# ANTIMICROBIAL RESISTANCE AND/OR SUSCEPTIBILITY OF BACTERIA IMPLICATED IN URINARY TRACT INFECTION

Mansour, F.A.\*; M. F. Ghaly\*\*; F. Elgenawy\*\*\* and A. A. Foda\*\*

- \* Botany Department, Faculty of Science, Mansoura University
- \*\* Botany Department, Faculty of Science, Zagazig University
- \*\*\* Microbiology and Immunology dept., Fac. of Medicine, Mans. Univ.

#### **ABSTRACT**

A total of 272 urine samples were collected from patients examined for urinary tract infections in Mansoura University hospitals or attending Mansoura University outpatients clinics. A positive bacterial growth on CLED, MacConkey and blood agars was detected in 36.77 % of the cases only. E. coli was the most predominant pathogen causing urinary tract infection in the examined positive samples (38.09%) followed by Klebsiella pneumonia , Klebsiella oxytoca Pseudomonas areuginosa . Staphylococcus aureus. Proteus mirabilis .Enterococcus Proteus vulgaris and Citrobacter freundii were present in the following frequencies 21.42, 9.52, 9.52, 7.14, 5.95, 4.76, 2.38, and 1.19 respectively. Antibiotic sensitivity tests for Gram negative bacteria and Gram positive bacteria showed that most of the Gram-negative bacteria were highly susceptible to meronem , nitofurantion, norfloxacin and Amikacin, While the Gram-positive bacteria were highly susceptible to meronem and nitofurantion. The effects of volatile oils against highly resistant bacteria isolates showed that marioram and dianthus were inhibitory to these isolates. Interleukin-8 was taken as a measure of UTI severity. The mean IL-8 levels in urine samples containing Proteus mirabilis, Proteus vulgaris, Klebsiella oxytoca, Klebsiella pneumoniae, E. coli and Pseudomonas areuginosa were 56,21±5,85 pg/ml. 102.95±47.97 pg/ml, 139.24±40.79 pg/ml, 259.58±61.08 pg/ml, 754.47±279.71 pg/ml and 1200.65±268.36 pg/ml respectively. While, the mean IL-8 levels in urine samples infected with Staphylococcus aureus and Enterococcus spp. were 100.09±27.95 pg/ml and 110.38±8.83 pg/ml respectively. A statistically significant differences (P ≤0.0001) in IL-8 levels among UTIs caused by Gram positive cocci, Gram negative bacilli, in either males and females were observed.

#### INTRODUCTION

Urinary tract infection (UTI) is one of the most important causes of morbidity in the general population, and is the second most common cause of hospital visits (Ronald and Pattulo, 1991). They are also among the most common bacterial infections in humans, both in the community and hospital settings (Hooton and Stamm, 1997), and have been reported in all age groups of both sexes. It is characterized by the evidence of uropathogens and pyuria and is accompanied by various clinical manifestations depending on the area of involvement. Although the detection of pyuria in patients with suspected UTI is readily available via laboratory test, but sometimes it is a non-specific indicator of UTI (Johnson and Stamm, 1999) and little is known about the mechanism of neutrophil recruitment into the urine (Ko et al., 2000). Nearly all UTIs are caused by bacteria that enter the urethral opening and

move upwards to the urinary bladders and sometimes to kidneys (Lerner, 1994).

UTIs occur in all populations and ages, however, infection is most common in women, especially sexually active women. About 20% of women experience a single episode of UTI during their lifetime, and 3% of women have more than one episode of UTI per year (Gebre-selassie, 1998). Recurrent infections are common and can lead to irreversible damage of the kidneys, resulting in renal hypertension and renal failure in severe cases (New, 1992).

The choice of antibiotics should depend upon the causative organism and their susceptibility pattern to various antibiotics (Goldstein , 2000). Proper management and prevention of bacteriuria can reduce the incidence of the life-threatening consequences of urinary tract infections. Together , physicians and laboratories must have guidelines and strategies that provide high quality treatment for the patient, while minimizing costs and preventing further emergence of antimicrobial drug resistance ( Lo and Smego, 2004; Higuchi et al., 2006).

