Trial For Preparation And Evaluation Of Inactivated Multicomponent Vaccine Containing BVD, IBR, PI-3 Viruses, Salmonella typhimurium And Salmonella Dublin For Calves

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ABSTRACT

An inactivaated multicomponent vaccine adjuvanted with aluminum hydroxide gel against Bovine Viral Diarrhea (BVD), Infectious Bovine Rhinotracheits (IPR), and Para influenza-3 (PI-3) viruses with *S. typhimurium* and *S. dublin* bacteria was prepared. The vaccine was pure and safe. Potency evaluation was applied on 12 adult apparently healthy calves divided into 4 groups 3 animals for each. The immune responses were monitored using serum neutralization, agglutination and mouse protection tests. The results revealed a lack of interaction and antagonistic effect among the component of the prepared vaccine. The humeral immune responses gave antibody titers over the permissible limit of protective level and persist for 24 weeks post vaccination (period of study).

INTRODUCTION

Pneumo-enteritis diseases complex syndrome are of paramount economic important because of high loses and deaths rate, so respiratory and enteric disorders are often cited as the significant cause of economic losses probably more costly than all other combined diseases (1).

Pneumo-enteritis diseases complex's syndrome is considered as multifactorial diseases syndrome caused by a number of infectious agents either viral and/or bacterial. Bovine viral diarrhea (BVD), Infectious bovine rhinotracheits (IBR) and parainflunza-3 (PI-3) viruses have been all incriminated in the etiology of bovine viral pnumo-enteritis complex syndrome among cattle and buffalo causing severe economic losses (2-4).

Bovine salmonellosis is of worldwide public health concern and is an economically important disease. Although cattle of all ages can be infected with salmonella, serious infections and deaths are most often seen in calves up to 10 weeks of age (5,6). The most common serotypes involved are salmonella typhimurium and salmonella dublin (7-10).

Numerous attempts have been made to protect calves by immunization (11-13).

Concurrent infection with salmonella and BVDV would be expected to cause more serious disease than infection with either agent alone. Therefore the two agents should be considered during disease outbreak (14-16).

Vaccination has proved to be by far, the most effective and cost effective method for controlling of infectious diseases. Vaccination is the undersigned, support immunization as the safest, most effective way to control infectious diseases. The two or more separate immunogens in a single product and administrated through the same way, have many potential advantages in the form of decreased of injecting risks, decreased of clinical visits and easier introduction of vaccine. So to facilitate the vaccination program, the multicomponent vaccines are being economic by reducing costs of storage transport and administration (17).

This investigation was designed to prepare and asses the new locally inactivated multicomponent vaccine named (Pneumo-Sal) containing pneumo-enteric viral and bacterial etiological agents (BVD, IBR, PI-3, S. typhimurium and S. dublin) for using in calves against respiratory and enteric diseases.

MATERIAL AND METHODS

1. Laboratory animals

1.1. Mice

A total of 100 Swiss albino mice (18-20 grams weight) were used in this work. Eighty mice were used for protection test in case of inactivated combined bacterial vaccine while twenty mice were used for safety test in locally prepared multicoponent (Pneumo-Sal) vaccine.

1.2. Guinea pigs

Ten adult male guinea pigs (300-400 grams weight) were classified into 2 groups (5 for each) and used in the safety test of the locally prepared multicoponent (Pneumo-Sal) vaccine.

1.3. Calves

A total of 18 cross breed apparently healthy male calves (6-9 months' old ages) were used for this study. Calves were housed in an isolation facility at Veterinary Serum and Vaccine Research Institute. All calves were tested to confirm that they are free from antibodies against BVD, IBR, PI-3, S. typhimurium and S. dublin prior to vaccination. Six calves were used in safety test while the other 12 were used for potency evaluation of the prepared vaccine.

All experimental animals used in this work were obtained from Veterinary Serum and Vaccine Research Institute (VSVRI) Abbassia, Cairo.

