Bacteriological Affections Of Livers And The Associating Serum Biochemical Changes In Buffaloes In Menoufiea Governorate

Elham I. Atwa* and Ola F. A. Talkhan**

*Bacteriology Dept., ** Biochemistry Dept., Animal Health Research Institute Shebin El- Kom-Menoufiea

ABSTRACT

Lesion specimens from 75 condemned buffaloes livers were collected in abattoirs at El-Menoufiea Governorate, from slaughtered buffaloes of different age for bacteriological examination, in Animal Health Research Institute – Shebin El-Kom, in trials to detect the possible bacterial agents. Liver appeared congestion, dark brown in colour, hard, firm, tough in consistency condemned and some samples affected with abscesses.

Out of 75 samples of examined liver of buffaloes, 63 samples (84%) were positive for bacterial isolation. Staph. aureus was the most predominant isolales. Staph. aureus, E. coli (0157), Streptococus pyogenes, C. perfringens type "A", Corynebacterium pyogenes, Klebsiella pneumoniae, Citrobacter freundi and Proteus vulgaris were isolated with incidence of (30.6%, 12%, 10.7%, 8%, 8%, 6.7%, 5.3% and 2.7%, respectively).

The serum biochemical analysis of *Staph. aureus* infected buffaloes revealed a significant increase in Malondialdehyde (MDA) concentration, on contrary the total protein, and albumin were significantly decreased with significant increase in total globulins. Moreover, the serum levels of AST, ALT, urea, uric and creatinine were significantly increased.

In vitro antimicrobial sensitivity test, of the predominant isolates indicated that Staph. aureus were highly sensitive to cephalocin, erythromycin, kanamycin, doxycycline, enrofloxacin, and trimethoprim & sulphamethoxazole. There is marked difference between the sensitivity to antibiotics between different bacterial isolates. Kanamycin, doxycycline, ciprofloxacin and trimethoprim & sulphamethoxazole were considered the antimicrobial agents of choice for treatment of bacterial liver affections. On the other hand penicillin G and streptomycin were an ineffective chemotherapy for treatment of any isolates.

INTRODUCTION

The Egyptian buffaloes, is named river or water buffaloes, are mainly distributed in the Nile valley and Delta in about 2.8 million animals, 90 % of which are located in private small herds (1). Buffaloes have been used in Egypt as agricultural work animals in the farms as well as the main source of both milk and meat necessary to fulfill the gap between the increased population and their demands from animal protein.

The liver is considered to be as one of the most important organs of mammalian metabolism and food conversion, which regulates, stabilizes, and protect the internal environment and the whole body. Any disturbance in this organ would have its

reflection on the general health and causes a great economic losses in animal production represented by liver condemenation at slaughter houses. Liver infection may lead to serious morbidity with damage of liver and pathological affections of liver may be attributed to a variety of causes including viruses, mycoses, parasites and bacteria (2). Many studies deal with the effect of bacterial pathogens on animal tissues particulary those used for human cosumption was performed (3 - 7).

Bacteria were considered to be one of the most serious infectious agents which causes liver affections in buffaloes. Several Gram positive and Gram negative bacteria were associated with liver affections and abscesses, these organisms orginated from the gastointestinal tract and arrived to liver by translocation via the blood stream (7).

Liver abscesses are commonly observed in fattened cattle, heavily and causes condemenation of approximately 10% of the liver (8). Numerous investigators evaluated the bacterial flora, both aerobic and anaerobic of bovine liver abscesses (8 - 10). Staph. aureus, E. coli, Proteus spp. Streptcoccus spp. and Corynebacterium spp. were isolated (5, 6, 7, 9, 11), and Clostridium spp. was recorded (12). The bovine liver was microbiologically contaminated as heavy as raw meat, when found more than 50% of liver samples were contaminated with Staph. Aureus (13). Corynebacterium spp., Staph. aureus, E. coli and Pseudomonas aerogenosa were isolated from the liver of sheep and goat (14, 15).

Liver function tests may be used to establish a diagnosis in an individual animal or to detect subclinical liver damage following the bacterial infection and its circulating toxins, hence determining its economic value (16).

The main purpose of this work was initiated to investigate the isolation and identification of possible causative bacterial agents of liver abscesses in buffaloes. On the other hand, antibiogram pattern of most bacterial isolates, as an aid to overcome this problem and reduce losses, and the biochemical changes associated with *Staph. aureus in* infected buffalo slaughtered in abattoir were our aim.

