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## INCIDENCE OF AEROBIC SPOREFORMERS IN UHT MILK

(With 4 Tables and 3 Figures)

By

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مدى تواجد الميكروبات المتحوصلة الهوائية في اللبن المعقم

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أجرى هذا البحث علي سبعون عينة عشوائية من اللبن المعقم (منها ٢٥ عينة لبن طبيعي كامل الدسم ، ١٥ عينة لبن منكه بالموز ، ١٥ عينة لبن منكه بالفراولة و ١٥ عينة أخرى لبن منكه بالشيكولاته) تم تجميعها من محلات البقالة والمحلات التجارية المختلفة في كفر الشيخ ، جمهورية مصر العربية. تم فحص جميع العينات للعدد الكلي للبكتريا الهوائية بعد تحضينها عند ٣٠ - ٣٥ م لمدة ٧ أيام وسجلت النتائج متوسطات ١٠,٤ ، ٢٧,٥ ، ١٩,٦ و ١٥,٢ خلية لكل ميلتر لبن من عينات اللبن الطبيعي كامل الدسم ، اللبن المنكه بالموز ، اللبن بالفراولة واللبن بالشيكولاته علي التوالي. كما أوضحت الدراسة أن ١٦ عينة (بنسبة ٢٢,٩%) قد تجاوزت الحد المسموح به في المواصفات القياسية المصرية للبن المعقم من حيث العدد الكلي للبكتريا (ألا يزيد علي ١٠ خلية/ملي بعد التحضين عند ٣٠ - ٣٥ م/٧ أيام). كما أظهرت نتائج فحص عينات اللبن المعقم أن ٦٤% من عينات اللبن الطبيعي كامل الدسم ، ٨٠% من اللبن بالموز ، ٧٣,٣% من اللبن بالفراولة وكذلك ٦٦,٦٧% من اللبن بالشيكولاته إيجابية للبكتريا الهوائية المتحوصلة (الباسيلس) بمتوسط عددي ٢٨,٦٧ ، ٧,٢٧ ، ٧,٧٠ ميكروب/ملي من عينات اللبن المعقم السابق علي التوالي. تم عزل وتصنيف البكتريا الهوائية المتحوصلة (الباسيلس) ووجد أن ميكروب الباسيلس إستيروثيروموفيلس هو الأكثر تواجد حيث تم عزله من ١٩ (٢٧,١%) من العينات. كذلك تم عزل أنواع أخرى مثل باسيلس سيرس ، باسيلس ليكنيفورمس ، باسيلس سيركيولانس ، باسيلس ميجاوتيريوم ، باسيلس ساتلس ، باسيلس كواجيولانس وباسيلس ألفي من العينات بنسب مختلفة كالآتي: (٢٠%) ، (١٤,٣%) ، (١٢,٩%) ، (١٢,٩%) ، (١٢,٩%) ، (١١,٤%) و (١,٤%) علي التوالي. وقد تم مناقشة مصادر تلوث الألبان الخام بهذه الميكروبات ، الأهمية الاقتصادية والصحية للمعزولات وكذلك الافتراضات الواجب إتباعها للحد من تلوث الألبان بها وتحسين جودة المنتج.

## SUMMARY

Seventy (25 unflavored full cream, 15 banana flavored, 15 strawberry flavored and 15 chocolate flavored) UHT milk samples were collected

randomly from different groceries and supermarkets in Kafrelsheikh, Egypt. All samples were examined for aerobic plate count and aerobic sporeformers. Results revealed that the mean aerobic plate count for unflavored full cream, banana flavored, strawberry and chocolate flavored UHT milk samples examined (after incubation of samples at 30-35°C/7days) were  $10.4 \pm 9.6$ ,  $27.5 \pm 39.4$ ,  $19.6 \pm 27.9$  and  $15.2 \pm 26.6$  cfu/ml respectively. Results declared that 16 (22.9%) out of 70 UHT milk samples examined, exceeded the Egyptian standards of total aerobic bacterial count (10 cfu/ml) for long life sterilized milk. Statistical analytical results of aerobic sporeformers count revealed that 16(64%) of unflavored full cream, 12 (80%) of banana flavored, 11(73.3%) of strawberry flavored and 10 (66.67%) of chocolate flavored UHT milk samples were positive for aerobic sporeformers with mean counts of  $13.19 \pm 28.7$ ,  $28.67 \pm 44.1$ ,  $7.27 \pm 4.3$  and  $7.70 \pm 4.6$  cfu/ml respectively. The isolated bacillus species were identified as *B. stearothermophilus* which recorded the highest incidence that it could be detected in 19 (27.1%) of the examined samples, followed by *B. cereus*, *B. Licheniforms*, *B. circulans*, *B. megaterium*, *B. subtilis*, *B. coagulans* and *B. alvei* that could be isolated from 14(20%), 10 (14.3%), 9(12.9%), 9(12.9%), 9(12.9%), 8 (11.4%) and 1 (1.4%) of the examined samples respectively. Source of these bacteria in milk, their economic and public health importance as well as the suggestive measures for control were discussed. This study was planned to throw light on aerobic plate count and the incidence of aerobic sporeformers in the examined UHT milk samples regarding the Egyptian Standards.

