NATURAL SAPONIN PLANT EXTRACT WITHOUT OR WITH FRESH BAKER'S YEAST IN LACTATING BUFFALOE COWS RATIONS IMPACT ON NUTRIENTS DIGESTIBILITY, MILK PRODUCTION, COMPOSITION, AND SOME BLOOD SERUM PARAMETERS.

Abdelmawla, S.M.S.

Animal Production Dept., Faculty of Agric., Ain Shams University, Hadaek Shoubra 11241. Cairo, Egypt.

ABSTRACT

In a trial lasted for 90 days to assess the impacts of ration supplementation with saponin containing plant extract (defaunator) (SCPE) without or with fresh baker's yeast culture Saccharomyces cerevisiae (Probiotic) (FSCYC) as feed additives to enhance rumen fermentation in lactating buffalo cow. Twenty lactating buffalo cows of an average weight of 591 Kg were allotted to four groups of five animals each according to age and assigned at random to receive one of four dietary treatments. The treatments were: T1, (Control) received the Basal ration BR that consists of concentrate feed mixture: Egyptian berseem: rice straw (50:25:25 %, on dry matter basis), T2, received the BR plus 800 mg, saponin from the SCPE, T3, received the BR plus 800 mg saponin plus 10 gm's FSCYC, and T4 received the BR plus 800 mg saponin plus 20 gm's of FSCYC respectively. Results of ration supplementation of saponin without or with 10 or 20 gm's FSCYC treatments T2, T3, and T4 increased (P<0.05) nutrients digestibility, milk yield, milk protein, fat, lactose, total solids and solids not fat contents. Additives supplementation also increased blood serum total proteins, albumin (A) and globulin (G); however it decreased (P<0.05) A/G ratio, cholesterol and GPT. Treatment (T4) exhibited the highest (P<0.05) nutrients digestibility coefficients, milk yield, milk constituents, protein and fat, and blood serum proteins, albumin and globulin, but the lowest (P<0.01) blood serum urea, cholesterol, GOT and GPT wre obtained the treatment T4. It could be concluded that ration supplementation with SCPE plus 10 or 20 gm's of FSCYC for lactating buffalo cows (especially T4) had beneficial effects on their productive performance with normal physiologic responses of the cows.

Key words: fresh baker's saccharomyces cerevicia yeast, natural saponin plant extract, lactating buffaloes, nutrients digestibility, milk yield, constituents, blood serum.

INTRODUCTION

It has been reported that the elimination of the rumen protozoa population (defaunation) lead to improved animal performance (Leng, 1990). Although, these microorganisms contribute to fiber digestion, they increase the availability of energetic substrates for the animal. Nevertheless, it has been shown that protozoa prey upon bacteria (200 cells/minute or 1% bacteria/minute) and that they are preferred retain in the rumen (Weller and Pilgrim 1974; Coleman, 1975).

As a result of protozoa activity, a significant reduction in the flow of microbial biomass to the small intestine has been documented (Bird and Leng, 1978; Bird et al., 1979; Hsu et al., 1991). Independent of the diet offered, a consistent effect of elimination of protozoa is a larger availability of

amino acids for absorption at the small intestine, amino acids of bacterial (Ushida et al., 1988) and sometimes dietary origin (Michalowsky, 1988).

There are however, no commercial alternatives to defaunate ruminants at the farm level. In this sense, tropical plants with high or medium content of secondary compound may be an alternative to eliminate protozoa from the rumen. Among these modifying compounds are the saponin and condensed tannins, which have been shown to exert a specific effect against rumen protozoa while the rest of the rumen biomass remains unaltered (Lu and Jorgensen, 1987; Getachew et al., 2000; Wang et al., 2000).

Wina et al. (2003) evaluated the effect of saponin containing plant materials such as Morinda citrifolia (fruit), Nothopanax scutellarium (leaves), Sesbania sesban (leaves) and Sapindus rarak (fruit) on in vitro fermentation and found that gas production, short chain fatty acids, acetate: propionate ratio and protozoa population were the lowest in treatment of Sapindus rarak and concluded that this saponin rich plant has a potential as a natural defaunating agent. Moreover, in vitro fermentation, supplementation with Sapindus saponaria can decrease protozoa count (by 54%) and daily methane release (by 20%) relative to the control, without saponin content, (Hess et al., 2003).

Plant saponin natural extract which has been classified as a natural feed flavoring material (Valdez et al., 1986) has also been evaluated for its anti-microbial effect and its potential to modulate rumen fermentation and improve nutrient utilization in ruminants, (Benchaar, 2006).

Wina et al., (2006) reported that protozoal counts were decreased only in the long-term trial with sheep, using saponins from different plant sources.

Probiotics are live microbial cultures fed to animals to alter the balance of intestinal organisms in a beneficial way. Yeast Culture is a live culture of yeast (a fungi) and the media on which it was grown and dried so as to preserve the yeast's fermenting capacity.

The main effect of yeast culture is to stabilize the rumen environment (Hutjens 2005). Yeasts are known as rich sources of vitamins, enzymes, nutrients and other important cofactors which make them attractive as a basic nutrient source by number of features: rich sources of vitamins, enzymes, nutrients and other important cofactors (Dawson, 1992). Yeast cells are able to maintain their metabolic activities under anaerobic conditions, and exposure to low pH (Dawson, 1992).

Inclusion of yeast culture (Saccharomyces cerevisiae) in the diets of ruminants has been shown to increase nutrient digestibilities (Dawson, 1993, El-Waziry et al., 2000, El-Ashry et al., 2001, El-Talty, 2001, and Marghany et al., 2006), shift bacterial populations (Harris et al., 1988), increase the number of rumen bacteria (Williams, 1988, Dawson 1993, and Edwards et al., 1991), alter the flow of nitrogen fractions to the duodenum (Erasmus et al., 1992, and Harrison et al., 1988), and increase milk yield (El-Ashry et al., 2001, and Marghany et al., 2006).

Moreover benefits of supplementing ruminant rations with SCYC are: neutralization of certain bacterial toxins (Castagliuolo et al., 1999), adherence of flagellate bacteria, due to the presence of mannose receptors, allow

pathogens to be eliminated by feces (Czerucka and Rampal 2002). Moreover, lactic bacteria, a beneficial flora, are increased, reinforcement of mucosal integrity and intestinal cells. Also, live yeasts have a documented efficacy on intestinal villi height and crypt depth, enhancing the assimilation of nutrients, and enhance modulation of the immune system by stimulation of IgA response to pathogens (Qamar et al., 2001, Cheeke and Otero (2005).

Therefore, the present experiment aimed to evaluate the effects of saponin and glyco components containing extract SCPE (as defaunator), (natural liquid saponin extract as 50: 50 (v/v) mixture from Yucca Shidigera and Quillaja Saponaria), alone, and its combined effects with two levels of the fresh Saccharomyces cerevisiae yeast culture FSCYC (as a probiotic) on feed intake, nutrients digestibility, milk production, constituents, and some blood serum parameters using lactating buffalo cows.

MATERIALS AND METHODS

This study was conducted at the Agricultural Experiment Farm Station in Shalakan, at Kaluobeia Provence which belongs to The Faculty of Agriculture, Ain Shams University. Cairo, Egypt.

Animals and feeding: Twenty lactating buffaloes, in their 4th or 5TH lactation seasons were used in 90 days trial started after two weeks of parturition. Buffalo cows were allotted to four groups of five animals each according to age and assigned at random to receive one of four dietary treatments supplemented with saponin (the steroid or triterpene glycoside as defaunator) with its companion glyco component (ammonia binder) from a saponin containin natural plant extract SCPE without or with the additive Fresh baker's *Saccharomyces cerevisiae* yeast culture FSCYC. The Fresh SCYC containing total cell count of 2.5-2.7×10¹⁰ and viable cell count of 1.3×10⁹-2×10¹⁰ per gram (Grand Cairo Bakeries Company, Yeast Factory, Alsalam City, Cairo, Egypt. The second additive the liquid saponin containing plant extract SCPE was a 50:50 (v/v) mixture of Yucca and Quillaja desert plants concentrated liquid extracts from Nor-Feed, Denmark.

