Acaricidal and Immunological Studies on Fowl Tick Argas persicus Infecting Commercial Balady Chickens Flock

Ramadan, M. Y.

Dep. Parasitol., Fac. Vet. Med. Moshtohor. Benha Univ.

The present work aimed to study the acaricidal effect of deltamethrin and ivermectin as well as immunization of chicken by whole crude Argas persicus larvae extract against A.persicus infestation in commercial balady chicken flock. Two experiments were carried out. The first experiment; 120 commercial balady chicken flock of 140 days old naturally infested by A. persicus belonged to private poultry farm in Meet- Kenena village Kalubyia Governorate were classified into 3 equal groups. Group! was adminstrated ivermectin orally at a dose of 100 mg/kg body weight. Group II was dipped in 0.05% deltamethrin. Group III was kept without treatment as a control group. Birds were kept for 4weeks and examined weekly at night for counting differe it stages of the infected ticks. In the second experiment; twenty of 120 days old naïve chickens were classified into two equal groups. The first group (group A) was was injected subcutaneously with initial dose of whole crude A. persicus larvae extract followed by booster dose after 12 days later. The second group (group B) was injected subcut ineously with physiological saline as a control group. After 7 days from the booster dose, all chicken groups were challenged by natural infection by A. persicus infection through keeping them in the same infected poultry house for 28 days. The total number of A. persicus larvae on each chicken was estimated weekly for 4 weeks. The present study showed that oral administration of ivermectin caused 100% reduction in the number of A. persicus burden in chicken in the 1st week post dosing while, dipping of chicken in deltamethrin 0.05% caused prolonged complete (100%) reduction in the number of ticks till the end of the 3rd week post dipping. Immunized chickens showed 85.6% reduction in the mean number of infective A. persicus larvae and the moulting percentage of larvae was significantly reduced (26.7%) compared to (80%) in non immunized control group. It was concluded that dipping birds in 0.05% deltan ethrin or immunization by using double doses of crude extract are better than oral administration of ivermectin

Key words: Argas persicus, Ivermectin, Deltamethrin, Immunization

Introduction

irgus persicus is an important ectoparasite of domestic and wild birds in the world. It causes sever irritation to poultry which has an adverse effect on bir is quality and cause heavy economic losses in poultry industry (21 and 24). Heavy infestation causes loss of blood leading to anemia and eventually

death (4). A. persicus also is an efficient reservoir and /or vector for many virses rickettsia, spirochaete and mycoplasma (17). As pullorum (27), Mycoplasma gallisepticum and Mycoplasm meleagrides (25), Salmonella gallinurum (9), Aegyptinella pullorum (29), avian encephalomyelitis and leucocytozoonosis (20), West nile virus (1) and Borrelia anserine (6). A. persicus harbours different types of bacteria including those of genus Salmonella, Aerobacer, Escherichia, Proteûs, Staphylococcus, Flavobacterium, Pseudomonas and Streptococcus (5). A. persicus infestation causes paralysis to heavily infested chickens (26). A. persicus larvae have been responsible for simultaneous occurrence of Infectious Bursal Disease and Spirochaetosis (2).

The control of A. persicus using chemical insecticides poses health risks and causes environmental problems (18). Additionally, the use of chemicals alone leads to rapid development of resistance (11 and 24)). So the present study aimed to evaluate the acaricidal effect of two new acaricides; deltaraethrin and ivermectin against fowl tick as well as immunization of the chicken by repeated injections of laboratory prepared crude A. persicus antigen.

Material and Methods

Acaricide

- 1-Butox (Deltamethrin) (1R 3R) (2, 3 dipromoving) 2-2- diethyl-cyclopropane carboxylate 5 * Cyano-3 phenoxy- benzyl. It is produced by Hoeckst Roussel Vet.
- 2- Ivermectin is a member of the ivermectin group of macrocyclic lactones.

Preparation of whole crude A. persicus larvae extract according to (31).

