

STIMULATION OF HEAT STRESS TOLERANCE OF BEAN PLANTS USING SA AND CPPU

Journal

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· ABSTRACT

Pot experiment was conducted in the Experimental Farm of Agricultural Botany Department, Faculty of Agriculture, Ain Shams Univ. during the two successive seasons of 2006 and 2007 to study the effect of Salicylic acid (SA at 10^{-2} , 4 x 10^{-2} μM) and N-(2-chloro-4pyridyl)-3-phenylurea (CPPU at 20 and 40 ppm), on bean plants (Phaseolus vulgaris) cv. Bronco grown under high temperature conditions (at average 35-40°C /22-25 °C day/night) as the growing period conducted from May to July in both seasons. Foliar application of SA and CPPU positively affected all growth characters. The best results were noted by CPPU at 20 ppm, followed by SA treatments. Moreover, the highest chlorophyll concentration was achieved by SA at 4 x 10⁻² µM and CPPU at 20 ppm. Furthermore, SA treatments as compared with CPPU were superior achieving the maximum number of pods per plant, pod fresh and dry weight. Meanwhile, both concentrations of SA significantly increased the concentration of total soluble protein. Most SA or CPPU or both treatments significantly improved carbohydrates, reduced and non-reduced concentration in seeds and reduced fibers percentage in pods especially with SA at 4 x 10⁻² µM. Exogenous application of SA or CPPU significantly suppressed superoxide dismutase (SOD) activities as compared to untreated plants. All the treatments caused a significant enhanced yield characters, except CPPU at 40 ppm.

Key words: *Phaseolus vulgaris*, SA, CPPU, Chlorophyll Concentration, Total Soluble Protiens, Carbohydrates, Reduced and non-reduced Sugars, Fibers, SOD.

INTRODUCTION

Bean plants (*Phaseolus vulgaris* L.) considered one of the important vegetable crops cultivated in Egypt for local market and for exportation. However, bean plants are relatively sensitive to environmental stresses that may occur in the field compared to most vegetable crops which negatively affect its growth, yield and even the quality of pods. Many investigations indicated that bean plants is very sensitive to different environmental stresses (abiotic stress) such as chilling (El-Tohamy et al., 2001), drought (Millar and Gardner, 1972; Halterlein, 1983; El-Tohamy et al, 1999) and heat stress (Dale, 1964). Freyer (1992) reported that the high incidence of temperature extremes caused by high concentration of ozone and the other pollutants subjected plants to oxidative stress which in turn negatively affects plant growth, productivity and quality. Two minutes of heat shock to the roots of *Phaseolus vulgaris* L. reduced cytokinin levels in both shoots and roots (Itai et al., 1973).

SA (Salicylic acid) belongs to plant phenolics group considered as a hormone-like endogenous regulator. SA affected several physiological processes either promoter or inhibitor depending on its concentration, plant species, development stages and environmental conditions (El-Mergawi and Abdel Wahed 2004). It plays an important role in the regulation of plant growth and development, seed germination, fruit yield, glycolysis, flowering and heat production in thermogenic plants (Raskin, 1992; Klessig and Malamy, 1994). Exogenous application of SA could regulate the activities of antioxidant enzymes and increase plant tolerance to the biotic and abiotic stress (He et al., 2002; Yalpani et al. 1994 and Szalai et al. 2000). Several investigators reported a positive effect of SA on photosynthesis and plant growth under stress. Treatment of bean and tomato plants with SA increased their tolerance against heat, chilling and drought stress (Senaratna at al., 2000).

CPPU (N-(2-chloro-4-pyridyl)-3-phenylurea) is a substance derived from phenylurea with cytokinin activity. It has a higher activity level than most adenine based cytokinins and metabolized even more slowly and thus last longer (Kapchina-Toteva and Stoyanova 2004). It influences cell division, morphogenesis in various species; increases fruit set in watermelon and persimmon and increases fruit size in many crops (Hayata, et al., 1995).

Cytokinins are antagonistic to many ABA mediated physiological responses such as stomatal closure, leaf senescence, leaf abscission, fruit abscission and seed germination inhibition (Kapchina-Toteva et al., 2000). However, to reduce senescence and improve retail holding, cytokinins could be applied pre-shipping to combat senescence.

Superoxide dismutase (SOD), within a cell, constitute the first line of defense against reactive O2 species (ROS) which may be due to a combination of the influences of cellular location of the enzyme and upstream sequences in the genomic sequence (Alscher et al 2002).