The experimental treatments were: (TI) -The control group which received the basal ration BR, that consisted of concentrate feed mixture (CFM): Egyptian berseem (EB): rice straw (RS); (50: 25: 25%, dry matter basis). (T2) received the BR+ 800 mg saponin; (T3) - received the BR+ 10gm's FSCYC+800 mg saponin and, (T4) which received the BR+ 20 gm's FSCYC+800 mg saponin. The CFM consisted of 25% undecorticated cotton seed cake, 35%, wheat bran, 30% com, 3% rice bran, 3% molasses, 2% limestone, 1% urea and 1% salt (Na CI).

The natural extracts, from the desert plants Mohave Yucca or Quillaja Saponaria, which contain saponins (defaunator) and glyco components (ammonia binder), have been classified as food grade materials and natural feed flavoring materials which are of course non-toxic and highly biodegradable, Valdez et al., (1986). This extract has long been used by the health foods industry in the USA as a healthy nutrient supplement product. The extract is approved for use in food and beverages by the FDA under CFR 172.510, FEMA number 3121. In the beverage industry, it is used to

•

prepare root beer, slush products, frozen carbonated beverages, beer, and juice.

The animals were hand- milked twice daily at 6.00 am and 16.00 pm., while milk samples were collected once biweekly for 90 days. Milk yield was recorded individually and milk samples were analyzed for percentages of fat, total solids (TS), solids-not fat (SNF), total proteins (TP), pH, acidity, and ash (Ling, 1963); lactose (Barnett and Abd El-Tawab, 1957).

The chemical composition of the ration ingredients is shown in Table (1).

Table (1): The experimental feed ingredients (Concentrate feed mixture CFM, Egyptian Berseem forage, Rice straw, (RS), and the fresh Saccharomyces cerevisia yeast culture (FSCYC) and the basal diet nutrients content.

item	Dry matter (DM)	Organic matter (OM)	Ash	Crude protein (CP)	Crude fiber (CF)	Ether Extract (EE)	Nitrogen free extract (NFE)
CFM ¹	92.6	90.1	9.9	14.1	13.4	2.7	59.9
Egyptian Berseem	12.3	88.2	11.8	13.8	27.3	2.6	46.9
Rice Straw	94.53	83.39	16.6	3.5	31.5	1.5	40.1
FSCYC	29.0	92.6	7.4	44.3	6.5	3.0	38.8
Basal ration ²	92.95	88.719	11.281	18.031	20.044	2.226	48.418
T2	92.95	88.719	11.281	18.031	20.044	2.226	48.418
T3	95.85	90.41	11.50	19.23	20.25	2.31	49.54
T4	95.95	94.09	11.71	20.60	20.32	2.39	50.67

CFM=Concentrate feed mixture. *Calculated chemical composition.

Management: Amounts of daily feeds were assessed to cover the maintenance and the production requirements (Shehata, 1970). The CFM was individually weighed for each animal and offered twice daily during milking times at 6.00 and 16.00 hr, while roughages were offered at 8.00, 15.00 and 21.00 hr. after the animals were allowed to drink fresh water at 7.00, 14.00 hr. and at 20.00 hr. The daily supplementary yeast was mixed with CFM twice daily just before feeding to ensure that each animal had consumed its own supplement. Saponin containing liquor doses were dissolved in drinking water for the specific groups (T2, T3 and T4) in the drinking pool at times of water drinking. The treatments were begun after 2 wk's after calving and extended till 90 days.

Feed and milk sampling and chemical analysis: Samples of CFM, Egyptian Berseem, Rice Straw and Fresh baker's yeast were analyzed for dry matter (OM), ash crude protein (CP), crude fiber (CF) and ether extract (EE) according to A. O. A. C (1995). Nitrogen-free-extract (NFE) was calculated by differences.

Sampling and analysis of blood serum: Blood samples were withdrawn from the jugular vein from each animal 4 hours (hrs.) post morning feeding (pmf.) in the same day of milk sample collection. Collected blood samples were centrifuged at 4000 r.p.m. for 20 min. and the blood serum was stored in clean glass vials at -20°C till analysis. Serum total proteins were

determined as described by Armstrong and Carr (1964), albumin (Doumas *et al.*, 1971), urea (Patton and Crouch, 1977), and creatinine was determined according to Husdan (1968), transaminases (GOT and ,GPT) activities (Reitman and Frankel, 1957), cholesterol (Kostner et al., 1979). Globulin and albumin/globulin ratio (A/G) were calculated.

Digestibility trials: Trial experimental periods were setup into three phases representing the first, second and third month's of lactation study. By the end of each of the three months of the experimental periods, three animals from each experimental group were used in the digestibility trial. Feces was hand collected at 10.00 a. m. employing and following the Grab sample method for three successive days from each animal. The acid insoluble ash as internal marker was measured for determining the digestibility (Van Keulen and Young, 1977). Proximate chemical analysis was carried out according to the A. O. A. C. (1995) procedures, which were employed for the digestibility coefficients of studied nutrients calculations according to Maynard and Loosli, (1957).

Statistical analysis: Statistica analysys was performed using the least square methods described by Snedecor and Cochran (1982). Significant differences among means were tested between treatments within each treatment using Duncan's new multiple range test was used to test the differences among means (Duncan, 1955). The General linear models (GLM) procedures of S A S (Statistical Analysis Systems, 1998) were used.

RESULTS AND DISCUSSION

Dry matter intake and digestibility coefficients:

Feed intake, and nutrients digestibility coefficients for tested rations with lactating buffaloes are presented in Table (2).

Table (2): Mean values of live body weight, dry matter intake (DMI), nutrients digestibility, (%) of the experimental rations fed to lactating buffalo cows.

	ating bu	Italo cows	٠.				
ITEM	Experimental rations						
	Control	T1+SCPE	T1 + SCPE+	T1 +SCPE+	±		
[T1	T2	10 g FSCYC T3	20 g FSCYC T4	ŞE		
LBWT (Kg)	593	592	585	595	i		
		DM inta	ke:				
Total DMI(Kg/head/day)	15.40	15.32	15.28	15.24	1		
Total DMI as % LBWT	2.59	2.59	2.60	2.56	ì		
From CFM	7.70	7.66	7.64	7.62	:		
From berseem	3.85	3.83	3.82	3.81	1		
From rice straw	3.85	3.83	3.82	3.81	1		
NUTRIENTS					1		
DIGESTIBILITY, (%):		-					
Dry matter	66.60"	68.38°	73.25	75.65°	0.109		
Organic matter	69.33	73.10°	77.35	79.38	0.134		
Crude protein	71.19°	73.65°	75.24"	77.09*	0.132		
Crude fiber	58.49	60.60°	66.14"	70.65°	0.040		
Ether Extract	77.01	79.18°	80.85°	82.30°	0.240		
Nitrogen free extract	64.08 ^u	66.00°	67.70	69.95°	0.099		
TDN	59.67	61.35	65.59	69.79	i		
DCP	13.52	13.52	15.17	15.24			

Each value is a mean of 27 samples from 3 animals;

SE= Standard error of means; CFM=Concentrate feed mixture;

^{*.} b, c, d Values in the same raw with different superscripts differ (P<0.05) significantly.

Averages of feed DM intake values ranged between 15.40 and 15.24 kg/day. The dry matter intake tended to decrease significantly (P>0.05) by the addition of SCPE (T2) and yeast culture with saponin (T3 & T4). Digestibility coefficients of dry matter and organic matter increased significantly as SCPE liquor (at 800 mg) without or with FSCYC were added at 10 or 20 g/d/head as compared by the control group.

