## INCIDENCE OF THEILERIOSIS, BABESIOSIS AND ANAPLASMOSIS IN CATTLE IN TRIPOLI - LIBYA.

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#### **SUMMARY**

Bovine theileriosis, babesiosis and anaplasmosis are a haemoparasitic disease caused by protozoans of Genus Theileria, Babesia and Anaplasma respectively. These haemoparasites are responsible for considerable losses due to mortality, weight loss and a reduction in milk beside the cost of prophylactic measures. The results showed that 51 (11.7%) and 13(2.9%) out of 443 cattle from different localities in Tripoli-Libya by Giemsa stained blood and lymph node smears were infected only with Theileria species respectively. The incidence of *Theileria* species by Giemsa stained blood films examination in different localities in Tripoli was 0.0%, 22.7% and 35.4% in Engeela, Algyran and Fum-Mulgha respectively. While the total incidence of T.mutans, B.bigemina and A.marginale in the examined cattle by different ELISA kits were 50.6%, 12.9, and 3.4%, respectively. Also, the present work recorded mixed infection.

**Keywords**: *Theileria*; *Babesia*; *Anaplasma*; blood films; ELISA; Cattle.

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#### INTRODUCTION

Theileriosis, Babesiosis and Anaplasmosis are a haemoparasitic diseases caused by protozoans of genus *Theileria*, *Babesia* and *Anaplasma* respectively and are present in all tropical and subtropical areas were the vectors occur. They are responsible for considerable economic losses due to mortality, weight loss and reduction in milk production, and result in a state of premunity, which make the host asymptomatic carrier and serving as a source of infection for the tick or insect vec-

tor. Also they have been cited, as constraints to the health and improved production of cattle (Young et al., 1988). The present study aims to clarify the incidence of blood parasites affecting cattle in some localities in Tripoli ñ Libya by blood films, and lymph node smears examination for detection of *Theileria* species schizonts and anaplasma morvlae serological examination by ELISA kits for diagnosis of *T.mutans.B.bigemina* and *A. marginalis*.

Anaplasma and other ehrlichiae are obligatory intracellular bacterial parasites that grow ineuokaryotic host cells form intracytoplasmic microcdonies (Dumler and Bakken,1998) Three genera are placed under family Anaplasmataceae namely genus Anaplasma, genus Ehrichia and genus Neorickettsia (Dumler et al.,2001).

#### MATERIALS AND METHODS

443 cattle (Friesian) 3 - 7 years old from farms located at Engeela (269), Elgyran (75) and Fum-Mulgha (99) districts in Tripoli, Libya were investigated in the present study for the presence of *Theileria*, *Babesia* and *Anaplasma* species by the use of the parasitological and serological methods.

#### 2.1. Blood films and serum samples:

Thin and thick blood films were prepared from the ear vein of each of the 443 cattle, fixed and stained with Giemsa stain and examined microscopically. Serum samples were prepared from the 443 investigated cattle. The serum was collected and divided into labeled tubes and stored at - 20 C until used for ELISA.

#### 2.2. Lymph node smears:

Lymph node smears(178) were prepared from each cattle by fixing the prescapular lymph node. The lymphatic juice was aspirated by sterile needle and smeared over a clean dry glass slide, air dried, fixed with methyl alcohol, stained with for detection of *Theileria* spp. schizont.

#### 2.3. Serological examination:

Antibody ELISA kits for diagnosis of *T.mutans*, *B.bigemina* and *Anaplasma marginale* were used for detection of the specific antibodies against *T.mutans*, *B.bigemina* and *Anaplasma marginale* in the 178 cattle sera. The kits were obtained from Svanova Biotech AB Uppsala, Sweden. ELISA procedure was done according to the procedure of the manufacture manual.

# 2.4. Studying the cross reaction between T. mutans, B. bigemina and A. marginale infecting cattle by ELISA.

This study was conducted during run of ELISA test for serodiagnosis *T. mutans*, *B. bigemina* and *A. marginale* by adding 100 µl of positive control serum of each of *T. mutans*, *B.bigemina* and *A. marginale* in each selected well contains the above mentioned blood organisms antigens.