Bacterial pathogens stimulate epithelial cells of interstitial tissue and macrophages to secrete proinflammatory cytokines viz: interleukin 1 (IL-1 beta), interleukin 6 (IL-6) and interleukin 8 (IL-8) (Hack et al., 1992; Hedges et al., 1992; Agace et al., 1993a&b). It has been demonstrated before that IL-8 levels were elevated in patient with PN and that the level of this chemokines may correlate with the virulence associated traits of the microorganisms (Jacobson et al., 1994). Amongst the many diseases in which IL-8 plays an important role as mediators of inflammation (Baggiolini et al., 2001). UTI were no exception (Agace et al., 1993 and Ko et al., 2000). The role of chemokines in the development and progression of inflammatory renal diseases, particularly pyelonephritis (PN) has not been thoroughly investigated.

This study investigated the resistance and susceptibility of bacteria implicated in urinary tract infection to some available antimicrobial drugs and evaluated the levels reliability IL-8 as a good marker for the severity of urinary tract infection .

#### MATERIALS AND METHODS

## Samples Collection

During the period between April 2007 to April 2008, a total of 272 urine samples were collected from UTIs patients at Mansoura University hospitals or outpatients clinics. Urine samples were collected in sterile wide mouth containers under complete aseptic condition and transferred immediately to the various laboratory tests or kept at 4° C for further examinations (Warren, 1996; NCCLs, 2001).

#### Isolation and Identification of Bacterial Isolates

Each sample was immediately tested for bacterial density on peptone water. They were inoculated and plated onto CLED agar, blood agar and MacConkey agar plates and incubated for 48 hours at 37 C. Positive culture

was defined if bacterial colony counts was  $\geq$  10 <sup>5</sup> colony forming units/ml. All bacteria were identified to species level according to standared procedures (Bergy's Manual, 1994); ( Mahon and Manuselis 1995; Collee *et al.*, 1996 and Zinsser, 1998).

### **Antibiotic susceptibility tests**

All bacterial strains isolated were tested for antimicrobial sensitivity by standardized disk diffusion technique as adapted and described by Bauer et al., (1986).

## Measurements of interleukin-8 (IL-8) level in urine samples:

The urinary IL-8 levels in urine samples from patients with UTI was determined by the Biosource IL-8 EASIA. of a solid phase Enzyme Amplified Sensitivity Immunoassay (EASIA) based on an oligoclonal system in which a blend of monoclonal antibodies (MAbs) directed against distinct epitopes of IL-8 are used.

#### RESULTS

Of the 272 urine samples cultured, only 100 (36.77%) were positive for UTI. Ages of positive cases ranged from 5 months to 83 years, they including 53 males and 47 females. Gram negative bacteria were the most common cause of UTIs which present in 88.07 % of patients while Gram positive bacteria were present in 11.9 % of patients. The frequency of bacterial pathogens isolated from positive cultures ( table.1) indicated that *E. coli* was the most predominant pathogen causing urinary tract infection in human (38.09%) followed by *Klebsiella pneumonia . Klebsiella oxytoca , Pseudomonas areuginosa , Staphylococcus aureus, Proteus mirabilis ,Enterococcus spp., Proteus vulgaris and Citrobacter freundii were present in low frequencies 21.42, 9.52, 9.52, 7.14, 5.95, 4.76, 2.38, and 1.19 respectively. Table (2) shows bacterial species isolated different ages and genders.* 

- Table 1: The frequency of bacterial pathogens isolated from positive cultures.

cultures.	<u> </u>			
Bacterial isolates	Number	%		
1-E. coli .	32	38.09		
2-Klebsiella pneumoniae .	18	21.42		
3-Klebsiella oxytoca .	8	9.52		
4-Citrobacter freundii .	1 1	1.19		
5- Pseudomonas areuginosa.	8	9.52		
6-Proteus vulgaris .	2	2.38		
7-Proteus mirabilis .	5	5.95		
8-Stphylococcus aureus .	6	7.14		
9-Enterococcus spp .	4	4.76		
Total	84	100		

Most of the Gram-negative bacteria were highly susceptible to meronem, nitofurantion, norfloxacin and Amikacin antibiotics. While the

Gram-positive bacteria were highly susceptible to meronem and nitofurantion (Table 3). *E. coli*, was susceptible to meronem (78.1%), nitofurantion (81.2%), norfloxacin (59.3%) and Amikacin (78.1%) but Klebsiella pneumoniae was highly susceptible to meronem (83.3%) and norfloxacin (77.7%) and *Staphylococcus aureus* was susceptible to meronem (83.3%) and nitofurantion (100%).