The aim of this study is to investigate the response of bean plants grown under heat stress to exogenous application of SA and CPPU.

MATERIALS AND METHODS

Pot experiment was conducted in the Experimental Farm of Agricultural Botany Department, Faculty of Agriculture, Ain Shams Univ. Three seeds per pot of bean (*Phaseolus vulgaris* L. cv. Bronco) were sown in 18th and 20th May in the two successive summer seasons of 2006 and 2007 respectively in PVC pots (25 cm in length and diameter) filled with 4 kg clay loamy soil (37% clay 36% silt and 27% sand), with 7.91 pH (organic matter 0.92%, Na⁺ 5.4, K⁺ 0.50, Ca⁺⁺ 4.00, Mg⁺⁺ 1.10, Cl⁻ 1.50, SO₄⁻⁻ 5.80 and HCO₃⁻⁻ 3.70 meq/1 for cations and anions). Thinning was performed after 1 weak from germination leaving one plant per pot. Basal fertilization was performed as following; Phosphorous as calcium super phosphate (15.5% P₂O₅) was mixed with soil before sowing at the rate of 3 g/pot. Nitrogen as ammonium nitrate (33.5% N) at the rate of 1.5 g/pot and potassium as potassium sulfate (48% K₂O) at the rate of 1 g/pot were added twice at 15 and 30 days after sowing. Other recommended agriculture practices for bean cultivation was followed.

Pots were arranged in complete randomized design with four replicates. Each replicate consisted of four pots with one plant per pot.

Bean seeds were sown at the rates of 3 seeds to each pot.

Plants were foliar sprayed two times (30 and 45 days from sowing) with CPPU at 20 and 40 ppm, salicylic acid at 10⁻², 4x10⁻²µM and control plants were sprayed with distilled water until complete covering of the plant foliage. Tween 20 at 0.05 ml/L was used as wetting agent. Temperature ranged 35-40°C /22-25 °C day/night

during the course of experiment and Relative humidity was approximately 90%. Samples were taken at 55 days after sowing for growth measurements (plant height, leaves number, shoots and roots fresh and dry weight) and chlorophyll analysis from each treatment and after 70 days for yield measurements (pods number, pods fresh and dry weights) and chemical analysis.

Chemical Analysis:

Chlorophyll Determination:

Chlorophyll a (Chl a), Chlorophyll b (Chl b) and total chlorophyll (Ch) concentration were determined in the second leaf (fully expanded leaves) from shoot tip. Samples of 0.1 g leaveswas ground and extracted with 5 mL of 80% (v/v) acetone in the dark according to the methods described by Holder (1965). The mixture was filtered then the absorbance values at 645 and 663 nm were measured. Using Jenway 6105 UV/VIS Spectrophotometer, concentrations of Chl a Chl b and Ch were estimated by the equations of Arnon (1949) as follows:

• C a+b = 20.2 D 645 + 8.02 D663

C: chlorophyll D: absorbance

Total Soluble Protein Determination:

Total soluble protein was determined (in a dry matter basis) in seeds using the method of Bradford (1976). The soluble protein concentration was calculated from the standard curve according to Read and Northcore (1931).

Total Carbohydrates Determination:

For extraction, one gram of seeds sample was randomly taken and added to 30 ml HCL 2N. Tubes were placed in boiling water bath for 6 h. after cooling, samples transferred into calibrated flasks (100 ml). Total carbohydrates were estimated by the alkaline potassium ferricyanide method (Shales and Schales, 1945).

Total Soluble Sugars Determination:

For extraction, one gram of seeds sample was ground in a mortar with ethanol 80% for 3 times. The extracts were combined and evaporated till dryness. The dried film was dissolved in 50 ml of 10

% aqueous isopropanol. Determination of total soluble sugars, reduced and non-reduced sugars were carried out according to the method by Shales and Schales (1945).

Fiber analysis:

The method of Van Soest and Wine (1968) was used to determine fibers in samples using Dosi-fiber extractor.

Superoxide Dismutase Determination:

Activity of SOD was determined by measuring its ability to inhibit the photoreduction of nitro blue tetrazolium (NBT) according to the methods of Giannopolitis and Ries (1977). The reaction solution (3 mL) contained 50 μ mol NBT, 1.3 μ mol riboflavin, 13 mmol methionine, 75 nmol EDTA, 50 mmol phosphate buffer (pH 7.8) and 20 to 50 μ L enzyme extract. The reaction solution was irradiated under a blank of fluorescent lights at 75 μ mol m-2 s-1 for 15 min. The absorbance at 560 nm was read against the blank (non-irradiated reaction solution) with a spectrophotometer (Spectronic Instruments, Rochester, NY). One unit of SOD activity was defined as the amount of enzyme that inhibited 50% of NBT photoreduction.

