

## **DIAZOTROPHY AND GROWTH OF BEANS (*PHASEOLUS VULGARIS*) GENOTYPES INOCULATED WITH RHIZOBIA AND LACTIC ACID BACTERIA**

**M.A. El-Howeity**

Environmental Studies & Research Institute (ESRI) , Minufiya Univ, Sadat City, Egypt

(Received: Sep. 2 , 2008)

---

**ABSTRACT:** *In a laboratory trial, the influence of inoculation with Rhizobium etli, Lactococcus lactis, and Rh. etli plus L. lactis, as well as, a control ( without bacterial inoculation) on root nodulation and plant growth of two genotypes of Phaseolus vulgaris was investigated. Results, obtained at 12-day plant age, revealed that inoculation with Rhizobium etli induced root nodulation, while Lactococcus lactis alone did not produce any nodules. Moreover, the co-inoculation with Rh. etli mixed with L. lactis exhibited salient superiority in enhancing the nodulation process and nitrogenase activity. Increases in number of nodules reached 18.4 and 45.3%, and in acetylene reduction activity (ARA) were 360 and 64.8% above the inoculation with Rh. etli alone for the two genotypes tested BAT477 and DOR364, respectively.*

*The results gained denoted that the bacterial inoculation increased shoot and root fresh and dry weights of the 12-day old plants, compared to the control. It was observed that the genotype BAT477 gave higher increases than those of DOR364 for such measures. Increases in shoot fresh weight, referring to, the control were (14.8, 6.1& 5.3 %) and (28.8, 7.6& 2.1 %) and those of shoot dry weights were (25.4, 24.5 & 21.5%) and (16.1, 14.8& 11.8 %) for the co-inoculation with both bacterial agents, for the bean genotypes BAT477 and DOR364, respectively. On contrast, DOR364 scored higher increases than BAT477 for root fresh and dry weights. The increases were more pronounced for the fresh weights, average increments, as compared with control, were (115, 86.4 & 5.9 % ) and (215.6, 155& 39.6 %) for the co-inoculation, Lactococcus lactis, Rhizobium etli, respectively .*

**Key words:** *Rhizobium etli, Lactococcus lactis, Co-Inoculation, Nodulation, Acetylene Reduction Activity , Phaseolus Genotypes .*

---

## INTRODUCTION

The beneficial effects of inoculating legumes with *Rhizobium* and *Bradyrhizobium* are well known and widely used in agriculture for crop improvement, due to their ability to fix molecular nitrogen, (Roughley et al., 1983). Co-inoculation of legumes with *Rhizobium* and plant growth promoting rhizobacteria (PGPR) has had an increasing attention in recent years. Plant growth –promoting bacteria are free-living in rhizosphere, rhizoplane, and phyllosphere, that under some conditions are beneficial for plants. PGPR affect plant growth in two different direct and indirect ways, for the metabolism of the plants by providing substances that are usually in short supply. These bacteria are capable of fixing nitrogen, solubilizing phosphorus and iron, and producing plant hormones, such as auxins, gibberellins, cytokinins and ethylene. Additionally, they improve plant tolerance to stresses such as drought, high salinity, metal toxicity, and pesticide load. A second way is biocontrol-PGPR, that indirectly improve plant growth by preventing the deleterious effects of phytopathogenic microorganisms (bacteria, fungi, and viruses) (Bashan, 2005). So, the benefits observed following inoculation with associative bacteria are mainly attributed to improved root development and enhanced water and mineral uptake, (Okon and Vanderliden, 1997). Mixed inoculation of *Azospirillum* with *Rhizobium* led to increase nodule stimulation and function, total number and weight of nodules, epidermal cell differentiation in root hairs, straw and grain yield and root surface area (Bashan, 2005). The lactic acid bacteria (LAB) family is composed of a heterogeneous group of Gram-positive, nonsporing, catalase and cytochrome negative, anaerobic or aerotolerant bacteria (Axelsson, 1998). Antimicrobial compounds produced by lactic acid bacteria (lactic and acetic acids, hydrogen peroxide and bacteriocins) play an important role in preventing the growth of food spoilage and food –borne pathogenic bacteria. Bacteriocins are of special interest due to their potential use as natural food preservatives (Klaenhammer, 1988). Bacteriocin is one of the important factors that affect proliferation of bacteria in rhizosphere as a cyst formation and siderophore and phytohormone production. Bacteriocins are proteic molecules synthesized for various lineages of Gram-positive and Gram-negative bacteria when exposed to stressful conditions. Bacteriocins have been characterized as molecules of high antimicrobial property even at low concentrations, provoking the microbial survival inhibition by antibiosis (Souza et al., 2005). Two bacteriocin-producing strains of *Lactococcus lactis* isolated from vegetables were active from pH 2 to 9 and inhibited species of *Listeria*, *Lactobacillus*, *Lactococcus*, *Pediococcus*, *Leuconostoc*, *Carnobacterium*, *Bacillus* and *Staphylococcus* (Uhlman et al., 1992). The

## Diazotrophy and Growth of Beans (*Phaseolus vulgaris*).....

potential application of antimicrobial –producing lactic acid bacteria as biopreservatives of ready to use vegetables is suggested (Vescovo et al., 1996).