MATERIAL AND METHODS

Samples

75 apparently diseased liver samples (condemned buffalo liver) were collected from different abattoirs at El- Menoufiea Governorate, from slaughtered buffaloes of different ages. Liver appeared congestion, dark brown in colour, hard, firm, tough in consistency condemned and some samples affected with abscesses. Each samples kept separately in a sterile plastic ice bag for bacteriological examination in Animal Health

Research Institute – Shebin El-Kom. Samples were divided into two portions and were submitted to aerobic and anaerobic examination.

Blood samples

Blood samples were collected in clean plastic centrifuge tubes and allowed to coagulate. The serum was separated by centrifugation at 3000 r.p.m for 10 minutes, Then the clear supernatant sera aspirated carefully into dry and sterile labeled vials.

Bacteriological examination

All collected samples were subjected to aerobic and anaerobic bacteriological examination.

1-Aerobic identification

Cultural methods were made from infected liver lesions by sterilizing the exposed surface of the liver and cutting down small piece from the deeper parts, and inoculated directly into nutrient broth, and aerobically incubated at 37°C for 24 hours, then subcultured onto nutrient agar, blood agar, MacConky bile salt lactose agar, crystal violet blood agar, mannitol salt agar media for isolation of Staph. aureus, and Eosin Methylene blue agar media for isolation of E. coli, all inoculated plates were incubated aerobically at 37°C for 24-48 hours.Suspected growing colonies onto the surface of these media were identified by studying characters of the colonies as well as Gram's stain, then identified morphologically according to the previously described methods (17, 18). One single colony showed typical colonial appearance and morphological characters was picked up and streaked into semisolid agar media and incubated at 37°C for 24 hours, for further identification.

The pure colonies were biochemically identified (19-22). The Gram negative bacteria included *Enterobacteriacae* family were typed (23).

2- Anaerobic identification

A loopful from a small piece of the deeper parts of affected liver, was inoculated into tubes of freshly prepared Robertson's cooked meat medium at 37°C for 24 hours. Loopful from each tubes was streaked onto the surface of 10% sheep blood agar, then incubated anaerobically at 37°C for 24 hours. The plates were examined for characteristic colonies of genus *Clostridium*. Subcultures from suspected colonies in cooked meat broth were made for further biochemical identification (20, 24, 25).

Serum biochemical analysis

The sera of Staph. aureus infected buffaloes were used for estimation of Malnodialdehyde (MDA) (26), total protein (27), albumin (28). While total globulins calculated mathematically by substacting albumin from total protein (29), AST and ALT (30), urea (31), uric acid (32), and creatinine (33) were determined.

Antibiogram sensitivity pattern of isolates

Antibiogram was applied on most isolated strains using disc diffusion technique (20, 34) with Mueller Hinton agar, using the following chemotherapeutic and antibiotic discs (Oxoid) cephalocin amoxycillin (10ug),(10ug),chloramphenicol (30ug), ciprofloxacin (5ug), (30ug), enrofloxacin doxycycline (5ug),(30ug), erythromycin (10ug), flumequine kanamycin gentamicin (10ug), (30ug), norfloxacin (10ug), pencillin G (10U),streptomycin (10ug), tetracycline (30ug), and trimethoprim & sulphamethoxazole (1.25ug). The results were interpretated according to the manual supplied by Oxoid Company.

Statistical analysis

All data were subjected to statistical analysis (35).

RESULTS

Table 1, showed the bacterial isolates from liver samples of buffaloes. Out of 75 samples of examined liver of buffaloes, 63 samples (84%) were positive. The bacteriological isolation of different types of bacteria from liver samples were Staph. aureus, E. coli (O157), Streptococus pyogenes, C. perfringens type "A", Corynebacterium pyogenes, Klebsiella pneumoniae, Citrobacter freundi

and *Proteus vulgaris* with incidence of 30.6%, 12%, 10.7%, 8%, 8%, 6.7%, 5.3% and 2.7%, respectively.

The biochemical studies illustrated in Table 2 and 3, Proved that serum of *Staph. aureus* infected buffaloes showed, a highly significant increase of MDA, total globulins. While total protein and albumin recorded high significant decrease. On the other hand, a highly significant increase was found in the serum AST, ALT, urea, uric and creatinine.

