# ANTAGONISTIC AND INSECTICIDAL ACTIVITIES OF SOME STREPTOMYCES ISOLATES

#### **ABSTRACT**

Fifteen local isolates of Streptomyces named, S01, S02, S03, S04, S05, S06, S07, S08, S09, S10, S11, S12, S13, S14, and S15; isolated from different soils and geographical areas in Egypt were used in this investigation. These isolates were propagated and subjected to screening studies to evaluate their efficiency as antagonistic agents against some phytopathogenic fungi such as, Rhizopus stolonifer, Aspergillus niger, Fusarium oxysporum, Helminthisporum gramenium and insect pest, cotton leaf worm (Spodopetra littoralis). The Streptomyces isolates were grown on starch nitrate broth medium under shaking condition at 28°C for 6 days. Culture supernatants were then tested against the five applied phytopathogenic fungi. Results revealed that most of the isolates were varied in their antagonistic activities. Isolate S08 was active against R. stolonifer, A. niger and F. oxysporum while isolates S01, S05, S11 and S14 were active against A. niger and F. oxysporum and isolates S04, S09 and S13 were active against R. stolonifer and A. niger. On the other hand, no antifungal activity was found against Helminthisporum gramenium. The insecticidal activity of both culture filtrates and cell pellets were tested against cotton leaf worm. The experimental results showed that the pellets of some Streptomyces isolates were more active against cotton leaf worm than culture filtrates. Generally, isolates S05, S08, S10 and S15 showed 80, 100, 70 and 80% mortality against cotton leaf worm, respectively. The protein(s) of isolate S08 cells was purified through ammonium sulfate saturation 40, 60 and 80%. Results of SDS-PAGE analysis showed that a 40 KDa protein was purified and showed high activity against four instars of the cotton leaf worm. This result demonstrated the ability of use such Streptomyces isolates as effective biopesticide agents.

**Key words**: *Streptomyces*, Antagonistic and insecticidal activity, Fungi, Cotton leaf worm, Toxicity, Mortality.

#### INTRODUCTION

Integrated pest management (IPM) programs create the need for novel fungicides or insecticides having more selective modes of action. Attempts have been made to develop *Streptomyces* species as fungal root disease control agents, since *Streptomyces* spp. are capable of producing a remarkably wide spectrum of antibiotics as secondary metabolites (Lechevalier and Waksman, 1962; Lechevalier, 1988 and Franklin *et al.*, 1989). Microorganisms produce many useful anthelmintic and insecticidal antibiotics (Pachlatko, 1998; Ghazal *et al.*, 2001 and Yuhui *et al.*, 2002).

In 2001, Bream et al. investigated the biological activity of the secondary metabolites of 41 Egyptian actinomycete strains on the cotton leaf worm Spodoptra littoralis. They found that 58% of the tested strains caused larval mortality ranging from 10-60%; Streptomyces and Streptoverticillum were the most potent actinomycetes affecting the biological and physiological criteria of the present insect species.

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Xiong et al. (2004) isolated some Streptomyces strains from seawater and sea sediments from Beidiahe and Dagang of the east coast of China. These strains were screened for their insecticidal activities using bioassay against Helicoverpa armigera. Results revealed that 40 out of the 331 (12.08%) isolates showed potential insecticidal activities. Of the 40 isolates, one isolate, designated Streptomyces sp.173, was found to have strong insecticidal activity against H. armigera.

Recently, Shiomi et al. (2005) isolated a new antimycin group antibiotic, antimycin A9 from a cultured broth of Streptomyces sp. K01-0031 together with antimycins A3a, A3b, A4, and A7, and flazin methyl ester. Antimycin A9 is the first antimycin having an aromatic 8-acyl residue. It showed potent nematocidal and insecticidal activities against Caenorhabditis elegans and Artemia salina, respectively. It inhibited bovine heart NADH oxidase at nanomolar level like other known antimycins.

Mourad et al. (2004) showed that the Egyptian cotton leaf worm (S. littoralis) is an important polyphagous insect pest attacking cotton, several cultivated crops and ornamental plants worldwide Therefore, the aim of this study was to determine the antifungal and insecticidal activities of fifteen soil-Streptomyces isolates against some phytopathogenic fungi and cotton leaf worm insect.