In this study, we evaluated the effect of inoculation with *Rhizobium etli* individually or mixed with *Lactococcus lactis* on diazotrophy represented by nodulation status (number and dry weight of plant root nodules) and nitrogenase activity, as well as shoot and root fresh and dry weights, of two *Phaseolus* genotypes under aseptic laboratory conditions.

## MATERIALS AND METHODS

### Bacterial strains

*Rhizobium etli* "CNPAF512 (Embrapa, Brazil)" isolated from root nodules of *Phaseolus vulgaris* plants and *Lactococcus lactis* isolated from coal rhizosphere cabbage, Oost-Vlaender, Belgium, (a PGPR that produces phytohormones and promotes lateral root and root hair formation) were employed in this study.

### Media

Trypton yeast extract (TY) medium, for *Rhizobium etli* (Berger, 1974) was introduced into one liter of distilled water (5g of trypton and 3g of yeast extract). After autoclaving the medium at 121°C,  $\text{CaCl}_2$  was added to each of a final concentration of 7mM. MRS medium was used for *Lactococcus lactis*.

### Plant hydroponic growth medium

The culture medium (based on Snoeck, 2001) contained the following: 0.70mM  $\text{MgCl}_2$ , 1.06mM  $\text{CaCl}_2$ , 0.48μM  $\text{MgSO}_4$ , 20.32 μM FeHEDTA, 14.08 μM  $\text{MnSO}_4$ , 0.198 μM  $\text{CuSO}_4$ , 0.602 μM  $\text{ZnSO}_4$ , 10.003 μM  $\text{H}_3\text{BO}_3$  and 0.02744 μM  $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24}$ . This medium was used for growing plants in a growth chamber.

### Preparation of cultures

*Rhizobium etli* (CNPAF512) stored at -80 ° C was grown for two days at 30 ° C on agar plates with Ty medium. The plated cells were suspended in 5 ml of Ty medium and cultured overnight at 30 ° C on a shaker. The optical density (OD) of the overnight culture was measured at 595nm using a spectrophotometer and OD was adjusted to 0.4, 1ml of the cell suspension having OD of 0.4 was centrifuged at 6000 rpm for 5 minutes, then the supernatant was discarded and the cells were washed two times with 1ml 10M  $\text{MgSO}_4$ . The supernatant was discarded and the cells were resuspended in 1ml 10M  $\text{MgSO}_4$ . The washed cells suspension was diluted to 10-12 in 10mM  $\text{MgSO}_4$ , and 200μl of the bacterial suspension,

corresponding to approximately  $10^3$  to  $10^4$  cells was used to inoculate each plant growing in plate as single inoculum and 100 $\mu$ l of the bacterial suspension from *Rh.etli* and *L. lactis* as mixed inoculum.

### **Plant materials and growth conditions**

*Phaseolus vulgaris* is a largely consumed grain legume. BAT477 and DOR364, used in this study, are two Meso-American phaseolus beans genotypes. Plant seeds were sterilized for 1min in 100%ethanol, then in 15% NaOCl for 12 min and finally rinsed 10 times in a sterile distilled water . Seeds were germinated on 10% water agar. Two days old seedlings were transferred to each of plates, (50 ml medium), and bottles (250 ml ) for N<sub>2</sub>-ase-activity assay. The plates were sealed with parafilm to avoid contamination and by sterile forceps and knife, then small holes were made, whereas the bottles were wrapped with aluminum foil to maintain darkness in the rooting environment. At 12-day plant age fresh and dry weights of the growing plants were determined and 21-dayplant age for N<sub>2</sub>-ase-activity assay.

