Studies on Mastitis in Cows at Kaliobia Governorate and the Public Health Importance of Their Milk

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Abstract

A total numbers of 395dairy cows at Kaliobia Governorate were clinically examined and milk samples of them were examined visually and by California mastitis test. The results were 12.9% clinical mastitic cows and 27.1% subclinical mastitic ones. The bacteriological examination of 89 mastitic milk samples proved that 85.4% were positive and 31.6% of them gave pure single cultures and 68.4% mixed ones . They yielded 175 isolates where Staphylococci (34.9%), mainly Coagulase positive Staphaureus (28.6%), and Streptococci (33.1%), mainly Streptagalactiae (17.1%), were the most isolated strains. While from 127 subclinical mastitic samples, 64.1% were positive with 29.1% containing pure single cultures and 70.9% mixed ones. They yielded 241 bacterial isolates where Staphylococci (33.6%), mainly Coagulase positive Staph, aureus (23.2%), Streptococci (30.7%), mainly Strept. agalactiae (15.4%) and Strept. dysagalactiae (11.2%), and E. coli (20.3%) were the main pathogens while other pathogens isolated with low incidence. Coagulasc activity of 142 isolated Staphylococci showed 106 Coagulase positive Staphaureus; 10 Coagulase negative Staph.aureus and 26 Coagulase negative Staph.epidermidis .Haemolysis and CAMP test for 132 isolated Streptococci showed 67 Strept.agalactiae were positive while 48 Strept.dysagalactiae and 17 Strept.uberis were negative. Eight different serotypes of 80 E.coli isolated strains were obtained and E.coli O111 was the most predominant (18.7%). Also, 7 different serogroups of 36 isolated Pseudomoneus aeruginosa and scrogroup K was the most one (25.0%). Most isolated strains were positive for mouse pathogenicity test. Enrofloxacin, lincomycine and gentamycin were the proper antibiotics with the highest in-vitro efficiency against most bacterial isolates. The chemical analysis of milk samples revealed a significant decrease in lactose and casein content. The haematological and biochemical analysis of blood samples showed a significant increase in total leukocyte counts; AST; ALT; AP; urea and creatinine. Meanwhile, a significant decrease in erythrocyte counts; haemoglobin contents; calcium; phosphorous and iron. Treatment of mastitic cows by intramuscular injection of enrofloxacin and intramammary infusion of lincomycine or gentarycin results in complete relief. The public health of isolated strains and controlled measured were also discussed.

Introduction

Mastitis is the most prevalent infectious disease affecting dairy cattle which have many adverse economic implications world wide represented by decrease in quantity and quality of milk components and shorten the reproductive life of affected animals, also feeding of the younger animals by masticic milk leads to retention of the growth; fall of immunity and pathologic states, sometimes with fatal end (2 and 18). Moreover, mastitis is considered of vital importance due to its association with many zoonotic diseases in which milk acts as a source of infection (58). Clinically, the disease occurs as per acute, acute or sub acute which is easily recognized by visual examination of the milk and udder. The subclinical mastitis is serous as it occurs without obvious signs and the secreted milk acts as an invisible potential source of spreading infection and may turn into the clinical form (3 and 22). Therefore, screening tests, bacteriological and biochemical

examinations are necessary for identification of affected quarters (34 and 50). Bovine mastitis is due to different factors but the most important is the invading microorganisms that mostly found in mixed infection. The predominant organisms are *Staphylococcus aureus*; Streptococci and *E.coli* (3, 11, 16, 18, 26, 55 and 59).

Mastitis is characterized by marked changes occur in the level of nearly all major and minor constituents of the milk. These compositional reflect the degree of damage caused to the cells and to the complex array of blood capillaries by the pathogen (5, 9 and 34). Some of these compositional changes are increase of sodium, chloride and phosphorous concentration, while, the casein content, lactose, fat, potassium, calcium and iron were lowered compared with normal ones (9, 15, 26 and 41). Moreover, there is ultration in the haematological and biochemical parameters in mastitic cows viz: Significant increase in total leukocytic counts; AST; ALT; AP; urea and creatinine and a significant decrease in total red blood cells and haemoglobin concentration (2, 51 and 56). As mastitis has sever adverse economic and public health effects, so, this study was carried out to estimate the prevalence of both clinical and subclinical mastitis in cows at Kaliobia Governorate; isolation and identification of the causative agents with bacteriological studies on pure isolates and antibiogram for them to

decide the available and specific treatment of mastitic cows .Moreover, some milk constituents; haematological and biochemical parameters associated with mastitic cows were estimated to be helpful in the diagnosis of mastitis. Also, public health importance of mastitic milk was discussed.

Materials and Methods

A total 398 dairy cows from two private farms and different veterinary clinics at Kaliobia Governorate ,were examined for mastitis .The animals were subjected to clinical examination by visual inspection; palpation of the udder for swelling, redness and pain; beside the physical changes in the milk secreted from such udders. California mastitis test was used for detection of the subclinical mastitis according to (52). With strict aseptic precautions and after discarding the fore milk, milk sample from each quarter was collected in sterile MaCartney bottles and transported to the laboratory in ice container for bacteriological examination and biochemical analysis. For bacteriological examination, a total of 89 clinically mastitic milk samples and 198 subclinically ones were activated by incubation for 12 hours at 37c° then centrifuged at 3000 rpm for 20 minutes .A loopful from sediment was streaked on the surface of Nutrient agar, MacConkey's agar, blood agar ,Mannitol salt agar; Baird Parker agar and modified Edward's media. All plates were incubated aerobically at 37c° for 24-72 hr. The developed colonies were picked up and subcultured for purification. The pure colonies were morphologically identified by Gram-stain and biochemically according to (32 and 46).

Further bacteriological studies on pure isolates were done:

- a- Coagulase activity of **Staphylococci** to different types of plasma (rabbit, sheep, horse and human) using slide coagulase test and confirmed by tube coagulase test following (32).
- b- Haemolysis and Christie-Atkins-Munoh-Peterson (CAMP) test for isolated Streptococci was carried out following (8 and 10).
- c- Serotyping of isolated E.coli strains following (14).
- d- Serotyping of Pseudomoneus aerugenosa following (27).
- N.B: Antisera of *E.coli* and *P.aerugenosa* were obtained from Denka Seiken Co.Ltd., Tokyo, Japan.

