Suppression of Bacterial Soft Rot Disease of Potato A.A. Hajhamed*; Wafaa M. Abd El-Sayed*; A. Abou El-Yazied** and N.Y. Abd El-Ghaffar*

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acterial soft rot disease of potato caused by Erwinia carotovora Bsubsp. carotovora can cause great losses to potato yield. especially during storage. This study aimed to management of the disease using salicylic acid and BION as resistance inducing factors. potassium sulfate, ammonium phosphate and calcium chloride as salt compounds and isolates of Pseudomonas fluorescens (Pf1 and Pf5), P. aeruginosa (Pal), Serratia marcescens (Sml and Sm2) and Bacillus subtilis (Bs3) as bioagents, under artificial inoculation condition. All tested agents decreased the disease compared with the control. Disease severity was completely reduced when salicylic acid was applied at 0.9 mM, before or at the same time or after inoculation with the pathogen. When potassium sulfate and calcium chloride were applied at 0.5 g/l, at the same time of inoculation by the pathogen and when P. fluorescens (Pf5), B. subtilis (Bs3) and S. marcescens (Sm2) were applied after or before or at the same time of inoculation with the pathogen, respectively. Efficiency of the inducer agent and salt compounds were increased against the disease by increasing their rates. Salicylic acid, calcium chloride and potassium sulfate were the most effective against the disease compared with BION and ammonium phosphate, Isolates of P. fluorescens (Pf1), S. marcescens (Sm1) and P. aeruginosa (Pa1) were moderately effective against the disease when they were applied before, after and at the same time of inoculation with the pathogen, respectively.

Keywords: Bioagents, *Erwinia carotovora*, resistance-inducing factors, potato, salt compounds, soft rot and *Solanum tuberosum*.

Bacterial soft rot, caused by *Erwinia carotovora* subsp. *carotovora* (Van Hall) Dye, is one of the most important and widespread bacterial diseases of a wide variety plants either in the field or during storage. Losses due to bacterial soft rot have been seriously complained by potato growers in Egypt. Currently, the control of this pathogen relies on (1) Seed technology, *i.e.* tuber pasteurization, sanitation and use of certified seeds, (2) Application of chemical pesticides in the field and (3) The use of potato cultivars resistant to *E. carotovora* (Cronin *et al.*, 1997).

A number of compounds that do not have direct antimicrobial activity increase resistance or at least decrease symptoms in some host-pathogen interaction (Hammerschmidt and Smith, 1997). From these compounds benzothiodiazole (BTH), which induces systemic resistance (Gorlach et al., 1996 and Bokshi et al., 2003) in many plants and against a broad spectrum of pathogens.

Increased resistance in potato tubers against *E. carotovora* subsp. *carotovora* was observed when tubers were dipped in acetylsalicylic acid (ASA) (Abd El-Sayed at al. 1996 and Bokshi et al., 2003). Salt treatments can inhibit plant pathogens or suppress toxin production (Olivier et al., 1998). Salts including calcium propionate and calcium chloride reduced tissue maceration of potato tubers caused by *E. carotovora* (McGuire and Kelman, 1986; Biggs et al., 1997 and Droby et al., 1997). The efficacy of salt treatments depends on the concentration, the target organism and the method of application (Punja and Gaye, 1993 and Olivier et al., 1998).

Application of fluorescent pseudomonads reduced soft rot incidence, improved plant growth and increased tuber yield in field trials (Xu and Gross, 1986 and Kloepper et al., 2004). Liao (1989) mentioned that an isolate of P. putida inhibited the growth of E. carotovora on potato slices. Also, P. putida isolate survived on the tubers and roots of potato plant for more than 5 weeks. Application of fluorescent pseudomonads to seed pieces to reduce infection of daughter tubers with soft rot bacteria has suppressed the population of E. carotovora on roots and daughter tubers (Kloepper, 1983). In the present work, it was planned to suppress bacterial soft rot disease on potato using different treatments, i.e. inducer compounds, salt compounds and bioagent under artificial inoculation conditions.