## **RESULTS AND DISCUSSION**

### **Diazotrophy**

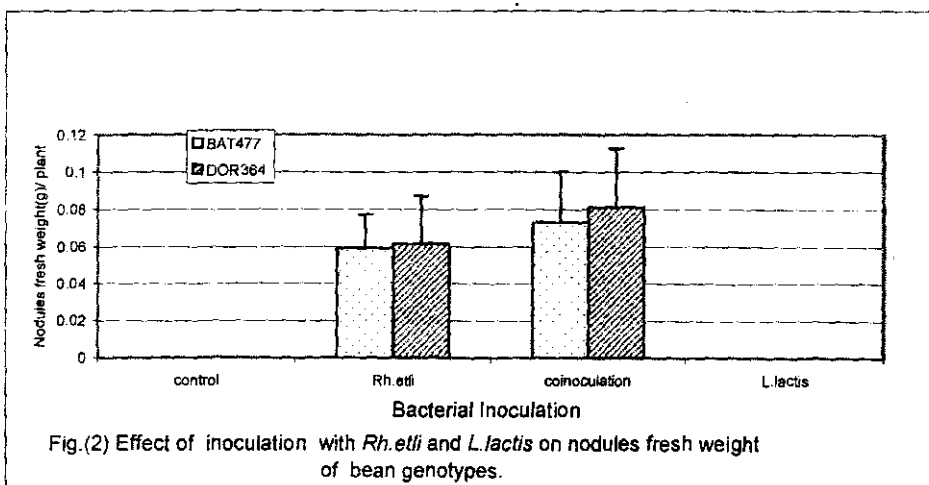
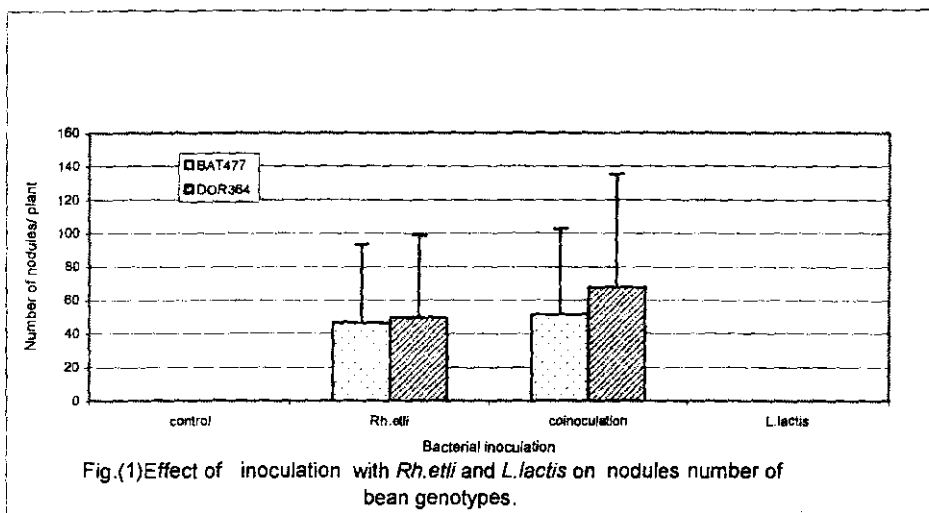
#### **A) Number and dry weight of root nodules of phaseolus beans**

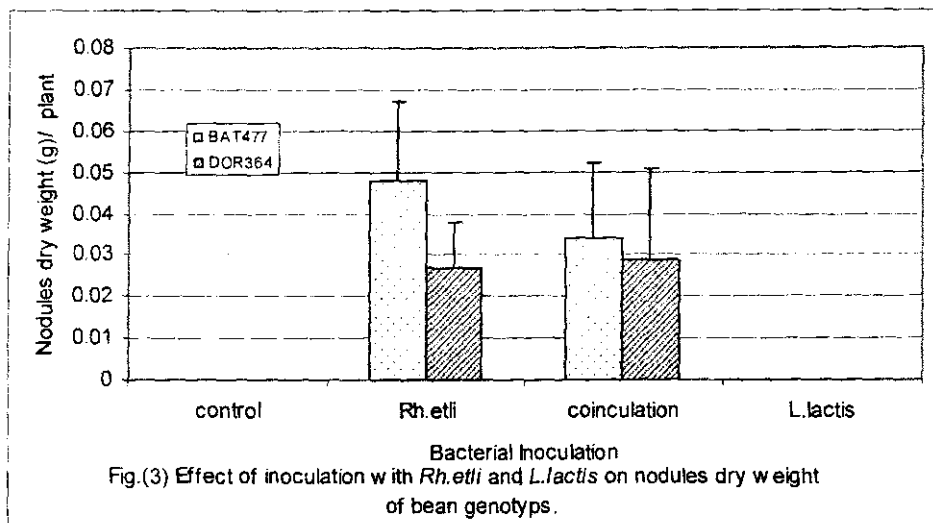
Numbers and dry weights of nodules developed on *Phaseolus* roots are illustrated in Figs.(1,2 and 3). It was apparent that nodulation status was affected by *Rhizobium etli* inoculation. Co-inoculation with *Rh. etli* and *L.lactis* led to increases in number and dry weight of nodules, while the control treatment, inoculated with *L. lactis* alone did not produce any nodules. The highest values of nodulation aspects were attained for the genotype DOR 364, compared to BAT477 .Increases in the number of nodules reached up to 18.4 and 45.3 % over those inoculated with *Rhizobium* alone. For instance, the values of nodular tissues accumulated due to sole inoculation with *Rhizobium* and co-inoculation with *Rh. etli* plus *L. lactis* were 365& 379 and 380 & 389 mg/plant for BAT477 and DOR364, plant genotypes, respectively. Also, the numbers of nodules were 43.4&46.5 and 51.4 & 67.6, respectively. These results demonstrated that inoculation of beans with *Rhizobium* enhanced nodulation process. However, co inoculation of *Phaseolus vulgaris* with *R.etli* together with *L. lactis* might be an efficient and effective approach for magnifying nodulation. Indeed, *L. lactis* may increase the nodulation status by widening the root surface area leading to create more infection sites on the root system( Srinivasan *et al.*,1997). Root exudates also provided the chance for a better nodulation

## Diazotrophy and Growth of Beans (*Phaseolus vulgaris*).....

process. On the other hand, *L.lactis* produced bioprotecting substances that could enhance plant healthy resulting in improved nodulation.