#### MATERIALS AND METHODS

### Streptomyces isolates:

Fifteen unidentified *Streptomyces* isolates (S01, S02, S03, S04, S05, S06, S07, S08, S09, S10, S11, S12, S13, S14, and S15) isolated from different soils in Egypt, were kindly obtained from, Department of Agricultural Microbiology, Soil, Water and Environment Research Institute (SWERI), ARC, Giza, Egypt.

# Isolates propagation and preparation:

Standard inoculums for each applied *Streptomyces* species was prepared by scraping the heavy spores from the surface of the growth of starch nitrate slant in the presence of 5 ml sterilized d.H<sub>2</sub>O. An aliquot of 2 ml of this standard inoculum (containing 1.5 ml spores/ml) was transferred aseptically to 50 ml of a broth medium (Not shown data) modified from starch nitrate broth medium **Waksman and Lechvalier** (1961) in a 250 ml conical flask. Inoculated flasks were incubated at 28±2°C for 6 days on a rotary shaker (160 rpm/min). Thereafter, growth was centrifuged at 10000 rpm at 4°C for 5 minutes. Then the supernatants and pellets were separately subjected to evaluation for their antagonistic or insecticidal activities.

**Pellets preparation:** The pellet of each *Streptomyces* isolates was re-suspended in equal volume of Tris buffer pH 7.5 then sonicated and spun at 14000 rpm.

# Antagonistic activities of Streptomyces isolates:

The antagonistic activities of the supernatants of the 15 Streptomyces isolates under investigation were tested against four phytopathogenic fungi, i.e., Rhizopus stolonifer, Aspergillus niger, Fusarium oxysporum and Helminthisporum gramenium, kindly provided by Cairo MIRCEN, Faculty of Agriculture, Ain Shams University. Test was carried out as described by Mohamed et al. (2001).

#### Source of insect:

Sets of the cotton leaf worm (S. littoralis) insect belonging to family Noctuidae were kindly provided by the insectary at AGERI, ARC, Giza, Egypt.

## Insecticidal activities of Streptomyces isolates:

The supernatants and pellets of the 15 Streptomyces isolates of this study were used to determine their insecticidal activities against the 1<sup>st</sup> instar larvae of cotton leaf worm. For each treatment, three bioassay cups 5-cm in diameter containing 2.5 ml of cooled-dried semi-artificial diet (Levinson and Novon, 1969) supplemented with 500  $\mu$ l of the supernatant or pellet-suspension and 10 larvae of cotton leaf worm were added. Cups were then kept at 28°C, and the mortality (%) was recorded after 3-5 days according to Finny (1962). As a control, 3 replicates of the bioassay cups with the same medium were also used.

# Protein purification:

The protein(s) of the *Streptomyces* isolate No. 8 were extracted and purified as follows: the pellet-suspension was separately treated with 40, 60 and 80% ammonium sulfate, incubated for three hours at 4°C and centrifuged at 14000 rpm for 15 min at 4°C. The supernatant was discarded and the pellet was then resuspended in a suitable volume of 20 mM Tris-HCl pH 7.5 and dialyzed against 1X phosphate buffer saline (PBS). The crude proteins prepared by 40, 60 and 80% ammonium sulfate treatments were subjected to evaluation of their insecticidal activities against 1<sup>st</sup> instar of cotton leaf worm larvae. Moreover, the protein of 40% was tested against 2<sup>nd</sup>, 3<sup>rd</sup> and 4<sup>th</sup> instars as mentioned above.

#### RESULTS AND DISCUSSION

Streptomycetes produce a wide variety of commercially important secondary metabolites, including antibiotics which exhibit antibacterial, antifungal, anthelmintic, antitumour and immunosuppressive activities (Hopwood, 1997; Katz, 1997 and Leadlay, 1997).

In this study, a set of fifteen soil-streptomycete isolates from Egypt were used in the present study to investigate the biological activities of their secondary metabolites against some phytopathogenic fungi as well as cotton leaf worm (S. littoralis). Results in Table (1) showed that culture filtrates of the applied streptomycete isolates were varied in their antifungal activities. Streptomyces isolate S08 was active against R. stolonifer, A. niger and F. oxysporum while isolates S01, S05, S11 and S14 were active against A. niger and F. oxysporum and S04, S09 and S13 isolates were active against R. stolonifer and A. niger. On the other hand, no antifungal activity was found against Helminthisporum gramenium. This finding confirmed the possibility of presence of secondary metabolites, i.e., fungicides in the culture filtrates of propagated isolates.