Statistical Analysis:

Data were subjected to statistical analysis of variance and were calculated according to SAS (1996).

RESULTS AND DISCUSSION

Growth parameters

Foliar application of SA and CPPU at 20 ppm significantly enhanced most of plant growth characters of bean plants. However, CPPU at 40ppm didn't give any significant increase comparing with untreated plants (Table, 1).

Maximum values of plant height, leaf numbers, and shoot fresh and dry weights were obtained by CPPU at 20 ppm followed by SA at $4 \times 10^{-2} \, \mu M$, and then SA at $10^{-2} \, \mu M$, these effects were significant except for shoots fresh weight with SA at 10^{-2} which failed to reach the level of 5 % significance. Meanwhile, roots fresh and dry weights showed the highest value with SA treatments. The same trend was found by Martín-Mex *et al* (2005) who reported that SA affected plant size, number of leaves and flowers of African violet plant. CPPU at 40

ppm obviously inhibited plant growth. These results were in agreement with Salisbury (1985), who reported that, high concentration of cytokinins can be inhibitory to shoot and root growth. On the other hand, the stimulatory effect of CPPU at 20 ppm can be explained by the major physiological function of cytokinins, which is promoting cell division and elongation (Salisbury, 1985).

Pod Yield and yield components

As shown from Data presented in Table (2), SA at 4 x 10⁻² recorded the highest values for all yield components (pod number per plant and pods fresh and dry weight) followed by SA treatment at 10⁻² µM and CPPU at 20 ppm. Similar findings were obtained by Cleland and Ajami, (1974), who mentioned that salicylic acid (SA) when applied to herbaceous plants, affected many physiological processes such as flowering induction which in turn increased yield components. On the other hand, CPPU treatment at 40 ppm didn't reach the values of untreated plants. That result may due to the reduction in vegetative growth caused by this treatment.

Table (1): Effect of SA and CPPU on growth characters of bean plants during two seasons of 2006 &2007

plant height (cm)	leaves No	shoot FW (gm)	shoot DW (gm)	root FW (gm)	root DW (gm)	plant height (cm)	Leaves No	shoot FW (gm)	shoot DW (gm)	root FW (gm)	root DW (gm)
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10.63	9.00	7.40	4.45	2.65	1.10	10.13	8.75	8.30	5.65	2.78	1.33
12.38	11.00	8.78	6.35	3.60	2.10	13.95	11.75	9.83	7.40		2.05
13.25	12.50	10.70	8.45	2.55	1.15	15.45	13,75	12.15	9.78		1.63
15.75	14.50	16.28	9.25	3.28	1.60	19.25	14.50	15.48	10.00		2.15
10.75	11.25	9.55	7.23	1.95	0.93	12.83	10.50	10.53	8.00		1.30
3.45	1.54	3.53	1.05	0.13	0.09	3.49	2.18	3.76	0.97	0.15	0.06
	height (cm) 10.63 12.38 13.25 15.75 10.75	height leaves (cm) No	height leaves FW (gm) 1" seas 10.63 9.00 7.40 12.38 11.00 8.78 13.25 12.50 10.70 15.75 14.50 16.28 10.75 11.25 9.55	height (cm) leaves No (gm) (gm) (gm) FW (gm) (gm) DW (gm) 1" season 1" season 4.45 12.38 11.00 8.78 6.35 13.25 12.50 10.70 8.45 15.75 14.50 16.28 9.25 10.75 11.25 9.55 7.23	height (cm) leaves (pm) FW (pm) DW (pm) FW (pm) 1" season 10.63 9.00 7.40 4.45 2.65 12.38 11.00 8.78 6.35 3.60 13.25 12.50 10.70 8.45 2.55 15.75 14.50 16.28 9.25 3.28 10.75 11.25 9.55 7.23 1.95	height (cm) leaves (pm) FW (pm) DW (pm) FW (pm) DW (pm) 1" season 10.63 9.00 7.40 4.45 2.65 1.10 12.38 11.00 8.78 6.35 3.60 2.10 13.25 12.50 10.70 8.45 2.55 1.15 15.75 14.50 16.28 9.25 3.28 1.60 10.75 11.25 9.55 7.23 1.95 0.93	height (cm) leaves (pm) FW (pm) DW (pm) FW (pm) DW (pm) height (p	height (cm) leaves (cm) FW (gm) DW (gm) FW (gm) DW height (cm) Leaves (cm) 10.63 9.00 7.40 4.45 2.65 1.10 10.13 8.75 12.38 11.00 8.78 6.35 3.60 2.10 13.95 11.75 13.25 12.50 10.70 8.45 2.55 1.15 15.45 13.75 15.75 14.50 16.28 9.25 3.28 1.60 19.25 14.50 10.75 11.25 9.55 7.23 1.95 0.93 12.83 10.50	height (cm) leaves (pm) FW (pm) DW (pm) FW (pm) DW height (pm) Leaves (pm) FW (pm) Leaves (pm) FW (pm) FW (pm) L	height (cm) leaves (pm) FW (pm) DW (pm) FW (pm) DW height (pm) Leaves (pm) FW (pm) DW (pm) Shoot shoot	height (cm) leaves (pm) FW (pm) DW (pm) FW (pm) DW height (pm) Leaves (pm) FW (pm) DW FW (pm) FW (pm) PW (pm) </td