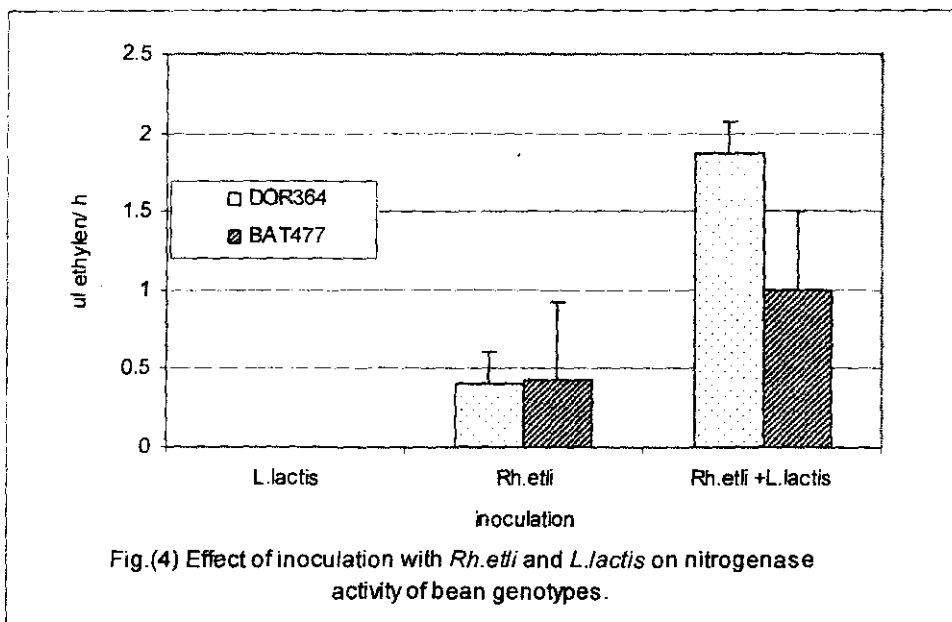
Similar finding, confirming that the co-inoculation increased nodulation of legume roots, was observed by Hassanein and Mekhemer(2003), El-Howeity (2004) and Abdel-Wahab et al.(2006) who reported that co-inoculation of some legumes with rhizobacteria exerted beneficial effects on nodulation of pea, faba bean, and peanut.





## B) Acetylene Reduction Assay (ARA)

N<sub>2</sub>-Fixation capacity was evaluated by ARA and presented in Fig. (4). Plants of both genotypes inoculated with *Rh. etli* alone or *Rh. etli* plus *L. lactis* showed an acetylene reduction, while the plants un-inoculated or inoculated with *L. lactis* alone were negative to ARA. Co-inoculation treatments gained higher values of ARA for the two genotypes, where gave 0.4058 & 0.4236 and 1.868 & 0.6983  $\mu\text{l/h}$  with DOR364 and BAT477 inoculated with *Rhizobium* only and *Rh. etli* + *L. lactis*, respectively. Increases in ARA reached up to 360 and 64.8% over *Rhizobium* alone for the two genotypes BAT477 and DOR364, respectively. These increases in ARA might be due to increases in number of nodules and dry weight of roots. Lactic acid bacteria could also stimulate the growth and activity of rhizobia. Sindhu et al. (1999) observed that a culture supernatant of *Pseudomonas* isolates contained a fluorescent compound which influenced the root flavonoid content of green gram (*Vigna radiate*) and concluded that the production of flavonoid-like compounds by plant roots augmented nodule formation by *Bradyrhizobium*. Chebotar et al. (2001) found that co-inoculation of *P. fluorescens* 2137 and *B. japonicum* A1017 increased the colonization of *B. japonicum* A1017 on soybean roots, nodule number, and acetylene reduction activity (ARA) at 10 and 20 days after inoculation. Thus, we suggest that *Lactococcus lactis* produced compounds which stimulated the growth of *Rhizobium etli*.



