

Antioxidant And Immunostimulant Effects Of *Peganum Harmala* Seeds In Stressed Broiler Chicken

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ABSTRACT

The present study was carried out to determine to any extent *Peganum harmala* seeds extract can antagonise the adverse effect of heat stress on antioxidant and immune status of broiler chicken. The results demonstrated a significant decrease in glutathione content (GSH), total protein, albumin, γ globulin, total leucocytic count and lymphocytes while increased catalase, glutathione-S-transferase (GST) and monocytes were recorded, in birds under heat stress (6 h/day of 39°C and 70% relative humidity). Treatment of heat stressed chicken with 100 mg/kg b.w./day of *P.harmala* seeds extract for 7 days resulted in a significant increase in GSH content, catalase and γ globulin with restoring most values of the resting parameters to the control ones. This study can substantiate the role of *P. harmala* as an antioxidant and immunostimulant for humoral immunity and in improving the status of broiler chicken under heat stress.

INTRODUCTION

Peganum harmala is the most ancient used plant in global flock medicine. The major alkaloids present in the seeds of the *peganum harmala* are beta-carbolines like harmaline and harmine (1). It is well established that *P. harmala* has antibacterial and antifungal effect (2), protective role against lipid peroxidation, potassium cyanide intoxication and whole-body lethal irradiation (3). In addition, it has hypothermic effect mainly through endogenous 5-HT (5-hydroxy traptamine) stimulation of 5-HT1A receptor (4). *P. harmala* was reported to be used in the treatment of haemosporidian infection and neoplasms (1,5, 6). Also, it is used as vasodilatory (7) and a disinfecting agent (8).

During summer months, heat stress is of great concern in all types of poultry production and this unfavourable or stressful environmental condition can negatively affect animal's immune system (9), feed consumption, growth rate, hatchability, mortality and the prosperity of the poultry industry (10). It has been shown that heat-induced reactive oxygen species (ROS) formation may be an additional factor that provides molecular changes in DNA, protein, lipid and other biological molecules (11). In addition, it could result in oxidative stress,

which in turn lead to cytotoxicity (12), decreased organ function and reduced thermotolerance in old animals (13).

The aim of the present study was to determine the effect of *Peganum harmala* on some anti-oxidant enzymes, total glutathione, serum total protein, albumin and α , β and γ globulins, in addition to the total and differential leucocytic count in broiler chickens exposed to heat stress.

MATERIAL AND METHODS

Experimental design

Eighty Hubbard broilers aged three weeks with 550-600 gm average weight were used. The birds were obtained from private farm at Sharkia governorate. Chickens were kept in wire-floored batteries within a thermostatically and humidistatically controlled environmental chamber and supplied with food and water *ad libitum*. After one week of adaptation chickens were divided into four groups of 20 birds each:

The 1st control group

Birds were kept under healthy and constant environmental conditions [27°C and 50% relative humidity (RH)].

The 2nd group

Birds of this group were subjected to a daily cyclic heat stress period (6 h of 39°C and 70% RH) for 7 days.

The 3rd group

This gp. was exposed to periods of heat stress as the previous group with supplying 100 mg/kg b.w./ day of *P. harmala* seeds extract (4) in drinking suspension for 7 days.

The fourth group

Birds were kept under healthy environmental condition as control with adding 100 mg/kg.b.w/day of *P. harmala* seeds extract in drinking suspension for 7 days.

Sampling: At the end of experimental period, 10 blood samples were collected from birds in each group by puncture of the brachial vein, 5 blood samples were collected with anticoagulant for blood picture and the other five samples were kept without anticoagulant to obtain clear sera that were kept at -20°C until used for biochemical analysis.

Biochemical analysis

- The total glutathione (GSH) content and catalase activity were determined colourimetrically (14, 15). While glutathione-S-transferase (GST) activity was estimated spectro-photo-metrically (16). Electrophoresis for protein fractionation (albumin, α , β and γ globulins) was performed (17).

Leucogram studies

Total and differential leukocytic count were performed using Natt and Herrick diluting solution and Wright stain (18).

Statistical analysis:

The obtained data were analysed using ANOVA computerized programme (19).

RESULTS

The present data showed a significant decrease in total glutathione content and increased both catalase and glutathione-S-transferase activity in broiler chicken under heat stress (Table 1). In groups receiving *P. harmala* the result exhibited a significant elevation in glutathione content and catalase activity while glutathione-S-transferase activity did not differ from the control groups.