Several investigators (Mohamed, 1998; Mohamed et al., 2000; Mahfouz and Mohamed, 2002; Abdel-Fattah (2005) and Mohamed et al., 2005) reported the ability of streptomycetes to inhibit the growth of different phytopathogenic fungi as well as soil-borne fungi.

Saroj et al. (1987) screened metabolites from 942 microbial isolates for insecticidal properties. The isolates included 302 streptomycetes, 502 novel actinomycetes including representatives of 18 genera, 28 unidentified aerobic

actinomycetes, 70 fungi and 40 bacteria other than actinomycetes. They showed that the metabolites from 55 isolates at a dilution of  $10^{-1}$  caused nearly one hundred percentage mortality in mosquito larvae (Aedes aegypti) within 24 h. These isolates included 27 isolates of Streptomyces, four of Actinoplanes, three isolates each of Actinomadura and Streptoverticillium, two isolates each of Micromonospora, Bacillus and Paecilomyces and one isolate each of Micropolyspora, Nocardiopsis, Streptosporangium, Oerskovia, Thermomonospora, Chainia, Pseudomonas, Fusarium, Monilia and Syncephalestrum.

In this investigation, the insecticidal activities of both culture filtrates and cell pellets had been tested against cotton leaf worm. Results in **Table** (2) which illustrated in **Figure** (1) revealed that the pellets of some *Streptomyces* isolates were more active against cotton leaf worm than culture filtrates. Streptomycete isolates S09; S14; S02 & S07; S12 & S13; S01; S11; S03 & S04; S06 showed percentages of mortalities against the 1<sup>st</sup> instar of cotton leaf warm as followed 32.5; 27.5; 25; 22.5; 20; 17.5; 12.5 and 7.5, respectively. On the other hand, four streptomycete isolates namely S05, S08, S10 and S15 were recorded as the most effective as they showed 80, 100, 70 and 80% mortality, respectively.

In a trial to define which protein is responsible for toxicity, the protein(s) of isolate S08 cells was purified through ammonium sulfate saturation (ASS) 40, 60 and 80%. Results of SDS-PAGE analysis showed that a 40 KDa protein was obtained from the 40% ASS (Data not shown). As interestingly, data in Table (3) showed 100% mortality for that purified protein against four instars of the cotton leaf (Figures 2 and 3).

In china, Streptomyces nanchangensis isolated from the soil in Nanchang, China (Ouyang et al., 1984) produces at least two kinds of insecticidal antibiotics (Ouyang et al., 1993), first resembles dianemycin and second resembles milbemycin (Takahashi et al., 1993). Both meilingmycin and nanchangmycin are very active against a broad spectrum of harmful nematodes and insects, and are non-toxic for mammals as well as plants (Ouyang et al., 1993).

Table (1): Antagonistic activities of the 15 Streptomyces isolates under investigation against four phytopathogenic fungi.

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Streptomyces isolates	Phytopathogenic fungi used						
	R. stolonifer	A. niger	F. oxysporum	H. gramenium			
S01	-	+	+	-			
S02	-	+		-			
S03	+	-	-	-			
S04	+	+	-	-			
S05	-	+	+	-			
S06	+	-	-	-			
S07	-	_	+	-			
S08	+	+	+	-			
S09	+	+	-	-			
S10	+	_	-	-			
S11	-	+	+	-			
S12	+	-	-	-			
S13	+	+	-	-			
S14	-	+	+	-			
S15	+	-	-	-			

<sup>-:</sup> Negative.

<sup>+:</sup> Positive.

Table (2): Insecticidal activities of the 15 *Streptomyces* isolates under investigation against 1<sup>st</sup> instar larvae of cotton leaf worm (*S. littoralis*).