One hundred and fifty of semi- engorged A. persicus larvae were collected from chicken, washed with physiological saline and used for preparation of crude extract. The washed ticks were disrupted and homogenized by using automatic tissue homogenizer in phosphate buffer saline (PBS) PH 7.2 at 4C then filtered by filter paper. The filtrate was centrifuged at 4000/min. in cooling centrifuge, the supernatant was collected. The protein concentration of the extract were estimated and stored at -20C till use according to (7).

Experiment I: Effect of Deltamethrin and ivermectin on tick burden of treated chicken.

One hundred and twenty commercial balady chickens of 140 days old naturally infested by A. persicus (some of them showed tick paralysis; Plate I, B) belonged to private poultry farm in Meet- Kenena village Kalubyia Governorate were classified into 3 equal groups each consists of 40 hen and treated as follow:

Group I was given ivermectin orally at a dose of 100 mg / kg body weight according to (16).

GroupII: were dipped once in Butox (Deltamethrin) 0.05% according to (19).

Group III. Kept without treatment as a control group. All bird groups were left in the same infected poultry house for 4 weeks post treatment and fedon balanced rations and clean source of water.

Birds were examined weekly at night for detection and counting of different stages of A.persicus starting from the day of treatment (0 day). Total number of ticks on each bird were estimated by counting the different stages of A.persicus ticks attached to the left side and the result was multiplied by two according to (14).

Experiment II: Effect of immunization of chicken by crude A. persicus larvae extract on tick burden:

Twenty of 120 days old naïve chickens obtained from clean private poultry farm and free from Apersicus infection were classified into two equal groups (group A and Group B) and treated as follow: group A were injected subcutaneously with initial dose of whole crude Apersicus larvae extract. Booster dose was injected after 12 days later. Group B were injected subcutaneously with physiological saline and used as control group. After 7 days from the booster dose, all chicken groups were challenged by A. persicus infection through keeping them in the same infected poultry house for 28 days according to (10). The total number of A. persicus larvae on each chicken was estimated every 7 days by counting the larvae attached to the left side and the result was multiplied by two according to (14). Thirty engorged larvae (felling off from chickens and wandering on feathers) were collected from immunized and control chicken groups at the end of the 3rd

week Larvae were incubated at 27C and 95% R.H for 12 days according to (12) to evaluate the effect of immunization upon its ability to moult.

Results

Results in table (1) showed that oral administration of ivermectin caused reduction in the number of A. persicus burden in chicken in the 1st week post dosing but tick burden increased again-at the 2nd, 3rd and 4th week post injection Dipping of chickens in Butox 0.05% caused prolonged and complete reduction in the number of ticks(starting from the first week till the end of the 3rd week post dipping.

Table (2) showed that ivermectin caused comparatively lower reduction in the mean number of larvae, nymphs and adults (62.89, 58.92, and 61.84%, respectively) compared to Butox which achieved high reduction (95.06, 98.30, and 97.34%) in the mean number of larvae, nymphs and adults respectively. The acaricidal efficacy of ivermectin against larvae was 100, 92.59, 36.01 and 10.08 during the 1st, 2nd, 3rd and 4th week respectively compared to 100, 100, 100 and 87.51% reduction caused by deltamethrin during the same weeks. The mean number of A. persicus nymphs moderately decreased (the reduction % was 100,100 29.18 and 0.00 % during the 1st, 2nd, 3rd and 4th week, respectively) under the effect of ivermectin administration but dipping in Butox 0.05% caused prolonged and high effect (100,100, 100 and 92.89 % during the same period, respectively). Simillarly, the acaricidal efficacy of ivermectin against adult A. persicus lasted for the first two weeks only post-treatment while Butox dipping lasted 4 weeks

Results in table (3) indicated that subcutaneous injection of two doses of whole crude A.persicus larvae extract resulted in 85.6% total reduction in the mean number of larvae. Tick burden in immunized chickens in the first week of challenge showed 95.09% reduction compared to non immunized control group (PlateI; C, D and E). Tick burden increased gradually again from the 2nd week till the end of the 4th week post injection. The moulting percentage of larvae collecting from immunized chickens was significantly reduced (26.7%) compared to (80%) in non immunized control group (Table, 4).