In vitro sensitivity of the most prevalent bacteria isolated from liver samples of buffalo were done against chemotherapeutic agents (15) as show in Table 4. All tested strains of Staph, aureus were highly sensitive to erythromycin. kanamycin, cephalocin, doxycycline, enrofloxacin, and trimmethoprim & sulphamethoxazole with activity percentage of 91.3%, 91.3%, 86.9%, 78.3% 73.9% 69.6% and 69.6%, respectively. Most of these strains were highly resistant to tetracycline, gentamicin, streptomycin, norfloxacin and penicillin G with activity percentage of 21.7%, 30.4%, 8.7%, 4.3% and 0%, respectively. Comparing the sensitivity of E. coli (O157) isolates, the majority of strains sensitive to were highly eryrhromycin, gentamicin, cephalocin, kanamycin, norfloxacin and doxycycline with activity percentage of 100%, 100%, 88.9%, 88.9%, 88.9% and 77.8%, respectively. The same strains were highly resistant to tetracycline, ciprofloxacin, enrofloxacin, chloramphenicol and flumequine with activity percentage of 33.3%, 22.2%, 11.1%, 0% and 0%, respectively. While the isolated strains of Streptococus Pyogenes were mainly highly ciprofloxacin. sensitive to amoxycillin, kanamycin, tetracycline, flumequine, and trimethoprim & sulphamethoxazole with activity percentage of 100%, 87.5%, 87.5%, 75%, 62.5% and 62.5%, respectively. As regards to C. perfringens type "A" isolates were highly sensitive to ciprofloxacin, enrofloxacin, chloramphenicol, flumequine, kanamycin and norfloxacin with percentage of 100%, 100%, 83.3%, 83.3%, 83.3% and 83.3%, respectively, but the same

strains were resistant to penicillin G, trimethoprim and sulphamethoxazole, cephalothin and tetracycline. While Corynebacterium pyogenes isolates were sensitive to amoxycillin, tetracycline, ciprofloxacin, kanamycin and enrofloxacin with activity percentage of 100%, 100%,

83.3%, 83.3% and 66.7%, respectively. Comparing the sensitivity of *Klebsiella pneumoniae* isolates, the majority of strains were highly sensitive to kanamycin, norfloxacin, ciprofloxacin, doxycycline and gentamicin with activity percentage of 100%, 100%, 80%, 80%, and 80%, respectively.

Table 1. Prevalence rate of different types of bacteria isolated from liver samples of buffalo.

Isolated microorganisms	Number of cases	%		
Staph. aureus	23	30.6		
E. coli (O157)	9	12		
Streptococcus pyogens	8	10.7		
C. perfringens type "A"	6	8		
Corynebacterium pyogens	6	8		
Klebsiella pneumoniae	5	6.7		
Citrobacter freundi	4	5.3		
Proteus vulgaris	2	2.7		
Total	63	84		

% was calculated according to the number of examined samples (75).

Table 2. Serum MDA (n mol/ml) total protein (gm/dl), albumin (gm/dl), and total globulins (gm/dl) in both Staph. aureus infected group and control one.

Parameters	Control	Staph. aureus infected buffaloes				
MDA	5.25 ± 0.15	7.72 ± 0.28*				
Total protein	6.32 ± 0.17	5.20 ± 0.16*				
Albumin	2.68 ± 0.053	1.27 ± 0.065*				
Globulins	3.66 ± 0.117	3.83 ± 0.126*				

Data are presented as mean + S.E.

Table 3. Liver and Kidney functions in both Staph. aureus infected group and control one.

Parameters	Control	Staph. aureus infected buffaloes				
AST (Iu/L)	57.5 ± 1.72	88.1 ± 2.04 *				
ALT (Iu/L)	18.8 ± 0.94	27.8 ± 1.7 *				
Urea (mg/dl)	27.89 ± 0.77	41.1 ± 1.96 *				
Uric (mg / dl)	0.95 ± 0.035	1.40 ± 0.079*				
Creatinine (mg / dl)	1.51 ± 0.079	20.4 ± 0.078*				

Data are presented as mean + S.E.

^{*} Highly significant (P < 0.001)

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Table 4. Antibiotic sensitivity test of the most prevalent bacteria isolated from liver samples of buffalo using disc diffusion method.