Table 2: Bacterial species isolated from cases of different ages and different genders.

	GIII	<u> </u>	iit ger	lucis.				
Patient No of Sex		Sex	Bacterial Isolates					
age	cases	M	F					
0 – 10	9	6	3	E. coli , Klebsiella pneumoniae , Klebsiella oxytoca , pseudomonas areuginosa , proteus mirabilis and Enterococcus spp.				
10-20	5	4	1	E. coli , Klebsiella pneumoniae , Klebsiella oxytoca and proteus vulgaris.				
20-30	4	3	1	E. coli.				
30-40	9	3	6	E. coli . Kiebsiella pneumoniae , Klebsiella oxytoca . staphylococcus aureus and Enterococcus spp.				
40-50	20	8	12	E. coli , Klebsiella pneumoniae , Klebsiella oxytoca . pseudomonas areuginosa , proteus mirabilis and Enterococcus spp.				
50-60	18	6	12	E. coli , Klebsiella pneumoniae , Klebsiella oxytoca . pseudomonas areuginosa , proteus vulgaris , staphylococcus aureus and Enterococcus spp.				
60-70	9	8	1	E. coli , Klebsiella pneumonia e, Klebsiella oxytoca , pseudomonas areuginosa , proteus mirabilis and Citrobacter freundli.				
70-80	5	3	2	E. coli . Klebsiella pneumoniae and staphylococcus aureus.				
80-90	2	1	1	Klebsiella pneumoniae and proteus mirabilis.				

Table 3: Susceptibility of isolated Gram-negative and Gram-positive bacteria

Bacterial isolates	Total	Susceptibility to antimicrobial drugs (%)									
	no.	CN	CTX	SXT	MEM	F	NA	NOR	AK	CEC	AMC
Gram-negative					1					[	1
bacteria.					1					ł	ŀ
Escherichia coli	32	62.5	25	25	78.1	81.2	40.6	59.3	78.1	21.8	21.812
Klebsiella oxytoca	8	50	25	-	75	62.5	37.5	50	· 75	12.5	.5
Klebsiella pneumoniae	18	55.5	22.2	22.2	83.3	55.5	44.4	77.7	61.1	11.1	22.2
Pseudononas areugnosa	8	50	_	12.5	62.5	25	12.5	50	75	-	37.5
Proteus vulgaris	2	50	100	-	50	50	50	50	50	50	-
Proteus mirabilis	5	60	40	40	80	40	-	100	60	60	40
Citrobacter freundii	1	100	<b>–</b>	<del> </del>	-	100	100	100	-	-	-
		CN	SXT	SAM	MEM	F	CL.	NOR	CIP	CEC	AMC
Gram-positive										}	
bacteria.			1	i		1	]	i		ļ	į
Sephylococus aueus.	6	66.6	16.6	50	83.3	100	33.3	50	50	16.6	66.6
Enterococcus spp.	4	50		50	75	75	25	75	50	-	

AK = Amikacin ( 30  $\mu$ g); AMC = Augmentin( 20  $\mu$ g); CEC = Cefaclor( 30  $\mu$ g); CTX = Cefotaxime( 30  $\mu$ g); CL = Cephalexin( 30  $\mu$ g); CIP = Ciprofloxacin( 5  $\mu$ g); CN = Gentamicin( 10  $\mu$ g); MEM = Meronem( 10  $\mu$ g); NA = Natidixic acid ( 30  $\mu$ g); F = Nitrofurantion( 300  $\mu$ g); NOR = Norfloxacin ( 10  $\mu$ g); SXT = Trimethoprim + sulphamethoxazole( 1.25  $\mu$ g); SAM = Unasyn( 10  $\mu$ g).