#### 3.1. Results of blood and lymph node smears:

Examination of 443 Giemsa stained cattle blood smears revealed that 50 (11.3%) were infected only with Theileria species (Table1). The incidence of *Theileria* spp. according to the localities was 0.0% in Engeela, 17(22.7%) in Algyran and

Lymph node smears showed schizont in 13 (2.9%) out of 443 cattle lymph node smears (Table 1). Examination of the blood films did not reveal infection with *Babesia spp*. And *Anaplasma spp*.

**Table (1)**: Incidence of Theileria species in cattle by examination of Giemsa stained blood and Lymph node smears.

| No. of Examined | Blood | d smear | Lymph node smear |     |  |
|-----------------|-------|---------|------------------|-----|--|
| samples         | +ve   | %       | +ve              | %   |  |
| 443             | 52    | 11.7    | 13               | 2.9 |  |

**Table (2):** Incidence of the *Theileria spp* by examination of Giemsa stained cattle blood films according to the localities.

| Localities | No. of samples | Results |      |  |  |
|------------|----------------|---------|------|--|--|
|            |                | +ve     | %    |  |  |
| Engeela    | 269            | 0       | 0    |  |  |
| Algyran    | 75             | 17      | 22.7 |  |  |
| Fum-Mulgha | 99             | 35      | 35.4 |  |  |
| Total      | 443            | 52      | 11.7 |  |  |

#### 3.2. Results of serological test

#### 3.2.1. Incidence of T.mutans

The results of examination of 178 cattle serum samples by *T.mutans* antibody ELISA kit showed 90 (50.6%) of animals had antibodies against T.mutans. 50 out of the 90 seropositive samples were *Theileria* species blood film positive while the other 40 seropositive samples were blood film negative (Table 3). The distribution of the infection with T.mutans according to the localities were 31(38.8%) in Engeela, 22 (51.2%) in Algyran, and 37(67.3%) Fum-Mulgha (Table 4).

**Table (3):** Distribution of the 91 T.mutans seropositive samples.

| Distribution T.mutans | blood | a species<br>I film<br>itive | Theileria species<br>blood film<br>negative |      |  |
|-----------------------|-------|------------------------------|---|------|--|
|                       | +ve   | +ve  %                       |   | %    |  |
| 90                    | 50    | 50 55.6                      |   | 44.4 |  |

**Table (4):** Seroprevalence of *T.mutans* by ELI-SA kit according to the localities.

| Localities | No.of samples | Res | sults |  |
|------------|---------------|-----|-------|--|
|            |               | +ve | %     |  |
| Engeela    | 80            | 31  | 38.8  |  |
| Algyran    | 43            | 22  | 51.2  |  |
| Fum-Mulgha | 55            | 37  | 67.3  |  |
| Total      | 178           | 90  | 50.6  |  |

#### 3.2.2. Incidence of B.bigemina:

The present study revealed that 23 (12.9%) out of the examined 178 cattle serum samples using *B.bigemina* antibody ELISA kit were positive for *B.bigemina*. All the 23 seropositive samples were negative for *B.bigemina* by blood film examination. In addition 6 out of the 23 seropositive were *T.mutans* seropositive (Table 5) The distribution of the infection with *B.bigemina* according to the localities were 11 (13.8%) in Engeela, 12 (21.8%) in Fum-Mulgha and 0.0% in other farm (Table 6).

**Table (5):** Distribution of 23 *B.bigemina* cattle serum samples.

| Distribution T.mutans seropositive | blood | a species<br>I film<br>itive | Theileria species<br>blood film<br>negative |      |  |
|------------------------------------|-------|------------------------------|---|------|--|
|                                    | +ve   | %                            | +ve   | %    |  |
| 23                                 | 6     | 26.1                         | 17  | 73.9 |  |

**Table (6):** Incidence of the seropositive *B.bigemina* among 178 cattle serum samples by *B.bigemina* antibody ELISA kit according to the localities.

| Localities | No.of samples | Res | sults |
|------------|---------------|-----|-------|
|            | ·             | +ve |       |
| Engeela    | 80            | 11  | 13.8  |
| Algyran    | 43            | 0   | 0     |
| Fum-Mulgha | 55            | 12  | 21.8  |
| Total      | 178           | 23  | 12.9  |

#### 3.2.3. Incidence of A.marginale:

The present investigation showed that examination of 178 cattle serum samples by A.marginale antibody ELISA kit showed that 6 (3.4%) had antibodies against A.marginale. All the 6 seropositive samples were negative for A.marginale by blood film examination. In addition, 3 out of 6 seropositive were T.mutans seropositive and the

other 3 were *B.bigemina* seropositive and the 6 seropositive samples were Theileria species blood film negative (Table 7). The distribution of the infection with *A.marginale* according to the localities were found 2 (2.5%) in Engeela, 4 (7.3%) in Fum-Mulgha and 0.0% in other farm (Table 8).