Table (1): Efficacy of ivermectin and Deltamethrin on the mean number of A. persicus burden in chicken for 28 days.

Days Grouop	Mean number Before	1 st week	Red.	2 nd week	Red.	3 rd week %	Red.	4 th week	Red.	Total	Red.
Ivermectin	Treatment. 276.65	0.00	100	14.52	94.71	189.78	31.40	267.59	3.27	471.9	61.3
Deltamethrin	281.42	0.00	100	0.00	100	0.00	100	49.73	71.67	49.7	95.9,
Control	284.37	290.9	0.00	282.1	0.00	288.23	0.00	351.33	0.00	1215.6	0.00

Red.% = Reduction%

Reduction % = Number of ticks before treatment – Number of ticks after treatment x 100

Number of ticks before treatment

Table (2): Effect of ivermeetin oral dosing and butex 0.05% immersion on different developmental stages of tick A. persicus infecting chicken for 28 days

of them 72. personal infecting enterior for 20 days											
week	Before	1stweek	Red.	2 nd wee	k Red.	3 rd week	Red.	4 th week	Red.	Total	Total
	Treatment		%		%		%		%	ticks	Red.%
Group										number	
GI											
Larvae	196.00	0.00	100	14.52	92.59	125.43	36.01	176.23	10.08	316.18	62.89
Nymphs	65.00	0.00	100	0.00	100	46.03	29.18	66.19	0.00	112.22	58.92
Adults	21.65	0.00	100	0.00	100	18.32	15.38	25.17	0.00	43.49	61.84
·											
GII											
Larvae	195.76	0.00	100	0.00	100	0.00	100	42.07	78.51	42.07	95.06
Nymphs	65.13	0.00	100	0.00	100	0.00	100	4.63	92.89	4.63	98.30
Adults	20.53	0.00	100	0.00	100	0.00	100	3.03	85.24	3.03	97.34
GIII											
Larvae	188.71	190.83	0.0	182.2 (0.00	193.15	0.00	199.47	0.00	852.22	0.00
Nymphs	67.35	63.49	0.0	70.8 (0.00	70.03	0.00	73.88	0.00	273.23	0.00
Adults	28.16	36.58	0.0	25.1 (0.00	25.05	0.00	28.25	0.00	113.98	0.00

GI: ivermectin treated GII: immersed in 0.005 deltamethrin solution GIII: control group

Red.

% = Reduction%

Reduction % = Number of ticks before treatment –number of ticks after treatment _x 100 Number of ticks before treatment

Table (3) Efficacy of whole crude A.persicus larvae extract on the mean number of larvae infesting chickens for 28 days

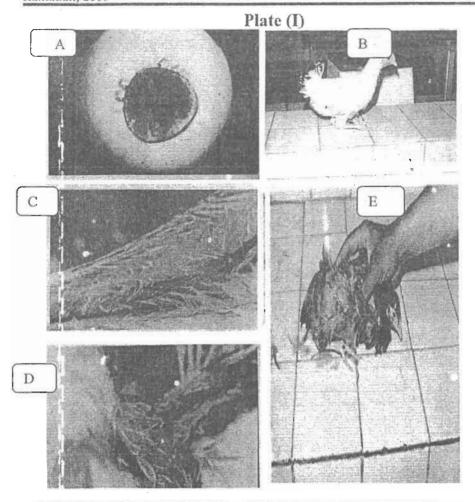
1st w. Red. Red. 3rdw. Red. 4th w. Red Total Red. % % % Weeks % groups GroupA 15.54 95.09 28.43 90.74 39.67 87.3 97.55 69.36 181.2 Immunized 316.72 0.00 307.21 0.00 312.60 0.00 318.41 0.00 1254.9 GroupB 0.0 control