Materials and Methods

1. Bacteria and inoculum preparation:

A virulent isolate of *E. carotovora* subsp. *carotovora* was isolated from potato (*Solanum tuberosum* L.) tubers showing bacterial soft rot and identified according to Fahy and Persley (1983). Two isolates of *Pseudomonas fluorescens* (Pf1 and Pf5), two isolate of *Serratia marcescens* (Sm1 and Sm2), isolate of *P. aeruginosa* (Pa1) and isolate of *B. subtilis* (Bs3) as bioagents were selected from the stock culture collection maintained in bacterial diseases laboratory, Department of Plant Pathology, Faculty of Agriculture, Ain Shams University. These isolates were evaluated against several diseases (Abd El-Ghaffar and Abd El-Sayed, 1997; Abd El-Ghaffar and Mosa, 2001; Abd El-Sayed *et al.*, 2003 and 2006). These isolates were grown on yeast extract peptone agar (YEPA) medium for 48 hrs at 28°C. The bacterial cells were suspended in sterilized distilled water (SDW) and adjusted to 10^7 colony forming units (cfu/ml) for the pathogen (Wang *et al.*, 1994) and to 10^8 cfu/ml for bioagents, spectrophotometrically (Abd El-Ghaffar *et al.*, 2006).

2. Treatments:

BION (acibenzolar S-methyl) was applied at 0.1, 0.3 and 0.5g/l and salicylic acid (2-hydroxybenzoic acid, $C_7 H_6 O_3$) was applied at 0.3, 0.6 and 0.9 mM as resistance inducing factors. Potassium sulfate, ammonium phosphate and calcium chloride as salt compounds were applied at 0.1, 0.3 and 0.5 g/l. Antagonistic bacteria mentioned previously, were applied as bioagents. These factors were applied at the same time of inoculation with the pathogen and before or after 24 hours. Certain tubers were only inoculated with the pathogen as a control. Entire potato tubers were weighted and surface sterilized by dipping in 70% ethanol fore one min. followed by two successive rinses in SDW and dried. Sterilized cork borer (0.5-cm-diam.) was used to make a hole (1cm, depth) in the middle of each sterilized tuber.

The pathogen (0.2 ml) and the tested factors (0.2 ml) were piptted into the hole. The holes were closed again with the same removed cylinders of tubers. Treated tubers were placed in sterilized plastic boxes (12X 9 X 8 cm). These boxes were closed with a lid as chamber room and incubated at 28 C for five days (Abd El-Ghaffar and Abd El-Sayed, 1997). Three boxes of tubers were used as replicates per each treatment and each box containing three tubers

3. Disease assessment:

Disease severity was estimated as percentage of rotted tissue weight according to change weight of tuber before and after treatment divided on weight of tuber before treatment (Schober and Vermeulen, 1999). Percentage of disease reduction (PDR) was calculated according to percentage of rotted tissue weight (Saettler *et al.*, 1989) as the following formula:

Whereas: Ack = disease severity in control and Atr = disease severity in treatment.

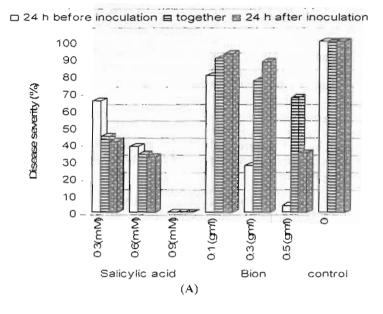
Results and Discussion

1. Effect of some resistance inducing factors against bacterial soft rot of potato:

Application of salicylic acid and BION as resistance inducing factors has significantly decreased severity of bacterial soft rot disease of potato compared with the control (Fig.1 A&B). Percentage of disease reduction was increased by increasing the rate of salicylic acid and BION. Efficiency of salicylic was more effective against the disease than BION. Also, salicylic acid was the most effective to reduce the disease, at 9 mM, where percentage of disease reduction was 100%. Applications of salicylic acid lead to decrease the disease, when this compound was applied at the same time or after inoculation with the pathogen. Meanwhile, application of BION tends to reduce the disease, when it was applied before inoculation with the pathogen. These results are in agreement with Abd El-Sayed et al. (2003). The effect of salicylic acid was not caused by direct action on the growth of pathogen, but the effect of was rather a consequence of induction of plant defence response (Malamy and Klessing, 1992). Many Biochemical changes occur during systemic acquired resistance (SAR), i.e. pathogenesis- related (PR) proteins. Acidic PR-proteins including acidic B-1.3 glucanase and chitinase are secreted in the intercellular spaces, where they would act against fungal and or bacterial pathogens at early stage of infection process. Basic B,1,3-glucanase and chitinase accumulate in the vacuole, may interact with pathogens, at a later stage of infection, during host cell deterioration (Ye et al., 1995 and Kuc, 1995).