**Table (7):** The result of serodiagnosis and blood examination for detection of *A.marginale*, *T.theileria*, *T.mutans* and *B.bigemina*.

| A.marginale        | Theileria species<br>blood film<br>positive |   | blood filn | a species<br>a negative<br>positive | Theileria species blood film negative and B.bigemina seropositive |      |  |
|--------------------|---|---|------------|-------------------------------------|---|------|--|
| seropositive +ve % |   | % | % +ve %    |                                     | +ve   | %    |  |
| 6                  | 0   | 0 | 3          | 50.0                                | 3   | 50.0 |  |

Table (8): Seropositive of the A.marginale in cattle serum samples by A.marginale antibody ELISA kit according to the localities.

| Localities | No.of samples | Res | sults |
|------------|---------------|-----|-------|
|            |               | +ve | %     |
| Engeela    | 80            | 2   | 2.5   |
| Algyran    | 43            | 0   | 0     |
| Fum-Mulgha | 55            | 4   | 7.3   |
| Total      | 178           | 6   | 3.4   |

#### 3.2.4. Results of mixed infection:

The results of examination of 178 cattle serum samples by ELISA kits for diagnosing *T.mutans*, *B.bigemina* and *A.marginale* revealed that 6 (3.4%), 3 (1.7%) and 3 (1.7%) had mixed antibodies against *T.mutans* with B.bigemina, *T.mutans* with *A.marginale* and *B.bigemina* with *A.marginale* respectively (Table 9).

Table (9): Results of the mixed infection of the blood parasites in the cattle serum samples by ELISA kits.

|                            |                             | Blood parasites |     |                   |                                |     |  |  |  |  |
|----------------------------|-----------------------------|-----------------|-----|-------------------|--------------------------------|-----|--|--|--|--|
| No. of animals<br>examined | T.mutans with<br>B.bigemina |                 |     | ns with<br>ginale | B.bigemina with<br>A.marginale |     |  |  |  |  |
|                            | +ve                         | %               | +ve | %                 | +ve                            | %   |  |  |  |  |
| 178                        | 6                           | 3.4             | 3   | 1.7               | 3                              | 1.7 |  |  |  |  |

The results revealed no cross reaction between *T.mutans*, *B.bigemina* and *A.marginale* infecting by ELISA. The total incidence of blood parasites in the examined cattle in some localities in Tripoli Libya by Giemsa stained blood films was 11.3% Theileria species (Table 11). While examination of serum samples by different ELISA kits (*T.mutans*, *B.bigemina* and *A.marginale*) were 50.6% (T.mutans), 12.9% (B.bigemina) and 3.4% (*A.marginale*) (Table 10).

was nearly similar to that obtained by Mohammed, Mohammed, 1980 (11%) in Egypt. Low incidence of Theileria species was detected in different countries. In Jordan 4.5% - 6% (Sherkov et al., 1976) and in Bangladesh 8.47% (Samad et al., 1983) and in Kashmir vally, 1.4% (Shaw, 1989). On the other hand, high incidence was recorded 32.6% to 85% Saidu et al., 1984) and 44% Lawal et al., 1998) in Nigeria. 45.5% Schoepf et al., 1984) in Somalia, 20.75% (Dar-

Table (10): Incidence of the blood parasites in the examined cattle by Giemsa stained blood smears and different ELISA kits according to the localities.