The Increase in DM & OM digestibility may be due to the stimulation of rumen cellulolytic bacteria by yeast addition (El-Ashry et al., 2001, El-Saadany et al., 2002, Abdel-Khalek et al., 2002, Hutiens 2005) and this might be attributed to the improvement of crude protein digestibility, due to suppression or elimination of protozoa and ammonia utilization as an effect of saponin with its accompanying glyco- components effects. (Benchaar, 2006; Lovett et al., 2006, Wina et al., 2006, Hutiens, 2005 and Wallace, 2007). Dann et al. (2000) and Robinson and Garrett. (1999) have reported significant increases in dry matter intake when yeast culture was fed to transition cows, resulting in higher milk yields and less weight loss postpartum. A reduction in rumen lactic acid concentrations has also been reported (Williams, 1989), Thalib, et al., (2001) reported that using probiotics improved the positive effects of defaunation on animal performance in which the performance of the treatment groups were significantly higher than the control treatment, when they used commercial probiotics (micro Bio. cellulolytic cocci, and rods) and a defaunator consisted of the extracted saponin from Sapindus rarak fruit with methanol.

Crude protein digestion results pointed out to an improvement (P<0.05) as the saponin and the saponin with FSCYC were added at 10 0r 20 g /h/d. Erasmus *et al.* (1992) and EL-Waziry et at. (2000) found that CP digestibility was significantly increased with yeast culture supplement. Wohlt *et al.*, (1998) and Wiedmeier *et al.* (1987) reported that significantly higher CP digestibility in dairy cattle was obtained by the addition of yeast culture to their rations.

The improvement of protein digestibility may be due to the increase of microbial protein production and the reduction in NH3-N production (Pen *et al.*, 2006), stimulation of rumen proteolytic bacteria (Williams, 1989, Newbold 1990 and Allam *et al.*, 2001) and/or may be due to increased degradation of protein and the flow of microbial nitrogen to the intestines, (Wiedmeier *et al.*, 1987).

Also as a result of saponin plant extract antimicrobial effects in elimination or reduction of protozoa numbers and count in the rumen (Benchaar, 2006; Lovett et al., 2006, Wina et al., 2006, Hutjens, 2005), that prey upon bacteria (200 cells/minute or 1% bacteria/minute), they are preferred retained in the rumen (Weller and Pilgrim 1974; Coleman, 1975). As a result of protozoa activity, a significant reduction in the flow of microbial biomass to the small intestine has been documented (Bird and Leng, 1978; Bird et al., 1979; Hsu et al., 1991).

Crude fiber digestibility increased significantly (P<0.05) as a result for the addition of saponin and fresh yeast culture (T2, T3, and T4) compared with T1. The improvement of crude fiber may be due to increasing the number of rumen cellulolytic bacteria by saponin and yeast addition (WIliams.

1989, Newbold, et al., 1990, and Allam et al., 2001, and it provides stimulatory factors to rumen bacteria (Piva et al., 1993, Putnam et al., 1997, and Wohlt et al., 1998). Ether extract and nitrogen free extract digestibility were increased significantly (P<0.05) by the addition of yeast. The increase in EE & NFE digestibility (P<0.05) in response to saponin and SCYC treatment (T2, T3, and T4) may be due to the increased energy utilization (instead of being lost as methane, CO2, or lactic acid, which might be saved as a cause of the additives effects on rumen medium and microflora) as indicated by the increase in the propionic acid in rumen (Wiedmeier et al., 1987, Harris and Lobo, 1988, Williams, 1989, Wohlt et al., 1998) or to the observed improvement of the digestibility of most nutrients (Robinson, 1997 and Al-Dabeeb and Ahmed. 2002).

Milk production and composition:

The effect of saponin without or with FSCYC supplementation on of milk composition and acidity (%) are presented in Table (3).

Table (3): Effect of experimental rations on milk yield (Kg/d) and

constituents of lactating buffalo cows.							
Experimental rations							
			Cont.+SCPE+				
			20 g FSCYC T4				
 	12	13	14				
16.06 ^d +0.15	16.81°+0.17	17 18 ⁶ +0 17	17.59°+0.16				
			17.47+0.13				
			17.56+0.11				
			17.54°±0.079				
10.23 10.010	10.05 10.001	17.20 10.013	17.07 30.070				
0.52540.44	0.880+0.43	10.06 ⁵ ±0.13	10.22 ^a +0.13				
			10.62+0.10				
			10.40+0.08				
			10 44°±0 062				
3.01 20.001	3.32 10.003	10.00 10.02	10.77 20.002				
8 85p+0 03	7 308+0 03	7.463+0.03	7 43°+0.03				
			7.50+0.02				
			7.44+0.02				
			7.46°±0.017				
0.00 20.010	1.42 10.011	7.90 20.011	1.40 20.011				
3 80°+0 06	3 930+0 07	4 625+0 07	4.71°+0.07				
			4.75+0.05				
			4.85+0.05				
			4.79°±0.037				
1.00 20.000	0,0.20.00.						
4.65°+0.05	4.78°+0.06	4.95°+0.06	5.23 ^a +0.06				
			5.27+0.05				
			5.16+0.04				
4.68°±0.031		4.99°±0.031	5.22°±0.031				
0.717°+0.002	0.724 ^b +0.002	0.728b+0.003	0.735°+0.003				
			0.738+0.002				
	0.719+0.002	0.719+0.002	0.731+0.002				
0.712°±0.002	0.722°±0.002	0.724°±0.002	0.734°±0.002				
 _		<u> </u>	1				
0.173°+0.001	0.175°+0.001	0.176 ^{ab} +0.001	0.1774+0.001				
0.171+0.001	0.174+0.001	0.174+0.001	0.175+0.001				
	0.172+0.001	0.174+0.001	0.175+0.001				
0.172°±0.0003	0.174°±0.0004		0.176°±0.0004				
	0.173°+0.001 0.171+0.001 0.171+0.001	Control (Cont.) No Supplement. T1 16.06 ^d +0.15 16.07+0.15 16.54+0.11 16.67+0.10 16.29 ^a ±0.078 16.63 ^a ±0.081 9.53 ^c +0.11 9.79+0.09 9.61 ^a ±0.061 6.82 ^b +0.03 6.84+0.03 6.84+0.03 6.76+0.02 6.80 ^a ±0.016 7.42 ^a ±0.017 3.89 ^c ±0.06 3.91+0.05 3.93+0.04 3.89 ^a ±0.03 4.65 ^d +0.05 4.66+0.05 4.70+0.04 4.68 ^a ±0.031 0.712 ^c ±0.002 0.712 ^c ±0.001 0.171 ^c +0.001 0.171 ^c +0.001 0.171 ^c +0.001 0.171 ^c +0.001 0.172 ^c +0.001	Control (Cont.) Cont.+SCPE Cont.+SCPE Cont.+SCPE+ 10 g FSCYC T3 16.06 ^d +0.15 16.81 ^e +0.17 17.18 ^b +0.17 17.18 ^b +0.17 16.07+0.15 16.41+0.15 17.09+0.13 16.54+0.11 16.63 ^e +0.081 17.23 ^e ±0.079 9.53 ^c +0.11 9.88 ^b +0.13 10.06 ^b +0.13 10.06 ^b +0.13 9.93+0.11 10.04+0.10 19.93+0.11 10.04+0.10 10.979+0.09 9.92*0.08 10.06+0.08 10.06+0.08 9.61*±0.061 9.92*±0.063 10.06*±0.02 7.39*40.03 7.46*±0.02 7.53+0.02 7.42±0.02 7.42±0.02 7.42±0.02 7.42±0.017 7.46*±0.017 3.93*±0.02 7.42±0.017 7.46*±0.017 3.86*±0.02 3.93*±0.036 3.93*±0.037 4.62*±0.037 4.62*±0.037 4.62*±0.037 4.62*±0.037 4.68*±0.037 4.48*±0.037 4.48*±0.037 4.68*±0.031 4.80*±0.032 4.95*±0.004 4.95*±0.004 4.95*±0.004 0.712*±0.002 0.724*±0.002 0.728*±0.002 0.728*±0.002 0.728*±0.002 0.728*±0.002 0.728*±0.002 0.728*±0.002 0.728*±0.002 0.719*±0.002 0.724*±0.001 0.174*±0.001				

Each value is the mean of 30 combined samples from 5 animals for each treatment.