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- e- Mouse pathogenicity test for isolated strains was carried out following (25 and 32).
- f- In-vitro sensitivity test for isolated strains against different antibiotic discs (Oxoid standerized discs) using the agar diffusion method following (32)

For chemical analysis of milk, milk gross composition (fat and lactose contents) was measured following (44); casein content was determined by standard method according to (39).

For haematological and biochemical analysis, blood samples with anticoagulant (heparin) were collected from all diseased and healthy cows for determination of total erythrocyte counts, total leukocyte counts and haemoglobin concentration according to

(29) Also, whole blood samples without anticoagulant were collected for determination of Aspartate aminotransferase and Alanine amino-transferase (49), urea (20), creatinine (21), calcium (23), phosphorous (17) and iron (43).

Data obtained were analyzed according to (57).

For trails of treatment, according sensitivity bases 11 clinically mastitic cows and 15 subclinically ones were treated with the most sensitive and effective antibiotics. Enrofloxacin 10% was used intramuscularly for each cow in a dose of 5mg/kg.body weight for 5 successive days and intramammary infusion of either lincocin forte (10ml syringe contain 330mg linconycine + 100mg neomycin + 10mg predinsolin anhydrous) or Gentamast (7.5 ml syringe contain 100 mg gentamycin as sulphate) once daily for mastitic teat for 7 days in clinical cases and 5 days for subclinically ones. After treatment, clinical; California mastitis test, bacteriological and biochemical examination were applied as described above.

Results

The results of clinical examination and California mastitis test (Table, 1) appeared that 51 (12.9%) were clinically mastitic cows. Affected udder was warm, swollen, doughy to firm and painful. The milk was watery, purulent or with thick clots and seven samples were tanged with blood. Most cases showed only one or two quarters affected. The California mastitis test revealed that 107 (27.1%) apparently healthy cows were affected with sub clinical mastitis.

The results of bacteriological examination of 89 mastitic milk samples and 198 subclinically ones (Table,2) showed that 76(85.4%) of clinically mastitic milk samples were positive with 24 (31.6%) containing pure single cultures (13 Staph.aureus;7 Strept.agalactiae; 1 Strept. dysagalactiae;

1 E.coli and 1 Ps. aerugenosa) and 52 (68.4%) mixed ones . While, 127 subclinical mastitic milk samples (64.1%) were positive with 37 (29.1%) containing pure single cultures (11 Staph. aureus; 4 Staph.epidermidis; 6 Strept agalactia; 5 Strept. dysagalactiae; 6 E.coli and 5 Ps. aerugenosa) and 90 (70.9%) mixed ones. Table (3) revealed that a total of 416 bacterial species were isolated from positive mastitic milk samples and both Staphylococci (142) 34.1% and Streptococci 132 (31.7%) were the most predominant pathogens, followed by E.coli 80(19.2%); Pseudomoneus aerugenosa 36 (8.7%); Actinomyces pyogenes 19 (4.6%) and Klebsilla pneumonae 7(1.7%). For clinical mastitic samples, 175 bacterial species were isolated and Staphylococci 61(34.9%), mainly Coagulase positive Staph. aureus 50 (28.6%) and Streptococci 58(33.1%), mainly Strept. agalactiae 30 (17.1%) and Strept. dysagalactiae 21(12.0%) were the most isolated strains, followed by E.coli 31(17.7%); Ps. aerugenosa 12 (6.9%); A. pyogenes 11(6.3%) and kl. pneumonae 2 (1.1%). Meanwhile, the positive milk samples of subclinical cases yielded 241 bacterial isolates where Staphylococci was the most predominant 81(33.6%), mainly Coagulase positive Staph. aureus 56 (23.2%) followed by Streptococci 74 (30.7%), mainly Strept. agalactiae 37 (15.4%) and Strept. dysagalactiae 27 (11.2%); E.coli 49 (20.3%); Ps. aerugenosa24 (10.0%); A.pyogenes 8 (3.3%) and kl. pneumonae 5 (2.1%).

The results of Coagulase activity of 142 isolated **Staphylococci** (Table, 3) revealed that, 106 isolates were Coagulase positive **Staph. aureus**; 10 Coagulase negative **Staph.aureus** and 26 Coagulase negative **Staph.epidermidis**.

The results of haemolysis and CAMP test for 132 isolated Streptococci proved that, 67 Strept.agalactiae showed large zone of β haemolysis on blood agar and were CAMP positive with large zone of β haemolysis at the junction of tested strains and Staph. aureus strains while, 48 Strept. dysagalactiae and 17 Strept.uberis strains were negative for both haemolysis and CAMP test. Table (4) showed that a total of 80 isolated E.coli strains were serotyped and O111 was the most isolated (18.7%) followed by O86; O44; O119; O126; O128; O125 and O55 (11.3%; 10.0%; 10.0%; 10.0%; 8.8%; 7.5% and 5.0% respectively). Beside 15 strains (18.7%) were untyped. Also, 36 strains of Ps. aerugenosa were serotyped and serogroup K was the most isolated (25.0%) followed by H (19.4%); M (13.9%); A, G (11.1% for each); C (5.6%) and B (2.8%). Beside 4 strains (11.1%) were untyped.

The results of pathogenicity test (Table, 5) showed that the most isolated strains were positive for mouse pathogenicity test where injected mice were either dead or showed lower physical activities and injected strains were reisolated from all mice. Mortality rates vary in between different strains. Coagulase positive Staph. aureus; Strept. agalactiae; A. pyogenes and Ps. aerugenosa H and K were the most virulent strains with mortality rate 100% followed by E.coli O55 and O111 (80% mortality rate) while other strains with 20% -60% mortality rates. The results of antibiotic sensitivity tests of 416 isolated strains (Table,6) revealed that enrofloxacin; linconycine and gentamycin were the most proper antibiotics with the highest in vitro sensitivity against most bacterial isolates. Mean while, most isolated strains were resistant to tetracycline; streptomycin and penicillin

The results of chemical analysis of milk (Table, 7) appeared that, there was a significant decreased in lactose and casein concentration and non significant increase in fat concentration.