| Parasites Theileria species * |               | ies * | T. Mutans** |               |     | B. bigemina** |               |     | A. marginale** |               |     |     |
|-------------------------------|---------------|-------|-------------|---------------|-----|---------------|---------------|-----|----------------|---------------|-----|-----|
| Localities                    | No of samples | (+)   | %           | No of samples | (+) | (%)           | No of samples | (+) | (%)            | No of samples | (+) | (%) |
| Engeeja                       | 269           | 0     | 0           | 80            | 31  | 38.8          | 80            | .11 | 13.8           | 80            | 2   | 2.5 |
| Algyran                       | 75            | 17    | 22.7        | 43            | 22  | 51.2          | 43            | 0   | 0              | 43            | 0   | 0   |
| Fum-Mulgha                    | 99            | 35    | 35.4        | 55            | 37  | 67.3          | 55            | 12  | 21.8           | 55            | 4   | 7.3 |
| Retail                        | 449           |       | lu a        | 1786-3        |     |               |               |     |                | 1 1           | 4   | 140 |

<sup>\*</sup> As indicated by Giemsa stained blood smears.

#### DISCUSSION

The present study revealed that 52 (11.7%) out of 443 examined cattle by Geimsa stained blood smear were found infected with Theileria spp in Tripoli. This low incidence might be due to low parasitaemia and the cattle were chronically infected with Theileria species. The present results

ghouth et al., 1996) in Tunisia, 63% (Rouina, 1984) and 53.7% (Ziam and Benaouf, 2004) in Algeria. This in agreement might be due to the variation in the environmental conditions (Temperature and humidity) which affect the biology of both parasite and vector. Also, the examined cattle might be sampled from an endemic region. It was worthy to mention that this variation in the

<sup>\*\*</sup> As indicated by ELISA kits.

incidence of Theileria infection among cattle from region to another might be regarded to presence of carrier population which serves as a reservoir of infection for ticks and variation of hygienic measures in the farms. In addition introduced susceptible livestock to farms and the potential for clinical relapse under serve nutritional and disease stress.

The results showed that the incidence of Theileria species by Giemsa stained blood smears in different localities in Tripoli were different (22.7% in Algyran, 35.4% in Fum-Mulgha, and 0.0% in Engeela). This variation in the incidence in different localities could be due to the difference in tick control and introducing susceptible chronically infected animals which can be infected easily.

The examination of 178 cattle serum samples for the detecting antibodies against *T.mutans* by *T.mutans* antibody ELISA kit showed that 90 (50.6%) were found positive. Low incidence was recorded in Kenya, 1.5% to 28% (Gitau et al., 1997). On the other hand high incidence was recorded in Ghana, 100% (Bell-Sakyi et al., 2004) By examination of 178 cattle serum samples for the detecting antibodies against B.bigemina by B.bigemina antibody ELISA kit revealed that 23 (12.9%) were found positive. This finding agreed with Todorovic and Carson (1981) who mentioned that negative microscopic examination does not exlude the possibility of Babesia infection. So it is necessary to detect specific antibod-

ies against Babesia species by serological tests (such as ELISA) rather than Babesia organisms. Nearly similar incidence was recorded in Morroco, 10 to 16% (EL Haj et al., 2002). The higher incidence were recorded in different countries. In Egypt, 27.5% and 22% at Giza and Ismailia (Nassar, 1992), and 64.3% to 100% (Ashmawy et al., 1998). In Tanzania, 88% (Woodford et al., 1990) and 27% (Swai et al., 2000); in Italy, 23.1% (Cringoli et al., 2002).

Also in the examination of 178 cattle serum samples for the detecting antibodies against A.marginale by A.marginale antibody ELISA kits 6 (3.4%) were found positive. Higher incidence was reported in many countries. In Egypt, 16.7% (Eid et al., 2001); in Uganda, 61.9% (Ssenyonga et al., 1991); in Italy, 55.6% (Ceci et al., 2002); in Brazil, 92.94% (Andrade et al., 2001).

The seropositivity of these blood parasites in cattle which were negative by blood smears examination indicated sub-clinical cases Sundar et al., 1993); (Soudarajan et al., 2000). These incidence is higher than that obtained by the blood films examination, this indicates that ELISA are capable for detection of antibodies as early as seven days post infection and for more than three years after infection (Bary et al., 1982). Also, ELISA had high sensitivity and specificity in the analysis of sera samples (Ashmawy et al., 1998) and (Morshedi et al., 2003). Mtshali et al. (2004) demonstrated that blood smears were negative but were

positive for *Anaplasma* by ELISA. Kang et al. (1992) referred the negative results with direct light microscope but positive by ELISA to the maternal immunity. The high incidence recorded in different countries might be due to that the examined cattle were sampled from an endemic region.

