentation System (Bio Rad). The molecular weights of the amplified fragments were estimated by running Kilo Base DNA marker (Amersham Pharmacia Biotech.) in gel as standard size marker.

Genetic analysis

The amplification profiles of the 12 different 'Barhy' samples were compared with each other by using software 'Diversity Data Base' (Bio Rad). The data was applied to estimate the genomic similarity among the plants on the basis of shared

amplified fragments (Nei, 1978; Nei and Li, 1979). Cluster analysis by the unweighted paired group of arithmetic means (UPGMA) was also performed and a dandrogram was constructed by Dice Coefficient Method with the help of the software 'Diversity Data Base'.

Morphology and microscopy of flowers

Ten to twenty fresh flowers each from six different offshoots and six different TC plants were fixed in FAA (10% formaldehyde, 5% acetic acid, 50% ethanol). The flowers were collected ten days before and 40 days after the pollination. Dehydration and embedding of fixed materials were done in paraffin wax. Thin transverse and longitudinal sections of flower buds were cut at 5 µm thickness on Reichert-Jung rotary microtome and permanent slides were prepared following Johansen (1940). The sections of flowers were examined under a compound microscope and images were documented with a digital camera. The shape and structure of carpels and stigmas were studied to detect abnormality.

RESULTS AND DISCUSSION

Genetical analysis of normal and abnormal fruit bearing plants

RAPD analysis was performed on the 30 RAPD profiles of the 12 DNA samples of 'Barhy' that include 6 normal fruit bearing offshoot derived plants and 6 abnormal, multiple carpel, fruit bearing TC derived plants with the aim to assess genetic variation if any exist among them. The 30 primers used in this experiment were prescreened and selected for date palm DNA fingerprinting in our laboratory (Askari et al., 2003). Amplified RAPD products were clearly detected with all the 30 primers but only 3 produced reproducible and distinguishable polymorphic bands which indicate low level of genetic variation among the offshoot and TC derived plants (Fig. 1a & b). The number of polymorphic bands per primer varied from 3 to 6, with a mean of 3 major bands per primer.

The pair-wise genetic similarity was estimated for the 12 samples of DNA on the basis of Nei and Li's (1979) similarity coefficients. A similarity matrix was also constructed (Table 2). Cluster analysis by the unweighted-paired group of arithmetic means (UPGMA) method showed close genomic similarity among the 12 DNA samples with the range of 0.486-0.904 Nei and Li's coefficients in the similarity matrix. A dangrogram by using Dice coefficient method for the 12 genotypes of Barhy by using 'Diversity Data Base software' (Bio Rad.) is presented in Fig. 2). The average similarity among the 12 samples of DNA was more than 50%, suggesting close genomic similarities between the samples. Maximum similarity was observed between offshoot derived plant (N 15/4 C) with normal unpollinated flowers and TC derived plant (P-

R2A2G1 6) with multiple carpel pollinated flowers (0.90). Offshoot derived plant (N 14/1 C) with normal unpollinated flowers showed 88% genomic similarity with them which is the second highest similarity value. Offshoot derived plant (N 14/3 O) with normal pollinated flowers and TC derived plant (P-R2A1G1) with multiple carpel pollinated flowers are more related genomically (0.86) while TC derived plant (C-R2A1G1) with multiple carpel unpollinated flowers is 76% genomically similar with them. Similarly offshoot derived plant (N 14/1 O) with normal pollinated flowers and TC derived plant (P-R2A2G1 3) with multiple carpel pollinated flowers are also genomically similar with similarity matrix value 0.81. TC derived plant with multiple carpel unpollinated flowers in general showed a minimum degree of similarity with all the other samples ranging between 0.48-0.72.