2. Effect of some salt compounds against bacterial soft rot of potato:

Application of potassium sulfate, ammonium phosphate and calcium chloride as salt compounds significantly decreased severity of bacterial soft rot disease of potato compared with the control (Fig. 2A&B). Efficiency of salt compounds was increased against the disease by increasing their rate. Severity of bacterial soft rot of potato was completely reduced, when potassium sulphate and calcium chloride were applied at 0.5 g/l and at the same time of inoculation with the pathogen. Tested



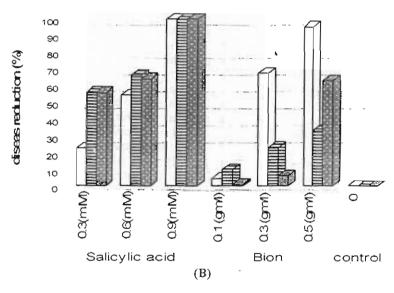
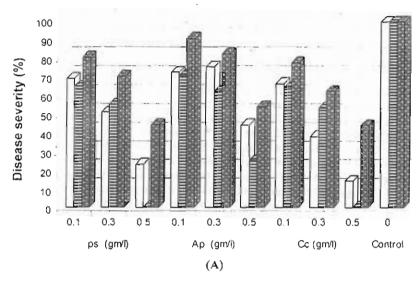


Fig. 1. Influence of salicylic acid and BION at different rates as inducer agents on percentage of rotted tissue weight (A) and percentage of disease reduction (B), using three treatments under artificial inoculation conditions.





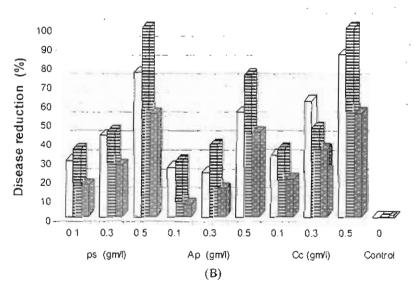


Fig. 2. Influence of potassium sulfate (PS), ammonium phosphate (AP) and calcium chloride (Cc), at different rats as salts on percentage of rotted tissue weight (A) and percentage of disease reduction (B), using three treatments under artificial inoculation conditions.

salts were most effective against the disease when they were applied at the same time of inoculation with the pathogen. Applications of salts before inoculation with the pathogen were moderately effective against the disease and were less effective when they were applied after inoculation with the pathogen.

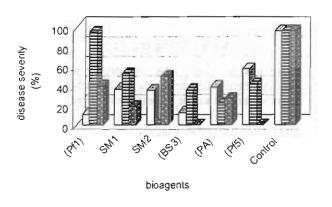
Free calcium ions in tissue stimulate pectate lyase (PL) activity but cause inhibition of polygalacturonase (PG) activity for virulent isolates of soft rot bacteria (Alghisi and Fravaron, 1995). The nitrogen content of the plant is positively correlated with the amount of bacterial soft rot (Wright, 1993). The ratio of chloride to potassium in plant tissues was found to be highly correlated with visual symptom development (Schneider, 1985). Increased chloride uptake can reduce the synthesis of malate in potato plant (Luttage and Higinbotham, 1979), a carbon substrate that might be utilized by plant pathogens. Schober and Vermeulen (1999) found no differences in the enzyme activity between the nitrogen and calcium treatment. Nitrogen levels in the crop were known to influence soft rot severity (Carballo et al., 1994). Nitrogen fertilization increases the dry matter content of broccoli (Everaarts, 1994) and has a direct effect on the cell wall composition, especially the esterification of pectate in cell walls. Nitrogen fertilization interacts with the production of plant defence substance as phenols and inhibited calcium uptake (Reerink, 1993), Perombelon and Salmond (1995) found a correlation between the resistance of potatoes against bacterial soft rot and the composition of the plant cell walls. Resistant potato cultivars had consistently higher amount of calcium in cell wall preparations than susceptible cultivars and they had increased levels of galacturonic acid in cell walls.