Our study revealed that the incidence of *T.mutans* was 38.8%. 51.2% and 67.3% in Engeela, Algyran and Fum-Mulgha respectively. *B.bigemina* and *A.marginale* occurred only in two localities with incidence 13.8% and 2.5% in Engeela, 21.8% and 7.3% in Fum-Mulgha respectively. The variation in the incidence between the localities could be due to difference in the management, tick control and veterinary service.

The present study demonstrated mixed infection as detected by serological examination, 6 (3.4%), 3 (1.7%) and 3 (1.7%) out of 178 cattle serum samples showed mixed infection of *T.mutans* with *B.bigemina*, *T.mutans* with *A.marginale* and *B.bigemina* with *A.marginale* respectively. The previous studies recorded a mixed infection in Nigeria (0.75%) between Babesia and Theileria species; Akinboade and Dipeolu, 1984); in Turkey 20% *T.annulata* and *T.mutans*, *T.mutans* and *B.bigemina* in one case and *T.annulata*, *T.mutans* and *B.bigemina* in another case Dumanli and Ozer, 1987); in Kashmir vally (0.69%) between anaplasmosis and babesiosis (Shaw, 1989); in Tanzania (45%) *B.bigemina* and *B.bovis* 

Woodford et al., 1990); in Turkey (13%) between B.bigemina with A.marginale or B.bovis (Acici, 1995); in Italy (49.4%)between B.bigemina and A.marginale (Cringoli et al., 2002); in India 3 out of 10 calves had mixed infection with Anaplasma, Babesia and Theileria species (Julie et al., 2005). These indicate that there is no cross reaction between the different genera of haemoparasites. This was also confirmed in the present study where their was no cross reaction was recorded between T.parva, T.mutans, B.bigemina and A.marginale. Madruga et al., 2001 reported that there is no cross reaction were verified with sera from calves inoculated three times with B.bovis for detection of B.bigemina by ELISA. Sundar et al. (1998) reported that their no cross reaction between T.annulata and B.bigemina.

#### REFERENCES

Acici, M., (1995): Prevalence of blood parasites in cattle in the Samsun region. Etlik Vet. Mikrob. Derg. 8 (1/2): 271-277.

Akinboade, O. A., and Dipeolu, O. O.,(1984): Comparison of blood smear and indirect fluorescent antibody techniques in detection of haemoparasite infections in trade cattle in Nigeria. Vet. Parasitol. 14 (2): 95-104.

Andrade, G. M. d.; Vidotto, O., Vidotto, M. C.; Yoshihara, E., Kano, F. S., and Amaral, C. H. S., (2001). Seroprevalence of Anaplasma marginale in dairy cattle and, studies on the dynamics of natural infection of Holstein calves in Southern Brazil. Semina Londrina 22 (2): 155-

- Ashmawy, K. E.; El-Wafa, S. A. A., and Fadly, R. S., (1998): Incidence of Babesia bigemina infection in native breed cattle, Behera Province, Egypt using different methods of diagnosis. Assiut Vet. Med. J. 39 (77): 110-120.
- Bary, D. N.; Rodwell, B. J.; Timm, P.,and McGreger, W., (1982): A microplate enzyme immunoassay for detecting and measuring antibodies to Babesia bovis in cattle serum. Aus. Vet. J. 59: 136-140.
- Bell-Sakyi, L.; Koney, E. B.; Dogbey, O. and Walker, A. R., (2004): Incidence and prevalence of tick-borne haemoparasites in domestic ruminants in Ghana. Vet. Parasitol. 124 (1-2): 25-42.
- Ceci, L.; Carelli, G.; Sasanelli; Tassi, P.; Paradies, P. and Caprariis, D. d., (2002): Clinical and epidemiological study on Tick-borne diseases in some regions of Southern Italy. Atti Soc. Ital. Buiatria 34: 249-257.
- Cringoli, G.; Otranto, D.; Testini, G.; Buono, V.; Di Giulio, G.; Traversa, D.; Lia, R.; Rinaldi, L.; Veneziano, V. and Puccini, V., (2002): Epidemiology of bovine tick-borne diseases in southern Italy. Vet. Res. 33 (4): 421-428.
- Darghouth, M. E.; Bouattour, A.; Miled, L. B. and Sassi, L., (1996): Diagnosis of Theileria annulata infection of cattle in Tunisia: comparison of serology and blood smears. Vet. Res. 27 (6): 613-621.
- Dumanli, N. and Ozer, E., (1987): Investigations on the blood parasites and their incidence in cattle in the Elazig region. Vet. Fak. Derg. Selcuk Univ. 3 (1): 159-166.
- Dumler, J.S. and Bakken, J.s. (1998): Human ehrlichioses: newly recognized infection transmitted by ticks. Annu.Rev.Med., 49: 201-213.
- Dumler, J.S.; Barbet, A.F.; Bekker, C.P.; Dasch, G.A.; Palm-