Volatile oils activity against highly resistant isolated bacteria shown in (Table 4) indicated that the marjoram inhibitory effect with Enterococcus spp. (Es.1), Klebsiella oxytoca (Ko.3) and Pseudomonas areuginosa (Pa.2) with a recorded inhibition zone (15mm, 12mm and 10mm respectively) and the other isolates gave lower inhibition zone. Dianthus showed inhibitory action against Klebsiella pneumoniae (Kp.3), Pseudomonas areuginosa (Pa.2), Enterococcus spp. (Es.1), Klebsiella oxytoca (Ko.3) and Klebsiella pneumoniae (Kp.8) and the highest inhibition zones were 13 mm which recorded with Klebsiella pneumoniae (Kp.3). Mean while, thyme, carawaya and eucalyptus show no inhibitory effect aganist all tested bacterial isolates.

Table 4. Diameter of inhibition zone(mm) of Bacterial isolates to various volatile oils.

Diameter of inhibition zone(mm)  Bacterial isolates	Eucalyptus	Cummin	Marjoram	Carawaya	Dianthus	Anise	Thyme
E. coli (Ec.9)	0	10	8	0	9	0	0
Klebsiella pneumoniae (Kp.3)	0	11	9	0	13	0	0
Klebsiella pneumoniae(Kp.8)	0	8	9	0	10	0	0
Klebsiella oxytoca (Ko.3)	0	11	12	0	10	0	0
Pseudomonas areuginosa (Pa.2)	0	9	10	0	12	0	0
Pseudomonas areuginosa (Pa.3)	0	0	0	0	0	0	0
Pseudomonas areuginosa (Pa.5)	0	0	0	0	8	0	0
Enterococcus spp. (Es.1)	0	11	15	0	12	10	0

# IL-8 levels in urine of patients with UTI due to Gram negative bacteria.

The results presented in figure (1) showed that the mean IL-8 levels in urine samples infected with *Proteus mirabilis, Proteus vulgaris, Klebsiella oxytoca, Klebsiella pneumoniae, E. coli* and *Pseudomonas areuginosa* were 56.21, 102.95, 139.24, 259.58, 754.47 and 1200.65 pg/ml respectively.

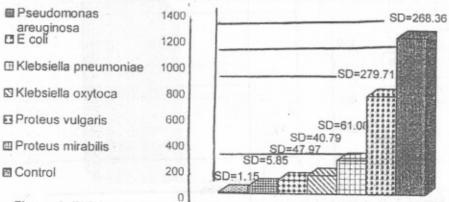


Figure 1. IL-8 levels in urine of patients with UTI due to Gram negative bacteria (pg/ml).

While the mean IL-8 levels were 5.81 pg/ml in control urine samples. A significant differences (P<0.05) between the mean values of IL-8 in urine samples infected with Gram negative bacteria and control urine samples is evident

# IL-8 levels in urine of patients with UTI due to Gram positive bacteria.

The results depicted in figure (2) revealed that there were significant differences between the mean values of IL-8 in urine samples infected with Gram positive bacteria (P<0.05) as compared to control urine samples, where the mean IL-8 levels in urine samples infected with Staphylococcus aureus and Enterococcus spp. were 100.09 pg/ml and 110.38 pg/ml respectively but in control was 5.81 pg/ml.

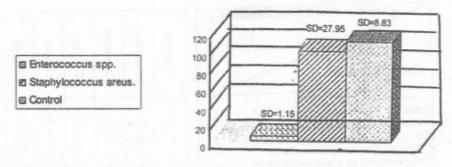


Figure 2. IL-8 levels in urine of patients with UTI due to Gram positive bacteria (pg/ml).

# IL-8 levels in urine of patients with UTI with different gender.

The results depicted in figure (3) revealed that the mean IL-8 levels in males and females were 285.59 pg/ml and 580.32 pg/ml. Meanwhile, there was a significant difference (P<0.05) between mean IL-8 level in males and females.