	Insecticidal activities						
Streptomyces isolates	Alive	(%)	Dead (%)				
	Supernatant	Pellets	Supernatant	Pellets			
S01	86.0	80.0	14.0	20.0			
S02	94.0	75.0	6.00	25.0			
S03	98.0	87.5	2.00	12.5			
S04	92.0	87.5	8.00	12.5			
S05	94.0	20.0	6.00	80.0			
S06	96.0	92.5	4.00	7.50			
S07	96.0	75.0	4.00	25.0			
S08	0.00	0.00	100	100			
S09	94.0	67.5	6.00	32.5			
S10	86.0	30.0	14.0	70.0			
S11	68.0	82.5	32.0	17.5			
S12	96.0	77.5	4.00	22.5			
S13	96.0	77.5	4.00	22.5			
S14	88.0	72.5	12.0	27.5			
S15	94.0	20.0	6.00	80.0			
Control	100	100	0.00	0.00			

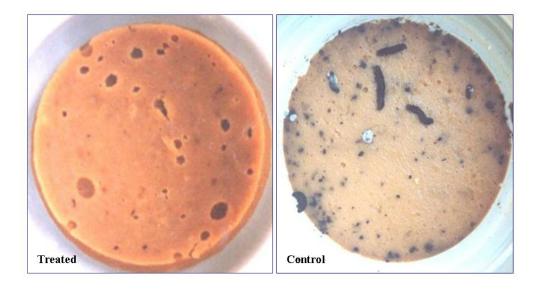


Figure (1): Insecticidal activities of *Streptomyces* isolate S08 under investigation against 1<sup>st</sup> instar larvae of cotton leaf worm (*S. littoralis*).

Table (3): Insecticidal activities of purified protein (40 KDa) of *Streptomyces* isolate S08 against four instars larvae of cotton leaf worm (S.littoralis) post 72 hours from treatment.

Larvae instars	Treatment	Insecticidal activities				
		Alive		Dead		
		No.	%	No.	%	
<b>1</b> <sup>st</sup>	Treated	00	00	30	100	
	Control	27	90	03	10	
2 <sup>nd</sup>	Treated	00	00	30	100	
	Control	28	93.3	02	6.7	
3 <sup>rd</sup>	Treated	00	00	30	100	
	Control	28	93.3	02	6.7	
4 <sup>th</sup>	Treated	00	00	30	100	
	Control	27	90	03	10	

Thirty larvae of cotton leaf worm in three replicates were used for treatment as well as control.



Figure (2): Insecticidal activities of purified protein (40 KDa) of *Streptomyces* isolate S08 against  $3^{\rm rd}$  instar larvae of cotton leaf worm (*S. littoralis*). Note, the control larvae of  $3^{\rm rd}$  instar grew up to  $6^{\rm th}$  instar.



Figure (3): Insecticidal activities of purified protein of *Streptomyces* isolate S08 against  $4^{th}$  instar larvae of cotton leaf worm (*S. littoralis*). Note, the control larvae of  $4^{th}$  instar grew up to  $5^{th}$  and  $6^{th}$  instars.