Haematological results (Table, 8) showed a significant increase in total leukocyte counts and a significant decrease in both erythrocyte counts and

haemoglobin concentration in mastitic cows when compared with normal ones. The results of serum biochemical analysis (Table, 9) showed a significant increase in AST; ALT; AP; Urea, and creatinine. Meanwhile, there were a significant decrease in calcium; phosphorous and iron in masticic cows when compared with normal control ones.

The results of treatment showed that all cows were clinically normal; milk samples were negative for both California mastitis test and bacteriological examination and the biochemical parameters were ameliorated.

Table (1): Results of clinical examination and California mastitis test.

No. of cows	Clinical	mastitis	Subclinic	al mastitis	Healthy cows		
	No.	9/2	No.	No. %		%	
395	51	12.9	107	27.1	237	60.0	

Table (2): Bacteriological examination of both clinical and subclinical mastitic milk of cows.

Animal status	No. of milk	nilk samples		Positive samples		Prevalence of single and mixed cultures Single Mixed				
	samples	No.	%	No.	%	No.	%	No.	%	
Clinical masticis cows	89	13	14.6	76	85.4	24	31.6	52	68.4	
Subcl nical mastific cows	198	71	35.9	127	64.1	37	29.1	90	70.9	

Table (3): Bacterial species isolated from positive milk samples of both clinical and subclinical mastitic cows.

Facterial isolates	m	inical astitis (76)	m	oclinical astitis (127)	Total (203)		
	No.	%*	No.	%*	No.	%**	
Total Staphylococci:	61	34.9	81	33.6	142	34.1	
Coag. Post. Staph. aureus.	50	28.6	56	23.2	106	25.5	
Coag. Neg. Staph.Aureus	3	1.7	7	2.9	10	2.4	
Coag. Neg. staph.epidermidis	8	4.6	18	7.5	26	6.2	
Total Streptococci:	58	33.1	74	30.7	132	31.7	
Strept, agalactiae	30	17.1	37	15.4	67	16.1	
Strept. dysagalactiae	21	12.0	27	11.2	48	11.5	
Strept. uberis	7	4.0	10	4.1	17	4.1	
Actinomyces pyogenes	11	6.3	8	3.3	19	4.5	
F. coli	31	17.7	49	20.3	80	19.2	
Kelbsiella pneumoniae	2	1.1	5	2.1	7	1.7	
Pesudomoneus aerugenosa	12	6.9	24	10.0	36	8.7	
Total .	175	100.0	241	100.0	416	100.0	

^{*} Percentage in relation to total No. of isolates in each case alone.

^{**} Percentage in relation to total No. of isolates in both mastitic milk

Table (4): Serological identification of *E.coli* and *Ps. aeruginosa* recovered from mastitic cows.

recovered from mastitic cows.									
E. coli			Ps. aeruginosa						
Serogroup	No.	%*	Serogroup	No.	% *				
O44	8	10.0	A	4	11.1				
O55	4	5.0	В	1	2.8				
U86	9	11.3	С	2	5.6				
0111	15	18.7	G	4	11.1				
O119	8	10.0	H	7	19.4				
O125	6	7.5	K	9	25.0				
O126	8	10.0	M	5	13.9				
O128	7	8.8	Untyped	4	11.1				
Untyped	-15	18.7							
Total	80	100.0		36	100.0				

Percentage in relation to total No. of each isolate.

Table (5): Pathogenicity of bacteria isolated from mastitic cow's milk.

Bacteria	No. of Total infected mortality		Bacteria	No. of infected	Total mortality		
	mice	No.	%		mice	No.	%
Total Staphylococci:				E.coli O44	5	2	40.0
Coag. Post. Staph. aureus.	5	5	100.0	O55	5	4	80.0
Coag. Neg.Staph.aureus	5	1	20.0	O86	5	-3	60.0
Coag. Neg. Staph Epidermidis	5	2	40.0	0111	5 .	4	80.0
Total Streptococci:				O119	5	2	40.0
Strept. agalactiae	5	5	100.0	O125	5	2	40.0
Strept. Dysagalactiae	5	. 3	60.0	O126	5	1	20.0
Strept. uberis	5	2	40.0	O128	5	2 ·	40.0
Actino myces pyogenes	5	5	100.0	Ps. aeruginosa A	5	1	20.0
				В	5	2	40.0
	{	}	1	С	5	2	40.0
Klebsilla pneumonae	5	2	40.0	G	5	3	60.0
Meosina piteumonae	3	-	40.0	H	5	5	100.0
			1	K	5	5	100.0
	<u> </u>			M	5	3	60.0

Table (6): In-vitro antibiotic sensitivity test of 416 isolated strains from clinical and subclinical mastitic cow's milk samples.

	An	ox.	Ar	np.	Chlor	omph.	Enr	oflo.	Ge	nta.	Lir	ico.	Pe	nc.	Str	ept.	Te	tra.
В	No	%	No	%	No.	%	No	%	No	%	No	%	No	%	No	%	No	%
Staph.aureus. (106)	85	80.2	81	76.4	65	61.3	90	84.9	87	82.1	91	85.8	58	54.7	44	41.5	32	30.2
Staph. aureus (10)	7	70.0	5	50.0	4	40.0	7	70.0	6	60.0	8	80.0	2	20.0	1	10.0	1	10.0
Staph.epiderm. (26)	14	53.8	12	46.2	9	34.6	15	57.7	13	50.0	15	57.7	8	30.8	4	15.4	3	11.5
Strept. Aglact. (67)	53	79.1	49	73.1	32	47.8	58	86.6	48	71.6	58	8 6.6	29	43.3	12	17.9	8	11.9
Strept.dysag (48)	37	77.1	33	68.8	25	52.1	40	83.3	37	77.1	39	81.3	17	35.4	9	18.8	7	14.6
Strept. uberis (17)	11	64.7	10	58.8	. 7	41.2	13	76.5	9	52.9	12	70.1	7	41.2	3	17.6	3	17.6
A. pyogenes (19)	15	78.9	13	68.4	10	52.6	15	78.9	16	84.2	16	84.2	8	42.1	. 2	26.3	3	15.8
E. coli (80)	49	61.3	50	62.5	56	70.0	71	88.8	73	91.3	65	81.3	22	27.5	28	35.0	27	33.8
Kl. .pneumoniae(7)	2	28.6	3	42.9	4	57.1	5	71.4	6	85.7	4	57.1	1	14.3	2	28.6	2	28.6
Ps. aerugenosa(36)	19	52.8	19	52.8	20	55.6	28	77.8	29	80.6	25	69.4	10	27.8	11	30.6	11	30.6