2. Vaccines

2.1.Preparation of inactivated combined viral vaccine (Pnemue-3)

The vaccine was prepared from 3 reference Egyptian viral strains. BVD (Iman strain) with a titer of 7 log 10 TCID₅₀ / ml. which was isolated and identified (18). IBR (Abo-Hammad strain) with a titer of 8 log 10 TCID₅₀ / ml. which was isolated and identified (19). While PI-3 with a titer of 8 log 10 TCID₅₀ /ml (Strain 45) which was isolated and identified (20). The 3 viruses (BVD, IBR and

PI-3) were initially adapted and propagated in Madin Darby Bovine Kidney (MDBK) cell culture. One ml of each virus was inoculated in roller bottles containing healthy and confluent sheet of MBDK cell culture using MEM as maintenance media. All roller bottles inoculated virus were kept in an incubator at 37 °C and examined daily for follow up the specific CPE related to each virus. After complete CPE showing 70 % of whole sheet of cell culture the harvesting was done by rapid freezing and thawing. The process of inactivation was made by adding 0.01 M of Binary Ethylene amine and mixed together by stirring at 37 °C for optimum times according to the described techniques (21-23). The stoppage action of BEI was performed by adding of Sodium thiosulphate 20 % with the final concentration of 2 %.

2.2.Preparation of inactivated combined bacterial vaccine

The vaccine was prepared from 2 local bacterial field isolates (S. typhimurium and S. Dublin). The bacterial isolates serologically identified by somatic flagellar standard antisera (24). Both bacterial strains (S. typhimurium and S. dublin) were grown in nutrient broth enriched with casein and yeast extract then incubated at 37 °C. The cultures were shaken at 120 CPM and harvested during the logarithmic growth phase. Washed saline growth suspension standardized optically to contain 2 mg dry weight of cells / ml (25). Cold ethanol was added (26). Equal volume of both inactivated culture suspensions were mixed well.

2.3.Preparation of inactivated multicomponent vaccine (Pneumo-Sal)

The inactivated viral and bacterial suspensions in equal volume were mixed together. Then Aluminum hydroxide gel 30 % (Al-hydrogel solution) was added as an adjuvant and mixed well by using of magnetic stirrer for obtaining of homogenized solution. Thiomersal was added as vaccine preservative (0.01%) and PH was adjusted to 7.8. The final vaccine product was distributed in sterile glass bottles of 100 ml capacity capsulated and labeled.

3.Evaluation of the locally prepared inactivated multicomponent vaccine (Pneumo-Sal)

3.1. Sterility test

It was performed according to American protocol (27). It was done to be proved that the vaccine is free from bacteria, mycoplasma, and fungi.

3.2.Safety test

3.2.1. Safety test in mice

A group of 20 Swiss albino mice were classified into 2 groups (10 for each). The first group were inoculated I/P with a dose of 0.2 ml of the prepared multicombonent vaccine (Pnumo-Sal.). The second group was used as control and inoculated I/P with 0.2 ml of physiological saline. Mice were kept under observation for 2 weeks to detect undesired post vaccinal side effects.

3.2.2. Safety test in guinea pigs

Ten adult male guinea pigs were used in this experiment allocated into 2 groups. The first group consists of 5 guinea pigs that inoculated I/P with 0.5 ml of the prepared vaccine (Pneumo-Sal) while the second group was left as control and inoculated with 0.5 ml of physiological saline. All animals were kept under observation for 2 weeks post inoculation.

3.2.3. Safety test in calves

It was achieved using six adult male calves. Three calves of them were inoculated I/M with ten times of the vaccinal dose (Pnumo-Sal.) 75 ml of the prepared vaccine were distributed to be inoculated in different places of animal body. The sofety test was applied (27). The other 3 calves were inoculated with the same dose (75 ml) at the same time with physiological saline solution and kept as a non-vaccinated control. All animals were kept under observation for 2 weeks post inoculation for detection of any abnormalities, also daily rectal temperature has been recorded.