Antibacterial	Symbol Con	Conc.	Staph. aureus (23)*		E. coli O157 (9)*		Streptococcus pyogenes (8)*		C. perfringens type "A" (6)		Coryne. Pyogens (6)*		Klebsiella pneumoniae (5)*	
agents			S.	Activity %	S.	Activity %	S.	Activity %	S.	Activity %	S.	Activity %	. S.	Activity%
Amoxycilln	AMLi	10ug	11/23	47.8	4/9	44.4	8/8	100	4/6	66.7	6/6	100	2/5	40
Cephalocin	KF	10ug	21/23	91.3	8/9	88.9	3/8	37.5	1/6	16.7	3/6	50	1/5	20
Chloramphenicol	C	30ug	16/23	69.6	0/9	0	3/8	37.5	5/6	83.3	3/6	50	1/5	20
Ciprofloxacin	CPR	5ug	13/23	56.5	2/9	22.2	7/8	87.5	6/6	100	5/6	83.3	4/5	80
Doxycycline	D	30ug	18/23	78.3	7/9	77.8	4/8	50	4/6	66.7	3/6	50	4/5	80
Enrofloxacin	ENR	5ug	17/23	73.9	1/9	11.1	3/8	37.5	6/6	100	4/6	66.7	3/5	60
Erythromycin	E	10ug	21/23	91.3	9/9	100	4/8	50	4/6	66.7	3/6	50	2/5	40
Flumequine	UB	30ug	12/23	52.2	0/9	0	5/8	62.5	5/6	83.3	4/6	66.7	3/5	60
Gentamicin	CN	10ug	5/23	21.7	9/9	100	1/8	12.5	3/6	50	0/6	0	4/5	80
Kanamycin	K	30ug	20/23	86.9	8/9	88.9	7/8	87.5	5/6	83.3	5/6	83.3	5/5	100
Norfloxacin	NOR	10ug	1/23	4.3	8/9	88.9	0	0	5/6	83.3	0/6	0	5/5	100
Penicillin G	P	10U	0/23	0	5/9	55.6	2/8	25	2/3	33.3	2/6	33.3	3/5	60
Streptomycin	S	10ug	2/23	8.7	5/9	55.6	0	0	3/6	50	1/6	16.7	3/5	60
Tetracycline	T	30ug	7/23	30.4	3/9	33.3	6/8	75	1/6	16.7	6/6	100	2/5	40
Trimethoprim& Sulphamethoxazole	SXT	1.25ug	16/23	69.6	4/9	44.4	5/8	62.5	2/6	33.3	4/6	66.7	1/5	20

^{*:} Number of isolates.

Conc.: Concentration of antibiotic disc.

S: Sensitive.

^{%:} Percentage of sensitive isolates in relation to tested isolates = Activity percentage.

DISCUSSION

Buffalo liver is an important edible meat byproduct, but the liver which condemned at slaughter was unfit for human consumption because they harbour small numbers of intrensic bacteria and potential human pathogens (7). Several causes of liver diseasae including ischemia induced by bacterial emboli, vitamin Eselenium deficiency and immune mediated disease (36, 37)

In this study, the bacteriological examination of diseased liver samples was done in an attempt to throw spotlights upon the incidence percentages of bacterial infection, in vitro chemotherapeutic susceptibility testing of the most prevalent bacterial isolates were detected and biochemical changes associated with Staph. aureus infected buffalo slaughtered in abattoir was also checked.

The obtained data in Table 1, proved that out of 75 diseased examined liver of buffalo, 63 samples proved to harbour bacteria with an incidence of 84%.

Previous studies (7,9), on bacteriological examination of apparently normal hepatic abscesses, and telangiectatic liver in cattle recorded that about 82% of examined diseased liver were positive for several types of bacterial infection.

The identification of isolates in Table 1. proved that Staph. aureus, E. coli (O157), Streptococus pyogenes, C. perfringens type "A", Corynebacterium pyogenes, Klebsiella pneumoniae, Citrobacter freundi and Proteus vulgaris were present with incidence of 30.6%, 12%, 10.7%, 8%, 8%, 6.7%, 5.3% and 2.7%, respectively. These results agreement with the work which indicated recorded Corvnebacterium pyogenes, Staphylococcus spp. and Streptococcus spp. were the most prevalent bacteria recovered from liver abscess. More than 50% of market bovine liver samples were contaminated with Staph. aureus and these results indicated that the liver was microbiologicaly contaminated as heavy as raw meat (13). Corynebacterium pyogenes was the most predominant isolates from liver abscesses of cattle, followed by Staphylococcus and Streptococcus spp (9). Staph. aureus, E. coli (O157), Streptococci and proteus were also isolated from liver of cattle (5). In addition Corynebacterium spp., Citrobacter and Clostridium spp. were isolated from diseased livers of cattle (7,38).