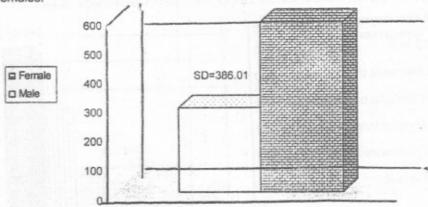


Figure 3. IL-8 levels in urine of patients with UTI with different gender (pg/ml).

IL-8 levels in urine of patients with UTI with different age.

The results presented in figure (4) showed that the mean IL-8 levels in the different age group as 1(0-20y), (>80y), (20-40y), (60-80y) and (40-60y) were 137.55, 142.77, 368.85, 580.62 and 589.11 respectively. So there were significant differences (P<0.05) between the mean values of urinary IL-8 in the different age groups.

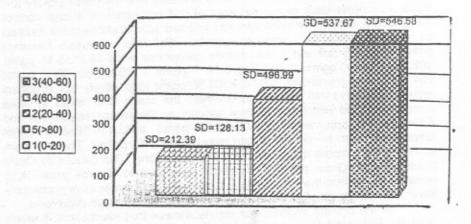


Figure 4. IL-8 levels in urine of patients with UTI with different age (pg/ml).

# DISCUSSION

This study revealed that Gram negative bacteria were the most common cause of UTIs which present in 88.07 % of patients while Gram positive bacteria were present in 11.9 % of patients. *E. coli* was the most predominant pathogen causing urinary tract infection in human (38.09%). The other pathogens namely: *Klebsiella pneumonia*, *Klebsiella oxytoca*, *Pseudomonas areuginosa*, *Staphylococcus aureus*, *Proteus mirabilis*, *Enterococcus spp.*, *Proteus vulgaris and Citrobacter freundii* were present in low frequencies 21.42, 9.52, 9.52, 7.14, 5.95, 4.76, 2.38, and 1.19 respectively. This study was consistent with other studies which showed that *E. coli* was the most frequent cause of UTIs at all ages (Garcia and Nager, 2002; Maherzi *et al.*, 1978; Abbott, 1972; Tamim *et al.*, 2003; Bergstrom *et al.*, 1972; Ginsberg and McCracken, 1982). Yuksel *et al.*, (2006) reported that the most causative agents was *E. coli* (87%of cases) followed by *Klebsiella pneumonia* (10%) and this agree with this study.

The results of the present study showed that the susceptibility rate of urinary isolates was highest for meronem (76.19 %), followed by amikacin (70.27 %), nitrofurantion (66.60 %), norfloxacin (64.28) and gentamicin (58.33 %). Meanwhile, the resistant rate of urinary isolates was highest for cefaclor (71.42 %) and trimethoprim-sulphamethoxazole (70.23 %). Adeyemo

and colleagues (1994) reported that all urinary isolates were poorly susceptible to trimethoprim-sulphamethoxazole and ampicillin, but exhibited good susceptibility to nalidixic acid, nitrofurantion, and ofloxacin. Yuksel et al., (2006) showed that nitrofurantion was the most active agent against *E. coli* (2.2% resistant isolates), followed by amikacin (4.9%), ceftriaxone (7.5%) and ciprofloxacin (12%) and this agree with this study.

The results of the present study revealed that the mean urinary IL-8 levels was remarkably higher in cases of UTI compared to normal control. The mean IL-8 levels in urine samples infected with Gram negative bacteria as Proteus mirabilis, Proteus vulgaris, Klebsiella oxytoca, Klebsiella pneumoniae, E. coli and Pseudomonas areuginosa were 56.21±5.85 pg/ml, 102.95±47.97 pg/ml, 139.24±40.79 pg/ml, 259.58±61.08 754.47±279.71 pg/ml and 1200.65±268.36 pg/ml respectively but in control group was 5.81±1.15 pg/ml. Mean while, the mean IL-8 levels in urine samples infected with Gram positive bacteria as Staphylococcus aureus and Enterococcus spp. were 100.09±27.95 pg/ml and 110.38±8.83 pg/ml respectively but in control group was 5.81±1.15 pg/ml. There statistically significant differences (P≤0.0001) among UTIs caused by Gram positive cocci, Gram negative bacilli as compared to control group. Also results showed that the mean urinary IL-8 levels in Gram positive cocci were lower than the mean urinary IL-8 levels in Gram negative bacilli organisms.