3.3. Potency test

Potency evaluation of the prepared multicomponent inactivated vaccine (Pnumo-

Sal) was carried out (27). Twelve adult male calves were used in this study and divided into four groups, 3 animals for each (Table 1).

4.Blood sampling

Blood samples were collected from each calve of all groups at different intervals. Before vaccination then 2 weeks post 1st vaccination and every 2 weeks after booster vaccination till 24 weeks later.

5. Serological investigations

5.1.Serum neutralization test (SNT)

SNT was applied in microtiter plate for detection of viral antibody titers following the procedures described (28,29). SNT antibody titers were expressed as log 10 TCID₅₀ of the reciprocal serum dilutions following the calculation formula (30) for viral antibody evaluation.

5.2. Serum agglutination test (SAT)

The test was used for detection of antibody titers against S. typhimurium and S. dublin (31).

5.3. Mouse protection test

The test was used for evaluation of both combined bacterial and multicomponent (Pnumo-Sal.) vaccines through challenge test. Eighty mice were used for this experiment and divided into 8 groups; ten mice were used for each group. All groups exposed to challenge with both virulent S. typhimurium (2656 V) and S. Dublin (2560 V) 2 weeks post vaccination following the reported technique (25). Dead and survived mice in every group were registered post challenge over a period of 14 days. The deaths of challenged mice were used as criteria for lack of immunity and the percentage of protection was calculated according to the formula:

= Survivors in vaccinated group - Survivors in the control group

Х	100	

10

Calves	No.	Vaccination protocol									
group	of calves	Туре	First second dose dose		Route	Interval period					
Group 1	3	Inactivated combined viral vaccine (Pneumo-3)	5 ml	5 ml							
Group 2	3	Inactivated combined bacterial vaccine	2.5 ml	2.5 ml	1						
Group 3	3	Inactivated multicomponent vaccine (Pneumo-Sal.)	7.5ml	7.5ml	I/M	2 weeks					
Group 4	3	Control non-vaccinated calves									

Table 1. Experimental Design for potency evaluation of pnumo - sal inactivated vaccine

- Inactivated combined viral vaccine (Pneumo-3) = Containing BVD, IPR and PI-3 viruses
- Inactivated combined bacterial vaccine = Containing S. Typhymurium & S. dublin.
- Inactivated multicomponent vaccine (Pneumo-Sal.) = Containing both viral and bacterial components.

RESULTS AND DISCUSSION

Salmonella infections in cattle are of world wide public health concern, therefore numerous attempts had been made to protect cattle (5,32). Perhaps the most frightening aspect of salmonellosis is the rapid strain resistance to drugs and despite the application of vaccine and availability of therapeutic agents; bovine salmonellosis remained a serious and wide spread problem (33). S. typhimurium and S. dublin appears to be the commonest serovars isolated from cattle, hence the great demand for preparation of vaccine which may have a greatest part to play in the control of disease outbreaks (34).

Acute BVDV infection of cattle was closely associated with temporary suppression of the immune function which predispose to endogenous bacteraemia (35). BVD infection can aggravate and possibly predispose to salmonella infection, this situation could assume greater significance in calves stressed by colostrum deprivation and also stressed by movement through markets, and hence the etiological interaction of the two should be considered during outbreaks of salmonella in cattle. Also the super infection or combined infection of salmonella and BVDV in calves is more severe than salmonella infection alone (14). In addition the severe disease was

observed in pregnant dairy heifers that had natural BVDV and S. typhimurium DT 104 infection (15).

So the use of vaccine will continue to be a basic tool used to reduce the risk of infection of viral disease alone (BVD, IBR, PI-3 viruses) or accompanied with bacterial infection (S. typhimurium or S. dublin). It will be necessary to produce of a broad spectrum of protection against a range of viral and bacterial agent (36) and high level of specific antibodies prior to infection may reduce the risk of disease (37).