**Key words:** UHT milk- APC- Aerobic sporformers – *Bacillus* Species

## INTRODUCTION

Ultra-heat treated (UHT) milk was developed to meet the demand for milk be stable for extended periods at room temperature. UHT was defined by Dr. Harold Burton- Pioneer of UHT milk processing as a treatment in which milk is heated to a temperature of 135-150 °C in continuous flow in a heat exchanger for sufficient length of time (1- 4 S) to achieve commercial sterility with an acceptable amount of change in the product (Varnam and Sutherland, 1994). In spite of this treatment, UHT milk was accounted for 1.5% of 69 outbreaks due to milk and dairy products as reported by De Buyser *et al.* (2001). Also, Hammer *et al.* (1995) recorded that heat resistant

mesophilic aerobic sporeformers have been detected in UHT milk in several European countries since 1985.

Members of genus *Bacillus* such as *B. licheniformis*, *B. cereus*, *B. subtilis*, *B. stearothermophilus*, *B. polymyxa*, *B. badius*, *B. coagulans*, *B. mycoides* and *B. pumilus* are the most important spoilage species isolated from UHT milk by many authors (Foschino *et al.*, 1990; Hasan, 1990; Aly, 1992; El-Shennawi *et al.*, 1995; Ezz-El-Din, 1999 and Taher, 2004). These bacteria contaminate milk supplies from water, udder and teat surfaces or from feed concentrate, dust, soil, milk stone deposits on farm tanks, pumps, pipelines and processing equipment and their endospores can survive UHT treatment (Meer *et al.*, 1991; Aman *et al.*, 1998; Huang *et al.*, 1999; Scheldeman *et al.*, 2002). Post processing contamination due to failure in equipment sterilization downstream thermal processing or most commonly during packaging was also recorded (Varnam and Sutherland, 1994).

*Bacillus spp.* are associated with flavor defects as fruity, sour, bitter, unclean also sweet curdling and bitty cream which caused by the action of proteolytic, lipolytic and phospholipase enzymes. Although the main concern about the presence of *Bacillus spp.* in milk is off-flavor production with consequent reduction of shelf life, it also represents a public health hazard. *B. cereus* food poisoning is a major concern world wide and it is responsible for two forms of human gastroenteritis, but the number of cells required for toxin production is high,  $10^6$  to  $10^7$ /ml (Ombui *et al.*, 2008; Meer *et al.*, 1991; Rangasamy *et al.*, 1993).

There are several reasons for the problems of spore forming bacteria in the dairy industry. First of all, it is impossible to completely avoid their presence in milk. Secondly their spores are very hydrophobic and will attach to surfaces of the pipelines of the dairy plant where they might multiply and will re-sporulate. A third problem is that spores are heat resistant (Andersson *et al.*, 1995). Therefore this work was planned to throw light on the incidence of aerobic sporeformers in UHT milk.

## **MATERIALS and METHODS**

### **Samples:**

Seventy random samples of market UHT milk were collected randomly from different groceries and supermarkets in Kafr El-Sheikh, Egypt. Twenty five samples were unflavored full cream UHT milk and forty five were flavored UHT milk (chocolate, banana and strawberry, 15 samples each). All samples were rapidly delivered to the laboratory, prepared and serially diluted according to A.P.H.A. (1992).

UHT milk samples were divided into to parts, first part was incubated at 30-35°C for seven days then examined for:

**Aerobic plate count:**

According the technique recommended by A.P.H.A. (1992).