Table (7): Chemical analysis of milk in mastitic cows (subclinical and clinical) before and after treatment cows.

Subclinical Clinical Groups After LSD Control Mastitic mastitic treatment *P*≤0.05 N=9 cows cows varameters N=26N = 15N=11 28.68 ab 31.22 b 31.10 b 30.46 ab Fat 2.539* gl^{I} ±0.659 ±0.676 ±0.793 ±0.810 26.31 a 23.23 b 24.22 ° 26.52 * Casein 3.081* g[¹ ±0.716 ±0.412 ±0.321 ±0.635 132.70 ° 140.80^b 144.22 b 123.90 * Lactose 16.900* $\sigma \Gamma^{I}$ ±0.980 ±2.416 ± 0.741 ±1.015

Values with different litters within the same raw differed significantly at P≤0.05

Table (8): Haematological studies in mastitic cows (subclinical and clinical) before and after treatment cows.

Group	Control	Subclinical	Clinical mastitic	After	LSD
Parameters	N=9	mastitic cows N=15	cows N=11	treatment N=26	P≤0.05
Tota! leucocytes counts (x10³/ml)	8.322 b ±0.255	10.27 a ±0.180	10.89 * ±0.172	7.77 b ±0.442	1.500
Total erythrocytes counts (x10³/ml)	8.088 b ±0.221	5.430 a ±0.240	6.2 0° a ±0.235	7.86 b ±0.334	2.430
Hb gm/dl	11.691 ±0.344	8.530 * ±0.242	9.23 ^a 0.189	10.923 b ±0.394	2.393

Values with different litters within the same raw differed significantly at P≤0.05

Table (9): Some biochemical changes of serum of mastitic cows (subclinical and

clinical) before and after treatment cows.

Group	Normal	Subclinical	Clinical	After		
Parumeters	Control	Mastitic cows N=15	mastitic cows N=11	treatment N=26	LSD P≤0.05	
.4ST	52.017 a	92.710 ^b	90.320 b-	59.13 a	33.58 *	
u/ml	±5.793	±0.670	±0.760	±3.390	33.36	
ALT	16.362 a	29.400 в	28.56 b	16.230 ª	13.037*	
u/ml	±0.327	±0.573	±0.653	±0.409	13.037	
AP '	12.150 a	15.54 в	16.120 b	11.88 a	3.390 *	
u/ml	±0.414	±0.462	±0.435	±0.476	3.390	
i/rea	15.557 a	20.332 в	19.673 b	15.420 a	4.774 *	
mg/dl	±0.233	±0.536	±0.464	±0.390	4.774	
Creatinine	1.358 a	2.101 b	1.998 b	1.464 a	0.637 *	
rng/dl	±0.183	±0.193	±0.187	±0.132	0.037	
Calcium	9.090 ^b	7.680 a	8218 a*	8.940 b	1.260 *	
mg/100ml	±0.288	±0.298	±0.253	±0.380	1.200	
Phosphorous	6.487 b	· 5.070 a	5.086 a	6.550 в	1.417 *	
mg/100ml	±0.244	±0.250	±0.268	±0.285	1.41/	
Tron	1.593 b	1.096 a	1.087 a	1.483 b	0.387 *	
mg/100ml	±0.070	±0.069	±0.087	±0.094	0.387	

Values with different litters within the same raw differed significantly at P≤0.05

Discussion

Mastitis is one of the most persistent and widely spread disease conditions of importance to milk hygiene and quality among dairy cattle and associated with many zoonotic diseases for human beings (7, 9 and 58).

The recorded results of clinical examination and California mastitis test (Table, 1) revealed that 237(60.0%) were normal cows; 51 (12.9%) clinical mastitis and 107 (27.1%) subclinically mastitic ones. These results are closed to that recorded by (3, 11, 22, 42 and 55).

The results of bacteriological examination of 89 clinically mastitic milk samples and 198 subclinically ones (Table,2) showed that 76 of clinical mastic samples (85.4%) were positive with 31.6% containing pure single cultures and 68.4% mixed ones. Meanwhile, for subclinical mastitic samples, 127 (64.1%) were positive 29.1% containing pure single cultures and 70.9% mixed ones. Similar results were obtained by (1, 11, 16, 28, 55 and 59). A total of 416 bacterial isolates were recovered from positive mastitic milk samples (Table, 3). Staphylococci (34.1%) and Streptococci (31.7) were the most predominant pathogens followed by *E.coli* (19.2%); Ps.aerugenosa (8.7%); A.pyogenes (4.6%) and Kl. pneumonae (1.7%). Nearly similar results were obtained by (16, 18, 22, 26, 28 and 59). For clinical mastitic samples, 175 bacterial species were isolated and Staphylococci was the most predominant pathogen (34.9%) mainly Coagulase positive Staph.aureus (28.6%) followed by Streptococci (33.1%) mainly Strept.agalactiae (17.1%) and Strept. dysagalactiae (12.0%), then E.coli (17.7%); Ps.aerugenosa (6.9%); A.pyogenes (6.3%) and K! pneumonae (1.1%). Similar results were recorded by (6, 11, 18, 22, 38, 47, 55 and 59). Meanwhile, the positive subclinical milk samples yielded 241 bacterial isolates where Staphylococci was the most predominant one (33.6%), mainly Coagulase positive Staph.aureus (23.2%) followed by Streptococci (30.7%), mainly Strept.agalactiae (15.4%) and Strepi. dysagalactiae (11.2%), then E.coli; Ps.aerugenosa; A.pyogenes