- er,G.H. Ray,S.C.; Rikihisa,Y. and Rurangirula ,F.R. (2001) Reorganization of genera in the families Rickett-siacea and Anaplasmataceae in the Order Rickettsiales: unification of some species Ö.. Int.J.Syst. Evol. Micriobiol., 51(6): 2145-2165.
- Eid, G. E.; Abd El Hamid, K. F. M. and Gad El Said, W. A., (2001): Comparison of DNA probe technique and C-ELISA for detection of Anaplasma marginale in cattle. Vet. Med. J. Giza. 49 (1): 45-57.
- El Haj, N.; Kachani, M.; Bouslikhane, M.; Ouhelli, H.; Ahami, A. T.; Katende, J. and Morzaria, S. P., (2002): Sero-epidemiology of Theileria annulata and Babesia bigemina infections in Morocco. Rev. Med. Vet. 153 (3):189-196.
- Gitau, G. K.; Perry, B. D.; Katend; McDermott, J. J.; Morzaria, S. P. and Young, A. S., (1997): The prevalence of serum antibodies to tick-borne infections in cattle in smallholder dairy farms in Murang'a District, Kenya; a cross-sectional study. Prev. Vet. Med. 30 (2): 95-107.
- Julie, B.; Harikrishnan, V. S. and Baby, P. G., (2005):
  Mixed haemoprotozoan infection in calves. Indian Vet.
  J. 82 (5): 543-544.
- Kang, Y. B; Wee, S. H.; Kim, J. S.; Hyun, K. J.; Fujisaki, K. and Kamio, T., (1992): Relationship between the intra-erythrocytic forms and the ELISA antibodies of Theileria sergenti in the overwintered cattle. Res. Rep. rural dev. Admin. Vet. 34 (2): 45-52.
- Lawal, I. A.; Folaranmi, D. O.; Asselbergs, ; Perie, N; Okoro, J. E.; Bale, J. S. and Musa, B., (1998): Studies on prevalence of bovine theileriosis in Negeria using the immunofluorescent antibody (IFA) test and microscopic detection technique. Nigerian Vet. J. 19: 53-60.
- Madruga, C. R.; Marques, A. P. C.; Araujo, F. R.; Miguita,

- M.; Carvalho, F. M. E.; Araujo, F. S.; Umaki, A. C. S.; Crocci, A. J. and Queiroz, R. A. (2001): Evaluation of an ELISA for detection of antibodies to Babesia bigemina in cattle and itis application in an epidemiological survey in Brazil. Pesqui. Vet. Bras. 21 (2): 72-76.
- Mohammed, A. E. M. (1980): Studies on the blood parasites of cattle in Quina, M.V.Sc.Thesis Cairo Uni.
- Morshedi, A.; Horr yadollahi, M. R.; Tavassoli, M. and Dalir Naghade, B. (2003). A seroprevalence survey of Theileria spp. infection by ELISA, compared with blood-smear observation in cattle. J. Fac. Vet. Med. Univ. Tehran 58 (4): 319-322.
- Mtshali, M. S.; De Waal, D. T. and Mbati, P. A. (2004): A sero-epidemiological survey of blood parasites in cattle in the north-eastern Free State, South Africa. Onderstepoort J. Vet. Res. 71 (1): 67-75.
- Nassar, A. M., (1992). Serological diagnosis of Babesia bigemina by Dot-Enzyme linked immnuosorbent assay and indirect fluorescent antibody test. Proc. 2nd Cong. Faculty Vet. Med. Cairo Uni. 31-33.
- Rouina, A. D. (1984). Clinical study of bovine Theileriosis based on the 327 cases in Algeria (North West region, Mascara), Maghreb Vet. 1: 23-27.
- Saidu, S. N. A.; Abbulkadir, I. A. and Akerejola, O. O. (1984): Theileria mutans infection in Nigerian cattle. Trop. Anim. Health. Prod. 16 (3): 149-152.
- Samad, A.; Dhar, S. and Gautam, O. P. (1983): Prevalence of Theileria annulata infection among cattle of Bangladesh. Indian J. Parasitol. 7 (1): 61-63.
- Schoepf, K.; Mustafa, M. H. A. and Katende, J. M. (1984): Observation on blood parasites of domestic livestock on the lower Juba region of Somalia. Trop. Anim. Health Prod. 16 (4): 227-232.