3. Effect of some bioagents against bacterial soft rot of potato:

All tested bioagents have significantly reduced severity of bacterial soft rot disease of potato compared with the control (Fig. 3): Severity of disease completely decreased when isolates of *P. fluorescens* (Pf5) and *B. subtilis* (Bs3) were applied after inoculation with the pathogen and when the isolate of *Serratia marcescens* (Sm2) was applied at the same time inoculation with the pathogen. Meanwhile, disease severity was decreased when *P. fluorescens* (Pf1) was applied before inoculation with the pathogen, *S. marcescens* (Sm1) after inoculation with the pathogen and *P. aeruginosa* (Pa1) at the same time inoculation with the pathogen.

Fluorescent pseudomonads are aggressive rhizosphere colonizes and produce a wide range of antimicrobial compounds (Kloepper, 1983 and Gross, 1988). Certain fluorescent pseudomonads can protect plants from diseases caused by root pathogens and often this biocontrol effect involves antimicrobial compounds such as siderophores (Bakker et al., 1987), hydrogen cyanide (Voisard et al. 1989) and antibiotics (Raaijmakers et al., 2002) like phenazine-1-carboxylate, pyroluteorin and 2,4-diacetyl phoroglucinol. P. aeruginosa produces a plethoro of extracellular lytic enzyme and secondary metabolites of low molecular weight known as auto-inducers (Pedersen, 1992), where auto-inducers are extracellular N-acyl-homoserine lactones synthesized by various Gram-negative bacteria. The blue phenazine pigment together with Ferripyochellin (a sidrophore of P. aeruginosa), and a reducing agent such as NADH, which catalyses the reaction of hydroxyl and superoxide radicals (Britigan, 1993).

□ 24 hours before inoculation ☐ togethere ☐ 24 hours after inoculation



(A)

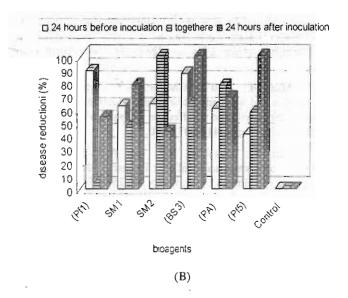


Fig. 3. Influence of Pseudomonas fluorescens (Pf1 and Pf5), P. aeruginosa (PA1), Serratia marcescens (Sm1 and Sm2) and Bacillus subtilis (Bs3) as bioagents, on percentage of rotted tissue weight (A) and percentage of disease reduction (B), using three treatments under artificial inoculation conditions.

Experiments screening of *Bacillus* spp. had elicited induced systemic resistance (ISR) on other crops (Jetiyanon and Kloepper 2002), where two strains of *B. pumilus* and a strain of *B. mycoides* enhanced peroxidase activity and increased production of one chitinase isozyme and two isozymes of B-1,3 glucanase (Bargabus *et al.*, 2002 and 2004). Another strain of *B. pumilus* had greatly increased levels of salicylic acid (Zhang *et al.*, 2002).