and KL pneumonae (20.3%; 10.0%; 3.3% and 2.1% respectively). Similar results were obtained by (1, 11, 22, 28 and 38). As shown in Table (3), 106 out of 142 isolated Staphylococci from both clinical and subclinical masticic milk samples were identified as Coagulase positive Staph.aureus, Coagulase negative Staph.aureus and 26 Coagulase negative Staph.epidermidis. These results came in harmony with (30). The results of haemolysis and CAMP test showed that all isolated Strept.agalactiae strains (67) were positive while, Strept. dysagalactiae (48) and 17, Strept. uberis strains were negative. The same results were obtained by (13, 25 and 48). This due to the co haemolysis by the concerted action of β-toxin from the Staph.aureus strains and CAMP factor (Protein B) from Strept.agalactiae strains (25). The serotyping of 80 isolated E.coli strains (Table, 4) showed that E.coli O111 was the most isolated (18.7%) followed by O86; O44; O119. O126; O128 and O55. These results came in accordance to some extent with that of (4, 19 and 28). Moreover, serotyping of 36 isolated Ps.aerugenosa appeared that serogroup K was the most isolated (25.0%) followed by serogroups H; M; A and B. Similar results obtained by (28). The results of pathogenicity test (Table, 5) showed that most isolated strains were positive and injected mice were either dead or showed lower physical activities and injected strains were re-isolated from all mice. Coagulase positive Staph.aureus; Strept.agalactiae; A.pyogenes and Ps.aerugenosa H and K were the most virulent strains with mortality rate 100% followed by E.coli O55 and O111(80% mortality rate) while other strains with mortality rates 20%-60%. These results coincided to some extent with those of (19 and 28). The results of antibiotic sensitivity tests of 416 bacterial isolates of both clinical and subclinical mastitic milk samples (Table,6) revealed that enrofloxacin, lincomycine and gentamycin were the most proper antibiotics with highest in-vitro efficiency against most isolated strains. Nearly similar results were obtained by (1, 6, 19, 42, 47 and 55).

The results of chemical analysis of milk (Table, 7) showed an insignificant change in fat concentration and significant decrease in casein and lactose concentrations in diseased (subclinical and clinical mastitis) cows when compared with normal control ones. This result came in

agreement with (9, 26, 35 and 36) who cited that the significant decrease of casein and lactose of mastitic milk may be due to activation of plasmin system resulting in proteolysis of casein. Also, enzymatic hydrolysis of casein liberates peptides this peptides reduce the out put of lactose and other osmotic components from the alveoli into the gland lumen (54).

Haematological results revealed that a significant increase in total leucocyte counts and significant decrease in erythrocyte counts and haemoglobin concentration in diseased (subclinical and clinical mastitis) when compared with normal control ones (Table, 8). This results were parallel with (51 and 56). The increase of total leucocyte counts were mainly due to increase the number of neutrophile as a results of inflammation (31) .The significant decrease of erythrocyte counts and haemoglobin concentration may be attributed to the decrease of glutathione peroxidase in the erythrocytes of the mastitic cattle may lead to an increase in the reactive oxygen species levels and decreased antioxidant status resulted in denaturation of Hb and disruption of red cell membrane which result in decreased life span of the erythrocytes and a state of anemia (51). The results of some serum biochemical analysis (Table, 9) showed a significant increase in AST, ALT, AP, urea and creatinine. Similar results were recorded by (2 and 51). The elevation of liver enzymes may be attributed to the inflammatory process and decrease of the immune activity (2). Meanwhile the level of urea and creatinine in mastitis sick cows blood serum were more than healthy ones this may be connect to proteases activation in affected udder and amine nitrogen enter to urea cycle (2). Moreover, there were a significant decrease in calcium, phosphorous and iron in diseased (subclinical and clinical mastitis) cows when compared with normal control ones. This results came in agreement with (9, 15, 26, 37. 41 and 42). This results were probably linked to the reduced secretory activities of the mammary cells and increased the permeability of the mammary epithelium which can lead to transfer of calcium ,phosphorous and iron from blood to milk so, reduced their levels (26). The decrease in serum trace mineral concentration in response to bacterial infection are - thought to act as a nonspecific hot defense mechanism sequestering micro

minerals, which are vital for normal bacterial growth (37). The results of biochemical analysis must be taken in consideration during treatment of masticic cases.

Regarding to trail of treatment, all cows were clinically normal and milk samples were negative for both California mastitis test and bacterial examination and the biochemical parameters were ameliorated. Same results were recorded by (42).

Regarding to the public health importance of mastitic cow's milk, the pathogens isolated from milk in the current study is associated with many diseases for human beings which milk act as a vehicle of infection. Staphylococcus aureus strains have the ability to produce one or more enterotoxins in milk and milk products resulting in food poisoning (7, 18 and 45). Also, it is responsible for endocarditis, osteomylities and septic arthritis (33 and 40). Strept.agalactiae remains one of the most prevalent cause of invasive neonatal infections with sever pneumonia, sepsis and meningitis (12 and 53). Moreover, Gram-negative bacteria in mastitic raw milk when consumed by human result in food poisoning and gasteroentric infection (18 and 58).

Finally, it could be concluded that, mastitis is a serious disease of cows with economic and public health importance at Kaliobia Governorate. Staphylococci, mainly Coagulase positive Staph .aureus; Streptococci, mainly Strept. agalactiae and Strept dysagalactiae, and E.coli are the most common causes of both clinical and subclinical mastitis. Enrofloxacin, Lincomycine and gentamycin are drugs of choice for treatment of mastitis.