Reduction % = Number of ticks in control group—Number of ticks immunized group x 100

Number of ticks in control group

Table (4): Efficacy of whole crude A.persicus larvae extract on moulting percentage of collected larvae from immunized birds

Stages of ticks Group	No. of collected larvae	Number. of moulted nymphs	% of moulting
GroupA Immunized	30	8	26.7
Group B control	30	. 24	80



A = Argus persicus larva B= Chichen showed tick paralysis
C & D= Immunized chichen showed ligh infestation
E= Non immunized chichen showed heavy infestation

Third Inter. Sci. Conf., 29 Jan. - 1 Feb./ 2009, Benha & Ras Sudr, Egypt Fac. V.t. Med. (Moshtohor), Benha Univ

Discussion

Most studies delt with control of ticks infesting birds were carried out in Vitro and few were carried out in vivo (28). In Egypt, control measures against were carried out traditionally by veterinarians depending their experience which lead to development of resistant strains of ticks to traditionally applied organophosphorous acaricides environmental problems. In the present study two synthetic acaricides of low mammalian toxicity; ivermectin and deltamethrin were applied separetely to chicken flocks of 140 days old naturally infected infected by A. persicus. Concerning the efficacy of oral administration of ivermectin against fowl tick, the present data indicated that it caused 100% reduction in the mean number of ticks infesting chicken during the first week post administration and 94.71% reduction at the end of the second week while during the 4th weeks its acaricidal efficacy was reduced to 3.27%. These results agreed with (16) who mentioned that population of engorged A. persicus showed 80, 80, 100, 90 and 75% paralysis in after 3, 15, 24, 48 and 72 hrs respectively when fed on chickens fed a single oral dose of 100 microgram ivermectin per kg body weight. Also (13) recorded that ivermectin had an acaricidal effect against A. persicus and achieved 82.96% for its control. Ivermectin also have an adverse effect on A.persicus by extensive alteration in its testes and prevent the development of new spermatid and break down of the cell membrane and cytoplasmic organelles of the spermatozoa so it can reduce the number of ticks in the long run (15). Deltamethrin is a synthetic chemical based on Pyrethrin with greater stability and low mammalian toxicity (22) and acts on the insect central nervous system to give a rapid knockdown but it is not used yet in destruction of tick A. persicus in commercial chicken flocks (30). The present study dealt with the acaricidal efficacy of deltamethrin dipping against A. persicus infection and showed that, it was more effective and had prolonged acaricidal effect compared to ivermectin as it achieved 100% reduction in A.persicus burden from the first week to the end of the third week post dipping. This result was agreed with (28) who reported in bioassays against A. persicus that deltamethrin was 106 times more effective than chlorvos and dipping of chicken legs in 0.05% deltariethrin resulted in death of 80,90, 100% of adult ticks nymphs and

larvae respectively within one day and its acaricidal effect lasted for 5 days. The more prolonged acaricidal effect of deltamethrin in the present study may attribute to change in the methods of application of the acaricide as (28) dipped the legs of chickensanly. The prolonged activity of deltamethrin was explained by (30) who noted that the microencapsulation of the pyrethroids prolonged their activity to weeks with greater stability as the microcapsules adhere to the insect skeleton and the pyrethroids absorbed through the chitin to produce its toxic effect.