The evaluation of the locally prepared inactivated multicomponent vaccine (Pnumo-Sal) for purity, safety and potency showed that the vaccine was free from any aerobic and anaerobic bacteria as well as mycoplasma and fungal contamination through inoculation on the specific media for 15 days. Belonging to the safety test of vaccine in mice, guinea pigs and calves, it was recorded to be safe as there were no post vaccinal reaction, no abnormalities or deaths during the period of observation (14 days). Also the vaccine was completely safe for calves as there were no local or systemic reaction has been observed during the period of the experiment (38).

Regarding to the potency evaluation of the prepared vaccine (Pneumo-Sal) through

immunization of adult calves with 2 immunizing doses at 2 weeks interval,: the immunogenicity of the inactivated vaccine was rapidly diminished when used a single dose, while 2 doses virtually eliminate this phenomena (39). The fact that the vaccine in an adjuvant was capable of inducing an immune response that revealed the active infection (26), as aluminium hydroxide gel exerts its effect by protracted release from the site of injection (40)

The results obtained in Tables 2, 3 indicated that each component of vaccinal strains either viral (BVD, IBR and PI-3 viruses) or bacterial (S. typhimurium and S. dublin) used in the preparation of inactivated multicomponent vaccine (Pnumo-Sal) proved that there was no antagonistic effect between each component of the prepared vaccine. Such

data was similar to that obtained in several studies (41,42).

The results of serum neutralization antibody titers illustrated in Table 2 and figures 1A, B, and C proved that calves vaccinated with Pnumo-Sal (group 3) have antibody titers nearly similar to that titers observed in calves vaccinated with Pnumo-3 (group 1). The level of the recorded antibody titers were effectively sufficient to combat such active infection which remained within minimum acceptable titer of protective level till 6 months post vaccination. BVD antibody level of 1: 8 dilutions (Log 0.9) was protective (43). Also it has been recorded that the minimum acceptable titers was 1: 4 dilutions or 0.6 Log was protective against PI-3 and IBR viruses (44,45).

Table 2. Mean of serum neutralizing antibody titers in calves vaccinated with inactivated multicomponent (Pneumo-Sal) and inactivated combined (Pneumo-3) vaccines.

		Me	an sei	rum n	eutra	lizing	antil	ody	titers	expre	ssed i	n log	10	
Type 0f Vaccine	Viral					•	week	s po	st-vac	cinat	ion			
	0f	antigens	* 0 day	**	4	6	8	10	12	14	16	18	20	22
	BVD	0.20	0.30	1.20	1.30	1.80	1.85	1.87	1.77	1.60	1.60	1.55	1.35	1.20
Pneumo- Sal Vaccine	IBR	0.25	0.40	1.40	1.45	1.90	2.00	2.00	1.95	1.75	1.65	1.55	1.30	1.25
	PI-3	0.26	0.30	1.20	1.35	1.86	1.90	1.93	1.90	1.70	1.57	1.50	1.35	1.20
Pneumo-3 Vaccine	BVD	0.25	0.30	1.45	1.55	1.85	1.95	1.87	1.75	1.70	1.70	1.60	1.40	1.30
	IBR	0.30	0.40	1.45	1.50	1.95	2.1	1.95	1.95	1.80	1.66	1.55	1.35	1.30
	PI-3	0.25	0.40	1.35	1.35	1.80	1.95	1.90	1.90	1.75	1.65	1.55	1.40	1.25
Control Non vaccinated	BVD	0.20	0.20	0.20	0.30	0.20	0.20	0.20	0.25	0.20	0.20	0.25	0.20	0.20
	IBR	0.20	0.20	0.25	0.20	0.30	0.20	0.26	0.30	0.30	0.30	0.20	0.30	0.20
	PI-3	0.50	0.30	0.30	0.30	0.20	0.20	0.20	0.20	0.20	0.20	0.30	0.20	0.20

⁻ Protective serum neutralizing (SN) antibody for BVD is 0.90 (43).

⁻ Protective serum neutralizing (SN) antibody for IBR is 0.60 (45).

⁻ Protective serum neutralizing (SN) antibody for PI-3 is 0.60 (44).