### Shoot and root fresh and dry weights

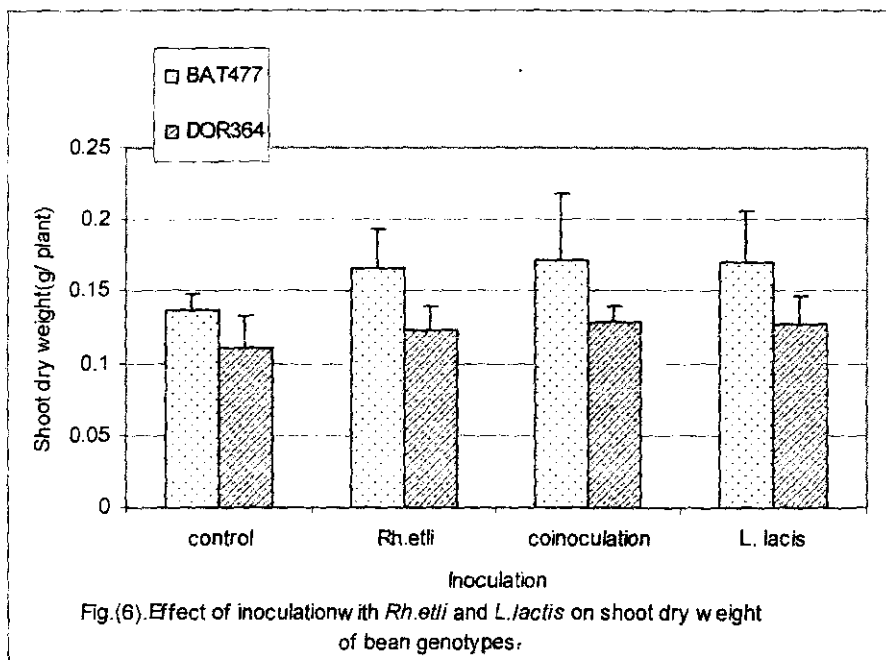
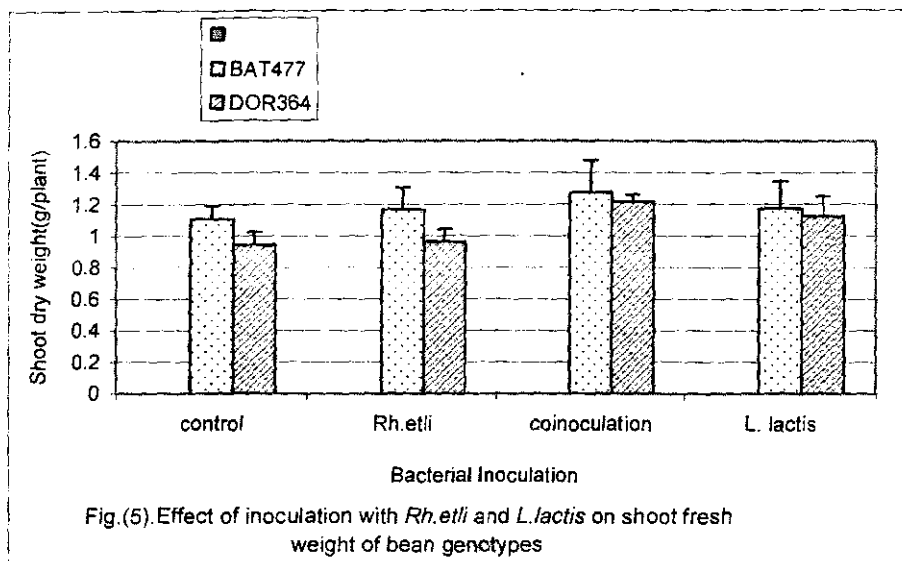
Results illustrated in Figs.(5 thru 8) show the effect of inoculation with *Rh. etli*, *L. lactis* and co-inoculation with both agents on the growth of the two genotypes of *Phaseolus vulgaris* BAT477 and DOR 364, under aseptic conditions. The results revealed that inoculation caused increases in shoot and root fresh and dry weights of both plant genotypes, but the effect was clearer with BAT 477 than with DOR 364 for shoot fresh and dry weights. However, the genotype DOR 364 showed higher values in root fresh and dry weights, compared to BAT477. The increases occurred in the fresh and dry weight of shoot due to aole rhizobial inoculation were 5.3 and 21.5 for c.v DOR 364, respectively and they were 2.1 and 11.8 % for c.v BAT 477 over control. The corresponding values attained as a result of inoculation with *L. lactis* were 6.1&24.5 % and 7.6 & 14.8 %, respectively.

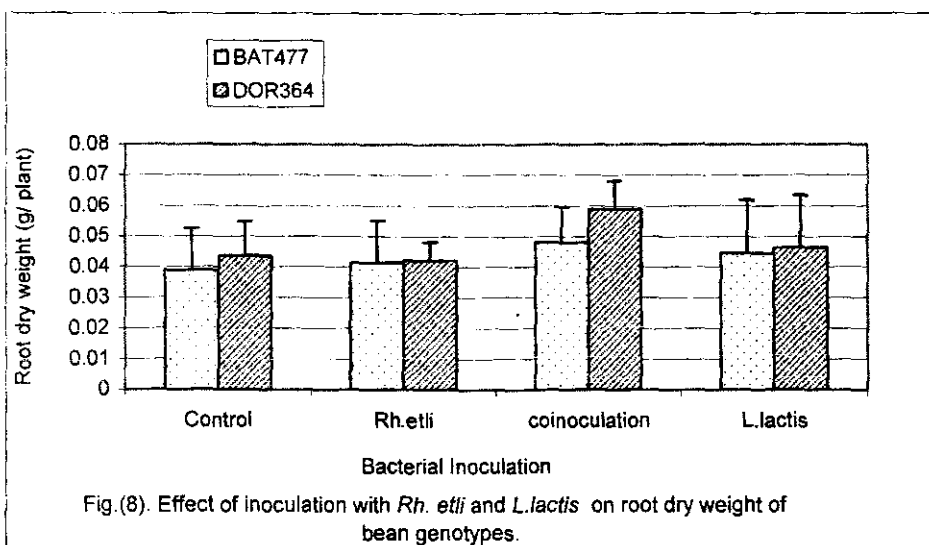
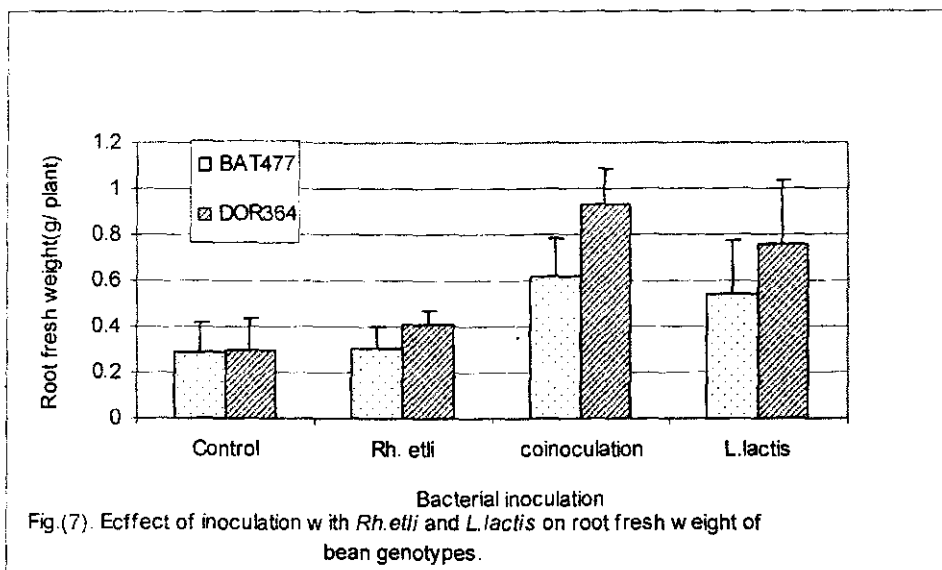
Co-inoculation with *Rh. etli* and *L. lactis* gave higher increases in the shoot fresh and dry weights, as compared with every single inoculation and control. Increases in the shoot fresh weight were(up to 14.8 &28.8%) and for the shoot dry weight (up to24.4& 16.1) for BAT 477 and DOR 364,bean genotypes, respectively. On the other hand, increases in the root fresh and dry weights were clearer for DOR364, compared to the BAT 477 cultivars.