The present study indicated that subcutaneous injection of two doses of whole crude A.persicus larvae extract resulted in total reduction (85.6%) in the mean number of A. persicus larvae. The evaluation of whole crude extract prepared from A. persicus larvae was carried out by challengeed the immunized chickens to natural infection by housing it in infested poultry house Such challenge permits the immune system of immunized chickens to react with the infested larvae resulting in its destruction. All previous studies dealt with immunization of chicken against A. persicus depend on using salivary gland extract or experimentally repeated infestation by adult ticks (23, 10, 7 and 8). Larvae burden infesting immunized chickens showed 95.09% reduction in the first week of challenge compared to non immunized control group. Our result was higher than that of (7) who recorded repeated infestation of spring fowl with A. persicus females stimulated 35.9% resistance to tick feeding compared to 20.4% in control. Such high reduction percentages may be attributed to the change of in immunization and challenge programs. The present data showed that, the premoult period in larvae collected from immunized chicken was prolonged compared to control group. In this respect, (12) mentioned that A. persicus larvae falling off after 5 days attachment, followed by 8 days before moulting to the first nymph. The recorded higher larval mortalities, prolonged premoult period and lowered moulting percentage in our results were explained by (3) who attributed such mortalities to gut damage and the digest cells were sloughed off leaving the basal lamina. Such damage allowed host leukocytes to enter the haemocael and attack tick muscles, malpighian tubules and blocking synthesis of vitellagenius by the gut cells.

Conclusion

It was concluded that:

- 1- Dipping in deltamethrin 0.05% were effective than oral dosing of 100 ug/kg.b.v/ ivermectin in control of A.persicus in naturally infested chickens.
- 2- Whole crude extract prepared from A. persicus larvae was effective in reduction of A. persicus larvae that infect immunized chickens and consequently reduce the number of developed nymphs and adult ticks in infested poultry house.

Further immunological studies are necessary to study the effect of other safe acaricides, prepare and evaluate a polyvalent vaccine prepared from larvae, nymphs and adults on all stages of *Argas persicus* to minimize the use of harmful acaricides

Aknowldgment: The author is grateful to Dr. Ashraf abdel-Hakeem El-Kouniy Professor of Veterinary Pharmacology, Fac. Vet. Medecine Moshiohor Benha University for his guidance in determination of the there utic doses and selection of the most suitable route of administration.

References

- 1. At basy, M. M.; Osman, M. and Marzouk, A. S. (1993): West nile virus (Flaviviridae: Flavirus). In: Experimentally infected *Argus* ticks (Acari: Argasidae). Amer. J. Trop. Med. Hyg., 48: 726-737.
- 2. Abdu, P.A. (1987): Infectious bursal disease in pullet chickens. Avian Dis., 31: 204-205.
- 3. Agbede, S.I.R. and Kemp, H.D. (1986): Immunization of cattle against *Boophilus microp:lus* using extracts derived from adult female ticks. Histopathology of ticks, feeding on vaccinated cattle. Int. J. Parasitology, 16 (1): 35-41.
- 4. Bergstrom, S.; Haemig, P. D. and Olsen, B. (1999): Increased mortality of black-browed albatross chicks at a colony heavily infested with the tick *ixòdes uriae*. Int. J. Parasitol., 29:1359-1361.
- 5. Buriro, S.N. (1983): Relative abundance of different species of bacteria isolated from Argus persicus. Pakistan Vet .J. 3: 126-128
- 6. Durden, L.A.; Oliver, J. H. Jr. and Kinsey, A. A. (2001): Ticks (Acari: Ixodidae) and spirochetes recovered from birds on a Gorgia Barrier island. J. Med. Entomol., 38: 231-236.
- 7. El Kammah, K. M. and Abdel Wahab, K.S.E. (2003): Studies on the immune defence of chickens against *Argus persicus* (Oken) 1818 and cattle against *Boophilus annulaius Say 1821* (Acari: Ixodoidea). Insect Science and its application, 23: 1, 75-83.
- 8. El Kammah, K. M., Oyoun .L. M. and Gabr, H. S. M. (2002): Studies on the feeding effects of *Argas persicus* and *A. hermanni (Acari: Argasidae)* on chicken and pigeon blood and plasma protein. Int. J. Acarology, 28: 3, 273-276.
- 9. Gyurov, B. (1983): Role of Argus persicus in Epidemiology of fowl typhoid. Veterinarna Sbirka, 81: 22-24.
- 10. He beeb, S.M.; Sayed, M.A. and El-Kammah, K.M. (2001): Studies on chicken acquired resistance to *Argus (Persicargus)*. persicus (Acari: Argasidae) due to repeated infestation. J. Egypt. Soc. Parasitol., 31: 467-477.
- 11. Hassanain, M.A.; El-Garhy, M.F.; Abdul-Ghaffar, F.A.; El-Sharaby, A. and Abdul Magee I, K.N. (1997): Biological control studies of soft and hard ticks in Egypt. I. The effect of *Bacillus thuringiensis* varieties on soft and hard ticks .Parasitolo. Res., 83: 209-113.
- 12. Kettle, D.S. (1995): Medical and Veterinary Entomology. 2nd Ed. CAB International, Wallingford Oxon Ox 10 8De UK.pp 453.
- 13. Khan, L.A.; Kham, M.N.; Iqbal, Z. and Qudoos, A. (2001): Comparative acaricidal efficacy of cypermethrin, ivermectin, trichlorphon and Azadirachta indica (neem) in layers naturally infested with Argus persicus. Pakistan J. Agric. Sci., 38:3/4, 29-31.
- 14. Kinsey, A.A.; Durden L.A. and Oliver, Jr. (2000): Tick infestation of birds in coastal Georgia and Albama. J. Parasitol., 86: 251-254.
- 15. Montasser, A.A., Gadelhak, G.G. and Tariq, S. (2005): Impact of ivermectin on the ultrastucure of the testes of *Argus (Pericargus) persicus (Ixodoidea: Argasidae)*. Experimental and applied Acarology . 36: 1/2: 119-129.