First vaccination

^{**} Second vaccination

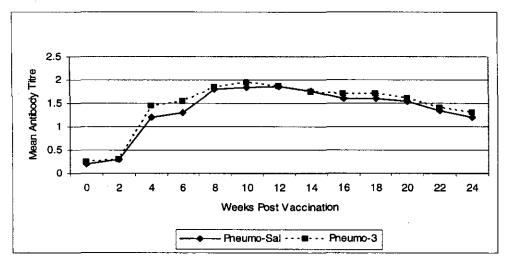


Fig. 1A. Mean serum neutralizing antibody titers against BVD in both pneumo-sal and pneumo-3 vacinated calves

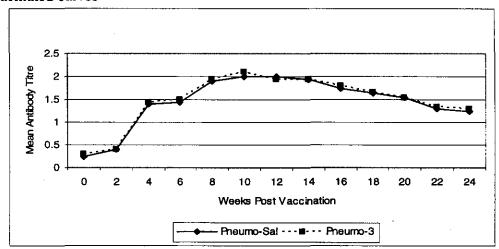


Fig. 1B. Mean serum neutralizing antibody titers against IBR in both pneumo-sal and pneumo-3 vacinated calves

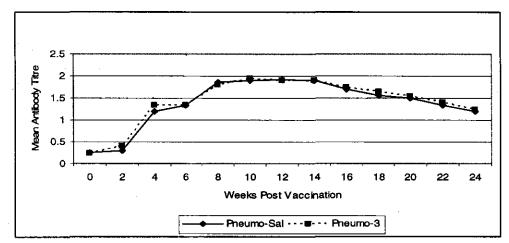


Fig. 1C. Mean serum neutralizing antibody titers against PI-3 in both pneumo-sal and pneumo-3 vacinated calves

The results of serum agglutination test presented in Table 3 and figures 2A and B revealed that there were higher agglutinating antibodies titers in calves vaccinated with Pneumo-Sal or inactivated combined bacterial

vaccine in comparison to the control non vaccinated group (39). Moreover the antibody titers were slightly similar in both Pneumo-Sal and combined bacterial vaccinated groups (25, 40).

Table 3. Mean of serum agglutinating antibody titers against S. typhimurum and S. dublin in calves vaccinated with inactivated multicomponent (Pneumo-Sal) and inactivated combined bacterial vaccines.

Tyme			Mea	an ser	um ag	glutin	ating a	antibo	dy tite	ers (ge	ometr	ic mea	nn)	
Type Of	Bacterial	*	* weeks post-vaccination											
Vaccine	antigens	0 day	**	4	6	8	10	12	14	16	18	20	22	24
Inactivated multicomponent	S. Typhimurium	4.6	160	320	640	1015	1015	1280	1280	1015	640	160	80	80
(Pneumo-Sal) vaccine	S. Dublin	4.6	201	640	1015	1280	1280	1015	1015	640	640	320	160	160
Inactivated Combined	S. Typhimurium	4.6	320	640	1015	1280	1280	1280	1280	1015	640	320	160	80
bacterial vaccine	S. Dublin	4.6	320	1015	1280	1280	1280	1280	1015	1015	640	320	160	160
Control non	S. Typhimurium S.	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0	10.0
vaccinated	Dublin	4.6	4.6	4.6	4.6	4.6	4.6	4.6	4.6	4.6	4.6	4.6	4.6	4.6

First vaccination

> 80 considered positive titer for salmonella (13).