These increases reached (up to 5.6& 39.6%) over the control for the root fresh weight and (up to 6.2& 2.4%) for the root dry weight under inoculation with *Rh. etli*. *Lactococcus lactis* gave increases in the root fresh weight (up to 85.8 & 155.9%) and its dry weight (up to 14.4& 11.7%). Co inoculation with *Rh. etli* plus *L. lactis* led to higher increases in the root fresh and dry weights, reaching (up to 114.7& 215.6%) over the control and for the root dry weight (up to 23.7& 43.4%) for BAT 477 and DOR 364, respectively. The promotive effect of *L.lactis* on the plant vigour might be elucidated by its ability to produce efficient bioactive substances, which acting to stimulate and regulate the plant growth. These growth promoting substances comprised several bacterial metabolites such as hydrogen peroxide, bacteriocins, siderophores, phytohormones and plant hormones regulators (Lowe and Arendt, 2004). Also, Ray (1996) noticed that lactic acid bacteria produce nisin and pediocin as examples of bacteriocins own antimicrobial spectrum which is able to exert inhibitory action on the growth of bacteria such as *Lactobacillus Plantarum*, *Pediococcus acidilactici*, *Leuconostoc mesenteroides*, *Listeria monocytogenes* and *Mirococcus luteus*. Souza et al.(2005) noted that bacteriocins are proteic molecules synthesized by various lineages of Gram-positive and Gram-negative bacteria when exposed to stressful conditions. Bacteriocins have been characterized as molecules of high antimicrobial property even at low concentrations, provoking the microbial survival inhibition by antibiosis. Antimicrobial compounds produced by lactic acid bacteria (lactic and acetic acids, hydrogen peroxide and bacteriocins ) play an important role in preventing the growth of food spoilage and food borne pathogenic bacteria . Bacteriocins are of special interest due to their potential use as natural food preservatives, (Klaenhammer, 1988). Such increases in the shoot and root fresh and dry weights might be due to amine groups produced by LAB. These results are in agreement with those obtained by El-Tarabily (2004) who found that growth promotion of beans (*Phaseolus vulgaris*) occurred by polyamine produced by isolate of *Streptomyces griseoluteus* . Polyamines are ubiquitous low molecular weight, aliphatic nitrogenous polycations that contain two or more amine groups. The suppressive effect of bacteriocin produced by lactic acid bacteria against various pathogenic bacteria has been reported by many investigators (Uhlman et al., 1992; Franz et al., 1997; Yildirim and Johnson 1998; Lowe and Arend 2004, Souza et al., 2005 and Sajur et al., 2007).



# Diazotrophy and Growth of Beans (*Phaseolus vulgaris*).....





## Diazotrophy and Growth of Beans (*Phaseolus vulgaris*).....

### Effect of bacterial inoculation on water absorption by the plants

Results presented in Table(1) demonstrated the effect of inoculation with *Rh.etli*, *L.lactis* and Co-inoculation with the two bacterial strains on water absorption by the two *Phaseolus vulgaris* genotypes BAT477 and DOR364 at 12-day age.

Table (1) Effect of the experimental treatments on absorption of water by the whole plants of phaseolus bean genotypes tested at 12-day age.