- 16. Mousa, S. M., Gad, A., Soliman. A., Sokkar, I. and Abdel-Raheem, M. (1988): Investigation on the efficacy of ivermectin for ectoparasites and nomatodes in chickens iii Effect of oral administration of ivermectin on different stages of *Argas*. *persicus* Assuit Vet Med. J 19; 190-194
- 17. Nemova, N.V., Alekeev, A.N., Kostiukov, M.A., Gordeeva, Z.E. and Kuima, A.U. (1990): An experimental study of the reproduction of the Hissar Bunyaviridae in *Argus persicus* ticks. Med. Parazitol. 1:35-36.
- 18. Oprescu, I. (2003): In vitro assay of *Argas persicus* susceptibility to doramectin (Dectornax Pfizer) and neocidol 600 EC (Ciba Geigy). Revista Scientia Parasitologica. 4: 1/1, 175-179.
- 19. Osman, I.M. (1997): The efficacy of deltamethrin and propoxur against the fowl tick *Argus versicus (Ixoddoidea: Argasidae)*. Sudan J. of vet. Science and animal husbandry. 63: 1/2. 102-108.
- 20. 20 Permin, A. and Hansen, J.W. (1998): Epidemiology, diagnosis and control of poultry parasites FAO, Rome, pp4-56.
- 21. Phulan, M,S., Bhatti, M.W. and Buriro, S.N. (1984): Incidence of Argus. persicus in poultry. Pakistan Vet. J. 4: 174-175
- 22. R::madan, M. R. (2004): Preliminary studies on caprine lice and its control in Kalubyia Governorate. Zag. Vet. J. (ISSN. 1110-1458) Vol. 32 (1): 110-119.
- 23. Sa (ena, V. K., Sharma, D.K., Singh, H. and Srivastava, S. C. (1996): Acquired resistance against *Argas persicus* and its association complement level in guinea fowl. J. of Vet. Parasitol. 10 (2): 191-195.
- 24. Shah, A.H.; Khan, M.N.; Iqbal, Z. and Sajid, M. S. (2004): Tick infestation in poultry Int. J. of Agriculure and biology 6: 1162-1165.
- 25. So iman, A.M., Mousa, S.A., Gad, N.; Desouky, U. and Sokkar I.M. (1988): Rodents and ticks as a reservoir of mycoplasma in poultry farms. Assuit Vet. Med. J 9: 184-190.
- 26. Sc ii , J.L. ((1979): An outbreak of of tick paralysis in white leghorn chicks due to Argus persicus. Indian Vet. J 56: 149-152.
- 27. St. fanov, V.; Matev, I. and Balimezov, I. (1975): Role of the tick of species Argus persicus in the eptizootology of nullorum disease in birds. Vet. Med. Nauki. 20: 41-46.
- 28. Tain, Q.Y. and Guan, Y. (1989: Sensitivity of Argus persicus to 4 acaricides and the use of deltamethrin dips to kill A. persicus on chicken. Chinese J.Vet. Med. 15: 24-25.
- 29. Ts.mov, T.S. (1983): Epizootiology of Aegyptianellosis in poultry. Vet. Med. Nauki, 20: 41-16.
- 30. Wall, R. and Shearer, D. (1997): In "Veterinary entomology" First edition T.J. International Ltd.
- 31. Wikel, K. S. (1981): The induction of host resistance to tick infestation with salivary gland. Immunology 30: 311-314.