Marititis causes alteration in milk components and haematological and bioch-mical parameters in mastitic cows. Mastitis and its adverse effects can be controlled by detection of diseased animal and its isolation for treatment; cleaning of teats and udder before milking and dipping the teats into a sutable disinfection after milking; Milkers must be healthy and their hands must be free from any lesions or wounds; excluding milk from diseased animals and the raw milk must be boiled or pasteurized before human consumption.

References

- 1-Abdel-Khalek, A. and El- Sherbini, M. (2005): Prevalence of contagious pathogen of bovine subclinical mastitis and relationship to bacteria and somatic cell counts.4th Int.Conf.Mansura:1-10.
- 2-Akopyan, A.S.; Haruthyunyan, H.A.; Gasparyan, V.K.; Haruthyunyan, H.D.; Khocanyan, V.V. and Kamalyan, R.G. (2007): The indices of nitrogen metabolism mastitis in cows blood. National Academy of Sciences of RA. Electronic J. Natural Sciences, 2 (9).
- 3-Almaw, G.; Zerihun, A. and Asfaw, Y. (2008): Bovine mastitis and its association with selected risk factors in small holder dairy farms in and around Bahir Dar Ethiopia. Trop Anim. Health Prod., 40(6):427-432.
- 4-Ali, A.R. (1999): Bacteriological and serological studies on *Escherichia coli* in mastitic milk in Gharbia Province .Egypt J.Agric.Res. 77:921-930.
- 5-Alocam, A.S.; Gibson, C.D.; Kirk, J.H.; Kaneene, J.B. and Ahl, J.G. (1989): Cost of mastitis and its prevention in four cattle herds on St Croix, US Virigin Islands. J. Am. Vet. Med. Assoc., 194:1418-1421.
- 6-Aydin, F.; Leli, N.; Saliin, M.; Colak, A. and Otlu, S. (1995): Identification and antibiotic sensitivity of microorganisms causing clinical and subclinical mastitis in dairy cows ir Kars District. Pendik. Vet. Mikrobiyoloji Dergis, 26(1): 55-57.
- 7-CenciGoga, B.T.; Karama, M.; Rossito, P.V.; Morgante, R.A. and Cullor, J.S.
- (2003): Enterotoxin production by Staphylococcus aureus isolated from mastitic cows. J. Food Prot .66(9):1693-1696.
- 8-Christie, R.; Atkins, N.E. and Munch-Petersen, E. (1944): Anote on alytic phenomenon shown by group B Streptococci. Aust. J. Exp. Biol. Med. Sci., 22: 197-200.
- 9-Coulon, J.B.; Gasqui, P.; Barnouin, J.; Othier, A.; Pradel, P. and Pomies, D. (2002): Effect of mastitis and related-germ on milk yield and composition during naturally occurring udder infections in dairy cows. Anim. Res., 51:383-393.
- 10-Darling, C.L. (1975): Standardization and evaluation of the CAMP reaction for the prompt, presumptive identification of *Streptococcus agalactiae* (Lancefield group B) in clinical material. J.Clin. Microbiol., 1:171-174.
- 11-Dego, O.K. and Tareke, F. (2003): Bovine mastitis in selected areas of Southern Ethiopia. Top Anim. Health Prod., 35(3):197-205.
- 12-Dmitriev, A.; Shakleina, E.; Tkacikova, L.; Mikula, I. and Totolian, A.
- (2002): Genetic heterogencity of the pathogenic potentials of human and bovine group B Streptococci. Folia Microbiol., 47:291-295.
- 13-Duarte, R.S.; Miranda, O.P.; Bellei, B.C.; Brito, M.V.P. and Teixeira, L.M. 2004): Phenotypic and molecular characteristics of *Streptococcus agalactiae* isolated from milk of dairy cows in Brazil. J. Clin. Microbiol., 42(9): 4214-4222.
- 14-Edwards, P.R. and Ewing, W.H. (1972): Identification of Enterobacteriaceae. Third Ed. Burgess Publishing Co. Minneapolis.

- 15-El-Khodery, S.A. and Hoedemaker, M. (2005): Incidence and type of mastitis in the livestock of north Germany concerning management factors.4th Int.Conf.Mansoura:973-987.
- 16-El-Marzabani, M.M.; El-Aaser, A.A. and Zakhary, N.I. (1977): Colorimetric method for phosphorous determination. J. Clin .Chem. Biochem., 15:715-718.
- 17-El Zubeir, I.E.; Elowni, O.A.and Mohamed, G.E. (2005): Effect of mastitis on macro-ninerals of bovine milk and blood serum in Sudan. J.S.Afr.Vet.Assoc.2005 Màr; 76(1):22-25.
- 18-Esron, D.K.; Lughano, J.K.; Robinson, H.M.; Angolwisye, M.K.; Calvin, S.
- and Dominic, M.K. (2005): Studies on mastitis, milk quality and health risks associated with consumption of milk from post oral herds in Dodoma and Morgoro region, Tanzania J. Vet. Sci., 6(3):213-221.
- 19-Essa, M.F. (2007): Some studies on bacterial causes of mastitis in buffaloes. J. Egypt Vet .Med. Assoc., 67(3):129-139.
- 20-Fawcett, J.K. and Scott, J.E. (1960): Colorimetric determination of urea. J. Clin. Path., 48:313-156.
- 21-Folin, O.Z. (1934): Estimation of creatining by the Jaffe reaction cited in creatinine kits of Bio Adwic. Pys. Chem., 268,228.
- 22-Gianneechini, R.; Concha, C.; Rivero, R.; Delucci, L. and Moreno Lopenz, J. (2002): Occurrance of clinical and suclinical mastitis in dairy herds in the West Littoral region in Uruguay Acta Vet. Scand., 43(4):221-230.
- 23-Gindler, E.M. and King, J.D. (1972): Rapid colorimetric method for determination of calcium in biological fluids with methyline blue. Am. J.Clin. Path., 48:367.
- 24-Hansen, A.K. (2000): Hand book of Laboratory Animal Bacteriology. Boca Raton, London, New York.
- 25-Hansen, S.M. and Sorensen, U.B.S. (2003): Methods for quantitative detection and presumptive identification of group B Streptococci on primary plating. J. Clin. Microbiol., 41(4):1399-1403.
- 26-Henry, O.; Anakola, S. and Jackin, N. (2007): Effect of mastitis on raw milk compositional quality .J. Vet. Sci, 8(3):237-242.
- 27-Homma, J.Y. (1982): Designation of thirteen o- group antigens of *Pseudomonas aeruginosa*. An amendment for the tentative proposal in 1976. Japan. J. Exp. Med., 52: 317-319.
- 28-Ibrahim, H.S. (2007): Bacteriological study on bovine mastitis in Assuit Governorate with special reference to virulence of some isolates. J. Egypt Vet. Med. Assoc., 67(2): 57-74.
- 29-Jain, N.C. (1986): Shalm's Veterinary Haematology. 4th Ed. PP. 55-69, Lee and Fibiger Philadelphia, U.S.A
- 30-Jarb, J. (1991): Classification of coagulase negative Staphylococci isolated from bovine clinical and subclinical mastitis. Vet. Microbiol, 27:515-518.
- 31-Klinkon, M., and Zadnik, T. (1999): Dynamics of red and white blood picture in dairy cows during the periparturient period. Comparative Haematology International 9:156-161.