MDA is an indicator of free radical production, and an increase in MDA may threrefore be due to oxidative stress. Also MDA is a major degradation production of lipid hydroperoxides and measurements MDA concentration is generally accepted as a marker for assessing the extent of lipid peroxidation in vivo (39, 40). Indeed MDA assay is the most popular and easiest methods used as an indicator of lipid peroxidation and free radical activity in biological samples (41). Staph. aureus infected buffaloes revealed a highly significant increase in serum MDA levels in comparison with control one which indicated oxidative stress. MDA produced by peroxidation can cause cross-linking and polymerization of membrane components. This can alter intrensic membrane properties such as a deformabilities, ion transport, enzyme activities and aggregation state of cell surface determinants. These lead to the removal from the circulation of cells exposed to the agents causing lipid peroxidation. MDA is diffusible, and react with nitrogenous bases of DNA. (42).

A highly significant drop in serum total protein and albumin which may be reffered to the state of inability of the liver to synthesis protein. The reduction of the protein level is attributed to the stress factors and the general unthriftiness which may affect worsely the hepatic parenchyma resulting in failure of protein synthesis (43). Certain bacteria and its toxin cause increased capillary permeability and permit the escape of plasma proteins in the tissue fluids and decrease the osmatic pressure (44, 45). There was a significant increase in the serum globulin level in the diseased buffaloes. Which may be due the stimulation of immune system infectious agents, to produce high amounts of immunoglobulins (46.47).

The obtained data concerning liver function tests, showed marked elevation of serum AST and ALT. Such elevation of the liver enzymes referred to the degnerative and necrotic changes of the liver following the bacterial infection and its circulating toxins (16). It has been shown that Staph. aureus affected buffaloes revealed focal area of necrosis (48).

Serum urea, uric and creatinine recorded highly significant increase. The increases may be due to protein catabolism (49).

The data presented in Table 4, indicated that, there was marked difference between the sensitivity to antibiotics between different bacterial isolates. Kanamycin, doxycycline, ciprofloxacin and trimethoprim sulphamethoxazole considered is the antimicrobial agents of choice for treatment of bacterial liver affections. While variable results were recorded with the remaining used antibiotics. On the other hand penicillin G and streptomycin were ineffective chemotherapy for treatment of any isolates, because 40% of these bacteria were penicillinase positive (50). Staph. aureus strains were highly sensitive to erythromycin, cephalocin, kanamycin. doxycycline and enrofloxacin. On the other hand Strept. pyogenes were highly sensitive to amoxycillin, ciprofloxacin, kanamycin, tetracycline, flumequine, and trimethoprim & sulphamethoxazole. Similar findings were recorded by several investigators (51 -53).

All tested strains of Corynebacterium pyogenes were sensitive to amoxycillin, tetracycline, ciprofloxacin, kanamycin and enrofloxacin. These findings nearly agreed with that previously reported (54 - 56), and revealed that the majority of Corynebacterium pyogenes isolated from diseased liver of buffaloes were highly sensitive to tetracycline and tylosin. On the other hand it has been found that C. perfringens strains were resistant to tetracycline, lincomycine but not to erythromycin (57).

REFERENCES

- 1.FAO/ RNE (2004): Buffalo Newsletter, Europe-Near east 1993-2003. Cairo Regional Office of the Near East (RNE) of the Food and Agriculture Organization of the United Nations, Rome.
- 2.Blood, D.C. and Radostitis, O.M. (1989): Veterinary Medicine. A text book of the disease of cattle, sheep, pigs, goats and horses, Six ed, Bailliere Tindall.
- 3.Gill, C. O. (1979): A review. Intrinsic bacteria in meat. J. Appl. Bacteriol., 47: 367-378.
- 4.Kriaa, H.; Arthand, J.F. and Fournaud, J. (1985): Contamination and bacterial retention capacity of beef carcasses at the abattoir. J. Appl. Bacterial. 55:23-28.
- 5.Szazados, I. (1991): Meat inspection of endocarditis cases (pigs, cattle). Maggar-Lapja, 46 (1):27-43.
- 6.Edward, J.F.; Simpson, R.B.; and Brown, W.C. (1995): Bacteriological culture and histological examination of samples collected from recumbent cattle at slaughter. J. Am. Vet. Med. Assoc., 207(9): 1174-1176.
- 7. Stotland, E.I.; Edwards, J.F.; Roussal, A.J. and Simpson, R.B. (2001): Bacterial microflora of normal and telangiectatic livers in cattle. J. Am. Vet. Med. Assoc. 219(1): 36-39.
- 8.Scanlan, C.M. and Hathcock, T.L. (1983):
 Bovine rumenitis-liver abscess complex. A
 Bacteriological Review. Cornell Vet. 73:
 288-297.
- 9.Lechtenberg, K.F.; Nagaraja, T.G.; Leipold, H.W. and Chengappa, M.M. (1988): Bacteriological and histologic studies of hepatic abscesses in cattle. Am. J. Vet. Res. 49 (1): 58-62.
- 10.Nagaraja, T.G.; Laudert, S.B. and Parrott, J.C. (1996a): Liver abscesses in feedlot cattle. Part 1: Causes, pathogenesis, pathology and diagnosis. Comp. Cont. Edu. Pract. Vet. 18: S230 -S256.
- 11.Berry, B.W.; Leddy, K.F. and Rothenbery, C.A. (1984): Survival and growth of