Results in Table 2 revealed a significant decrease in total protein, albumin and γ globulin in birds under heat stress, while groups given *P. harmala* showed no significant changes in the concentration of both total protein and albumin whereas a significant increase in of β and γ globulins were noticed in these groups in comparison to the control.

Heat stress resulted in a significant decrease in total leucocytic count and lymphocytes while monocytes increased significantly. At the same time, the use of *P. harmala* resulted in elevation of total leucocyte and lymphocytes with decreasing monocytes to simulate nearly the control group (Table 3).

Table 1. Total glutathione, catalase, glutathione-S-transferase of 21 days old broiler chicken under heat stress treated with *Peganum harmala* extract for 7 days (n= 10).

Parameters Treatments	Total glutathione $\mu\text{mol/g protein}$	Catalase $\mu\text{mol/g protein}$	Glutathione-S- transferase $\mu\text{mol/min./mg protein}$
Control (C)	29.3 \pm 1.6 b	27.0 \pm 2.8 b	0.51 \pm 0.01 b
Heat stress (H)	20.5 \pm 1.9 c	32.1 \pm 2.1 a	1.7 \pm 0.07 a
Heat stress + <i>P.harmala</i> (H + P)	38.7 \pm 1.7 a	34.1 \pm 2.5 a	0.47 \pm 0.02 b
<i>P.harmala</i> (P)	40.1 \pm 1.8 a	35.7 \pm 1.8 a	0.49 \pm 0.01 b

Mean \pm S.E

Means in the same column followed by different letters are significantly different (P<0.05).

Table 2. Total protein, albumin, total globulin and its fractionation of 21 days old broiler chicken under heat stress treated with *Paganum harmala* extract for 7 days. (n= 10).

Parameters Treatments	Total protein (gm/dl)	Albumin (gm/dl)	α globulin (%)	β globulin (%)	γ globulin (%)
Control	4.10 \pm 0.31 a	1.50 \pm 0.07 a	0.51 \pm 0.05 a	0.35 \pm 0.02 b	1.70 \pm 0.14 c
Heat stress	3.30 \pm 0.11 b	0.8 \pm 0.07 b	0.55 \pm 0.03 a	0.41 \pm 0.02 b	1.48 \pm 0.07 d
Heat stress + <i>P.harmala</i>	4.80 \pm 0.15 a	1.40 \pm 0.10 a	0.58 \pm 0.04 a	0.54 \pm 0.02 a	2.28 \pm 0.05 a
<i>P.harmala</i>	4.3 \pm 0.26 a	1.35 \pm 0.08 a	0.50 \pm 0.06 a	0.48 \pm 0.03 a	2.00 \pm 0.07 b

Mean \pm S.E

Means in the same column followed by different letters are significantly different (P<0.05).

Table 3. Total and differential leucocytic count of 21 days old broiler chicken under heat stress treated with *Peganum harmala* extract for 7 days (n= 10).

Parameters Treatments	Total leucocytic count 10 ³ /μl	Lymphocyte (%)	Heterophils (%)	Monocytes (%)	Eosinophils (%)	Basophils (%)
Control	26.0 ±1.14 a	29.0±0.70 a	64.0±1.58 a	3.0±0.70 b	2.0±0.31 b	2.0±0.31 a
Heat stress	15.0±0.70 b	21.8±0.80 b	63.2±0.70 a	10.0±0.70 a	3.0±0.44 a	2.0±0.44 a
Heat stress + P.harmala	25.4±0.1.20 a	29.0±0.70 a	65.2±0.86 a	2.0±0.54 b	1.8±0.20 b	2.0±0.31 a
P.harmala	27.0±0.70 a	30.0±0.70 a	65.0±1.14 a	2.0±0.31 b	2.0±0.31 b	1.0±0.31 b

Mean ± S.E

Means in the same column followed by different letters are significantly different (P<0.05).

DISCUSSION

The harmful effects of hyperthermia on organisms have become a matter of concern. ROS resulted from heat exposure are controlled *in vivo* by a wide spectrum of enzymatic and non-enzymatic defense mechanisms. The present data revealed a significant decrease in the level of GSH in heat stressed chickens, which is consistent with the previous results (20) which attributed this decrease to the increased oxidation to oxidized glutathione (GSSG), increased degradation or decreased synthesis. Also it was recorded that hyperthermia may reduce body concentrations of antioxidants required for homeostasis (21).