The second part of the samples was serially diluted and examined for:

**Aerobic sporeformers count:**

The already prepared serial dilutions and the original samples were heated at 80°C/10 minutes to destruct all vegetative cells then cooled to 10°C. One milliliter quantities from each dilution as well as original samples were plated in sterile duplicate pertri dishes using dextrose tryptone agar (Oxoid, 1980) and polymyxin pyruvate egg yolk manitol bromothymol blue agar (PEMBA) (Holbrook and Anderson, 1980). Following incubation at 32°C for 72 h and 30°C for 48 h respectively, the total number of colonies were calculated and recorded. Separate colonies of acid producing (yellow halo) and non acid producing were picked up and purified on nutrient slants for further identification.

**Identification of isolated *Bacillus* species:**

Isolated colonies were confirmed by microscopic examination and biochemically according to Krieg and Holt (1984).

## RESULTS

**Table 1:** Statistical analytical results of aerobic plate count in the examined UHT milk samples (after incubation of samples at 30-35°C/7 days)

UHT milk samples	No. of the examined samples	Positive samples		APC ( cfu/ml)			Samples exceeding *E.S.	
		No.	%	Min.	Max.	Mean $\pm$ SD	No.	%
Unflavored full cream	25	20 <sup>a</sup>	80	4	39	10.4 $\pm$ 9.6	6	24
Banana flavored	15	8	53.3	3	100	27.5 $\pm$ 39.4	3	20
Strawberry flavored	15	10	66.67	2	90	19.6 $\pm$ 27.9	3	20
Chocolate flavored	15	10	66.67	2	50	15.2 $\pm$ 26.6	4	26.7
Total	70	48	68.57	2	100	16.67 $\pm$ 22.6	16	22.9

\* Egyptian standards (2005)

**Table 2:** Statistical analytical results of aerobic sporeformers count in the examined UHT milk samples

UHT milk samples	No. of the examined samples	Positive samples		Aerobic sporeformers count (cfu/ml)		
		No.	%	Min.	Max.	Mean $\pm$ SD
Unflavored full cream	25	16	64	3	120	13.19 $\pm$ 28.7
Banana flavored	15	12	80	4	140	28.67 $\pm$ 44.1
Strawberry flavored	15	11	73.3	3	16	7.27 $\pm$ 4.3
Chocolate flavored	15	10	66.67	1	14	7.70 $\pm$ 4.6
Total	70	49	70	1	140	14.53 $\pm$ 27.96

**Table 3:** Frequency distribution of positive UHT milk samples according to aerobic sporeformers count

Aerobic sporeformers count (cfu/ml)	Unflavored Full cream positive UHT milk samples	flavored positive UHT milk samples			Total	
		Banana	Strawberry	Chocolate	No.	% *
< 10	12	4	7	6	29	59.19
10 : < 20	3	5	4	4	16	32.65
20 : < 40	0	1	0	0	1	2.04
40 : < 80	0	0	0	0	0	0
80 : $\leq$ 100	0	1	0	0	1	2.04
> 100	1	1	0	0	2	4.08

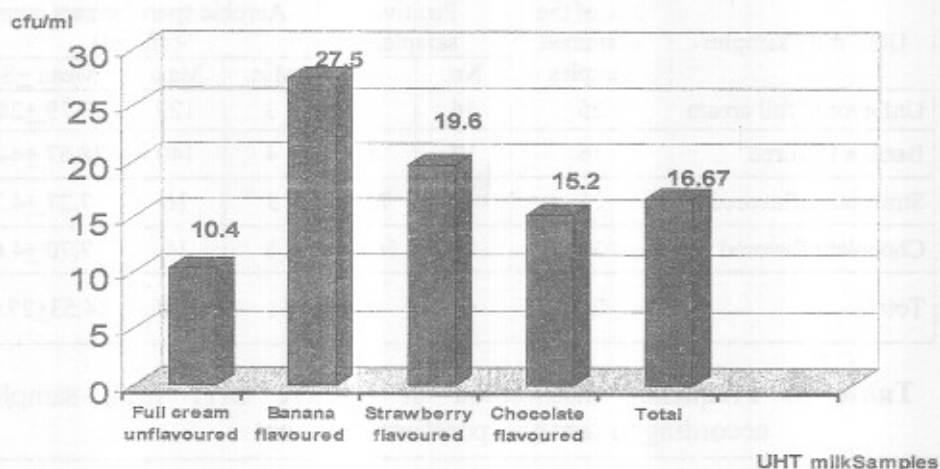
\* Percentages were calculated to the number of positive samples (n=49)