Elham and Ola

- Staphylococcus aureus on temperature abused beef livers. J. Food Protect., 47 (4): 260-262.
- 12.Simon, P.C., and Stovell, P.L. (1971): Isolation of Sphaerophorus necrophorus from bovine hepatic abscesses in British Columbia. Can. J. Comp. Med. 35: 103-106.
- 13. Okamato, K.; Yoshimitu, F. and Amemiya, J. (1985): Bacteriological studies on hygienic status of the market bovine liver. Memoirs of the Facul. Agricult., Kagoshima University, 21:175-182.
- 14. Charles, M.S. and John, F.E. (1990): Bacteriological and pathological studies of hepatic lesions in sheep. Am. J. Vet. Res. Vol. 51(3): 363-366.
- 15.Samya, F. Ahmed (2001): Morphological studies on some liver affections of goat in Assiut Governorate with special reference to parasitic lesions. Thesis M.Sc. Pathology and Clinical pathology, Fact. Vet. Med. Assiut University.
- 16.Haziroglue, R. and Kul, O. (2004):
 Associated Pasteurella hemolytica,
 multocida and and Haemophillus
 sommnus with pneumonia in calves. DTW
 Dtsch Tieraztl Wochnschr, 104 (4): 105153.
- 17.Kloss, W. E. and Schleifer, K. H. (1986):
 Bergey's Manual of Systematic
 Bacteriology. Vol. II. Williams and
 Wilkins, London.
- 18. Barrow, G. L. and Feltham, R. K.(1993):
 Cowan and Steel's "Manual for the identification of Medical Bacteria, 3rd Ed., University Press Cambridge.
- 19. Cruickshank, R.; Duguid, J.P.; and Swain, R. H. (1975): Medical Microbiology, the practice of microbial, Chucohill Livingstone 12th ed. Vol.11 Edinburg. London, and New York.
- 20.Koneman, K. W.; Allen, S. D.; Dowell, V. R. and Sommers, H. M. (1992): "Color atlas and text book of Diagnostic Microbiology." 2nd ed, J. B. Lippicott Co., London.

- 21. Baily, E. R. and Scott, E. G. (1994): "Diagnostic Microbiology, A Text Book for the isolation and identification of pathogenic Microorganism" 4th Ed. The C.V. Mosby Company, Saint-Louis.
- 22.Quinn, P. J.; Markey, B. K.; Carter, M. E.; Donnelly, W.J. and Leonard, F. C. (2002): Vet. Microbiol. and Microbial Disease. Black well Sc., U.K.
- 23.Krieg, N. R. and Holt, J. G. (1984):
 Bergey's Manual of Systematic
 Bacteriology. Vol. (1) Williams &
 Wilkins, Baltimore.
- 24. Smith, L.D. and Holdeman, L.V. (1968): The pathogenic Anaerobic Bacteria. Pp.201-205, 1st Ed., Charles C. Thomas, Springfield, USA.
- 25.Collee, J.G.; Duguid, J.P.; Fraser, A. G.;
 Marmion, B.P.; and Simmons, A. (1996):
 Mackie and McCartney Practical Med.
 Microbiol. 14th ed., Churchill Livingstone,
 N.Y. London.
- 26.Esterbauer, H.; Cheeseman, K. H.; Danzani, M. U.; Poli, G. and Slater, T.F. (1982): "Separation and characterization of the aldehyde products of ADP/Fe²+ stimulated lipid peroxidation in rat liver microsomes." Biochem. J. 208: 129 140.
- 27. Henry, R. J. (1964): Determination of total protein: In Clinical Chemistry. Harper and Row publishers. New York.
- 28.Drupt, F. (1974): "Colorimetric determination of albumin." Pharm. Biol., 9: 777.
- 29. Doumas, B. T., and Biggs, H. G. (1972):
 Determination of serum globulins in standard methods of Clini. Chem. 7. Med. By G. R. Copper. New York, Academic press, pp. 175.
- 30.Reitman, S. and Frankel, S. (1957): "A colorimetric determination of serum glutamic oxaloacetic and glutamic Pyruvic transaminase." Am. J. Clin. Path., 28: 56-58.
- 31. Patton, G. J. and Crouch, S. R. (1977): "Determination of urea (urease modified