Treatments	Whole plant weights( g / plant)							
	BAT 477				DOR364			
	Fresh	Dry	Difference *	Moisture ** %	Fresh	Dry	Difference *	Moisture ** %
control	1.31	0.189	1.121	593.12	1.23	0.206	1.077	703.92
<i>Rh.etli</i>	1.47	0.206	1.264	613.59	1.37	0.165	1.205	730.30
<i>L.lactis</i>	1.71	0.213	1.497	702.81	1.76	0.172	1.588	923.25
Coinoculation	1.89	0.219	1.671	763.01	2.14	0.186	1.954	1050.53
Sum	6.38	0.837	5.553	2672.53	6.50	0.729	5.824	3408.00

Each value is a mean of six replicates.

\* Difference between fresh and dry weights of the plants represents the amounts of water absorbed at the assigned age of vegetative growth.

\* \*Moisture content of the plants is based on the dry weight.

Results showed that, inoculation with *Rh.etli* increased moisture content in the plants by 613.59 % compared with control (593.12 %), followed by *L.lactis* and Co-inoculation which gave 702.81 and 763.01 %, respectively for BAT477. The same trend was observed for DOR364, but with higher values, i.e. 730.30, 923.25 and 1050.53 %, respectively. These results coincided with those obtained by Vessey (2003), Dobbelaere et al.(2003), Bashan(2005) and Karlidag et al.,(2007) who suggested that plant growth promoting rhizobacteria stimulate plant growth by facilitating the uptake of minerals and microelements, by the plant. The present study revealed the superiority of the *Phaseolus* cultivar DOR364 above the counterpart BAT477 in regard to water absorption. On the other hand, the bacterial inoculation with both examined genera together greatly favored water absorption and consequently uptake of nutrients for a better plant growth.

## REFERENCES

- Abdel-wahab, A.F., G.A.A. Mekhemar, Heba Shehata and Awaref A. Hanafi (2006). Effect of plant growth bioprotecting and promoting rhizobacteria and compost on the healthy and productivity of peanut crop in sandy soil. *Minufiya J. Agric. Res.*, 31 (5):1323-1348.
- Axelsson L. (1998). Lactic acid bacteria: classification and physiology. In *Lactic Acid Bacteria: Microbiology and Functional Aspects*. (S. Salminen and A. von Wright, eds) pp. 1-72 Marcel Dekker, Inc., New York, USA.
- Bashan, Y. (2005). Plant growth –promoting substances. *Encyclopedia of Soils in the Environment*, Elsevier, Oxford, U.K. Vol. 1., pp. 103-115.
- Beriger JE. (1974). R-factor transfer in *Rhizobium leguminosarum*. *J. Gen Microbiol.*, 120: 421-429.
- Dobbelaere, S., J. Vanderleyden and Y. Okon (2003). Plant growth-Promoting effects of Diazotrophs in The Rhizosphere. *Critical Reviews in Plant Sciences*, 22(2):107-149.
- Chebotar, V.K., C.A. Asis and S. Akao (2001). Production of growth –promoting substances and high colonization ability of rhizobacteria enhance the nitrogen fixation of soybean when coinoculation with *Bradyrhizobium japonicum*. *Biol Fertil Soils*, 34:427-432.
- El-Howeity M.A. (2004). Colonization patterns of diazotrophs associated with legume and non-legume crops. Ph.D., Dep. Soil Sci., Fac. Agric., Minufiya Univ., Egypt.
- El-Tarabily, K.A. (2004). Growth promotion of bean (*Phaseolus vulgaris* L.) by a polyamine-producing isolate of *Streptomyces Griseoluteus*. The Fourth Annual U. A.E. University res. conference. SCI -49-53.
- Franz, C.M, M. Toit, A. Holy, U. Schillinger and W.H. Holzapfel (1997). Production of nisin-like bacteriocins by *Lactococcus lactis* strains isolated from vegetables. *J. Basic Microbiol.* 37(3):187-96.
- Hassanein, A.M. and G. A. A. Mekhemar (2003). Biological control of soybean damping-off caused by *Sclerotium rolfsii* using *Pseudomonas fluorescens* and *Pseudomonas putida*. *Egypt. J. Appl. Sci.*, 18:73-86.
- Klaenhammer, T.R. (1988). Bacteriocins of lactic acid bacteria. *Biochimic* 70: 337-349
- Karlıdag, H., A. Esitken, M. Turan and F. Sahin (2007). Effect of root inoculation of plant growth promoting rhizobacteria on yield, growth and nutrient element contents of leaves of apple. *Scientia Horticulturae*, 114:16-20.
- Lowe D.P. and E.K. Arendt (2004). The use and effects of lactic acid bacteria