Ab.c.dMeans in the same raw with different superscripts differ (P<0.05) significantly.

Total solids (TS) % content was higher (P<0.05) for T4 than T3, T2, and the control which was the lowest (P<0.05) as the level of yeast increased, which agree with that results reported by Allam *et al.*, (2001) and EL-Ashry *et al.*, (2001).

Solids-not fat (SNF) %, and ash % for T4 followed a trend being higher (P<0.05) than both T3, T2, where all supplemented treatment were higher (P<0.05) than the control.

Fat content, %, did not differ significantly for supplemented treatments, however, all of T4, T3, and T2 were higher (P<0.05) than the control, that agrees with Jin et al., (2007) findings. Also, these responses, for the treatment groups, might be a result of increased utilization of saved energy and protein because of the saponin and yeast additives effects on ruminal protozoa (defaunation), methanogenes suppression, and the activation of hydrogen and lactic acid utilizing bacteria, and enhancement of other bacteria genera for multiplication and producing more microbial protein, (Benchaar, 2006).

Milk protein content, %, was the highest (P<0.05) for T4, followed by T3, where both were higher than T2 which didn't differ from the control group. Which might be a result of the combined addition for saponin +FSCYC which might helped utilizing the microbial protein escaped from being consumed by protozoa (Benchaar, 2006; Pen et al., 2006, Agrawal et al., 2006) that might be reduced or eliminated by saponin supplementation in T4 and T3 and utilizing ruminal ammonia saved from being converted to urea (Table 5), (Thalib et al., 2001; Hutjens, 2005; Pen et al., 2006). Improvement of milk protein and fat content may be due to the stimulation of the rumen microbes multiplication, which cause a change in microbial protein synthesis lead to an increased postruminal protein passage and lead to an increased milk protein vield, as explained by Dawson (1993) and that the Supplementation with FSCYC might supply some stimulatory factors to rumen cellulolytic bacteria (Putn. al., 1997; Wohlt et al., 1998 and Allam et al., 2001). Results reflected that milk acidity (%) insignificantly increased vfor treatment rations than the control one, which agree with the results obtained by Dawson, (1993) and EL-Ashry et al. (2001).

Milk production and components yields are presented in Table (4).

Results of Milk yield and milk constituents (Table 3) reflected an increase for all parameters studied (P<0.05) as saponin without or with FSCYC (T2, T3, and T4) was added to T3 or T4 as compared to the control group during trial different periods. The 4% Fat corrected-milk daily yield, total solids and fat yields were significantly (P<0.05) increased for groups T4, T3 and T2 repectively, than those values for the control group during the 3 pahases of the trial. These results are in accordance with those reported on animals fed diets supplemented with, SCYC (Piva et al., 1993, Robinson and Garrett, (1999), Dann et al., (2000), Allam et al. (2001), EL-Ashry et al., (2001), and Jin et al., (2007). The T4 ration group produced more milk yield with higher (P<0.05) total protein, solids not fat and lactose yields than other treatments. Results which are in accordance with those reported by Wohlt et al. (1991), Kobayasli et al. (1995), Putnam et al. (1997) and EL-Ashry et al. (2001). Hutjens, (1991) reported that utilization of yeast by cows lead to

J. Agric. Sci. Mansoura Univ., 33(5), May, 2008

increased milk production averaged 25.1 kg of 4% FCM compared to control cows at 23.5 kg. The increase in milk total solids yield exhibits positive improvement which is very important to milk and cheese industry.

Table (4): Impact of SCPE without or with FSCYC supplementation on

lactating buffalo's milk yield and constituents¹.

	Experimental rations						
	Control	T1+SCPE T2	T1+SCPE+	T1+SCPE+	±		
ITEM	T1		10 g FSCYC	20 g SCYC	SE		
			T3	T4			
Total milk yield, Kg/d:							
1 ST. month	6.96⁴	7.49 ^c	7.79 ^b	8.35 ^a	0.056		
2 nd. month	7.00	7.59	7.86	8.45	0.056		
3 rd. month	7.1	7.63	8.77	8.48	0.056		
Overall mean	7.01 ^d	7.57 ^c	8.14 ^b	8.43 ^a	0.032		
Fat yield, g/d:							
1 ST. month	472.18°	553.2°	581.6 ⁶	619.6°	4.312		
2 nd. month	480.8	570.2	593.6	635.3	4.312		
3 rd. month	476.5	563.3	648.0	632.9	4.312		
Overall mean	476.48 ^a	562.21°	607.74 ^b	629.24°	2.489		
		ted milk yield.	Kg/d:				
1 ST. month	9.9⁴	11.3°	11.8 ⁸	12.6 ^a	0.086		
2 nd. month	10.0	11.6	12.0	12.9	0.086		
3 rd. month	10.0	11.5	13.2	12.9	0.086		
Overall mean	9.95⁰	11.46c	12.37°	12.81°	0.049		
		lids yield, g/d					
1 ST, month	1134.2 ^d	1247.3°	1340.0 ⁶	1465.9°	11.74		
2 nd. month	1111.8	1247.9	1335.8	1464.8	11.74		
3 rd. month	1180.5	1283.8	1533.5	1503.7	11.74		
Overall mean	1142.2 ^d	1259.7°	1403.32 ^b	1478.13 ^a			
	Solids not fat yield, g/d:						
1 ST. month	645.4 ^d	740.7°	784.8°	853.1	7.57		
2 nd. month	642.5	752.3	789.2	895.9	7.57		
3 rd. month	702.9	761.3	882.5	891.1	7.57		
Overall mean	673.6 ^d	7514°	818.8 ⁰	880.0°	4.373		
Total protein yield, g/d:							
1 ST. month	269.5°	293.2°	346.8 ⁰	397.3°	6.01		
2 nd. month	270.1	298.9	353.3	405.7	6.01		
3 rd. month	279.8	302.7	395.1	408.5	6.01		
Overall mean	273.1ª	298.3°	365.1 ^b	403.8°	3.470		
	Lactose yield, g/d:						
1 ST. month	322.6°	358.7°	384.1 ^b	429.7°	4.41		
2 nd. month	328.2	364.8	391.2	439.8	4.41		
3 rd. month	333.3	368.1	442.9	449.4	4.41		
Overall mean	328.03°	363.87°	405.1 ⁸	439.6ª	2.543		
Ash yield, g/d:							
1 ST. month	49.90	54.2°	56.7 ^b	61.4ª	0.43		
2 nd. month	49.6	54.8	57.0	62.4	0.43		
3 rd. month	50.2	54.9	60.1	62.1	0.43		
Overall mean	49.9ª	54.7°	58.8⁵	61.9ª	0.248		

Each value is the mean of 30 combined samples from 5 animals of each treatment.

a.b.c.d Means in the same raw with different superscripts differ (P<0.05) significantly.

^{4%} FCM was calculated as @ 0.4×milk weight(Kg)) + (15×fat weight(Kg)), (Ensminger, 1978).

Blood serum parameters and physiological performance of buffalo

Blood serum levels of total protein and albumin (g/dL) presented in table (5), showed that the total proteins values for the supplemented groups were higher (P<0.05) for T4 than T3 and T2 where also were which is higher (P<0.05) than the control which agree with El-Ashry et al., (1994), Soliman et al., (1997) and El-Ashry et al., (2001) findings, however our results contradict with those results reported by Piva et al., (1993) who found that blood plasma total proteins were adversely affected by adding yeast culture.