In the present investigation results showed that the mean IL-8 levels of males and females were 285.59 $\pm$ 386.01 and 580.32 $\pm$ 570.34 respectively there was a significant difference (P $\leq$ 0.0001) between mean IL-8 levels in males and females. The mean IL-8 levels in the different age group as 1 (0-20y), 5 (>80y), 2 (20-40y), 4 (60-80y) and 3 (40-60y) were 137.55 $\pm$ 212.39, 142.77 $\pm$ 128.13, 368.85 $\pm$ 496.99, 580.62 $\pm$ 537.67 and 589.11 $\pm$ 546.58 respectively. Mean while there were significant differences (P $\leq$ 0.0001) between the mean values of urinary IL-8 in the different age groups.

#### REFERENCES

- Abbott, G.D. (1972). Neonatal bacteriuria: a prospective study in 1,460 infants. Br Med J. 1:267-269.
- Adeyemo, A.A.; Gbadegesin R.A.; Onyemenem, T.N. and Ekweozor C.C. (1994). Urinary pathogens and antimicrobial sensitivity patterns in children in Ibadan, Nigeria. Ann Trop Paediatr; 14:271-274.
- Agace, W.; Hedges, S.; Ceska, M. and Svanborg, C. (1993a). Interleukin-8 and neutrophil response to mucosal gram-negative infection. J. Clin. Invest. 92: 780-785.
- Agace, W.; Hedges, S.; Andersson, U.; Andersson, J.; Ceska, M. and Svanborg, C. (1993b). Selective cytokine production by epithelial cells exposure to E coli. Infect. Immun., 61: 602-609.
- Baggiolini, M.; Dewald, B. and Moser, B. (2001). IL-8 and related cytokines. Advances in immunology. Vol. 55: 97-179.
- Bauer, A.W.; Kirby, W.M.; Sherris, J.C. and Turk, M. (1986). Antibiotics susceptibility testing by a standerized single disk method am. J of C1. pathology, 45:493-496.