Table (2): Effect of SA and CPPU on yield components of bean plants during two seasons of 2006 & 2007

Treatments	Pod No/plant	pod FW/plant (gm)	pod DW/plant (gm)	pod No/plant	pod FW/plant (gm)	pod DW/plan (gm)
		1st season			2 nd season	
control	1.75	3.78	2.10	2.00	3.95	2.20
SA 10 ⁻² μm	2.75	5.50	3.35	3.25	5.35	3.85
SA 4× 10 ⁻² μm	4.50	5.73	3.95	5.00	6.25	4.80
CPPU 20 ppm	2.00	4.28	2.75	3.25	5.00	2.85
CPPU 40 ppm	1.50	3.10 ^	1.70	5.50	4.78	3.45
LSD	0.50	0.39	0.26	0.66	0.22	0.30

Chlorophyll concentration

The superiority of chlorophyll concentration observed with SA at $4x10\text{-}2~\mu$ M and CPPU at 20ppm. Meanwhile, SA at $10\text{-}2~\mu$ M and CPPU at 40 ppm didn't give any significant increase (Table 3).These results were in agreement with those of Türkyılmaz et al., (2005 on bean and Zhou et al., (1999) on corn plants. They found that, foliar spray with salicylic acid increased Chl a, Chl b and other photosynthetic pigments under normal field conditions.

It is also possible that the anti-stress substances were effective in maintaining the membrane integrity to reduce the leakage of electrolyte forough its positive effect on the antioxidant enzymes system as has been demonstrated by Pinhero and Fletcher (1994) in corn seedlings. It is generally known that most of environmental stresses have in common a similar mechanism in affecting plant growth and performance. Under stress conditions, the generation of free radicals and low nutrient uptake are believed to be the main cause for damaging and dis-functioning of plant cells (Bohnert et al., 1995; Sreenivasulu et al., 2000). Regarding CPPU, it could be noticed that with low concentration (20 ppm) chlorophyll concentration

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water deficit and high temperature stress. Cytokinins application (ZR) retarded the decline in photosynthetic rate for plants exposed to prolonged periods of high temperature stress, which could have been due to the effects of ZR in suppressing the decrease in photochemical efficiency and chlorophyll degradation (Liu and Huang, 2000). In contrary, Carimi et al (2004) mentioned that, the cytokinin 6-benzylaminopurine (BAP), if added at high dosage to plants and cultivated cell suspensions of Arabidopsis thaliana and Daucus carota, induce programmed cell death by accelerating senescence.