The control treatment group results values for blood serum parameters, A/g ratio, urea-N, creatinine, cholesteroland and GPT were higher (P<0.05) than the corresponding values in the treatments (T2, T3 and T4) throughout the periods of the experiment.

Results reflect that dietary treatments had no harmful effects but they were within the normal physiological levels. These results agree with the results of Soliman et al., (1997) and El-Ashry et al., (2001).

Blood serum level of urea nitrogen (mg/dL) and creatinine are presented in Table (5).

It was noticed that serum urea nitrogen values were decreased in the supplemented groups T2, T3, and T4, through the three experimental periods. Serum level of creatinine was lower in both T2 and T3 groups than other ones. These results indicate state of normal functioning of the kidneys.

Blood serum values of cholesterol (mg/dL) are presented in Table (5).

The present results revealed that cholesterol level was significantly (P<0.01) decreased in the supplemented treatments as an effect of saponin extract action which agree with a number of studies have shown that saponins from different sources lower serum cholesterol levels in a variety of animals including human subjects as reported by Southon *et al.* (1988); Harwood *et al.* (1993); Potter *et al.* 1993; Matsuura, (2001); Al-Habori and Raman, (1998).

On way for reduction of blood cholesterol is through the large mixed micelles formed by the interaction of saponins with bile acids account for their increased excretion when saponin-rich foods such as soyabean, lucerne and chickpea are consumed Oakenfull, (1986); Oakenfull and Sidhu, (1990). The resulting accelerated metabolism of cholesterol in the liver causes its serum levels to go down Francis et al., (2002) and Cheeke et al., (2006). Another way for decreasing cholesterol is through the decreased intestinal cholesterol absorption induced by some saponins, however, was seen to be without interference with the entero-hepatic bile acid recirculation (Harwood et al. 1993). Saponins also reduced the more harmful LDL- cholesterol selectively in the serum of rats, gerbils and human subjects (Potter et al. 1993; Harris et al. 1997; Matsuura, 2001).

J. Agric. Sci. Mansoura Univ., 33(5), May, 2008

Table (5): Effects of SCPE and FSCYC supplementation on some blood serum parameters¹.

Serum parameters . Experimental rations							
	Control .	Cont.+	Cont.+	Cont.+	±		
Item	No	SCPE	SCPE+	SCPE+	SE		
	Supplement.	0 FSCYC.	10 g FSCYC	20 g FSCYC			
	T1	T2	T3	T4			
T.Proteins, g/dL							
1st. month	7.18d	7.52c	7.66b	8.38a	0.072		
2nd. Month	7.21	7.51	7.67	8.63	0.072		
3rd. month	7:34	7.56	7.66	8.47	0.072		
Overall mean	7.25c	7.53b	7.66b	8.5a	0.041		
Albumin, g/dL				i	T		
1st, month	3.33c	3.51b	3.39c	3.82a	0.018		
2nd. Month	3.28	3.46	3.43	3.83	0.018		
3rd, month	3.39	3.41	3.45	3.85	0.018		
Overall mean	3.34c	3.40b	3.42b	3.84a	0.010		
Globulin, g/dL	† · · · · · · · · · · · · · · · · · · ·			ì			
1st. month	3.85d	4.01c	4.27b	4.56a	0.068		
2nd. Month	3.93	4.05	4.24	4,79	0.068		
3rd. month	3.95	4.15	4.21	4.62	0.068		
Overall mean	3.91d	4.07c	4.24b	4.66a	0.039		
A/G Ratio	†						
1st. month	0.87a	0.87a	0.79b	0.84ab	0.013		
2nd, Month	0.83	0.85	0.80	0.79	0.013		
3rd, month	0.85	0.82	0.81	0.84	0.013		
Overall mean	0.85a	0.85a	0.81b	0.83ab	0.007		
Urea-Nitrogen, mg/dL			1				
1st, month	36.97a	35.88b	34.69cd	34.83d	0.182		
2nd, Month	36.68	35.77	34.50	34.88	0.203		
3rd, month	35.96	35.59	34.46	34.83	0.181		
Overall mean	36.53a	35.75b	34.35c	34.84d	0.107		
Creatinine, mg/dL		_ <u></u>					
1st. month	i 1.19a	1.14b	1.116	1,22a	0.115		
2nd, Month	1.22	1.16	1.14	1.21	0.115		
3rd. month	1.19	1.14	1,13	1.14	0.115		
Overall mean	1.2a	1.15b	1.13b	1.19a	0.009		
Cholesterol,mg/dL			77.00	1	10000		
1st. month	153.6a	143.7b	144.6b	144.1b	1.488		
2nd. Month	154.0	141.9	144.6	144.0	1.488		
3rd. month	153.4	142.1	141.2	141.9	1.488		
Overall mean	153.7a	142.6b	143,4b	143.3b	0.859		
GOT, U/L				1	1		
1st. month	127.9bc	129.0b	132.8a	124.1d	1.559		
2nd. Month	127.6	129.6	1.33.0	123.3	1.559		
3rd, month	128.3	128.4	132.6	126.4	1.559		
Overall mean	127.9bc	129.0b	132.8a	124.6d	0.908		
GPT, U/L					3.550		
1st. month	53.95a	46.84b	37.10c	35.34c	1.196		
2nd. Month	56.33	45.82	36.72	34.99	1.196		
3rd. month	57.78	44.85	36.03	34.86	1.196		
Overall mean	56.02a	45.8b	36.6c	35.1c	0.690		

Each value is the mean of 30 combined samples from 5 animals for each treatment. ab.c.d Means in the same raw with different superscripts differ (P<0.05) significantly.

Mean activity values of GOT in T4 was the lowest (P<0.05) while T3 was the highest (P<0.05). The GPT (U/L) results showed a decreasing trend with introducing additives to rations of the lactating buffaloes.

Results indicate no harmful effects for the saponin or the FSCYC additives on liver function of the supplementd animals (Comelius 1970) and that biosynthesis of albumin and globulin in the liver were normal. Physiological levels of blood serum parameters studied in this trial were within the normal range for blood constituents of buffaloes reported by El-Ashry *et al.*, (1994) and El-Ashry, *et al.*, (2001).

The economic efficiency:

Table (6) shows the effect of supplementing lactating buffalo cows rations with saponin from SCPE and FSCYC on the economic efficiency expressed as price of milk per cost of feed consumed. The prices considered here were (CFM 1700 LE/ton, Egyptian berseem, 280 LE/ton; Rice straw, 350 LE/ton; FSCYC, 20 LE, and SCPE, \$18.47/L). The results indicated that using the supplement of saponin, yeast and different combinations were better economically than control. The FSCYC supplemented groups were more beneficial than the control and the saponin only group. The T4 (saponin 800 mg+ 20g FSCYC) was the best and the superior (P<0.05) of all treatments.

The results revealed that the introduction of the additives to rations decreased the feed cost of 1 Kg milk produced and improved the economic efficiency by 54.6, 84.5, and 202.9 %. It is very important to notice that the increase of the milk total solids with the introduction of the additive in rations of lactating buffalo cows indicates to the efficiency of treatments in increasing the industrial value (cheese industry) as the net total solids yield increased with treatment with the study additive additives.

It could be concluded that the addition of saponin at 800 mg with 10 or 20 g FSCYC/h/d to the lactating buffalo cows, especially T4, is recommended for the improvement of nutrients digestibility, milk yield and the total solids components (which is very important to milk and chees industry) and the maximization of the economic return.