The results of genetic variations suggest that RAPD analysis could be efficiently used for such assessments. Although low level of DNA polymorphism was observed, some genetic variation was detected between offshoot and TC derived plants. The present RAPD data so far generated suggest narrow genetic diversity among the offshoot derived normal flowering plants and TC derived abnormal flowering plants indicating that most of the plants of cv. 'Barhy' are true-to-type and have nearly the same genetic makeup. Saker et al. (2000) have also detected low level of genetic variation in young TC derived plantlets (4% of 70 analyzed plantlets) by using RAPD markers. Only those plantlets that were phenotypically different at TC level were detected as genetically variant. Gurevich et al. (2005) recently applied AFLP on normal offshoots and abnormal flower and fruit bearing TC derived plants of cv. 'Barhy' detecting low level of genetic variation among the plants but were unable to. link any AFLP marker to this abnormal flower and fruit setting character. We can therefore suggest that the abnormal flower and fruit bearing trait in TC derived plants of cv. 'Barhy' are due to the epigenetic changes that occur during the TC stages, and not due to any major genomic changes. Most of the abnormal fruit bearing TC plants become normal after 8-10 years of flowering but in some plants increase in the percentage of abnormal fruits also occur indicating epigenetic effects that are expressed under stress conditions during tissue culture. The stress conditions may cause DNA methylation, DNA amplification and or activation of transposable elements in the plants (Kaeppler et al., 2000).

In our study limited number of plant DNA samples and RAPD primers were used. It could be one of the reasons for detecting low level of genetic variation and not detecting any linked RAPD marker to this particular phenotype. RAPD markers should be of high value for assessing the genetic variance in date palm. It is therefore

desirable to increase the size of the population and also the number of RAPD primers to better assess the genetic diversity and detect linked markers to this phenotype.

Morphology and microscopy of flowers

Flowers and fruits in plants propagated from offshoots and TC originated orchards were observed (Fig. 3a-e). Low fruit setting has close relationship with structural floral abnormalities in TC derived plants such as abnormal development of carpel, stigma and ovule (Fig. 3c & e). A longer twisted stigma attached with narrow carpel head rather than clear joint between stigma and carpel was observed (Fig. 3c). Because of these abnormalities pollen tube looses its path that leads to failure of fertilization. Same results were also found by different workers during their studies (Al-Wasel, 2000; Djerbi, 2000; Cohen et al., 2004). Our observation revealed that many TC derived plants have major fruit setting problems. Beside normal single fruits many abnormalities were recorded such as single carpel parthenocarpic fruits, parthenocarpic fruits with two or three carpels originated from tri-carpellary flowers (Fig. 3a & d) and abnormal fruit lets with additional supernumerary carpels (Fig. 3a). A close correlation between structural floral abnormalities, altered pollen tube growth and low level of fruit setting was also reported by Cohen, et al. (2004).

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Table 1. List of 12 plants originated from offshoot and or tissue culture that were used in this study.

Accession #	Mode of cultivation	Phenotype
N-14/1 (O)	Offshoot	Normal
N-14/3 (O)	Offshoot	Normal
N-15/4 (O)	Offshoot	Normal
N-14/1 (C)	Offshoot	Normal
N-14/3 (C)	Offshoot	Normal
N-15/4 (C)	Offshoot	Normal
P-R2A1G1 (O) (1)	Tissue culture	Multiple carpel flowers
P-R2A1G1 (O) (3)	Tissue culture	Multiple carpel flowers
P-R2A2G1 (O) (6)	Tissue culture	Multiple carpel flowers
C-R2A2G1 (C) (1)	Tissue culture	Multiple carpel flowers
C-R2A2G1 (C) (3)	Tissue culture	Multiple carpel flowers
C-R2A2G1 (C) (6)	Tissue culture	Multiple carpel flowers

Note: N= normal flower, O=opern flower, C= closed flower

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		1	2	3	4	5	6	7	8	9	10	11	12	
14/3 (C)	1	100.0												
C-R2A2G1 (6)(C)	2	81.9	100.0											
N 15/4	3	79.5	77.5	100.0										
N 15/4 (C)	4	85.7	81.0	78.4	100.0									
N-14/1 (C)	5	81.1	71.6	76.1	88.0	100.0								
N-14/1 (O)	6	52. 9	61.3	55.4	55.1	51.5	100.0							
N-14/3 _. (O)	7	63.6	63.0	60.3	68.7	56.3	75.9	100.0						
P-R2A1G1 (O)	8	62.3	52.9	58.6	67.7	57.6	64.2	86.3	100.0					
P-R2A1G1 (O) (3)	9	58.7	70.7	52.8	65.8	57.5	80.6	80.0	70.0	100.0				
P-R2A2G1 (6)	10	83.3	75.9	72.5	90.4	88.6	50.0	61.3	66.7	59.2	100.0			
R2A1G1 (C) (1)	11	62.3	52.9	51.7	58.1	54.2	79.2	74.5	78.3	70.0	59.6	100.0		
R2A1G1 (C) (3)	12	60.3	65.0	48.6	59.5	50.7	70.8	69.8	58.6	72.2	52.2	62.1	100.0	