**Table 4:** Statistical analytical results of the identified *Bacillus* species isolated from the examined UHT milk samples

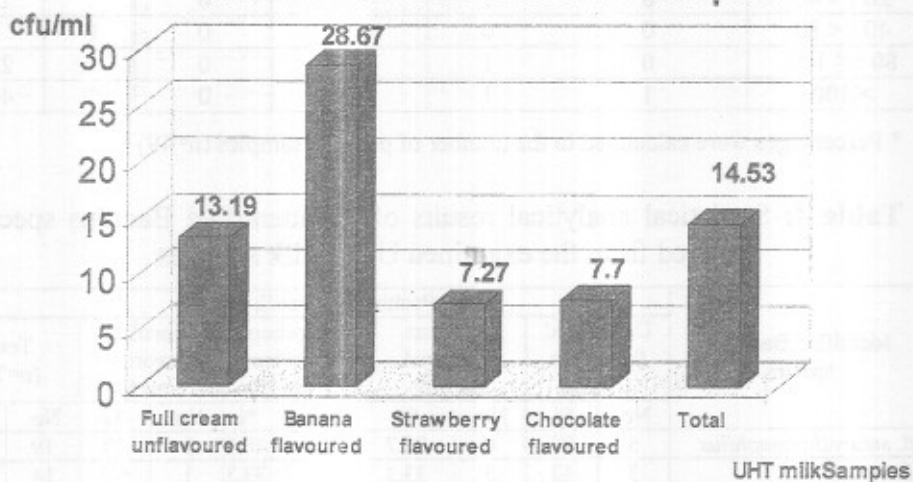
Identified <i>Bacillus</i> species	Positive UHT milk samples									
	Unflavored full cream (n=25)		Banana flavored (n=15)		Strawberry flavored (n=15)		Chocolate flavored (n=15)		Total (n=70)	
	No.	%	No.	%	No.	%	No.	%	No.	%
<i>B. stearothermophilus</i>	5	20	4	26.7	7	46.7	3	20	19	27.1
<i>B. cereus</i>	3	12	5	33.3	5	33.3	1	6.7	14	20
<i>B. Licheniformis</i>	4	16	2	13.3	3	20	1	6.7	10	14.3
<i>B. circulans</i>	3	12	4	26.7	1	6.7	1	6.7	9	12.9
<i>B. megaterium</i>	4	16	1	6.7	1	6.7	3	20	9	12.9
<i>B. subtilis</i>	3	12	2	13.3	1	6.7	3	20	9	12.9
<i>B. coagulans</i>	2	8	3	20	1	6.7	2	13.3	8	11.4
<i>B. alvei</i>	0	0	1	6.7	0	0	0	0	1	1.4



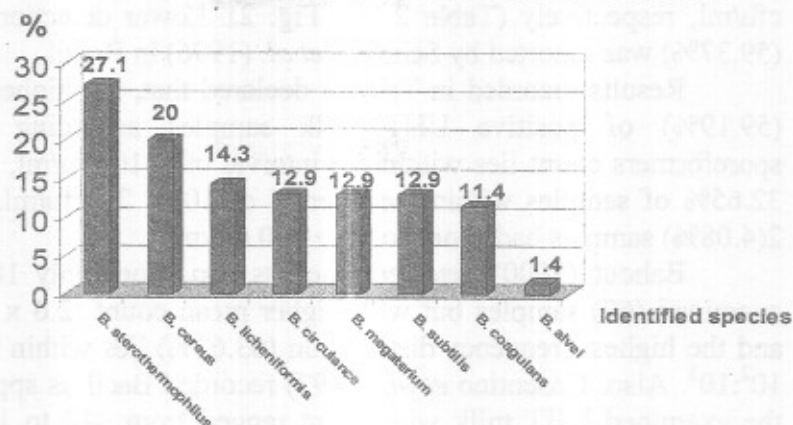
**fig. (1) The mean aerobic plate count of the examined UHT milk samples**



**Fig.(2) The mean aerobic sporeformers count in the examined UHT milk samples**



**fig.(3) Results of the identified Bacillus species isolated from the examined UHT milk samples**



## DISCUSSION

The results presented in Table 1 revealed that aerobic bacteria could be detected in 68.57% of the examined UHT milk samples after incubation at 30-35°C for 7 days (80%, 53.3%, 66.67% and 66.67% of the examined unflavored full cream, banana flavored, strawberry flavored and chocolate flavored UHT milk samples respectively). The mean aerobic plate counts recorded for the previous samples were  $10.4 \pm 9.6$ ,  $27.5 \pm 39.4$ ,  $19.6 \pm 27.9$  and  $15.2 \pm 26.6$  cfu/ml, respectively (Table 1 and Fig. 1). Ezz El-Din (1999) recorded higher aerobic mesophilic count ( $1.29 \times 10^4$  cfu/ml as mean value) for 15 UHT milk samples.