#### REFERENCES

- **Abdel-Fattah, H.I. (2005).** Cultural, morphological, physiological and molecular studies on some streptomycete isolates. Arab Univ. J. Agric. Sci., Ain Shams Univ., Cairo 13(2): 249-268.
- Bream, A.S.; Ghazal S.A.; Abd el-Aziz, Z.K. and Ibrahim, S.Y. (2001). Insecticidal activity of selected actinomycete strains against the Egyptian cotton leaf worm *Spodoptera littoralis* (Lepidoptera: *Noctuidae*). Meded. Rijksuniv. Gent. Fak. Landbouwkd Toegep. Biol. Wet. 66(2a):503-12.
- Finny, D. (1962). Probit Analysis. Cambridge University Press, Cambridge.
- Franklin, T.J.; Snow, G.A., Barrett-Bee, K.J. and Nolan R.D. (1989). Antifungal, antiprotozoal and antiviral agents, p.137–161. In Biochemistry of rhizosphere with bacteria antimicrobial action, 4th ed. Chapman & Hall Ltd., New York.
- Ghazal, S.A.; Bream, A.S.; Abd el-Aziz, Z.K. and Ibrahim, S.Y. (2001). Preliminary studies on insecticidal activities of actinomycete strains propagated on solid and broth media using *Musca domestica* (Diptera: *Muscidae*). Meded. Rijksuniv Gent. Fak. Landbouwkd Toegep Biol. Wet. 66(2b):559-570.
- **Hopwood, D.A. (1997).** Genetic contributions to understanding polyketide synthases. Chem. Rev. 97: 2465-2497.
- **Katz, L. (1997).** Manipulation of modular polyketide synthases. Chem. Rev. 97: 2557-2576.
- Leadlay, P.F. (1997). Combinatorial approaches to polyketide biosynthesis. Curr. Opin. Chem. Biol. 1: 162-168.
- Lechevalier, H.A. and Waksman, S.A. (1962). The actinomycetes. III. Antibiotics of actinomycetes, p.430. The Williams & Wilkins Co., Baltimore.
- Lechevalier, M.P. (1988). Actinomycetes in agriculture and forestry, p.327–358. *In* M. Goodfellow, S. T. Williams, and M. Mordarski (ed.), Actinomycetes in biotechnology. Academic Press, Inc., New York.
- Levinson, Z.H. and Novon, A. (1969). Ascorbic acid and unsaturated fatty acids in the nutrition of the Egyptian cotton leaf worm. J. insect Physiol. 15: 591-595.
- Mahfouz H.T. and Sonya H. Mohamed (2002). Physiological, antagonistic and fingerprinting studies on some haloterant *Streptomyces* strains. Arab Journal of Biotechnology 5(1): 103-120.
- Mohamed, Sonya H. (1998). Role of actinomycetes in the biodegradation of some pesticides, p.151. Ph.D. Thesis, Agric. Microbiol., Dept. Agric. Microbiol., Faculty of Agric., Ain Shams University.
- Mohamed, Sonya H.; Abdel-Fattah, H.I.; Selim, Sh.M. and Sharaf, M.S. (2001). Identification and molecular studies on some halotolerant actinomycetes isolated from Sinai sandy soil. Arab Journal of Biotechnology 4: 179-196.
- Mohamed, Sonya H.; E.A. Saleh and M.M. Zaki (2005). Identification of eight halotolerant streptomycete isolates using a suggested numerical taxonomy. Arab Univ. J. Agric. Sci., Ain Shams Univ., Cairo 13(3): 641-668.

- Mohamed, Sonya H.; Sh.M. Selim and E.A. Saleh (2000). Taxonomical and biochemical studies on some halotolerant actinomycetes isolated from sandy soil in Egypt. Arab Univ. J. Agric. Sci., Ain Shams Univ., Cairo 8(1): 41-61.
- Mourad, A.K.; Saad, A.S.; Esawy, M.M. and Hassan, S.M. (2004). Influence of the nonsteroidal ecdysone agonist, tebufenozide, on certain biological and physiological parameters of the cotton leaf-worm, Spodoptera littoralis (Boisd.) (*Noctuidae*: Lepidoptera) in Egypt. Commun. Agric. Appl. Biol. Sci. 69(3):119-39.
- Ouyang, L., Tu, G., Gao, Y., Zhang, P. and Xie, X. (1993). Two insecticidal antibiotics produced by *Streptomyces nanchangensis*. J. Jiangxi. Agricul. Univ. 15: 148-153.
- Ouyang, L., Wan, S., Tu, G., Chen, X., Zhen, Y. and Gao, Y. (1984). A new species of *Streptomyces* producing insecticidal antibiotics. Chin. J. Microbiol. 24: 195-199.
- Pachlatko, J.P. (1998). Natural products in crop protection. Chimia 52: 29-47.
- Saroj K. Mishra; James E. Keller; James R. Miller; Rod M. Heisey; Muraleed-haran G. Nair and Alan R. Putnam (1987). Insecticidal and nematicidal properties of microbial metabolites. Journal of Industrial Microbiology and Biotechnology 2(5): 267-276.
- Shiomi, K.; Hatae, K.; Hatano, H.; Matsumoto, A.; Takahashi, Y.; Jiang, C.L.; Tomoda, H.; Kobayashi, S.; Tanaka, H. and Omura, S. (2005). A new antibiotic, antimycin Ag, produced by *Streptomyces* sp. K01-0031. J. Antibiot. (Tokyo) 58(1):74-78.
- Takahashi, S., Miyaoka, H., Tanaka, K., Enoktta, R. & Okazaki, T. (1993). Milbemycin all, al2, al3, al4, and al5, a new family of milbemycins from *Streptomyces hygroscopicus* ssp. aureolacrimosus, taxonomy, fermentation, isolation, structure elucidation and biological properties. J. Antibiot.49:1364-1371.
- Waksman, S.A. and Lechevalier, H.A. (1961). The actinomycetes. Vol. II-Classification, identification and description of genera and species. The Williams and Wilkins, Co., Baltimore, USA, p. 340.
- Xiong, L.; Li, J. and Kong, F. (2004). Streptomyces sp. 173, an insecticidal microorganism from marine. Lett. Appl. Microbiol. 38(1):32-37.
- Yuhui Sun, Xiufen Zhou, Jun Liu, Kai Bao, Guiming Zhang, Guoquan Tu, Tobias Kieser and Zixin Deng. (2002). Streptomyces nanchangensis, a producer of the insecticidal polyether antibiotic nanchangmycin and the antiparasitic macrolide meilingmycin, contains multiple polyketide gene clusters. Microbiology 148:361-371.