Table 2. Similarity matrix for Nei and Li's coefficients of 6 normal flower producing offshoots and 6 multiple carpel flower (shees) producing TC derived genotypes of cultivar 'Barhy' obtained from RAPD markers.

a) M 1 2 3 4 5 6 7 8 9 10 11 12

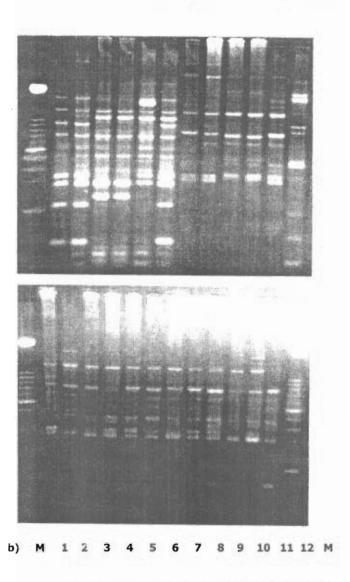


Figure 1. RAPD profiles of 6 normal flower producing offshoot plants and 6 multiple carpel flower (*shees*) producing TC derived plants of cultivar 'Barhy' using OPA7 (a) and OPB16 (b) primers. *Lanes: M* Molecular weight markers. *1,* 2 ,3, 4, 5 & 6 multiple carpel flower (*shees*) producing TC derived plants. 7, 8, 9, 10, 11 & 12 normal flower producing offshoot plants.

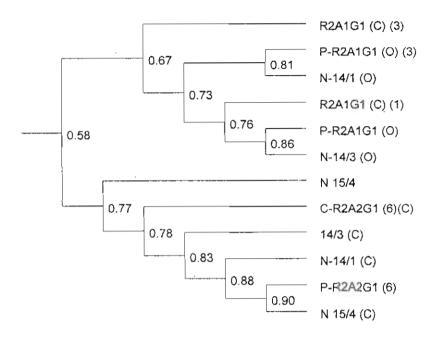
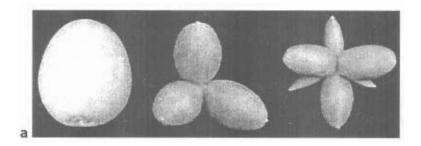


Figure 2. A dendrogram of phylogenetic relationships among 6 normal flower producing offshoots and 6 multiple carpel flower (*shees*) producing TC derived genotypes of cultivar 'Barhy' based on Nei and Li's similarity coefficients generated from 30 RAPD profiles.



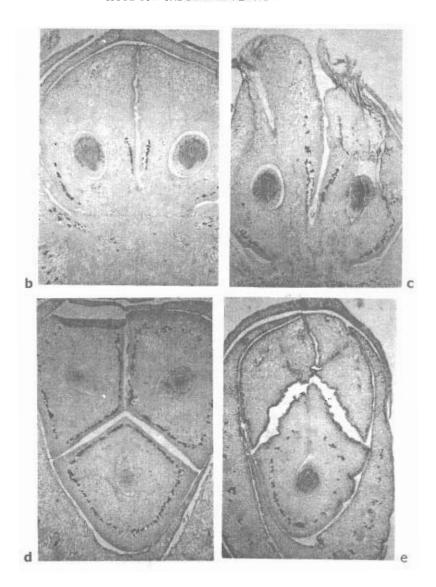


Figure 3. Fruit morphology and microphotographs of date palm flowers from offshoot and TC derived plants. **a.** A comparison of the normal and multi-carpel fruits. **b.** Longitudinal section (LS) of closed flower of offshoot derived plant showing normal development of carpel and ovule. **c.** LS of open flower of TC plant showing abnormal stigma and style in both carpel and abnormal ovule in only one carpel. **d.** Transverse section (TS) of closed flower of TC plant showing development of all the three carpels and ovules that led to shees formation. **e.** TS of closed flower of TC plant showing abnormal development of two carpels without any ovule.