Table (3): Effect of SA and CPPU on chlorophyll (chl a, chl b and total chlorophyll) contents (mg/ml) in bean leaves during two seasons of 2006 &2007

	Chl a	Chl b	total chlorophyt	ca	cb	total chlorophyl
Treatments		1st season	•		2 nd season	
control	0.005	0.010	6.58	0.004	0.010	6.89
SA 10 ⁻² μm	0.006	0.013	9.52	0.005	0.012	8.80
SA 4× 10 ⁻² μm	0.006	0.013	11.19	0.006	0.013	10.96
CPPU 20 ppm	0.007	0.015	10.39	0.007	0.016	10.19
CPPU 40 ppm	0.004	0.009	9.71	0.004	0.010	9.96
LSD	0.00014	0.00047	0.99	0.00024	0.00061	1.2

Total soluble protein

It was clearly shown that CPPU at 20ppm and both concentrations of SA treatments significantly increased total soluble protein concentration as compared with untreated plants (Table 4). However, opposite trend was noticed with CPPU at 40ppm treatment which didn't give any significant increment as compared with control plants. Similar trend was obtained in both tested seasons. Such results may be attributed to proteins produced in plants in response to biotic and abiotic stress and many of these proteins are induced by phytohormones such as salicylic acid (Hoyos and Zhang 2000). Regarding plants exposure to high CPPU concentration, stressed plants recorded less of total soluble protein concentration than control plants.

Table (4): Effect of SA and CPPU on total soluble protein concentration (mg/g) in bean seeds during two seasons of 2006 & 2007.

T44-	Pro	tein	
Treatments —	1st Season	2nd season	
Control	53.69	55.12	
SA 10 ⁻² μm	61.95	63.61	
SA 4 ×10 ⁻² μm	62.44	65.10	
CPPU 20 PPM	57.76	59.22	
CPPU 40 PPM	52.8	53.95	
LSD	6.24	6.55	

Total soluble sugars and carbohydrates:

Table (5) demonstrated seeds concentrations of reduced and nonreduced sugars and total carbohydrates in response to SA and CPPU treatments. It could be concluded that both SA treatments increased concentrations of reduced and non-reduced sugars and total carbohydrates followed by treatment of CPPU at 20 ppm. Khodary, (2004) suggested that SA treatment of stressed plants could stimulate their tolerance via accelerating their photosynthesis performance and carbohydrate metabolism. Concerning CPPU at 40 ppm, seeds concentration of reduced and non-reduced sugars and total carbohydrates was less than untreated plants which explained by Häggman and Haapala (2006) who found that very high amount of 6-Benzylaminopurine caused a great increase in amylase activity which was accompanied by a relatively high concentration of starch in the chloroplasts (leaf). The induction of amylase activity was thus not high enough to diminish the starch concentration. This is supposed to be due to the lack of a sink organ (seeds) since the growth of roots and shoots was greatly inhibited in concentrated solutions.

Table (5): Effect of different concentrations of SA and CPPU on total soluble carbohydrate, total soluble sugar, reduced sugar and non-reduced sugar contents (mg/g FW) in bean seeds during two seasons of 2006 & 2007.

Treatments	Total carbohydrates	Total soluble sugars	Reduced sugars	Non-reduced sugars	Total carbohydrates	Total soluble sugars	Reduced sugars	Non-reduced sugars
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control	15.38	5.44	1.90	3.54	16.52	5.13	1.95	3.18
SA 10 ⁻² µ M	28.71	8.76	3.06	5.70	30.61	10.34	3.22	7.11
SA 4×10 ⁻² μ M	31.79	12.65	4.71	7.95	34.38	14.61	4.88	9.72
CPPU 20 ppm	24.89	8.59	2.71	5.88	27.01	9.12	2.96	6.16
CPPU 40 ppm	20.88	5.79	2.03	3.75	22.56	6.18	2.12	4.07
LSD	12.04	2.56	0.39	0.98	14.52	4.27	0.42	2.05

Fibers concentration

The percentage of fibers in bean pods is one of the most important characters indicating the quality of bean pod (Silberngel et. al., 1991). It could be observed the decrease in fiber concentration with both SA treatments and with CPPU at 20 ppm compared with control plants (Table 6). Sistrunk et al, (1989) pointed that high temperatures affect fiber concentration in pods of bean plants. This effect of high temperature could be reversed by SA treatments and low concentration of CPPU. However, CPPU at high concentration gave the same trend of untreated plants.

Table (6): Effect of different concentrations of SA and CPPU on % Fiber in bean pods during two seasons of 2006 & 2007.

Treatments	% Fiber	% Fiber
	1 st season	2 nd season
control	16.4	16.57
$SA~10^{-2}~\mu M$	10.34	10.96
SA 4×10 ⁻² μM	9.45	10.03
CPPU 20 ppm	12.08	11.14
CPPU 40 ppm	15.65	14.12
LSD	2.05	1.98

Supercially Dismanus activity

Activity of SOD in all treated plants increased significantly when compared with untreated plants (Fig. 1). Applying CPPU increased SOD activity especially at 20 ppm. Meanwhile in plants treated with $4\times10^{-2}~\mu M$ SA, SOD activity gave the highest value followed by both concentrations of CPPU and SA at $10^{-2}~\mu M$.