- Bergstrom, T., Larson, H., Lincoln, K. and Winberg, J. (1972). Studies of urinary tract infections in infancy and childhood. XII. Eighty consecutive patients with neonatal infection. J pediatr., 80:858-866.
- Bergy's Manual of Determinative Bacteriology (1994). Facultatively anaerobic gram-negative rods (eds.). Holt, J.G.; Krieg, N.R.; Sneath, P.H.A.; Stalety, J.T. and Williams, S.T.9<sup>th</sup> ed.., William and Wilkins, Baltimore, pp.189-252.
- Collee, J.G.; Duguid, J.P.; Fraser, A.G.; Marmion, B.P. and Simmons, A. (1996). Laboratory strategy in the diagnosis of the infective syndromes. In: Collee, J.G.; Fraser, A.G.; Marmion, B.P. and Simmons, A. (eds.): Mackie and MacCarthey Practical Medical Microbiology, pp.53-94. Churchill Livingstone, New York, London, Tokyo.
- Garcia, F.J. and Nager, A.L. (2002). Jaundice as an early diagnostic sign of urinary tract infection in infancy. Pediatrics., 109:846-851.
- Gebre-Selassie, S. (1998). Asymptomatic bacteriuria in pregnancy: epidemiological, clinical and microbiological approach. Ethiop Med J., 36:185-192.
- Ginsberg, C.M. and McCracken, G.H., J.R. (1982). Urinary tract infections in young infants. Pediatrics., 69:409-412.
- Goldstein, F.W. (2000). Antibiotic susceptibility of bacterial strains isolated from patients with community acquired urinary tract infections in France. Multicentre Study Group. Eur. J. Clin. Microbiol. Infect. Dis., 19: 112,. M. Abu Setteh.
- Hack, CE.; Hart, M.; Strack, R.J.M.; Eerenberg, A. j. M.; Nuijens, J.H.; Thijs, L. G. and Aarden, L.A.(1992). Interleukin-8 in sepsis: relation to shock and inflammatory mediators. Infect. Immun., 60:2835-2842.
- Hedjes, S.; Stenquist, L.; Lidin-Janson, G.; Martinell, J.; Sandberg, T. and Svanborg, C. (1992) Comparison of urine and serum concentrations of interleukin-6 in women with acute pyelonephritis or asymptomatic bacteriuria. J. Infect. Dis., (166): 653-656.
- Higuchi, C.K.; Correa, C.R.; Rall, V.L.M. and Junior, A.F. (2006). Evaluation of the diramic system for urine cultures. Brazilian J. Microbial., 37(3): 122-129.
- Hooton, T.M. and Stamm W.E. (1997). Diagnosis and treatment of uncomplicated urinary tract infection. Infect. Dis. Clin. North Am., 11: 551-581.
- Jacobson, S.H.; Hylander, B.; Wretlind, B. and Brauner, A. (1994). Interleukin-6 and interleukin-8 in serum and urine in patients with acute pyelonephritis in relation to bacterial-associated traits and renal function. Nephron. 67: 172–179.
- Johnson, J.R., and Stamm, W.E. (1999): urinary tract infection in women, diagnosis and treatment. Ann. Intern. Med., 111:906.
- Ko, Y. C., Mukaida, N., and Ishiyama, S. (2000). elevated IL-8 levels in urine of patient with UTIs. Infect. And Immunol. 61:1307-1341.
- Lerner, G.R.(1994). Urinary tract infections in children Pediatr. Ann.,23; 463-473.
- Lo, T.S. and Smego, R.A. (2004). Avoiding laboratory pitfalls in infectious diseases. Postgraduate Med. J., 80: 660-662.

#### Mansour, F.A. et al.

- Maherzi, M.; Guignard, J.P. and Torrado, A. (1978). Urinary tract infection in high-risk newborn infants. Pediatrics., 62:521-523.
- Mahon, C.R. and Manuselis, J.R.G. (1995). Textbook of diagnostic Microbiology (ed.) W.B. Saunders Co., Philadelphia., pp. 1134.
- NCCLs (2001). Approved Guideline, 2<sup>nd</sup> ed. Routine urinanalysis and collection, transportation and preservation of urine specimens. Document GP16-A2, Nov. 2001.
- New, H.C.(1992). Urinary tract infections. Am. J. Med., 92(Suppl. 4A): S63-S70.
- Ronald, A.R. and Pattulo, M.S. (1991). The natural history of urinary infection in adults. Med Clin North Am., 75:299-312.
- Tamim, M.M.; Alesseh, H. and Aziz, H. (2003). Analysis of the efficacy of urine culture as part of sepsis evaluation in the premature infant. Pediatr Infect Dis J., 22:805-808.
- Warren , J. W. (1996). Clinical presentation and epidemiology of urinary tract infections. P.3-27. In H.L.T. Mobley and J. W. Warren (ed.), Urinary tract infections. Molecular pathogenesis and clinical management. American Society of Microbiology, Washington, D.C.
- Yuksel, S.; Ozturk, B.; Kavaz, A.; Ozcakar Z.B.; Acar, B.; Guriz, H, Aysev, D. Ekim, M. and Yalcinkaya, F. (2006). Antibiotic resistance of urinary tract pathogens and evaluation of empirical treatment in Turkish children with urinary tract infections. Int. J. Antimicrob. Agents., 5: 413-416.
- Zinsser, H. (1998). Zinsser Microbiology, 20th ed. Appleton & Lange, USA., Pp:420-693.