**Diazotrophy and Growth of Beans (*Phaseolus vulgaris*).....**

- in malting and brewing , with their relationships to antifungal Activity , mycotoxins and gushing :Review. J. of The Institute of Brewing, Vol. 110, No. 3.
- Okon Y., J. Vanderleyden (1997). Root- associated *Azospirillum* species can stimulate plants, ASM New 63, 366-370.
- Ray B.(1996). Fundamental Food Microbiology. Wash.:CRC Press.
- Roughley, R.S.;J.T.Sprent and J.M.Day(1983). Nitrogen Fixation in Faba Bean. Ed.P.D. Hebblethwaite, Pub., Butterworths, pp.233-360.
- Sajur S.A., F.M. Saguir and M.C.Manca de Nadra (2007). Effect of dominant species of lactic acid bacteria from tomato on natural microflora development in tomato puree. Food control, 18 (5): 594-600.
- Sindhu S.S., S.K Gupta and K.R. Dadarwal (1999). Antagonistic effect of *Pseudomonas spp.* on pathogenic fungi and enhancement of growth of green gram (*Vigna radiate*).Biol fert. Soils, 29:62-68.
- Snoeck C(2001). Host specificity determinants of *Rhizobium* sp.BR816 for early signaling in symbiotic interactions.PhD thesis, K.U.Leuven.
- Souza, E.L., C.A. Silva and C.B. Sousa(2005). Bacteriocins:molecules of fundamental impact on the microbial ecology and potential food biopreservatives. Brazilian Archives of Biology and Technology, 48, (4): 559-566.
- Srinivasan,M., D.J.Peterson and F.B.Hall(1997). Nodulation of *Phaseolus vulgaris* by *Rhizobium etli* is enhanced by the presence of *Bacillus*. Can.J.Microbial.,43:1-8.
- Uhlman L.,U.Schillinger, JR. Rupnow and WH. Holzapfel (1992). Identification and characterization of two bacteriocin- producing strains of *Lactococcus lactis* isolated from vegetables . Int. J. Food Microbial , 16 (2): 141-151.
- Vescovo M, S.Torriani, C.Orsi, F.Macchiarolo and G.Scolari (1996) . Application of antimicrobial producing lactic acid bacteria to control pathogens in ready -to- use vegetables.J. of Appl. Bact., 81 (2):113-119(7).
- Vessey, J.K.(2003). Plant growth promoting rhizobacteria as biofertilizers. Plant and Soil 255: 571- 586
- Yildirim,Z. and M.G.Johnson (1998). Detection and characterization of bacteriocin produced by *Lactococcus lactis sub sp.* isolated from radish. Letters in Applied Microbiology , 26:297-304.

## تثبيت النيتروجين الجوى فى أصناف نباتات الفاصوليا الملقة بالريزوبيا وبكتيريا حمض اللاكتيك

محمد أحمد الحويطى

معهد الدراسات والبحوث البيئية - مدينة السادات - جامعة المنوفية

### الملخص العربى

فى تجربة معملية تم دراسة أثر تلقيح نباتات الفاصوليا النامية على بيئة اصطناعية بالريزوبيا (ريزوبيم اتلى) وبكتيريا حمض اللاكتيك (لاكتوكوكس لاكتس) على عملية تكوين العقد الجذرية ( عدد العقد ووزنها )، وكذلك نشاط إنزيم النيتروجينيز بالجذور والنمو الخضرى للنباتات النامية .

وقد أظهرت النتائج أن تلقيح نباتات الفاصوليا بالريزوبيا أعطى عقدا بكتيرية على الجذور بينما لم تظهر أى عقد على جذور النباتات النباتات الملقة فقط ببكتيريا اللاكتوكوكس لاكتس وكذا الكنترول. وأعطى التلقيح المشترك بين الريزوبيا وبكتيريا اللاكتوكوكس لاكتس تفوقا واضحا فى عدد العقد ووزنها مقارنة بالريزوبيا منفردة، حيث ووصلت الزيادة الى ١٨,٤ %، ٤٥,٣ %، وكذا وصلت الزيادة فى النشاط الإنزيمى الى ٣٦,٥ و ٦٤,٨ % مقارنة بالريزوبيا منفردة لكلا الصنفين المستخدمين فى هذه الدراسة وهما BAT477 & DOR364 على التوالى

وأدى التلقيح البكتيرى إلى زيادة الوزنين الرطب و الجاف للنباتات مقارنة بالكنترول. وكان الصنف BAT477 أعلى استجابة عن الصنف DOR364 فى القياسات الخضرية، حيث كانت متوسطات الزيادة عن الكنترول فى المجموع الخضرى هى ( ١٤,٨ ، ٦,١ و ٥,٣ %). ( ٢٨,٨ ، ٧,٦ ، ٢,١ %). والزيادة فى الوزن الجاف هى ( ٤,٢٥ ، ٢٤,٥ ، ٢١,٥ %). ( ١٦,١ ، ١٤,٨ ، ١١,٨ %) للمعاملات الآتية: التلقيح المشترك للريزوبيا مع بكتيريا اللاكتوكوكس لاكتس يليها اللاكتوكوكس لاكتس منفردة ثم الريزوبيا منفردة لكلا الصنفين على التوالى و على العكس تفوق الصنف DOR364 على الصنف BAT477 فى مجموع

### **Diazotrophy and Growth of Beans (*Phaseolus vulgaris*).....**

الاوران الرطبة والجافة للجنور , حيث سجلت متوسطات زيادة عن الكنترول بمقادير ( ١١٥.٠ و ٨٦,٤ , ٥,٩%) , ( ١٥,٦ , ١٥٥,٠ , ٣٩,٦%) عند استخدام التلقيح البكتيري المشترك ثم بكتيريا اللاكتوكوكس لاكتس ثم الريزوبيا فقط على الترتيب .