Superoxide dismutase considered as key enzyme in the active-

SOD activity, this activity was elevated by exogenous SA and CPPU application, especially at the higher concentration of SA. Therefore, alleviation of heat injury in bean plants by SA and CPPU could be related to the maintenance of the scavenging ability of antioxidants to super oxygen free radicals at high temperatures (Yordanova and Popova 2007).

Results in this study suggested that SA and CPPU (at low concentration) could alleviate heat stress injury in bean plants (the most effective treatment was SA at 4 x $10^{-2}~\mu M$). They improved vegetative growth, yield characters, chlorophyll concentration, total soluble protein concentration, sugar and carbohydrates concentration and SOD activity in bean plants and minimize fiber concentration. SA protecting the endogenous anti-oxidant system which is often correlated with increased resistance to oxidative stress and/or controlling the level of free radicals within plant tissues (Sreenivasulu et al., 2000). SA and CPPU may mediate the acclimation of plants to environmental stress by maintaining active antioxidants, and interact with other cellular metabolites and environmental factors in the regulation of stress responses.

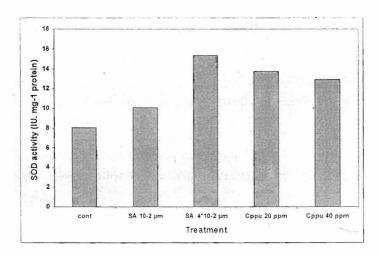


Fig.1: Superoxide dismutase (SOD) activities in bean plants as affected by foliar applications of Salicylic acid (SA) and N-(2-chloro-4-pyridyl)-3-phenylurea (CPPU)

REFERENCES

- Alscher R. G.; N. Erturk and L. S. Lenwood 2002. Role of superoxide dismutases (SODs) in controlling oxidative stress in plants. 53(372) 1331-1341.
- Arnon, D.I., 1949. Copper enzymes in isolated chloroplasts, polyphenoloxidasein *Beta vulgaris* plants. Plant Physiol, 24: 1-12.
- Asada, K. 1992. Ascorbate peroxidase a hydrogen peroxide scavengingenzyme in plants. Physiol. Plant. 85:235–241.
- Bohnert, H., E.D. Nelson and R.G. Jensen, 1995. Adaptation to environmental stresses. The Plant Cell, 7: 1099-1111.
- Bowler, C.,M.Van Montagu, and D. Inze. 1992. Superoxide dismutase and stress tolerance. Ann. Rev. Plant Physiol. Plant Mol. Biol. 43: 83–116.
- Bradford, M.M., 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein untitlizing the principle of protein Dye binding. Anal. Biochem, 72: 248-254.
- Carimi F., M. Terzi, R. De Michele, M. Zottini and F. Lo Schiavo 2004. High levels of the cytokinin BAP induce PCD by accelerating senescence Plant Science (166), 4, 963-969.
- Dale, J.E., 1964. Some effects of alterating temperature on the growth of French bean plants. Ann. Bot, 28:127-135.
- El-Mergawi, R.A. and M.S.A. Abdel-Wahed, 2004. Diversity in salicylic acid effects on growth criteria and different indole acetic acid forms among faba bean and maize. Egypt. J. Agron., 26:49-61.
- Elstner, E.F. 1982. Oxygen activation and oxygen toxicity. Ann. Rev Plant Physiol. 33:73–96.
- El-Tohamy, W.A., S.M. Singer, U.A. El-Behairy and A.F. Abou-Hadid, 2001. Effects of low tunnels, plastic much and mineral nutrient treatments on chilling tolerance of snap bean plants. Acta Horticulturae, 559: 127-134.
- El-Tohamy, W.A., W.H. Schnitzler, U.A. El-Behairy and S.M. Singer, 1999. Effect of long-term drought stress on growth and yield of bean plants (Phaseolus vulgaris L.). Journal of Applied Botany-Angewandte Botanik, 73: 173-177.
- Freyer, M.J., 1992. The antioxidant effects of thylacoid vitamin 8