دراسات على حساسية ومقاومة البكتريا المتورطة فسى عدوى القنساة البوليسة للمضادات الميكروبية

فتحى عواد منتصور "، محمد فساروق غسالى " "، فسؤاد القنساوى " " و عبد السميع عزيز عبد السميع فوده " "

- قسم النبات \_ كلية العلوم جامعة المنصورة
- قسم النبات \_ كلية الطوم جامعة الزقازيق
- \*\*\* قسم الميكروبيولوجي والمناعة \_\_ كلية الطب جامعة المنصورة

أجريت هذه الدراسة على ٢٧٢ عينة بول تم جمعها من المرضي المقيمين بمستمشيات جامعة المنصورة والمترددين على العيادات الخارجية بجامعة المنصورة. ووجد أن ٣٦،٧٧ ٪ فقط من عينات البول التي تم جمعها أعطوا نمو بكتيري ايجابي على وسط مغذي من الكليدأجار. كمـــا اتُبِنَتُ نَتَائِج هذه الدراسة أن إشيريشياكو لاي "Escherichia coli" هي أكثر أنواع البكتيريا التي تصيب الجهاز البولي حيث أنها تمثل ٣٨٠٠٩% من اجمالي العينات التي اعطبت نمو بكتيسري ابجابي ٠ اما بقبة البكتريا و هي كليب سيلانو مو نيا Klebsiella pneumonia، كليبسيلااو كسمي تركا Klebsiella oxytoca ، سودوموناس إيروجينوزا Klebsiella oxytoca ، ستافیلو کو کاس او ریوس Staphylococcus aureus ، بروتیسوس میسر ابیاس Proteus mirabilis انتير و كو كاس سبيشن Enterococcus spp ، بروتيوس فالجارز Proteus vulgaris، ستروباكتر فرونداي Citrobacter freundii تمثل المعدلات الأتية على التوالي (۲۱,٤٢ ، ۹,۵۲ ، ۹,۵۲ ، ۷,۱٤ ، ۵,۹۵ ، ۲۷,٤ ، ۲۰۳۸ ، ۱،۱۶). وتم عمل اختبار حــساسية لجميع المعزولات البكتيرية فأوضحت هذه الدراسة أن البكتريا العصوية السالبة تبدى حساسية عالية تجاه الميرونام النيتروفيورانتوين النورفلوجساسين وأميكاسين ولكس البكتريسا الكرويسة الموجبة تبدى حساسية عالية تجاه الميرونام والنيتروفيورانتوين. وبعمل اختبار حسساسية لـبعض البكتريا التي اعطت مقاومة عالية تجاة المضادات الحيوية المستخدمة باسستخدام بعسض الزيسوت العطرية فأثبتت نتائج الدراسة ان هذه البكتريا اعطت حساسية تجاه القرنفل والبردقوش. وبدر اســـة تأثير العامل المناعي انترليوكين- ٨ (B-IL) في عينات البول التي أعطت نمو بكتيري إيجسابي ، أظهرت الدراسة أنَّ المعزولات البكتيرية العصوية السالبة وهي بروتيوس فالجـــارز ، بروتيـــوس مير ابيلس ، كليبسيلالوكسي توكاو ، كليبسيلانومونيا ، إشير يشياكو لاى ، سودوموناس ايروجينوز ا كان متوسط تركيز الانترليوكين هو ٥٦،٢١ ، ٥٠٩٠٠ ، ١٣٩٠٢ ، ٢٥٩٠٥٨ ، ٧٥٤٠٤٧ ، ١٢٠٠٠٦٥ بيكوجرام لكل ملي على التوالي . ولكن متوسسط تركيـــز الإنترليـــوكين فــــي حالــــة المعزو لات البكتيرية الكروية الموجبة وهي ستافيلوكوكاس أوريوس ، إنتيروكوكاس سبيــشز هـــو ١٠٠٠٠٩ ، ١٠٠٠٣٨ بيكوجرام لكل ملى على التوالي ومن خلال هذه الدراسسة نجــد ان هنـــاك فروق معنوية (P≤٠٠٠٠١) بين عنوى الجهاز البولي" Urinary tract infection" الناتجة عن المعزولات البكترية العصوية السالبة و المعزولات البكترية الكروية الموجبة عند مقارنتها بعينات الكونترول ، واذلك يمكن اعتبار تركيز الإنترليوكين دليل جيد لتحديد مدى الإصابة بعدوى الجهار اليولى للانسان.