The data present here showed increased activity of both catalase and GST in chicken under heat stress. In agreement with this results, it was found that heat stress caused a significant increase in catalase and GST in rats (20) and increased endogenous catalase activity in myocardium after heat stress (22). The elevated activity of catalase may be due to hyperthermia enhanced the cytotoxicity of H₂O₂ in cells and catalase has important role in reduction of H₂O₂ to H₂O and O₂ (23). Elevation of GST activity after heat stress may be derived from the increased lipophilic substances released during

oxidative damage to polyunsaturated lipids, such as short-chain aldehydes and alkenals, which are substrates for GST (24).

The obtained data showed increased the GSH content and catalase activity in groups receiving P.harmala. In accordance with this study glutathione, glutathione peroxidase and glutathione reductase were recorded to be significantly higher in melatonin treated rats (25), suggesting that melatonin may effectively normalize the impaired antioxidant status. Where melatonin is active metabolite to 6methoxy-tetrahydro-beta-carboline (26). Furthermore, beta-carboline had potent antioxidants effects and evaluated for cerebral protection against lipid peroxidation (26). Harmaline and harmine was reported to reduce the rate of vitamin E disappearance and exhibited a significant free radical scavenging capacity.

In the heat stressed chickens, the present data exhibited a significant decrease in the concentration of total protein, albumin and γ globulin. Similarly the total protein and albumin were lowered significantly in broiler chickens under toxic stress (26). Heat stress may cause liver injury through increasing the production of ROS levels and oxidative damage to hepatocellular macromolecules

(29) where liver is the main site for plasma protein synthesis.

The present finding revealed a significant elevation of γ globulin in both *P.harmala* treated groups. Similarly γ globulin in rats showed a significant elevation after oral administration of *Rhazya stricta* extract (harmal) and reported no deaths or even toxic symptoms from this extract at dose ranging from 400-4000 mg/kg B.W, indicating the presence of immunopotentiating factor in the plant extract (30).

In heat stressed chicken receiving *P.harmala*, the total protein and albumin concentration returned to their normal values, this may be due to hypothermic effect of *P.harmala* that antagonized the adverse effect of heat stress on these parameters (4).

Total leucocytic count and lymphocytes of chicken under heat stress were lowered significantly in this study. In agreement with this result lymphocytes decreased significantly in the blood of adrenocorticotrophic hormone (ACTH)-injected and heat stressed birds (9). Such leucocytopenia and lymphocytopenia may be attributed to the fact that heat stress increases the activity of the hypothalmo-pituitary adrenal axis in chickens (31) causing increase glucocorticoids which induce a mechanism of depressing the lymphopoiesis, arresting lymphocyte formation and producing lysis of these cells, so that in severe stress, the lymphatic tissues are depleted (32). *P.harmala* receiving groups revealed leucocytic picture around the control level, this may be related to melatonin which is metabolized from beta carboline induces metosis and correction of DNA damage (26).

From the obtained results, it was concluded that *P. harmala* treatment can help to restore the altered antioxidant and immunosuppressive effects on broiler chicken under heat stress.

Acknowledgements

The author expresses her appreciation to Prof. Dr. Mona M. Abd El-Hady

(Physiology Dept., Faculty of Vet. Med. Zagazig University) for her assistance during this study.

REFERENCES

1. Sobhani, A.M.; Ebrahimi, S.A. and Mahmoudian, M. (2002): An in vitro evaluation of human DNA topoisomerase I inhibition by *Peganum harmala* L. seeds extract and its beta carboline alkaloids. J. Pharm. Sci. 5 (1): 19-23.
2. El-Rifaie, E.S.M. (1980): *Peganum harmala*: its use in certain dermatoses. Int. J. Dermatol. 19 (4): 221-222.
3. Kawashima, Y.; Horiguchi, A.; Taguchi, M.; Tuyuki, Y.; Karasawa, Y.; Araki, H. and Hatayama, K. (1995): Synthesis and pharmacological evaluation of 1,2, 3, 4-tetrahydro-beta-carboline derivatives: chem pharm bull (Tokyo) 43: 5: 783-787.
4. Abdel-Fattah, A.F.; Matsumoto, K.; Gammaz, H.A. and Watanabe, H. (1995): Hypothermic effect of harmala alkaloid in rats: involvement of serotonergic mechanism. Pharmacol. Biochem. Behav. 52 (2): 421-426.
5. Hu, T.; Fan, B.; Liang, J.; Zhao, S.; Dang, P.; Gao, F. and Dong, M. (1997): Observations on the treatment of natural heamosporidia infections by total alkaloid of *Peganum harmala* L in cattle. Trop. Anim. Health. Prod. 29 (4): 725-765.
6. Lamchouri, F.; Settaf, A.; Cherrah, Y.; Zemzami, M.; Lyoussi, B.; Zaid, A.; Atif, N. and Hassar, M. (1999): Antitumor principles from *Peganum harmala* seeds. Therapie 54 (6): 753-758.
7. Berrougui, H.; Herera-Gonzalez, M.D.; Marhuenda, E.; Ettaib, A. and Hmamouchi, M. (2002): Relaxant activity of methanolic extract from seeds of *Peganum harmala* on isolated rat aorta. Therapie. 27 (3): 236-241.
8. Shahverdi, A.R.; Monsef-Esfahani, H.R.; Nickavar, B.; Bitarafan, L.; Khodaei, S. and Khoshakhlagh, N. (2005): Antimicrobial activity and main chemical