- Giannopolitis, C.N., and S.K. Ries. 1977. Superoxide dismutase. I. responses to cytokinins involve the cyanide-resistant respiratory. Occurrence in higher plants. Plant Physiol. 59:309–314.
- Häggman J. and H. Haapala 2006. The Effect of 6-Benzylaminopurine on the Starch Metabolism of Stellaria media. Physiologia Plantarum (24) 3, 548 551.
- Halterlein, A.J., 1983. Bean. In: Teare, I.D. and Peet, M.M. (eds). Crop-Water Relations. John Wiley, New York.
- Hayata, y., y. Niimi and N. Iwasali. 1995, Synthetic cytokinin-1-(2-chloro-4-pyridyl)-3- phyenylurea (CPPU)- promotes fruit set and induces parthencocarpy in watermwlon. J.Amer.Soc.Hort.Sci. 120: 997-1000
- He, Y.L., Y.L. Liu, Q. Chen, and A.H. Bian. 2002. Thermotolerance related to antioxidation induced by salicylic acid and heat acclimation in tall fescue seedlings. (In Chinese, with English abstract.) J. Plant Physiol. Mol. Biol. 28:89–95.
- Hoyos, M. E. and S.Q. Zhang. 2000. Calcium-independent activation of salicylic acid- indeued protein kinase and a 40-kilodalton protein kinase by hyperosmotic stress. Plant Physiology 122: 1355-1363.
- Itai, C., A. Ben-zioni, and L. Ordin. 1973. Correlative changes in endogenous hormone levels and shoot growth induced by short heat treatments to the root. Physiol. Plant. 29:355–360.
- Kapchina-Toteva, V., H.J.Van Telgen and E. Yakimova. 2000. Role of phenylurea cytokinin CPPU in apical dominance release in invitro cultured *rosa hybrid* L. J. Plant Growth Regul. 19(2), 232-237.
- Kapchina-Toteva, V and D. Stoyanova. 2004. Effect of Cytokinins and Cytokinin Antagonists on in vitro Cultured *Gypsophila paniculata* L. Biologya plantarum. 46: 337-341
- Khodary S.E.A 2004. Effect of Salicylic Acid on the Growth, Photosynthesis and Carbohydrate Metabolism in Salt Stressed Maize Plants. International Journal of Agriculture & Biology, 1560–8530/06–1–5–8.
- Klessig, D.F. and J. Malamy, 1994. The salicylic acid signal in plants. Plant Mol. Biol., 26: 1439-1458.
- Liu, X., and B. Huang. 2000. Heat stress injury in relation to membrane lipid peroxidation in creeping bentgrass. Crop Sci. 40:503-510.

- Martín-Mex, R., E. Villanueva-Couoh, T. Herrera-Campos and A. Larqué-Saavedra, 2005. Positive effect of salicylates on the flowering of African violet., 103(4): 499-502.
- Millar, A.A. and W.R. Gardner, 1972. Effects of soil and plant water potential on the dry matter production of snap bean. Agron. J., 64: 559-562.
- Pinhero, R.G. and R.A. Fletcher, 1994. Paclobutrazol and ancymidol protect corn seedlings from high and low temperature stress. Plant Growth Reg., 15: 47-53.
- Raskin, I., H. Skubatz, W. Tang and B.J.D. Mense. 1990. Salicylic acid levels in thermogenic and non-thermogenic plants. Annals of Botany 66: 376-373.
- Raskin, I., 1992 .Role of salicylic acid in plants, Ann .Rev .Plant Physiol . and Plant Mol .Biol., 43, 439-463.
- Read, S.M. and D.H. Northcote, 1981. Minimization of variation in the response to different proteins of the coomassie blue G.dye binding assay for protein. Anal. Biochem., 116: 53-64.
- SAS, 1996. Statistical analysis system, SAS user's Guide: statistics. SAS Institute Inc. Editors, Cary, NC.
- Senaratna, T., D. Touchell, E. Bunn, K. Dixon, 2000. Acettyl salicylic acids) Aspirin (and salicylic acid induce multiple stress tolerance in bean and tomato plants, Plant Growth Regulation, 30, 157-161.
- Scandalios, J.G. 1993. Oxygen stress and superoxide dismutases. Plant Physiol. 101:7–12.
- Scandalios, J.G. 1994. Regulation and properties of plant catalases. p. 275-315. In C.H. Foyer and P.M. Mullineaux (ed.) Cau e of photoxidative stress and antelioration of defense system in plants. CRC Press, Borg Roton, bl...
- Shales, O., and S.S. Schales. 1945 A sample method for determination of glucose in blood. Arch. Biochem. 8:285.
- Silbernagel, MJ.; J. Wilhelmus and H.C. Jeremy, 1991. Snap bean production in the tropics: implications for genetic improvement (Chapter16).). UK by Redwood Press Ltd, Melksham, Wiltshire. pp.980.
- The to WA A D Generalez and VI Moor 1989 (Green beans