- Shaw, A. A. (1989). Investigation on some infection in the exotic, pure and cross cattle of Kashmir valley. J. Comp. Microb. Immunol. Infec. Dis. 10 (1): 33-38.
- Sherkov, S. N.; EL Rabie, Y.; and Kokash, L., (1976): A survey of parasitic blood diseases tick borne fever in domestic animals in Jordan. Egyptian J. Vet. Sci. 13 (1): 22-35.
- Soudarajan, C.; Rajavelu, G.; Anadan, R. and Ramacass, P. (2000). Enzyme-linked immunosorbent assay for detection of Theileria annulata infection in cattle and buffaloes. J. vet. Parasitol. 14 (2): 165-166.
- Ssenyonga, G. S. Z.; Kakoma, I.; Montenegro James; S., Nyeko, P. J. and Buga, R. (1991): Anaplasmosis in Uganda. II. Prevalence of bovine anaplasmosis in Uganda. Ann. Trop. Med. Parasitol. 85 (3): 305-308.
- Sundar, N.; Balasundaram; S.,and Anandan, R., (1998): Sensitivity of capillary-tube agglutination test for detection of theileriosis in cattle. Cheiron 27 (1/2): 13-14
- Sundar, N.; Balasundaram, N.; Ramadass, P. and Anadan, R., (1993): Useful of enzym-linked immunosrbent assay for the diagnosis of Theileria annulata infection. Indian J. Anim. Sci. 63 (12): 1219-1221.
- Swai, E.; Kambarage, D.; French, N. P.; Ogden, N. H.; Fitzpatrick, J.; Bell, C.; Karimuribo, E. and Bryant, M., (2000): Antibody responses to B. begimina, B. bovis and T. parva in dairy cattle in Tanga, Tanzania: geographic variation. Tanzanian Vet. J. 20: 170-177.
- Todorovic, R. A. and. Carson, C. A. (1981): Methods for measuring the immunological response to Babesia. In Babesiosis (ed.M.Ristic and J.P. Kreier): 381:410.
- Woodford, J. D.; Jones, T. W.; Rae, P. F.; Boid, R. and Bell-Sakyi, L. (1990): Seroepidemiological studies of bovine babesiosis on Pamba Island, Tanzania. Vet. Para-

sitol. 37 (3-4): 175-184.

Young, A. S.; Groocock, C. M. and Kariuki, D. P. (1988): Integrated control of ticks and tick - borne diseases of cattle in Africa. Parasitol.96: 403 - 432. Ziam, H. and Benaouf, H., (2004): Prevalence of blood parasites in cattle from wilayates of Annaba and El Tarf east Algeria. Arch Inst. Pasteur Tunis 81 (1-4): 27-30.

### نسبة حدوث الإصابة بمرض التيليرياو البابيزيا والأنايلازها في الأبقار بهنطقة طرابلس - ليبيا

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اجرى هذا البحث على 443 مصل ابقار ووجد ان 11 و 2.9 % من هذه الأبقار مصابة بطفيل التيليريا وذلك بعد فحصها بواسطة استخدام صبغة الجمسا ومسحات الغدد الليمفاوية.

باستخدام اختبار الأيليزا وجد ان 50.6 و 12.9 و 3.4% من الأبقار التى فحصت مصابة بطفيل التيليريا ميوتانز و البابيزيا بيجيمنا و الأنا لازما مارجينال على التوالى . كما أوضح البحث وجود اصابات في الأبقار باكثر من طفيل من الطفيليات المذكورة.