REFERENCES

- AOAC. (1990). Association of Official Analytical Chemists. 1990. Official methods of analysis, 15th edition. AOAC Inc., Arlington, VA.
- Abdel-Khalek, A.E; A.F. Mehrez and E.A. Omar; (2000). Effect of culture (Lacto-Sacc) on rumen activity, blood constituents and growth of suckling Friesian calves. Proc.Conf. Anim. Prod. In the 21st Century, Sakha, Kafr El-Sheikh, Egypt, 18-20 April, 2000: 201.
- Agrawal, N., Kamra, D. N., Chaudhary, L. C., Patra, A. K. (2006). Effect of Sapindus mukorossi extracts on In Vitro methanogenesis and fermentation characteristics in buffalo rumen liquor. Journal of Applied Animal Research. 30 (1): 1-4.

- Al-Dabeeb, S.N. and S.M. Ahmed (2002). Effect of dry yeast: (Saccharomyces cerevisiae) in sheep rations differing in their roughage to concentrate ratio on digestion, nitrogen balance and rumen fermentation. Egyptian J. Nutrition and Feeds, 5:1
- Al-Habori M. and Raman A. (1998) Antidiabetic and hypocholesterolaemic effects of fenugreek (Review). Phytotherapy Research 12, 233–242.
- Allam, A.M; K. El-Shazly; B.E.A. Borhami and M.A. Mohamed; (2001). Effect of Baker's yeast)Saccharomyces cerevisiae(supplementation on digestion in sheep and milk response in dairy cows. Proc. of the 8th Conf. on Anim. nutrition, Sharm El-Sheikh, Egypt, 23-26 October 2001, Egyptian J. Nutrition and Feeds, 4(Special Issue): 315.
- Armstrong, W. D. and C. W. Carr. (1964). Physiological Chemistry Lab.boratory Direction, 3rd. Edition. P75. Burges Publishing Co. Minneapolis.
- Barnett.; A. J.G. and G. Abd El-Tawab. (1957). Determination of lactose and cheese. J. Sci. Food Agric., 8: 437441.
- Benchaar, C. (2006). Feed intake, digestion, milk production, and milk composition in dairy cows fed natural plant extracts. WCDS Advances in dairy Technology. Vol., 18, Abstract, page 354.
- Bird, S. H., and R. A. Leng. (1978). The effect of defaunation of the rumen on the growth of cattle on low protein high energy diets. Br. J. Nutr. 40: 163-167.
- Bird, S. H., M. K. Hill, and R. A. Leng. (1979). The effect of defaunation of the rumen on the growth of lambs on low - protein high-energy diets. Br. J. Nutr. 42: 81-87.
- Castagliuolo I., Riegler M. F., Valenick L., Lamont J. T. and Pothoulakis C. (1999). Saccharomyces boulardii protease inhibits the effects of clostridium difficile Toxins A and B in Human Colonic Mucosa. Infect. Immun. 67: 302-307.
- Anti-inflammatory and anti-arthritic effects of yucca schidigera: A review.
- Cheeke, P. R. and R. Otero. (2005). Yucca, quillaja may have role in animal nutrition. Feedstuffs, (Suppl. 3):11-14.
- Cheeke, P. R.; S. Piacente; and W. Oleszek. (2006). Anti-inflammatory and anti-arthritic effects of yucca schidigera: A review. *Journal of Inflammation* 2006, 3:6 doi:10.1186/1476-9255-3-6.
- Coleman, G. S. (1975). The interrelationship between rumen ciliate protozoa and bacteria. IN: Digestion and Metabolism in the Ruminant (Eds. I W McDonald and A C I Warner) pp149-164. University of New England Publishing Unit, Armidale.
- Comelieus, C.E. (1970). Liver function. In: clinical biochemistry of domestic animals P. 161 (EDS) Kanekof. New York Acad.
- Crutzen, P. J. (1995). The role of methane in atmospheric chemistry and climate. Pages 291–315 in Ruminant Physiology: Digestion, Metabolism, Growth and Reproduction. Proc. 8th Int. Symp. Ruminant Physiol. W. v. Engelhardt, S. Leonhard-Marek, G. Breves, D. Giesecke, eds. Ferdinand Enke Verlag, Stuttgart, Germany.

- Czerucka, D., and Rampal, P. (2002). Experimental effects of Saccharomyces boulardii on diarrheal pathogens. Microbes and infection, 4: 733-739.
- Dann, H. M., J. K I. Drackley, G. C. McCoy, M. F. Hutjens, and J.E. Garrett. (2000). Effects of yeast culture on prepartum intake and postpartum intake and milk production of Jersey cows. J. Dairy Sci. 83: 123.
- Dawson, K.A. (1992). Current and future role of yeast cultures in animal production: A review of research over the last six years. In: supplement to the proceedings of Alltech's 8'h Annual Symposium. I: 23.
- Dawson, K. A. (1993). Current and future role of yeast culture in animal production: a review of research over the last seven years. Page 269 in Biotechnology in the Feed Industry. T. P. Ly-ons, ed. Alltech Tech. Publ., Nicholasville, KY.
- Duncan, D. B. (1955). The Multiple Range and F. Test. Biometrics, 11:1.
- Doumas, B.; W. Wabson and H. Biggs (1971). Albumin standards and measurement of serum with bromocresol green. Clin. Chem Acta, 31:87.
- El-Ashry, M. A.; A. Z. El-Basiony, A. M. El-Serafy, and M. F. Sadek. (1994). Probiotic (LBC) in buffalo heifers ration: 2- effect on som blood parameters. Egyptian J. Anim. Prod. 32 (1): 15-25.
- El-Ashry, M. A; A. M. Kholif; H. A. El-Alamy; H. M. El-Sayed and T. A. El-Hamamsy. (2001). Effect of different yeast cultures supplementation to diet on the productive performance of lactating buffaloes. Egyptian J. Nutrition and Feeds, 4 (1): 21.
- El-Saadany, S. A; M. A. Hanafy; Y. I. El-Talty and A. A. M. El-Mekass. (2002). Performance of lactating Friesian cows fed different rations with or without Baker's yeast) Saccharomyces cerevisiae (supplement. Proc., 1st Ann. Sc. Conf. on Anim. & Fish Prod. Mansoura, Egypt, 24-25 Sep 2002, 107.
- El-Talty, Y.I.; G.M. Abdul-Aziz and AM. Ali (2001). Calcium hydroxide treatment of some organic wastes. 2- Effect on performance of growing goats without or with yeast supplementation. Egyptian. J. Nutrition and Feeds.; 4:415.
- EI-Waziry, AM.; HEM. Kamel and M.H.M. Yacout (2000). Effect of Baker's yeast (Saccharomyces cerevisiae) supplementation to berseem (Trifolium alexandrinum) hay diet on protein digestion and rumen fermentation of sheep. Egyptian J. Nutrition and Feeds, 3:71.
- Ensminger, M. E. (1978). The Stockman's Handbook. (Animal Agriculture Series). 5^{th.} Edition. The Interstate Printers & Publishers, Inc., Danville, Illinois..
- Erasmus L. J., P. M. Botha and A. Kistner. 1992. Effects of yeast culture supplement on production, rumen fermentation, and duodenal nitrogen flow in dairy cows. J. Dairy Sci. 75:3056.
- Francis, George, Zohar Kerem, Harinder P. S. Makkar and Klaus Becker. (2002). The biological action of saponins in animal systems: a review. British Journal of Nutrition, 88, 587–605