- Berthelot reaction)." Anal. Chem., 49: 464 469.
- 32. Wilding, P. and Health, R. (1975): "Annals of clinical Biochemistry." 12:142 in "Wooton and freeman" (1982): Microanalysis in medical biochemistry. 6th Ed., Churchill Livingstone, Edinburgh, London and N.Y., Pp 79.
- 33. Henry, R. J.; Cannon, D.C. and Wikelaman, J. W. (1974): "Clinical chemistry principle and techniques." 2nd Ed., pp. 543, Harper and Row, Hagerstown.
- 34. Finegold, S.M. and Martin, W.J. (1982):
 Bailey and Scott's "Diagnostic Microbiology" 6th ed. the C.V. Mosby Co., St., Lowis, Tornoto, London.
- 35. Sendecor, w. C. and Cochran, W. G. (1982): Statistical methods. 7th ed. The Ioua Univ. Press, Ames. Lowa, U.S.A.
- 36. Jensen, R.; Johnson, L. W. and Lauerman, L.H. (1982): Ischemia, a cause of hepatic telangiectasis in cattle. Am. J. Vet. Res. 43: 1436-1439.
- 37. Krook, L. and Todd, G.C. (1966): Nutritional hepatic necrosis in beef cattle. "Sawdust liver." Pathol. Vet. 3: 379-400.
- 38.Nagaraja, T.G. and Chengappa, M.M. (1998): Liver abscesses in feedlot cattle: A review. J. Anim. Sci. 76: 287-298.
- 39. Kishiada, E. j.; Kamara, A. and Tokumaru, S. (1993): "Re-evaluation of malondialdhyde and thiobarbituric acid reactive substances as indices of outoxidation based on oxygen cosumption". J. Agric. Food Chem. 41:1-40.
- 40.Zhang, S. S.; Noordin, M. M.; Omar, S. and Rahman, A. (2000): Effect of copper overload on hepatic lipid peroxidation and antioxidation defense in rats. Vet. Human. Toxicol, 42 (5): 261-264.
- 41. Romero, F. J.; Bosch-Morell, F.; Romera, M. J.; Martin, N. M. and Roma, J. (1998): "Lipid peroxidation products and antioxidant in human disease." Environ. Health Perspect, 106(5): 1229-1234.
- 42. Esterbauer, H.; Dieber Rotheneder, M. and Striegl, G. (1991): "Role of vitamin

- E in perventing the oxidation of low-density lipoprotein." Am. J. Clin. Nutr., 53: 3145-3152.
- 43.Coles, E. H. (1986): Veterinary Clinical Pathology. 4th ed. W.B. Saunders company Lodon.
- 44.Hoe, C. (1969): "Text book of Veterinary Clinical Pathology" Eds. By Midway, W.; Prier, J. E. and Wilkinson, J. S., The Williams and Wilkins Company, Baltimore.
- 45.Doxey, D. (1971): "Veterinary Clinical Pathology" 1st Ed. Bailliere, Tindall, London.
- 46. Varley, H. (1976): "A Text book in practical Clinical Biochemistry" 4th Ed. Indian Vrzirari for Arnold Heimann.
- 47. Abd EL-latif, M. M. and EL-Dessouky, S. A. (2006): Studies on some bacterial causes and blood serum biochemical changes of respiratory affections in lambs." Assiut Vet. Med. J., 52 (8): 170-182.
- 48. Soliman, A. S. Refaat, M. and Gobran, R. A. (2005): Pathological and Bacteriological studies of liver affections in buffaloes at Kaluobia Governorate. "Comp. Path. and Clinic. Path. 18 (1): 199-210.
- 49. Radostitis, O. M.; Blood, D. C. and Gay, C. C. (2002): Veteriany Medicine. 10th Ed. Bailliere Tindal.
- 50. Sobiraj, A.; Kron, A.; Schollmeyer, U. and Failing, K. (1977): Federal investigations on the distribution and in vitro resistance of udder pathogenic bacteria in the milk of cows with subclinical mastitis. Tierarztl Prax., 15(2): 108-115.
- 51. Havelka, B. (1983): Antibiotic resistance of the commonest bacterial agents causing bovine mastitis in the period 1979-1981. Biologizace Chemizace Zivocisne Vyroby Veterinaria, 19 (3): 269-275.
- 52. Hazarika, R. A.; Mahanta, P.N.; Dutta, G.N. and Devriese, L.A. (1991): Drug susceptibility and treatment of