The Egyptian standards (E.S) (2005) for long life sterilized milk reported that the total aerobic bacterial count should not exceed 10 cfu/ml after incubation of milk at 30-35°C for 7 days. Table 1 showed that 16(22.9%) samples out of 70 UHT milk samples examined, exceeded the Egyptian standards; 6 (24%) samples of unflavored full cream UHT milk, 3(20%) samples of both banana and strawberry flavored UHT milk and 4(26.7%) of chocolate flavored UHT milk samples.

Statistical analytical results of aerobic sporeformers count of the examined UHT milk samples revealed that, 49(70%) of the total examined samples, 16(64%) of unflavored full cream, 12(80%) of

banana flavored, 11(73.3%) of strawberry flavored and 10(66.67%) of chocolate flavored UHT milk samples examined were positive with mean counts of  $13.19 \pm 28.7$ ,  $28.67 \pm 44.1$ ,  $7.27 \pm 4.3$  and  $7.70 \pm 4.6$  cfu/ml, respectively (Table 2 and Fig. 2). Lower detection percentage (59.37%) was reported by Schoken *et al.* (1996) in Brazil.

Results recorded in Table 3 declared that, the highest frequency (59.19%) of positive UHT milk samples according to aerobic sporeformers count lies within the interval of  $< 10$  cfu/ml, followed by 32.65% of samples within the interval of  $10: < 20$  cfu/ml, while only 2(4.08%) samples had spore count  $> 100$  cfu/ml.

Bahout (2000) detected *Bacillus* spp. from only 18.3% of the examined (60) samples but with higher mean count ( $2.6 \times 10^2$  cfu/ml) and the highest frequency distribution (63.63%) lies within the range of  $10^2:10^3$ . Also, Cosentino *et al.* (1997) recorded *Bacillus* spp. in 30% of the examined UHT milk with count ranged from  $<10$  to 1200 cfu/ml. and Schoken *et al.* (1996) detected *Bacillus* spp. in 19(59.37%) of 32 long life milk samples, of which 2 samples contained  $> 100$  cfu/ml.

The most frequent isolated *Bacillus* species was *B.stearothermophilus* that isolated from 19(27.1%) of the examined samples. It is very heat resistant thermophilic strain that causes flavor spoilage (Lewis, 1999). The other species that could be isolated were *B. cereus*, *B. Licheniformis*, *B. circulans*, *B. megaterium*, *B. subtilis*, *B. coagulans* and *B. alvei* that could be isolated from 14(20%), 10 (14.3%), 9(12.9%), 9(12.9%), 9(12.9%), 8 (11.4%) and 1 (1.4%) of the examined samples respectively (Table 4 and Fig.3 ). The same species were identified by Aly, 1992; El-Shennawi, 1995; Cosentino *et al.*, 1997; Bahout, 2000; Mayr *et al.*, 2004 and Al-Leboudy *et al.*, 2007 at different percentages.

These bacteria can contaminate milk supplies from water, udder, teat surfaces or from feed, dust and soil (Huang *et al.*, 1999 and Scheldeman *et al.*, 2002). Moreover, milking equipment can act as reservoirs for the spores into raw milk (Scheldeman *et al.*, 2005). Furthermore, *Bacillus* species endospores can survive UHT milk production process (Huang *et al.*, 1999; Vyletelova *et al.*, 2002).

The main concern about the presence of these species in milk is not only the production of flavor defects by the action of proteolytic, lipolytic and phospholipase enzymes, but also they represent a public health hazard, as *B. cereus* that is responsible for two form of human gastroenteritis (Meer *et al.*, 1991; Rangasamy *et al.*, 1993).



In conclusion it is suggested that the quality of UHT market milk could be improved by paying more attention to the quality of raw milk and to high standards of plant hygiene to avoid post-processing contamination. In addition, the dairy industry needs a simple rapid, sensitive, reliable and economical method for assessing the presence of heat resistant spores or their enzymes in milk.

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