- Getachew, G., H. P. S. Makkar, and K. Becker. 2000. Effect of polyethylene on *in vitro* degradability of nitrogen and microbial protein synthesis from tannin-rice browse and herbaceous legumes. British Journal of Nutrition. 84: 73-83.
- Harris, B. and R. Lobo (1988). Feeding yeast culture to lactating dairy cows. J. Dairy Sci., 71: Suppl., 1:276.
- Harris, W. S.; Dujovne C. A.; Windsor S. L.,; Gerrond L. L. C.; Newton F. A. and Gelfand R. A. (1997). Inhibiting cholesterol absorption with CP-88,818 (beta-tigogenin cellobioside; tiqueside): Studies in normal and hyperlipidemic subjects. Journal of Cardiovascular Pharmacology 30, 55–60.
- Harrison, G. A., R. W. Henken, K. A. Dawson, R. J. Harmon, and K. B. Barker. 1988. Influence of addition of yeast culture supplement to diets of lactating cows on ruminal fermentation and microbial populations. J. Dairy Sci. 71:2967.
- Harwood, H. J, Jr. Chandler C. E., Pellarin L. D., Bangerter F. W., Wilkins R.W., Long C. A., Cosgrove, P. G., Malinow M. R., Marcetta C. A., Pettini J. L, Savoy Y. E. and Mayne J. T. (1993). Pharmacologic consequences of cholesterol absorption inhibition: alteration in cholesterol metabolism and reduction in plasma cholesterol concentration induced by the synthetic saponin b-tigogenin cellobioside (CP-88818; tiqueside). Journal of Lipid Research 34, 377–395.
- Hess, H. D., Monsalve L M, Lascano C E, Carulla J E, Diaz T E and Kreuzer M. 2003 Supplementation of a tropical grass diet with forage legumes and Sapindus saponaria fruits: effects on in vitro ruminal nitrogen turnover and methanogenesis. Australian Journal of Agricultural Research 54: 703-713.
- Hsu, J. T.; G. C. Jr, Fahey; L. L. Berger, R. I. Mackie, and N. R. Merchen. 1991. Manipulation of nitrogen digestion by sheep using defaunation and various nitrogen supplementation regimens. Journal of Animal Science 69:1290-1299.
- Husdan,H. (1968). Chemical determination of creatinine with deproteinization; Clin. Chem, 14:222.
- Hutjens, M. F. 1991. Feed additives. Vet Clinics North Am.: Food Animal Practice. 7:2:525.
- Hutjens, M. F. 2005. Managing variation in feed additives. Proc. Southwest Nutrition Conference, 32-42.
- Jin, G. L.; Choi, S. K.; Choi, SH; Song, M. K. (2007). Effect of microbial additives on metabolic characteristics in sheep and milking performance of lactating dairy cows. Journal of Animal Science and Technology. Korean Society of Animal Sciences and Technology. Korea Science & Technology Center, Seoul, Korea Republic: 2007. 49: 6, 819-828.
- Kobayasli, T S.Oda; A. Takenaka and H. Itabashi (1995). Effects of yeast culture supplement on milk protein yield, ruminal fermentation, and blood components in early to mid lactation dairy cows. Bulletin of National Institute of Animal Industry, 55:13.

- Kostner, G.M., P. Avogaro, G. Bittolo Bon,G. Cazzolato and G.B. Quinci (1979). Determination of high-density lipoproteins: Screening methods compared. Clin. Chern. 25/6: 939.
- Leng, R. A. 1990. Factors affecting the utilization of poor-quality forages by ruminants particularly under tropical conditions. Nutrition Research Reviews 3:27-91.
- Ling, E.R. (1963). "Text Book of Dairy Chemistry". Vol. 11. Practical Champan and Hall, L.T.D. London, 4th ed. pp. 140.
- Lovett, D. K., Stack, L., Lovell, S., Callan, J. Flynn, B., Hawkins, M., O'mara, F. P. (2006). Effect of feeding yucca schidigera extract on performance of lactating dairy cows and ruminal fermentation parameters in steer. Livestock Science. 102 (1-2):23-32.
- Lu, C. D. and Jorgensen, N. A. 1987. Alfalfa saponins affect site and extent of nutrient digestion in ruminants. *Journal of Nutrition* 117: 919-927.
- Marghany, M., M. A. Sarhan, A. Abd El-Hey, and A. A. H. El-Tahan. (2005). Performance of lactating buffaloes fed rations supplemented with different levels of baker's yeast Saccharomyces cerevisiae. Egyp. J. Nutrition and Feeds. 8(1), Special Issue:21-34.
- Matsuura M (2001) Saponins in garlic as modifiers of the risk of cardiovascular disease. Journal of Nutrition 131, 1000S–1005S.
- Maynard, L.A. and G.K. Loosli. (1957). Animal Nutrition 6th Ed.. McGraw-Hill, Inc. New York.
- Michalowsky, T. 1988. Importance of protein solubility and nature of dietary nitrogen for the growth of rumen ciliates *in vitro*. IN: The role of protozoa and fungi in ruminant digestion. Proceedings of a seminar held at the University of New England, Armidale, N.S.W. Australia. pp223-232.
- Newbold, C. J.; P. E. V. Williams; N. Mckain; A. Walker and R. J. Wallace. (1.990). The effects of yeast culture on yeast numbers and fermentations in the rumen of sheep. Proc. Nutr. Soc., 49:474.
- Oakenfull DG (1986) Aggregation of bile acids and saponins in aqueous solution. Australian Journal of Chemistry 39, 1671–1683.
- Oakenfull DG & Sidhu GS (1990) Could saponins be a useful treatment for hypercholestero-lemia? European Journal of Clinical Nutrition 44, 79– 88.
- Patton, C.J. and S.R. Crouch (1977). Spectrophotomitic and kinetics investigation of the berthelotteaCffon for the determination of ammonia. Anal. Chern., 49:464.
- Pen. B., Sar, C., Mwenya, B., Kuwaki, K., Morikawa, R., Takahashi, J. (2006). Effect of yucca schidigera and Quillaja saponaria extracts on In Vitro ruminal fermentation and methane emission. Anim. Feed Sci., and Technol. 129 (3-4):175-186.
- Piva, G.; S. Belladanna; G. Fusconi and F. Sicbaldi (1993). Effects of yeast on dairy cow performance, ruminal fermentation, blood components and milk manufacturing properties. J. Dairy SCi., 76:2117.
- Potter SM, Jimenez-Flores R, Pollack J, LoneTA&Berber-Jimenez MD (1993)
 Protein saponin interaction and its influence on blood lipids. Journal of
 Agricultural and Food Chemistry 41, 1287–1291.