- composition of two smoke condensates from *Peganum harmala* seeds. *Z Natur Forsch [C]*. 60 (9-10): 707-710.
9. Trout, J.M. and Mashaly, M.M. (1994): The effects of adrenocortico-tropic hormone and heat stress on the distribution of lymphocyte populations in immature male chickens. *Poultry Science* 73: 1694-1698.
 10. Bartlett, J.R. and Smith, M.O. (2003): Effects of different levels of zinc on the performance and immunocompetence of broilers under heat stress. *Poult. Sic.*, 82: 1580-1588.
 11. Bruskov, V.I.; Malakhova, L.V.; Masalimov, Z.K. and Chernikov, A.V. (2002): Heat-induced formation of reactive oxygen species and 8-oxoguanine, a biomarker of damage to DNA. *Nucleic Acids Research* 30 (6): 1354-1363.
 12. Bernabucci, U.; Ronchi, B.; Lacetera, N. and Nardone, A. (2002): Markers of oxidative status in plasma and erythrocytes of transition dairy cows during hot season. *Journal of Dairy Science* 85 (9): 2173-2179.
 13. Zhang, H.; Xu, L.; Drake, V.J.; Xie, L.; Oberley, L.W. and Kregel, K.C. (2003): Heat induced liver injury in old rats is associated with exaggerated oxidative stress and altered transcription factor activation. *FASEB J* 17: 2293-2295.
 14. Akerboon, P.T. and Sies, H. (1981): Assay of glutathione, glutathione disulfide and glutathione mixed disulfides in biological samples. In *Methods in Enzymology* (Jakopy, B.W., ed) Academic Press Inc, 77: pp. 373-382.
 15. Ponting, J.D. and Joslyn, M.A. (1948): Ascorbic acid oxidation and browning in apple tissue extracts. *Arsh. Biochem.*, pp. 19-47. Porpova, M.P. and Nauch, M.R. (1955): *Thru Chem. Abst.*, 55: 22642e.
 16. Habig, H.W.; Micheal, J.P. and William, B.J. (1974): Glutathione-S-transferase, the first enzymatic step in mercapturic acid formation. *J. Biochemical Chem.*, 25: 7130-7139.
 17. Laemmli, V.K. (1970): Structural protein during the assembly of the head of bacteriophage T₄. *Nature*, 227, 15: 680.
 18. Coles, E.H. (1986): *Veterinary Clinical Pathology*, 4th ed., W.B. Saunders Company Philadelphia and London.
 19. Snedecor, G.W. and Cochran, W. (1980): *Statistical Methods*, 7th ed., Iowa State Univ. Press. Ames., Iowa, USA.
 20. Ozturk, O. and Gumuslu, S. (2004): Age-related changes of antioxidant enzyme activities, glutathione status and lipid peroxidation in rat erythrocytes after heat stress. *Life Sciences* 75: 1551-1565.
 21. Lakritz, J.; Leonard, M.J.; Eichen, P.A.; Rottinghous, G.E.; Johnson, G.C. and Spiers, D.E. (2002): Whole-blood concentrations of glutathione in cattle exposed to heat stress or a combination of heat stress and endophyte-infected tall fescue toxins in controlled environmental conditions. *Am. J. of Vet. Res.* 63 (6): 799-803.
 22. Kingman, Jr., J.G.; Simard, D.; Rouleau, J.R.; Tanguay, R.M. and Currie, R.W. (1996): Effect of 3-aminotriazole on hyperthermia-mediated cardioprotection in rabbits. *American Journal of Physiology* 270 (4/2), H1165- H1171.
 23. Ceballos-Picot, I.; Trivier, J.M.; Nicole, A.; Sinet, P.M. and Thevenin, M. (1992): Age-correlated modifications of copper-zinc superoxide dismutase and glutathione-related enzyme activities in human erythrocytes. *Clinical chemistry* 28 (1): 66-70.
 24. Galli, F.; Rovidati, S.; Benedetti, S.; Buoncrisiani, U.; Covarelli, C.; Floridi, A. and Canestrari, F. (1999): Overexpression of erythrocyte glutathione-S-transferase in uremia and dialysis. *Clinical Chemistry* 45 (10): 1781-1788.