- Staphylococcus hyicus dermatitis in cattle. Ind. Vet. J., 68 (2): 163-166.
- 53. Muhammed, G.; Khan, M.Z.; Athar, M. and Shakoor, A. (1996): Clinical. microbiological and epidermiological features of an outbreak of ulcerative blepharitis associated with Staph. epidermidis in buffalo herds. Preventive Vet. Med., 26(1): 47-52.
- 54.Brown, H.; Elliston, N.G.; McAskill, J.W.; Muenster, O.A and Tonkinson, L.V. (1973): Tylosin phosphate (TP) and tylosin urea adduct (TUA) for prevention of liver abscesses, improved weight gain and feed efficiency in feedlot cattle. J. Anim. Sci. 37: 1085-1091.
- 55.Potter, E.L.; Wary, M.I.; Muller, R.D.; Grueter, H.P.; McAskill, J. and Young,

- D.C. (1985): Effect of monensin and tylosin on average daily gain, feed efficiency and liver abscess incidence in feedlot cattle. J. Anim. Sci. 61: 1058-1065.
- 56. Specht, H.; Gedek, W. and Dirksen, G. (1988): Minimum inhibitory concentrations of different antibiotics for Pasteurella multocida, Pasteurella haemolytica and Corvnebacterium pyogenes of bovine origin and therapeutic considerations. Bovine Pract. 23: 35-41
- 57.Rood, J.I.; Marker, E.A.; Samero, E.B.; Campos, E. and Duncan, C.L. (1978): Isolation and characterization of multiple antibiotic resistant to C. perfringens strains faeces. Antimicrobial Chemoth. 13(5):871-880

الملخص العربي

الإصابات البكتيرية للكبد والتغيرات البيوكيميائية المصاحبة لها في الجاموس في محافظة المنوفية

الهام ايراهيم عطوة* علا فؤاد عبد الغني طلخان** *قسم البكتريولوجي * وقسم الكيمياء الحيوية بمعهد بحوث صحة الحيوان ـ معمل شبين الكوم

في هذة الدراسة تم فحص (٧٥) عينة من الكبد المصاب للجاموس من المجازر المختلفة بمحافظة المنوفية فوجد أن (٢٦) عينة إيجابية للفحص البكتيريولوجي بنسبة (٨٤%). وبالفحص البكتريولوجي للأكباد المصابة في الجاموس ثم عزل الميكروب العنقودي الذهبي والأشريشيا القولونية والمكور السبحي الصديدي والكلوستريديم بيرفرنجينز نوع (أ) والكوريني الصديدي والكليبسيلا الرنوية والمستروباكتر والبروتيوس فيلجارس بنسبة (٢٠,٦% ،١٠,٧ ، ١٠,٧ ، ١٠,٥ ،

٣,٥% ، ٢,٧ %) على الترتيب). وباجراء الفحوصات البيوكيمپائية لعينات مصل الدم للحيوانات المعزول منها الميكروب الذهبي العنقودي ومقارنتها بالعينات السلبية إتضَّحت النتائج التألية.

١- وتَجوُّد زّيادةٌ عالليةُ المّعنوية جداً في المالون داي ألدهيد والجلوبيولين الكلي مع نقص عالى المعنوية جدا في البروتين الكلى والالبيومين .

٢- وجود زيادة عالية المعنوية جدا في انزيمات الكبد (ALT, AST).
 ٣- وجود زيادة عالية المعنوية جدا في بولينا الدم وحمض اليوريك والكرياتنين.

وتم ايضًا دراسة مدى حساسية العترات البكتيرية المعزولة للمضادات الحيوية فكانت معظم العترات من الميكروب العنقودي الذهبي أكثر حساسية للكيفالوسين والأريثروميسين والكاناميسين والدوكسي سين والدوكسي سيكلين و الأنروفلوكساسين و التراي ميثوبريم مع السلفاميثوكسازول. كما أننا نلاحظ أن حساسية المصادات الحيوية تختلف باختلاف البكتريا المعزولة فنجد أن الكاناميسين

و الدوكسي سيكلين والسبر وفلوكساسين و التَرَّاي ميثوبريم مع السلفاميثُوَّكساز وَلَ من أفضل المَصْـاَداتُ البكتيرية لعلاج الأصابات البكتيرية للكبد بينما البنسيلين والاستربتومايسين كانت غير مؤثرة كعلاج للبكتريا