REFERENCES

- Al-Khalifah, N.S. and E. Askari. 2003. Molecular phylogeny of date palm (*Phoenix dactylifera* L.) cultivars from Saudi Arabia by DNA fingerprinting. Theor. Appl. Genet. 107: 1266-1270.
- Al-Khalifah, N.S., F.A. Khan, E. Askari and S. Hadi. 2004. In Vitro culture and genetic analysis of male and female Date Palm (*Phoenix dactylifera* L.). 5th IVCHB Symp. I on In Vitro culture and Horticulture Breeding: Biotechnology, as Theory and Practice in Horticulture. Debercen, Hungry 12-17 Sep. p-55.
- 3. Al-Khayri, J.M. 2003. In vitro germination of somatic embryos in date palm: Effect of auxin concentration and strength of MS salts. Current Science. 84 (5): 680-683.
- 4. Al-Wasel, A.S. 2000. Vegetative and fruiting comparison of tissue culture derived and conventionally propagated date palm (*Phoenix dactylifera* L.) cv. Barhi trees. In Vitro Biol. Cellular Dev. Biol. 36: P-1010.
- Al-Wasel, A.S. 2001. Field performance of somaclonal variants of tissue culturederived date palm (*Phoenix dactylifera* L.). Plant Tissue Cult. 11 (2): 97-105.
- Askari, E., N.S. Al-Khalifah, T. Ohmura, Y.S. Al-Hafidh, F.A. Khan, A. Al-Hindi and R. Okawara. 2003. Molecular phylogeny of seven date palm (*Phoenix dactylifera* L.) cultivars by DNA fingerprinting. Pak. J. Bot. 35 (3): 323-330.
- Azeqour, M., K. Majourhat and M. Baaziz. 2002. Morphological variations and isozyme polymorphism of date palm clones from in vitro culture acclimatized and established on soil in South Morocco. Euphytica. 123: 57-66.
- 8. Bashah, M.A. 1996. Dates varieties in the Kingdom of Saudi Arabia. p 1225-1319. In: Guidance booklet, palms and dates. King Abdulaziz University press, Riyadh.
- 9. Bhansali, R., R. Kaul and H. Dass. 1988. Mass cloning of date palm plantlets through repetitive somatic embryogenesis. J. Plant Anat. Morphol. 5: 73-79.
- Brar, D.S. and S.M. Jain. 1998. Somaclonal variation: mechanisms and applications in crop improvement. p. 17-37. In: S.M. Jain, D.S. Brar, and B.S. Ahloowalia (eds.), Somaclonal variation and induced mutations in crop improvement. Kluwer Acad. Pub. Boston, USA.
- 11. Brettell, R.I.S. and E.S. Dennis. 1991. Reactivation of a silent Ac following tissue culture is associated with heritable alterations in its methylation pattern. Mol. Gen. Genet. 229: 365-372.

- Cohen, Y., R. Korchinsky and E. Tripler. 2004. Flower abnormalities cause abnormal fruit setting in tissue culture-propagated date palm (*Phoenix dactylifera* L.). J. Hort. Sci. & Biotech. 79(6): 1007-1013.
- Corley, R.H.V., C.H. Lee, I.H. Law and C.Y. Wong. 1986. Abnormal flower development in oil palm clones. Planter KL, 62: 23-240.
- 14. Corniquel, B. and L. Mercier. 1994. Date palms (*Phoenix dactylifera* L.) cultivar identification by RFLP and RAPD. Plant Sci. 101: 163-172.
- Cullis, C., S. Rademan and K.J. Kunert. 1999. Method for finding genetic markers of somaclonal variation. International publication number WO 99/53100.
- Dellaporta, S.L., J. Wood and J.B. Hicks. 1983. A plant DNA minipreparation: version II. Plant Mol. Biol. Rep. 1: 19-21.
- Djerbi, M. 2000. Abnormal fruiting of the date palm derived from tissue culture.
 Proc. Date Palm Intl. Symp. Windhoek, Namibia. P-73.
- Gurevich, V., U. Lavi and Y. Cohen. 2005. Genetic variation in date palms propagated from offshoots and tissue culture. J. Amer. Soc. Hort. Sci. 130(1): 46-53.
- Hirochika, H., K. Sugimoto, Y. Otsuki, H. Tsugawa and M. Kanda. 1996.
 Retrotransposons of rice involved in mutations induced by tissue culture. Proc. Natl. Acad. Sci. USA. 93: 7783-7788.
- 20. Johansen, D.A. 1940. Plant Microtechnique. McGraw Hill, New York.
- 21. Kaeppler, S.M., H.F. Kaeppler and Y. Rhee. 2000. Epigenetic aspects of somaconal variation in plants. Plant Mol. Biol. 43: 179-188.
- 22. Kunert, K.J., M. Baaziz, and C.A. Cullis. 2003. Techniques for determination of true-to-type date palm (*Phoenix dactylifera* L.) plants: A literature review. Emirates J. Agric. Sci. 15 (1):1-16.
- 23. Kunert, K.J., J. Vorster, C. Bester and C.A. Cullis. 2002. DNA microchip technology in the plant tissue culture industry. In: K. Rajasekaran, T.J. Jacks and J.W. Finley (eds.), Crop Biotechnology, ACS Symp. series 829. ISBN 0-8412-3766-2. American Chemical Society, Washington DC. p. 86-96.
- 24. McCubbin, M.J., J. Van Staden and A. Zaid. 2000. A South African survey conducted for off-types of date palms produced using somatic embryognesis. Proc. Date Palm Intl. Symp., Windhoek, Namibia. P-68-72.