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References

- Abd El-Ghaffar, N.Y. and Mosa, A.A. 2001. Integration between biological and chemical treatment to control bacterial spot disease of tomato. *Egypt. J. Phytopathol.*, **29**: 33-45.
- Abd El-Ghaffar, N.Y. and Abd El-Sayed, Wafaa M. 1997. Biological control of bacterial soft rot of potato. *Arab Univ. J. Agric. Sci.*, Ain Shams Univ., Caire, 5 (2): 419-431.
- Abd El-Sayed, Wafaa M.; Gado, E.A.M. and Abd El-Ghaffar, N.Y. 2006. Utilization of combining biotic and abiotic treatments to control bacterial angular leaf spot disease of cucumber. Arab Univ. J. Agric. Sci., Ain Shams Univ., Cairo, 14: 381-394.
- Abd El-Sayed, Wafaa M.; Abd El-Ghaffar, N.Y. and Shehata, A.M. 1996. Application of salicylic acid and aspirin for induction of resistance to tomato plants against bacterial wilt and its effect on endogenus hormones. *Annals Agric. Sci.*, Ain Shams Univ., Cairo, 41:1007-1020.
- Abd El-Sayed. Wafaa M.; Bayoumi, R.A. and Abd El-Ghaffar, N.Y. 2003. Biological control of potato bacterial wilt disease under Egyptian conditions. *Annals Agric. Sci.*, Ain Shams Univ., Cairo, 48: 353-364.
- Alghisi P. and Fravaron, F. 1995. Pectin degrading enzymes and plant-parasite interactions. Eur. J. Plant Pathol., 101: 365-375.
- Bakker, P.A.H.M.; Bakker, A.W.; Marugg, J.D.; Weisbeek, P.J. and Schippers, B. 1987. Bioassays for studying the role of siderophores in potato growth stimulation by *Pseudomonas* spp. in short potato rotations. *Soil Biol. Biochem.*, 19: 443-449.
- Bargabus, R.L.; Sherwood, N.K. and Jacobsen, B.J. 2002. Characterization of systemic resistance in sugar beet elicited by a non-pathogenic, phyllospherecolonizing *Bacillus mycoides*, biological control agent. *Physiol. Mol. Pathol.*, 61: 289-298.

- Bargabus, R.L.; Sherwood, N.K. and Jacobsen, B.J. 2004. Screening for the identification potential biological control agent that induce systemic acquired resistance in sugar beet. *Biol. Control*, 30: 342-929.
- Biggs, A.R.; El-Kholi, M.M.; El-Neshawy, S. and Nickerson, R. 1997. Effect of calcium salts on growth, polygalacturonase activity, and infection of peach Fruit by *Monilinia fructicola*. *Plant Dis.*, 81: 399-403.
- Bokshi, A.I; Morris, S.C. and Deverall, B.J. 2003. Effects of benzothiadiazole and acetylsalicylic acid on b-1,3-glucanase activity and disease resistance in potato. *J. Plant Pathol.*, **52**: 22-27.
- Britigan, B.E. 1993. Role of reactive oxygen species *Pseudomonas* infection in *P. aeruginosa*: the opportunist pathogenesis and disease. Pages: 113-140. Fick, R.B. (ed.). Boca Roton CRC Press.
- Carballo S.J.; Blankenship, S.M.; Anders, D.C.S. and Ritchie, D.F. 1994. Drip fertilization with nitrogen and potassium and post-harvest susceptibility to bacterial soft rot of bell peppers. J. Plant Nutr., 17: 1175--1191
- Cronin, D.; Loccoz, Y.M.; Fenton, A.; Dunne, C.; Dowling, D.N. and Gara, F.O. 1997. Ecological interaction of a biocontrol *Pseudomonas fluorescens* strain production 2, 4-diacetylphloroylucinol with the soft rot potato pathogen *Erwinia carotovora* subsp. *atroseptica*. *FEMS Microbiology Ecology*, 23: 95-106.
- Droby, S.; Wisniewski, M.E.; Cohen, L.; Weiss, B.; Touitou, D.; Eilam, Y. and Chalutz, E. 1997. Influence of CaCl2 on *Penicillium digitatum* grapefruit peel tissue and biocontrol activity of *Pichia guilliermondii*. *Plant Dis.*, 87: 310-315.
- Everaarts, A.P. 1994. Nitrogen fertilization and head rot in broccoli. *Neth. J. Agric.* Sci., 42: 195-201
- Fahy, P.C. and Persley, G.J. 1983. *Plant Bacterial Diseases. A Diagnostic Guide*. American Press, Sydney, New York, London.
- Gorlach, J.; Sandra, V.; Gertrud, K.; Georges, H.; Uli, B.; Karl-Heinz, K.; Oostendrop, M.; Staub, T.; Ward, E.; Kessmann, H. and Rayals, J. 1996. Benzothiadiazole, a novel class of inducers of systemic acquired resistance in wheat. *Plant cell*, 8: 629-643.
- Gross, D.C. 1988. Maximising rhizosphere populations of fluorescent pseudomonads on potatoes and their effects on *Erwinia carotovora*. *Amer. Potato J.*, **65**: 697-710.
- Hammerschmidt, R. and Smith, J.B. 1997. Acquired resistance to disease in plants. Hort. Rev., 18: 247-289.
- Jetiyanon, K. and Kloepper, J.W. 2002. Mixture of plant growth- promoting rhizobacteria for induction of systemic resistance agents multiple plant diseases. *Biol. Control*, 24: 285-291.