- salt-tolerant and salt-sensitive seedlings of foxtail millet (Setaria italica). Physiologia Plantarum, 109: 435-440.
- Szalai G, Tari I, Janda T, Pestenácz A, Páldi E (2000) Effects of cold acclimation and salicylic acid on changes in ACC and MACC concentrations in maize during chilling. Biol Plant 43:637-640.
- Türkyılmaz, B., L.Y. Aktaş and A. Güven. 2005. Salicylic acid induced some biochemical and physiological changes in *Phaseolus vulgaris* L. Science and Engineering Journal of Firat Univ. 17(2): 319-326.
- Van Soest, P. J., Wine, R. H., 1968. Determination of lignin and cellulose in acid-detergent fiber with permanganate. J Assoc of Anal Chem, 51:780-785.
- Yalpani N, Enyedi, AJ, León, J, Raskin I (1994) Ultraviolet light and ozone stimulate accumulation of salicylic acid and pathogenesis related proteins and virus resistance in tobacco. Planta 193:373-376.
- Yordanova R. and L. Popova, 2007 Effect of Exogenous Treatment With Salicylic Acid on Photosynthetic Activity and Antioxidant Capacity of Chilled Wheat Plants Gen. Appl. Plant Physiology, 33 (3-4), 155-170.
- Yordanov, I., T. Tsonev, V. Goltsev, M. Merakchiiska-Nikolova, and K. Georgieva. 1997. Gas exchange and chlorophyll fluorescence during water and high temperature stresses and recovery. Probable protective effect of carbamide cytokinin 4-PU30. Photosynthetica (Prague) 33:423–431.
- Zhou X.M., A.F. MacKenzie, C.A. Madramootoo and D.L. Smith. 1999. Effects of stem-injected plant growth regulator, with or without sucrose, on grain production, biomass and photosynthetic activity of field-grown corn plants. J. Agronomy and Crop Science 183: 103-110.

تشجيع نباتات الفاصوليا على تحمل الإجهاد الحراري باستخدام حامض الساليسيليك و CPPU

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ادت المعاملة بحامض الساليسيليك بالتركيزين (2 و 2) الى تحسين صفات النمو و زيادة من المحصول يليها معاملة CPPU بتركيز 20 جزء في المليون بينما خفضت معاملة CPPU بتركيز 2 و CPPU بتركيز 2 و CPPU جزء في المليون المحصول مقارنة بالنباتات غير المعاملة أدت معاملة SA بتركيز 2 و CPPU جزء في المليون الى زيادة البروتينات الكلية الذائبة الذائبة الكلوروفيل بينما ادت معاملة SA بكلا التركيزين الى زيادة البروتينات الكلية الذائبة و السكريات المختزلة ، السكريات الكلية الذائبة و الكربوهيدرات وخفضت تكوين الألياف مقارنة بمعاملة CPPU جزء في المليون . بينما لوحظ نشاط انزيم سوبر اوكسيد ديس ميوتيز في النباتات المعاملة بحمض الساليسيليك بتركيز 2 بتركيز 2 بتركيز 2 من المليون 3 ما المعاملة 4 (2 بالنباتات غير المعاملة 2 (2 بالنباتات غير المعاملة 2 (2 بالنباتات غير المعاملة 2

أوضحت نتائج الدراسة إلى أن حامض الساليسيليك و CPPU (التركيز المنخفض) ادوا الى تخفيف تاثير الإجهاد الحراري على الفاصوليا، وأن افضل معاملة كانت لحامض الساليسيليك بتركيز (4 × 10-2 ميكرومتر) من حيث تحسين النمو الخضري و المحصول و زيادة تركيز الكلوروفيل و البروتينات الكلية الذائبة و السكريات و الكربوهيدرات ونشاط انزيم سوبر اوكسيد ديس ميوتيز وانخفاض نسبة الألياف. و تقترح هذة الدراسة أن مقاومة النباتات المعاملة بحامض الساليسيليك و CPPU لارتفاع درجة الحرارة يعزى الى النباتات المعاملة بحامض الساليسيليك و للعوامل البيئية غير الملائمة.