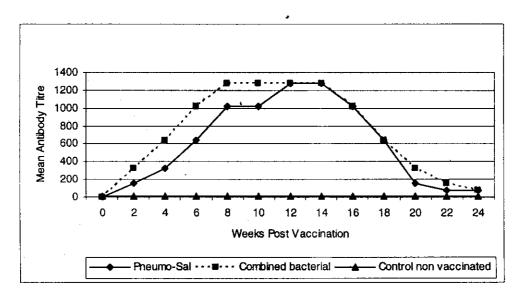


Fig. 2A. Mean serum agglutinanting antibody titers against S. typhimurium in both pneumo-sal and pneumo-3 vacinated calves

^{**} Second vaccination

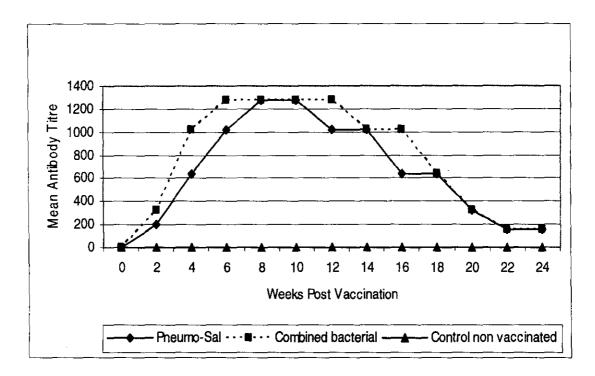


Fig. 2B. Mean serum agglutinanting antibody titers against S. dublin in both pneumo-sal and pneumo-3 vacinated calves

The results depicted in Table 4 showed that the protection percentage in mice vaccinated with inactivated combined bacterial vaccine and challenged with *S. typhimurum and S. Dublin* were 80 % and 70 % while it was 70 % in case of mice vaccinated with Pnumo-Sal vaccine respectively.

The humeral antibody response induced by Pnumo-3 (group 1), combined salmonella (group 2) and Pnumo-Sal (group 3) vaccines, revealed that there is no interference or competition observed between each fraction consisting the prepared vaccine. The combined vaccine of BVD, S. typhimurium

and S. dublin gives adequate protection against such component and recommended for field use (25).

From the previous results it could be concluded that, the locally prepared inactivated multicomponent (Pnumo-Sal) vaccine containing BVD, IBR, PI-3, S. typhimurium and S. dublin has been proved to be pure, safe and potent in calves against respiratory and enteric illness inducing immune response lasting for 24 weeks against such viral or bacterial causative agents and it is recommended practically to be used effectively in the field application.

Table 4. Results of challenge test for S. typhimurum and S. dublin in mice vaccinated either with inactivated combined bacterial or multicomponent (Pneumo-Sal) vaccines

Days	Inactiv	vated combi	ned bacterial		Inactivated multicomponent (Pneumo-Sal) vaccine					
post Challenge	Mice challenged with S. Mic			enged ublin	Mice chall with S. typh	enged	Mice challenged with S. dublin			
	No. of survived mice N		No. of surviv	No. of survived mice		ved mice	No. of survived mice			
	Vaccinated	Vaccinated Control		Control	Vaccinated	Control	vaccinated	Control		
	group	group	group	group	group	group	group	group		
1	10/10	10/10	10/10	9/10	10/10	10/10	10/10	9/10		
2	10/10	9/10	10/10	9/10	10/10	9/10	10/10	9/10		
3 _	10/10	9/10	9/10	8/10	10/10	9/10	10/10	8/10		
4	9/10	7/10	9/10 8/10		10/10	7/10	10/10	8/10		
5	9/10	7/10	9/10	8/10	10/10	7/10	9/10	8/10		
6	9/10	7/10	8/10 7/10		10/10	7/10	9/10	7/10		
7	9/10	5/10	8/10	7/10	10/10	5/10	9/10	7/10		
8	9/10	5/10	8/10	5/10	9/10	5/10	9/10	5/10		
9	9/10	5/10	8/10	2/10	9/10	5/10	9/10	2/10		
10	9/10	2/10	7/10	2/10	9/10	2/10	8/10	2/10		
11	9/10	2/10	7/10	2/10	9/10	2/10	8/10	2/10		
12	8/10	1/10	7/10	1/10	9/10	1/10	8/10	1/10		
13	8/10	0/10	7/10	0/10	8/10	0/10	8/10	0/10		
14	8/10	0/10	7/10	0/10	7/10	0/10_	7/10	0/10		
Protection										
%	80 %	0 %	70 %	0 %	70 %	0 %	70 %	0 %		