# نشاط تضادي ونشاط كمبيد حشري لبعض عزلات من الأستربتوميستات جمال عثمان ، صلاح مصطفى ، سونيه حموده محمد ،

ا: معهد بحوث الهندسة الوراثية الزراعية- مركز البحوث الزراعية- ص.ب. ١٢٦١٩ - جيزة-مصر.

٢: قسم الميكروبيولوجيا الزراعية- معهد بحوث الأراضي والمياه والبيئة- مركز البحوث الزراعية
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في هذه الدراسة تم استخدام ١٥ عزلة من الأستربتوميسس وهي -S01-S02-S3-S04-S05-S06 S07-S08-S09-S10-S11-S12-S13-S14-S15 معزولة من تربه مختلفة ومناطق جغرافية في مصر. وبعد إكثار العزلات خضعت لحصر لتقييم فعاليتها كعوامل ذات نشاط تضادي ضد بعض الفطريات الممرضة Fusarium oxysporum Aspergillus niger Rhizopus stolonifer . Helminthisporum gramenium واقه حشرية هي دودة ورق القطن المعروفة بـ Pelminthisporum gramenium. littoralis . وللوصول الى هذا الغرض تم تنميه العز لات على بيئة مرق النشا تحت ظروف رج على درجة حرارة ٢٨ درجة منوية لمدة ٦ أيام. ثم تبع ذلك اختبار القدرة التضادية ضد الفطريات سالفة الذكر للراشح المتحصل عليه من العز لات. وقد أوضحت الدراسة أن معظم العز لات قد تباينت في قدرتها التضادية. وكانت العزلة S08 نشطه ضد F. oxysporum ، A. niger ،R. nigricans بينما كانت العزلات S05 -S01 العزلة F. oxysporum، A. niger والعز لات S14 -S11 كانت نشطه ضد A. niger 'nigricans . وعلى الجانب الآخر لم تلاحظ أية قدرة تضادية ضد فطر الـ .Helminthisporum sp. وبالنسبة للنشاط التضادي ضد الحشرة فقد تم استخدام الراشح والخلايا المترسبه للعز لات. وقد ثبت أن الخلايا لبعض العز لات كانت أكثر نشاطا ضد حشرة دودة ورق القطن مقارنة بالراشح. وبصفة عامة فإن العزلات SOS - SOS - SOS - SOS أوضحت ۸۰، ۲۰۰، ۲۰، شببة موت ضد الحشرة على التوالي. كما تم تنقية البروتين من خلايا العزلة SO8 باستخدام تركيزات كبريتات الأمونيوم هي ٤٠، ٦٠، ٨٠ % ثم تبع ذلك تقدير الوزن الجزيئي للبروتين بواسطة التفريد الكهربائي SDS-PAGE وقد ثبت تواجد بروتين ذو وزن جزيئي ٤٠ كليودالتون ذو فعالية وصلت الى درجة ١٠٠% ضد حشرة دودة ورق القطن في أربعة أطوار حشرية من الأول الى الرابع مما يوضح إمكانية استخدام الأستربتوميسيس في الزراعة كمبيد حيوي.