- 25. Nei, M. 1978. Estimation of average heterozygosity and genetic distance from a small number of individuals. Genetics. 89: 583-590.
- 26. Nei, M. and W. Li. 1979. Mathematical model for studying genetic variation in terms of restriction endonucleases. Proc. Natl. Acad. Sci. USA. 76: 5269-5273.
- 27. Reuveni, O. 1986. Date palm pollen germination and tube elongation on pistillate flowers cultured at different temperatures. Acta Hort. 175: 91-95.
- 28. Saker, M.M., S.A. Bekheet, H.S. Taha, A.S. Fahmy and H.A. Moursy. 2000. Detection of somaclonal variations in tissue culture-derived date palm plants using isoenzyme analysis and RAPD fingerprints. Biologia Plantarum. 43 (3): 347-351.
- 29. Sala, F., A. Arencibia, S. Castiglione, P. Christou, Y. Zheng, and Y. Han. 1999. Molecular and field analysis of somaclonal variation in transgenic plants. p. 259-262. In: A. Altman, M. Ziv, and S. Izhar (eds.), Plant Biotechnology and In Vitro Biology in the 21st Century. Kluwer Acad. press. The Netherlands.
- 30. Tisserat, B. 1979. Tissue culture of the date palm. J. Heridity. 70: 221-222.
- 31. Tisserat, B. 1982. Factors involved in the production of plantlets from date palm tissue cultures. Euphytica. 31: 201-214.
- 32. Zaid, A., P.F. *de Wet.* 1999. Date palm propagation. p. 74-106. In: Zaid, A. (ed.) Date palm cultivation. FAO, Rome.
- 33. Zaid, A., P.F. *de Wet.* 1999. Origin, geographical distribution and nutritional values of date palm. p. 29-44. In: Zaid, A. (ed.) Date palm cultivation. FAO, Rome.

التحليل الجيني والتشريحي للزهور الطبيعية والشاذة في نخيل البلح المأخوذة من البراعم ونباتات ناتجة من زراعة الأنسجة

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التحليل العشوائي للحمض النووي دى ان اية لستة أزهار طبيعية وأخرى غير طبيعية تنتج براعم ومتعددة الكربلاء لأشجار ناتجة من زراعة الأنسجة للصنف البرحى، قد استخدمت لتحليل الاختلاف الوراثي في أي مستوى لل دى ان اية. عينات دي إن أي استخلصت من الزهور المُلَقَّحة والغير المُلَقَّحة من كلتا مجموعات النباتات.

أظهرت النَتائِجَ بأنَ التركيبَ وحالاتَ شذوذ الزهورِ في حالة زراعة الأنسجة قَدْ يُنْسَبانِ إلى epigenetic الذي يَحْدثانِ في المراحلِ المختلفةِ مِنْ زراعة الأنسجة وليس بسبب تغييراتِ رئيسيةِ في مستوى دي إن أي.