- 32-Koneman, E.W.; Allen, S.D.; DAnda, M.W.; Sohrechenberger, P.C. and Winn, W.C. (1997): Colour Atlas and Text book of Diagnostic Microbiology. Fifth Edition, Lippincott, Philadelphia, New York.
- 33-Korakaki, E.; Aligizakis, A.; Manoura, A.; Hatzidaki, E.; Saitakis, E.;
- Anatoi iotaki, M.; Velivasakis, E.; Maraki, S. and Giannakopoulou, C. (2007): Methicillin resistant *Staphylococcus aureus* osteomylitis and septic arthritis: diagnosis management Jpn. J. Infect. Dis., 60(2-3): 129-131.
- 34-Korhonen, H. and Kartinen, L. (1995): Changes in the composition of milk induced by mastitis, in: Sandholm M., Honkanen-Buzalski T., Karrtinen L., Pyorala S. (Eds). The bovine udder and mastitis. Gummerus, Jyvaskyla, Finland, pp. 76-82.
- 35-Leitner, G.; Krifucks, O.; Merin; U. Lavi, Y. and Silanikove, N. (2006): Interactions betwee 1 bacteria type, proteolysis of casein and physico-chemical properties of bovine milk. International Dairy J., 16:648-654.
- 36-Leitner, G.; Chaffer, M.; Shamay, A.; Shapiro, F.; Merin, U.andEzra, E. (2004): Changes in milk composition as affected by subclinical mastitis in sheep. J. Dairy Sci., 87:46-32.
- 37-Lohuis, J.A.C.M.; Schukken, J.H.M.; Verhijden, A. Brandand Van Miert, A.S.J.P.A.M. (1990): Effect of severity of systemic sings during the acute phase of experimentally induced *Echerichia coli* mastitis on milk production losses J.Dairy Sci.73:333-341.
- 38-Matinowski, E.; Lassa, H.; Kllossowska, A.; Smulski, S.; Markiewicz, H. and Kaczmarowski, M. (2006): Etiological agents of dairy cows mastitis in Western part of Poland Pol. J. Vet .Sci., 9(3):191-194.
- 39-Marshall, R.T. (Ed). (1992): Standard methods for the examination of dairy products (16thed). Washington, DC, USA: American Public Health Association.
- 40-Martinez, O.A.; Gracia, S.P.; Pueo, E. and Chopo, J.M. (2006): Septic arthritis as initial manifestation of bacterial endocarditis caused by *Staphylococcus aureus*. An. Med. Intern., 23(4): 184-186.
- 41-Middleton, J.R.; Luby, C.D.; Viera, L.Tyler; J.W. and Casteel, S. (2004): Influence of *Staphylococcus aureus* intramammary infection on serum copper, zinc, and iron concentrations. J. Dairy Sci., 87:976-979.
- 42-Mobarak, M.G.; El-Bealawey, M.A.; Maarouf, A.A. and Abd El-Raof, Y.M. (2007): Correlation between cattle mastitis and serum iron, zinic and capper. Proceeding of the 2nd .Sci.Conf.Fac.Vet.Med.Benha Univ.:334-346.
- 43-Osheim, D.L. and Ross, P.F. (1985): Atomic absorption spectrophotometric determination of liver capper. J.AOAC., 68: 44-45.
- 44-Pearson, D. (1981): Chemical analysis of milk. 8th Ed., Churchill Livingstone, Edhinburgh, London.
- 45-Peles, F.; Wagner, M.; Varga, L.; Hein, I.; Rieck, P.; Gutser, K.; Kéres Zturi, P.; Kardos, G.; Turcsany, I.; Beri, B. and Szabo, A. (2007): Characterization of