- Kloepper, J.W. 1983. Effect of seed piece inoculation with plant growth promoting rhizobacteria on population of *Erwinia carotovora* on potato roots and daughter tubers. *Phytopathology*, 73: 217-219.
- Kloepper, J.W.; Ryu, C.M. and Zhang, S. 2004. Induced systemic resistance and promotion of plant growth by *Bacillus* spp. *Phytopathology*, 94: 1259-1266.
- Kuc, J. 1995. Introduction Systemic Resistance. An Overview. Kluwer Academic Publishers. Netherlands.
- Liao, C.H. 1989. Antagonism of *Pseudomonas putida* strain PP22 to phytopathoginic bacteria and its potential use as a biocontrol agent. *Plant Dis.*, 73: 223-226.
- Luttage, U. and Higinbotham, N. 1979. Transport in Plants. Springer-Village, New York.
- Malamy, J. and Klessing, D.E. 1992. Salicylic acid and plant disease resistance. Plant J., 2: 643-654.
- McGuire, R.G. and Kelman, A. 1986. Calcium in potato cell wall in relation to tissue maceration by *Erwinia carotovora*. *Phytopathology*, 76: 401-406.
- Olivier, C.; Halseth, D.E.; Mizubuti, E.S.G. and Loria, R. 1998. Postharvest application of organic and inorganic salts for suppression of silver scurf on potato tubers. *Plant Dis.*, 82: 213-217.
- Pedersen, S.S. 1992. Lung infection with alginate signal-producing, mucoid *P. aerugenosa* in cystic fibrosis. *APMIS Suppl.*, **100**: 7-79.
- Perombelon, M.C.M. and Salmond, G.P. 1995. Bacterial soft rots. Pages: 1-17. In: *Prokaryotes*. U.S. Singh (ed.). Vol. I. Pergamon, Oxford.
- Punja, Z.K. and Gaye, M.M. 1993. Influence of post harvest handling practices and dip treatments on development of black root rot on fresh market carrots. *Plant Dis.*, 77: 989-995.
- Raaijmakers. J.M.; Vlami, Maria and de Souza, J.T. 2002. Antibiotic production by bacterial Biocontrol agents. *Antonie Van Leeuvwenhoek*, 81: 537-547.
- Reerink, J.A. 1993. Onderzoek naar factoren en processen die de produktie en kwaliteit van witlof be invloeden. Eindversl Pr 729 CAB-DLO, Wageningen, the Netherlands. 174 pp. (English summary).
- Schneider, R.W. 1985. Suppression of Fusarium yellows of celery with potassium. chloride, and nitrate. *Phytopathology*, 75: 40-48.
- Schober, B.M. and Vermeulen, T. 1999. Enzymatic maceration of witloof chicory by the soft rot bacteria *Erwinia carotovora* subsp. *carotovora*: the effect of nitrogen and calcium treatments of the plant on pectic enzyme production and disease development. *Eur. J. Plant Pathol.*, **105** (4): 341-349.