دراسات على بعض مبيدات الاكروسات والمناعة ضد قراد الدجاج (ارجس بيرسكس) الذي يصيب قطعان الدجاج البلدي التجارية

محمد يوسف رمضان قسم الطفيليات كلية الطب البيطري بمستهر جامعة بنها الملخص العربي

أجريت هذه الدراسة بهدف تقييم كل من الإيفرمكتين والبيوتوكس كمبيدي أكروسات وكذك تقييم المناعة المكتسبة المستحدثة في الدجاج ضد قراد الدجاج. ولهذا الغرض تم اجراء تجريتين:

- التجربة الأولى وفيها تم اختيار عدد ١٢٠ دجاجة عمر ١٤٠ يوم من قطيع واحد مصاب بالقراد الناعم تم تقسيمهم إلى ثلاث مجموعات متساوية؛ المجموعة الأولى: تم علاجها بمادة الافرمكتين عن طريق الفم بمعدل ١٠٠ مج /كجم وزن حي مرة واحدة .

المجموعة الثانية: تم تغطيسها في محلول ٠٠٠٠ % من مادة البيوتوكس مرة واحدة . المجموعة الثالثة تركت بدون علاج كمجموعة ضابطة .

- التجربة الثانية وفيها تم إنتاج وتقييم لقاح مستخلص من يرقات قراد أرجس برسكس وقد تم اختيار ٢٠ دجاجة عمر ١٢٠ يوم لم يسبق اصابتها بالقراد الناعم وتم تقسيمهم إلي مجموعتين ؟ المجموعة الأولى حقنت بمستخلص اليرقات تحت الجلد جرعة اولى ثم اعيد حقنها مرة أانية بعد ١٢ يوم . المجموعة الثانية تركت كمجموعة ضابطة. وتم تعرض المجموعتين للعدوي الطبيعية بالقراد وذلك بتربيتهما في حظيرة مصابة بالقراد الناعم و تم عد يرقات القراد بعد كل اسبوع ولمدة اربع اسابيع .

وجدت الدراسة أن العلاج بالايفرمكتين أدي إلي انخفاض عدد القراد بنسبة ١٠٠ % في الاسبوع الاول فقط أما تغطيس الدجاج في محلول ٠٠.% من مادة البيوتوكس فقد أدي انخفاض بنسبة ١٠٠ % في عدد القراد من الاسبوع الاول إلي نهاية الاسبوع الثالث. ما وجدت الدراسة أن حقن مستخلص يرقات القراد الخام تحت الجلد في الدجاج أدي إلي انخفاض عدد يرقات الآراد التي اصابت الدجاج المحصن بنسبة ٢٠,٦ % . كما أدي استخدام هذا المستخلص إلي انخفاض نسبة انسلاخ هذه اليرقات إلى ٢٧,٦ % مقارنة بنسبة ٨٠% في الدجاج غير المحصن .