- Staphylococcus aureus strains isolated from bovine milk in Hungary.Int.J.Food Microbial.,118(2):186-193.
- 46-Quinn, P.J.; Markyl, B.K.; Carter, M.E.; Donnelly, W.J. and Leonard, F.C. (2002): Veterinary Microbiology and Microbial Disease. Blackwell, U.K.
- 47-Rahman, H. and Boro, B. R. (1990): Isolation and antibiogram of bacterial pathogens producing mastitis. Ind. J. Animal Health, 1:49-52.
- 48-Ranner, H.B.; Weeks, L.S. and Stratton, C.W. (1986): Evaluation of Spot CAMP test for identification of group B Streptococci. J. Clin. Microiol., 24 (2): 296-297.
- 49-Rei man, S. and Frankel, S. (1957): Colorimetric determination of GOT and GPT activities. Am.J.Clin.Path. 28-56.
- 50-Ruegg, P.L. and Reinemann, D.J. (2002): Milk quality and mastitis tests J.Bov. Pract. 36:41-54.
- 51-Sayed, A.S. (2003): Studies on some biochemical and hematological changes in cattle mastitis. Assiut Vet.Med.J.Vol.49 No. 98.
- 52-Schalm, O.W. and Noordlander, D.O. (1957): Experiments and observations leading to development of California mastitis test. J.Am. Vet. Med. Ass., 130:199-204.
- 53-Schuchat, A. (2000): Neonatal group B Streptococcal disease: Screening and prevention. N. Engl. J. Med., 343:209-210.
- 54-Silanikove, N.; Shamay, A.; Sinder, D. and Moran, A. (2000): Stress down regulates milk yield in cows by plasmin induced β-casein product that blocks K⁺ channels on apical membranes. Life Science, 67:2201-2212.
- 55-Singh, K.B.; Baxi, K.K. and Joshi, D.V. (1988): Studies on incidence, diagnosis and etiolog of clinical mastitis. J. Res. Punjab, Agr. Univ., 25(2):287-293.
- 56-Smith, G.W.; Constable, P.D.; Mcrin, D.E. (2001): Ability of haematologic and serum piochemical variable to differentiate Gram-negative and Gram-positive mastitis in dairy cows. J. Vet. Intern. Med. 15(4): 394-400.
- 57-SPSS for Windows, Version: 11 (19 September, 2001). Copyright SPSS Inc. 1989 2001. All rights reserved.
- 58-Straley, B.A.; Donaldson, S.C.; Hedge, N.V.; Sawant, A.; Srinivasan, V.; Olivr, S.P. and Jayarao, B.M. (2006): Public health significance of anti microbial resistant gramnegative bacteria in raw bulk tank milk. FoodBorne Pathog. Dis., 3(3): 222-233.
- 59-Tenhagen, B.A.; Koster, G.; Wallmann, J. and Heuwieser, W.(2006): Prevalence of mastitis pathogens and their resistance against antimicrobial agents in dairy cows in Brandenburg, Germanay. J. Dairy Sci., 89(7):2542-2551.

دراسات على إلتهاب الضرع في الأبقار بمحافظة القليوبية والأهمية الصحية لألبانها

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الملخص العربي

تم الفحص الإكلينيكي لعدد ٣٩٥ بقرة حلاب بمحافظة القليوبية وأخذت عينات من البانها حيث تم فحصها ظاهريا ويواسطة إختبار كاليفورنيا. أظهرت النتائج أن ١٢,٩ % من الأبقار مصابة بالتهاب الضرع السريري و ٢٧,١% بالتهاب الضرع تحت السريريّ . أوضح الفحص البكتريولوجي ل٨٩ عينة لبن من أبقار مصابة بالتهاب الضرع السريري أن ٨٥،٤% كانت موجبة وتم عزل الميكروبات بصورة منفردة ٣١,٦ % أو مختلطة ٦٨,٤ % . ويم عزل ١٧٥ عترة وكان أعلى نسبة للعزل هي المكورات العنقودية (٩٤,٩%) وبخاصة المكور العنقودي الذهبي الموجب للتجلط (٢٨,٦%) والمكورات السبحية (٣٣٨١) وبخاصة المكور السبحي أجالاكتيا (٧٠٠١%) بينما أظهر الفحص البكتريولوجي ل ١٢٧ عينة لبن من أبقار مصابة بالتهاب الضرع السريري أن ١ ﴿ ١٤. % كانت موجبة و ٢٩٠١ % منه: تحتوى على الميكروبات منفردة و ٩٠, ٥٠% مختلطة وتم عزل ٢٤١ عترة وكانت المكورات العنقودية (٣٣,٦%) وبخاصة المكور العنقودي الذهبي الموجب للتجلط (٢٣.٢%) والمكورات السبحية (٣٠.٧%) ويخاصةُ المكور السبحي أجالكتيا (٤,٥١ %) والديس أجالكتيا (١١,١ %) وكذلك الميكروب القولوني (٣٠٠٣) هم الأكثر عزلاً بينما بقية الميكروبات تم عزلها بنسب بسيطة. اظهرت إختبارت التجمع للمكورات العنقونية المعزولة (١٤٢) أن ١٠٦ منهم كانت المكورات العنقودي الذهبي الموجب للتجلط و١٠ المكور العنقودي الذهبي السالب للتجلط و ٢٦ المكور العنقودي البشري السالب التجلط. أظهر إختبار كامب للمكورات السبحية المعزولة (١٣٢) أن ٦٧ مكور سبحي أجالاكتيا كانت موجبة بينما ٤٨ مكورسبحي ديس أجالكتيا و ١٧ مكور سبحي ابريس كانت سالبة. التصنيف السيرولوجي للميكروب القولوني المعزول (٠٠) أظهر وجود ٨ مجموعات سيرولوجية والمجموعة 0111 الأعلى تواجدا (١٨.٧). والتصنيف السير ولوجي لميكروبات سيدوموناس ايروجينوزا المعزولة (٣٦) اظهر وجود ٧ مجموعات سيرولوجية والمجموعة k هي الأعلى (٥,٠). أظهرت اختبارت الضراءة في الفنران أن جميع العترات المعزولة كانت مميتة او ممرضة للقنران. أظهر التحليل الكيمياتي لعينات اللبن وجود تقص معنوي في نسب اللاكتوز والكازبين .كما أظهرت نتاتج الفحص البيوكيمياني وصورة الدم وجود ارتفاع معنوى في عدد كرات الدم البيضاء وإنزيمات الكبد والبوريا والكرياتينين. بينما لوحظ انخفاض معنوى لعدد كرات الدم الحمراء ونسبة الهيمه جلوبين والكالسيوم والفسفور والحديد في الحيوانات المريضة بالمقارنة مع الحيوانات السليمة. وقد تم العلاج بحقن الاتروفلوكساسين عضليا مع اللينكومايسين موضوعيا (الأكثر كفاءة) نتج عنة شفاء تام للأبقار المصابة بالتهاب الضرع . وأخيرا تم مناقشة طرق السيطرة على التهابات الضرّع في الأبقار والأهمية الصحية للميكروبات المعزولة من ألبان هذه الأيقار بالنسبة للانسار