- Voisard, C.; Keel, C.; Haas, D. and Deèfago, G. 1989. Cyanide production by *Pseudomonas fluorescens* helps suppress black root rot of tobacco under gnotobiotic conditions. *EMBO J.*, 8: 351-358.
- Wang, J.F.; Jones, J.B.; Scott, K.W. and Stall, R.F. 1994. Several genes in Lycopersicon escolentum control hypersensitivity of Xanthomonas campestris pv. Vescatoria. Phytopathology, 84: 702-706.
- Wright, P.J. 1993. Effects of nitrogen fertilizer, plant maturity at lifting and water during field-curing on the incidence of bacterial soft rot of onions in store. N. Z. J. Hort. Sci., 21: 377-381.
- Xu. G.W and Gross, D.C. 1986. Field evaluation of the interactions among fluorescent pseudomonads and *Erwinia carotovora* and potato yields. *Phytopathology*, 76: 423-430.
- Ye, X.C., Storbel, N. and Kuc, J. 1995. Induced systemic resistance (ISR), activation of natural defence mechanisms for disease control as part of integrated pest management (IPM). Pages: 95-113. In: Novel Approaches to Integrated Pest Management. Plenum Press, New York.
- Zhang, S.: Moyne, A.L.; Reddy, M.S. and Kleopper, J.W. 2002. The role of salicylic acid in induced systemic resistance elicited by plant growth promoting rhizobacteria against blue mould of tobacco. *Biol. Control*, 25: 288-296.

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مكافحة مرض العفن الطري البكتيري في البطاطس عبد السلام عبدالله الحاج حامد - وفاء محمد عبد السيد -احمد أبواليزيد * - ناجي بسين عبد الغفار *

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يسبب مرض العفن الطري البكتيري خسائر فادحة في مصر والعالم، وعلى عوائل نباتية عديدة من أهمها محصول البطاطس يتسبب هذا المرض عن البكتريا Erwinia carotovora subsp. carotovora.

جرت هذة الدراسة بهدف مكافحة هذا المرض باستخدام وسائل مكافحة عديدة تتضمن كل من حمض الساليسيليك والبيون كعومل حث كيمانية ، وكبريتات البوتاسيوم وفوسفات الأمونيوم وكلوريد البوتاسيوم بالتركيز (١٠٠ ، و ٥٠٠ جم / ليتر) كاملاح ، وعز لات من بكتيريا مضادة كعوامل مكافحة حيوبة من كل من الأتواع (Pfl and Pf5) وعز لات من بكتيريا مضادة (Bs3) Serratia marcescens (Sm1 and Sm2), Bacillus subtilis (Bs3) فعالية هذه العوامل في قدرتها على خفض شدة المرض في ثلاثة مواقيت للمعاملة فعالية هذه العوامل في قدرتها على خفض شدة المرض في ثلاثة مواقيت للمعاملة بعد مرور على المعاملة بعد مرور المعاملة في نفس الوقت مع العدوى بالبكتريا الممرضة. (٢) المعاملة بعد مرور عليها في هذه الدراسة :

- ١- خفضت جميع المعاملات الشدة المرضية للعفن الطري بشكل معنوي المصائيا بالمقارنة بمعاملة المقارنة.
- ٧- اتخفضت الشدة المرضية للمرض كليا عند المعاملة بحمض الساليسيليك بالتركيز ٩٠٠ ملكي مول/ ليتر بالتواقيت الثلاثة للمعاملة ، وكذلك عند المعاملة بكل من كبريتات البوتاسيوم وكلوريد الكالسيوم بالتركيز ٩٠٠ جم / ليتر عند تطبيقها بنفس وقت العدوى بالبكتريا الممرضة.
- B. subtilis (Bs3) S. marcescens (Sm2) من كليا في المواقبت الثلاثة
 P. fluorescens (Pf5) الشدة المرضية للعفن كليا في المواقبت الثلاثة للمعاملة.
- ٤- تزداد فعالية كل من عوامل الحث الكيمانية (حمض السليسيلك ، البيون)
 والأملاح الثلاثة في تخفيض الشدة المرضية مع زيادة التركيز المستتخدم لكل
 منعا .
- حمض الساليمبيلك وكلوريد الكالسيوم و كبريتات البوتاسيوم كانت أكثر العوامل فعالية ضد المرض بالمقارنة بكل من البيون وفوسفات الأمونيوم .
- آ- باقي العزلات الحيوية المستخدمة خفضت المرض بشكل معتدل مقارنة بالعزلات الأكثر فعالية وذلك في المواقيت الثلاثة للمعاملة.

وعلى ضوء هذه النتائج لابد من إختبار وتقييم العوامل الأكثر فعالية في هذا البحث تحت الظروف الطبيعية للمرض للإستفادة منها في مكافحة المرض.