25. Meki, A.R.; Esmail Eel-D.; Hussein, A.A. and Hassanein, H.M. (2004): Caspase and heat shock protein-70 in rat liver treated with aflatoxin B₁: effect of melatonin. *Toxicol* 43 (1): 93-100.
26. Frederiksen, T. and Pless, G. (1998): Antioxidantive potential of melatonin and pinoline in lipid peroxidation of brain tissue. Master's thesis 1-52, App.
27. Berrougui, H.; Isabelle, M.; Cloutier, M.; Hmamouchi, M. and Khalil, A. (2006): Protective effects of *Peganum harmala* L. extract harmine and harmaline against human low-density lipoprotein oxidation. *J. Pharm. Pharmacol.* 58 (7): 967-974.
28. Abo-Norag, M.; Edrington, T.S.; Kubena, L.F.; Harrey, R.B. and Philips, T.D. (1995): Influence of a hydrated sodium calcium aluminosilicate and virginiamycin on aflatoxicosis in broiler chicks. *Poult. Sci.* 74 (4): 626-632.
29. Zhang, H.J.; Doctrow, S.R.; Larry, W.O. and Kregel, K.C. (2005): Chronic antioxidant enzyme mimetic treatment differentially modulates hyperthermia-induced liver HSP 70 expression with aging. *J. Appl. Physiol.* 100: 1385-1391.
30. Abdel-Aziz, S.A.; Omar, A.A.; Haroun, E.M. and Farah, M.O. (1992): Clinicopathological and toxicological studies of oral administration of *Rhazya stricta* extract "Harmal" in rats. *J. King Saudi Univ.* 4: 117-123.
31. Edens, F.W. and Siegel, H.S. (1975): Adrenal responses in high and low-response lines of chickens during acute heat stress. *Gen. Comp. Endocrinol.* 25: 64-73.
32. Isobe, T. and Lillehoj, H.S. (1992): Effects of corticosteroids on lymphocyte subpopulation and lymphokine secretion in chickens. *Avian Dis.* 36: 590-596.

الملخص العربي

التأثير المضاد للأكسدة والمحفز للمناعة لبذور نبات الحرمل في بدارى التسمين المجهدة

هدى محمد لطفى عبد الله

معهد بحوث صحة الحيوان بالزقازيق

يعتبر الاجهاد الحرارى من أكثر المشاكل الحقلية خطورة. ويرتبط ارتفاع درجة الحرارة أثناء شهور الصيف بزيادة قابلية الطيور للإصابة بالأمراض وزيادة الوفيات.

قد أجريت هذه الدراسة لتحديد إلى أى مدى يمكن لبذور نبات الحرمل أن تضاد التأثير الضار للإجهاد الحرارى على مضادات الأكسدة وعلى الحالة المناعية لبدارى التسمين وقد أثبتت هذه الدراسة نقص مستوى الجلوتاثيون والبروتينات الكلية والزلال والجاما جلوبيولين والعدد الكلى لكرات الدم البيضاء والخلايا الليمفاوية بينما ازداد معدل انزيم الكتاليز والجلوتاثيون اس ترانسفيراز فى الطيور المعرضة للإجهاد الحرارى (6 ساعات يوميا عند درجة حرارة 39°م و 70% من الرطوبة النسبية).

وعند معالجة الطيور المعرضة للإجهاد الحرارى ببذور نبات الحرمل بنسبة 100 مجم/كجم من وزن الجسم يوميا لمدة سبع أيام كانت هناك زيادة معنوية فى معدل الجلوتاثيون وانزيم الكتاليز والجاما جلوبيولين مع رجوع معظم القيم الباقية إلى مستوى المجموعة الضابطة.

وهذه الدراسة تعزز دور بذور نبات الحرمل كمضاد للأكسدة وكمحفز للمناعة (المناعة اللاخلوية) وفى تحسين الحالة العامة لبدارى التسمين المعرضة للإجهاد الحرارى.