- Putnam, DE; C.G. Schwab; M.T. Socha; N.L. Whitestone; N.A. Kierstead and D. Gaithwaite (1997). Effect of yeast culture in the diets of early lactation dairy COWS on ruminal fefmentation and passage of nitrog, R-fraction and amino acids to the small intestine. J. Dairy SCi., 80:374.
- Reitman, S. and S. Frankel (1957). Colorimetric method forthe determination of serum glutamic-oxaloacetic and glutamic-pyrovate transaminase. Am.J. Clin. Path. 28:56.
- Qamar A., Aboudola S., Warny M., Michetti P., Pothoulakis C., Lament J.T. and Kelly C.P. 2001. Saccharomyces boulardii Stimulates IntestinalImmunoglobulin A Immune Response to clostridium difficile Toxin A in mice. Infect. Immun. (am. society for microbiology).69: 2762-2765.
- Robinson, P. H. (1997). Effect of yeast culture (Saccharomyces cerevisiae) on adaptation of cows to diets postpartum. J. Dairy Sci., 80: 1119.
- Robinson, P.H.; and J. E. Garrett. (1999). Effect of yeast culture on adaptation of cows to postparturn diets and on lactational performance. J. Animal Sci. 77:988.
- SAS. (1992). SAS user's Guide, S.A.S. loc., Cary, N.S. U.S.A.
- Shehata, O. (1970). Lectures in Animal Production. Fac., Agric. Ain Shams Univ. (in Arbic).
- Snedecor, G. W. and W.G. Cochran. (1JI82). Statistical Methods. 7^{th.} ed. lowa, lowa University Press. Ames, USA
- Soliman, M.A.; F. A. Salem; D.M. EL-Kerdawy and S.H. Hassanin (1997). Yucca shidigera extract and 'yeast in sheep rations. Egypt. J. Appl. Sci.,12(11): 1-20.
- Thalib, M. A.,B. Haryanto,H. Hamid, D. Suheman, and Mulyani. (2001). Effect of combination of defaunator and probiotic on rumen ecosystem and sheep performance. Jurnal Ilmu Ternak dan Veteriner. Pusat Penelitian dan Pengembangan Peternakan, Bogor, Indonesia: 2001. 6: 2, 83-88.
- Southon S, Johnson IT, Gee JM and Price KR (1988). The effect of Gypsophylla saponins in the diet on mineral status and plasma cholesterol concentration in the rat. British Journal of Nutrition. 59, 49– 55.
- Ushida, K., J. P. Jouany, C. Kayouli, and D. I. Demeyer. (1988) Effect of defaunation on fibre digestion in sheep fed ammonia-treated straw based diets. IN: The role of protozoa and fungi in ruminant digestion. Proceedings of a seminar held at the University of New England, Armidale, N.S.W. Australia. pp307- 308.
- Valdez FR, Bush LJ, Goetsch AL & Owens FN (1986) Effect of steroidal sapogenins on ruminal fermentation and on production of lactating dairy cows. Journal of Dairy Science 69, 1568–1575.
- Van Keulen, J. and PAYoung (1977). Evaluation of acid insoluble ash as a natural marker in ruminant digestibility studies. J. Anim. Sci., 44: 282, . . .
- Wallace, R. J. (2007). Plant extracts as antimicrobials in ruminants. 41st
 University of Nottingham Feed Conference. Rowett Research Institute,
 Bucksburn, Aberdeen, AB21 9SB. UK.

- Wang, Y., McAllister, T.A., Yanke, L.J., Cheeke, P.R. (2000). Effect of steroidal saponin from Yucca schidigera extract on ruminal microbes. Journal of Applied Microbiology 88:887-896.
- Weller, R. A., and A. F. Pilgrim. (1974). Passage of protozoa and volatile fatty acids from the rumen of the sheep and from continuos in vitro fermentation system. British Journal of Nutrition 32:341-351.
- Wohlt, J.E.; A.D. Finkelstein and C.H. Chung (1991). Yeast culture to improve intake, nutrient digestibility performance by dairy cattle during early lactation. J. Dairy Sci., 74: 1395.
- Wohlt, J.E.; T.T. Carcione and P.K. Zajak (1998). Effect of yeast on feed intake and performance of cows fed diets based on corn silage during early lactation. J. Dairy Sci.. 81:1345.
- Wiedmeier, R.D.; M.J.Arambel and J.L. Walters (1987). Effect of yeast culture and Aspergillus oryzae fermentation extract on ruminal characteristics and nutrient digestibility. J. Dairy Sci., 70:2068.
- Williams, P.EV (1988). Understanding the biochemical mode of action of yeast culture. In Biotechnology in the Feed Industry (ed. T.P. Lyons), pp. 79, Alltech Technical publications. Nicholasville. Kentucky, USA.
- Williams, P.EV (1989). The mode of action of yeast culture in ruminant diets:

 A review of the effect on rumen fermentation patterns. pp. 65 in
 Biotechnology in the Feed Industry. Alltech Tech. Publ., Nicholasville,
 Kentucky, U.S.A.
- Wina, E., S. Muetzal, E. Hoffman, H. P. S. Makkar, and K. Becker. (2003). The effect of secondary compounds in forages on the rumen microorganisms quantified by 16S and 18S rRNA.
- Wina, E; Muetzel S; Becker K. (2005). The impact of saponins or saponincontaining plant materials on ruminant production--a review. J Agric Food Chem. 53(21):8093-105. Institute for Animal Production in the Tropics and Subtropics (480b), University of Hohenheim, Stuttgart 50793, Germany.
- Wina E, Muetzel S, Becker K. (2006). The dynamics of major fibrolytic microbes and enzyme activity in the rumen in response to short- and long-term feeding of Sapindus rarak saponins Journal of Applied Microbiology 100 (1): 114-122.

تأثير مستخلص الصابونين النباتى الطبيعى بدون أو مع خميرة الخباز الطرية الطازجة فى علائق الجاموس الحلاب عل هضم العناصر الغذائية ، وإنتاج اللبن وتركيبه ، وعلى بعض عوامل سيرم الدم.

سليمان محمد سليمان عبد المولى

قسم الإنتاج الحيواني - كلية الزراعة - جامعة عين شمس - حدائق شبرا ١١٢٤١ .

فى تجربة استمرت ٩٠ يوما لتقدير أثار استخدام مستخلص الصابونين النباتي الطبيعي بمفرده أو مع خميرة الخباز الطرية الطازجة فى علائق الجاموس الحلاب. استخدم ٢٠ جاموسة حلابة بمتوسط وزن حى ٩١٥ كجم. وزعت الحيوانات على ٤ مجموعات من خمس حيوانات تبعا للعمر ثم وزعت عشوائيا على أربعة معاملات العلائق التجريبية. وكانت المعاملات التجريبية متمثلة فى: Τ1 مجموعة المقارنة وغنيت على العليقة الأساسية (تكونت من علف مركز مكعبات: برسيم مصرى:قش أرز بنسبة ٢٥٠٠:٥٠٠% على أساس المادة الجافة)، Τ2 التي غنيت على العليقة الأساسية + ٨٠٠ ملليجرام (مجم) صابونين ، Τ3 التي غنيت على العليقة الأساسية + ٨٠٠ مجم صابونين + ٢٠٠ جم خميرة خباز طرية طازجة ، ۲4 التي غذيت على الأساسية + ٨٠٠ مجم صابونين + ٢٠٠ جم خميرة خباز طرية طازجة.

أظهرت النتائج أن إمداد الحيوانات بمستخلص الصابونين بدون أو مع ١٠ أو ٢٠ جم من خميرة الخباز الطرية أدى إلى تحسن (P<0.05) معاملات هضم العناصر الغذائية ، محصول اللبن ، بروتين ودهن اللبن ، اللاكتوز ، الجوامد الكلية ، والجوامد غير الدهنية للمعاملات , 72 اللبن ، مجموعة المقارنة. أدت الإضافات إلى زيادة (P<0.05) تركيز قياسات الدم: البروتين الكلى ، الألبيومين ، الجلوبيولين. وأدت إلى نقص (P<0.05) الكوليستيرول وال . GPT

أظهرت المعاملة الرابعة 74 أعلى (P <0.05) القيم لمعاملات هضم العناصر الغذائية ومحصول اللبن ومكوناته ، والبروتين والدهن، كما أنها أظهرت أعلى تركيز (P <0.05) لقياسات سيرم الدم : البروتين الكلى ، والألبيومين والجلوبيولين ، وأقل تركيز (P <0.05) لليوريا والكوليستيرول وال GPT وال GOT مقابل المجموعات الأخرى.

أظهرت المعاملة T2 (العليقة القاعدية+٨٠٠ مجم صابونين) استجابات أعلى (٣٥٥.٥%) من المجموعة T1 المقارنة المعاملات الهضم ، محصول اللبن ومكوناته ، واللا كتوز وبروتينات البن والجوامد الكلية والجوامد اللادهنية ، ولكنها لازالت أقل (٣<0.0%) من قيم أداء المجموعات T3, T4.

تظهر الدراسة أن العلائق التي أغنيت بإضافة الصابونين مع ١٠ أو ٢٠ جم خميرة الخباز الطرية الطازجة وخصوصا المجموعة الرابعة T4 والتي أعطت أعلى قيم هضم العناصر الغذائية وإنتاج اللبن وتركيبه مع عدم وجود أى أثار غير طبيعية أو مرضية كما ظهر من نتائج دراسة قياسات سيرم الدم.