REFERENCES

- 1. Jensen R, Grinner L A, Chow T L and Brown W W (1978): IBR infected Lot cattle 1. Pathology and symptoms, Proc. M. S., Livestock, Saniti-Assoc., 59: 189-199.
- 2.Durham P J K and Hassard, L E (1990):
 Prevalence of antibodies to IBR, PI-3,
 BRSV and BVDV in cattle in
 Sakatchewas and Alberta, Canada.
 Canadian Vet. J. 31: 815-820.
- 3. Houe H (1999): Epidemiology features and economical importance of bovine viral diarrhea virus (BVDV) infections. Vet. Microbiology, 64: 89-107.
- 4.Aly N M, Shehab C G and Abdel-Rahim I H A (2003): Bovine viral diarrhea, Bovine Herpes virus and Para Influnza-3 virus infections in three cattle herds in Egypt. Rev. Sci. Off. Epiz., 2 (3): 879-892.

- 5.Smith B P, Habasha F G, Reina-Guerra M and Hardy A J (1980): Immunization of calves against salmonellosis. Am. J. Vet. Res. 41: 1947-1951.
- 6.Hoiseth S K and Stocker B A D (1981):

 Aromatic-dependent Salmonella typhimurium are non-virulent and effective as live vaccines. Nature (London) 291: 238.
- 7.Peterson K J and Coon R E (1978): Salmonella typhimurium infection in dairy cows. J. Am. Vet. Assoc. 151: 344-350.
- 8. Hughes L E, Gibson E A and Roberts H E (1971): Bovine salmonellosis in England and Wales. Br. Vet. J. 127: 225-237.
- 9.Moor G R, Rothenbacher H, Bennett M V and Barner R D (1982): Bovine salmonellosis. J. Am. Vet. Med. Assoc. 141: 841-844.

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10.Anonymous (1992): Salmonella in animal and poultry production. MAFF, CVL, Weybridge, England.

- 11.Bailey, M. H. (1978): Immunization of calves against salmonellosis. J. Am. Vet. Med. Assoc. 173: 610-613.
- 12.Aitken M M, Jones P W and Brown G T H (1982): Protection of cattle against experimentally induced salmonellosis by intraderml injection of heat-killed Salmonella dublin. Res. Vet. Sci. 32: 368-373.
- 13.Barrow P A and Wallis T S (2000): Vaccination against salmonella infection in food animals: rational, theoretical basis and practical application, 323-339.
- 14. Wray C and Roeder P L (1987): Effect of BVD mucosal disease infection on salmonella infection in calves. Res. Vet. Sci., 21 (2): 184-189.
- 15.Penny CD, Low J C, Nettleton P F, Scott P R Sargisor N D, Strachan W D and Honeyman P C (1996): Concurrent bovine viral diarrhea virus and S. typhimurium D T 104 infection in a group of pregnant dairy heifers. Vet. Rec., 18: 485-489.
- 16. Wells S J, Ott S L and Seitzinger A H (1998): Symposium: emerging health, issues. J. Dairy Sci., 81: 3029-3035.
- 17. Parkmann P O (1995): Combined and simultaneously administrated vaccines. (a brief history in combined vaccines and simultaneously administration, current issues and perspective). "Eds. Williams, J. C.; Goldenthal, K. L; Burns, D. L. and Lewis, B. P". The New York Academy of Science, New York pp. 1-9.
- 18.Baz Thanaa I (1976): Isolation, characterization and serological studies on BVD-Md virus in Egypt. Ph D thesis, Fac. Vet. Med. Cairo University.
- 19. Hafez S M, Thanaa I B, Mohsen A Y and Monera H (1976): Infectious bovine Rhinotracheits in Egypt, isolation and

- serological identification of the virus. J. Egypt. Vet. Med. Assoc. 36 (1): 129-139.
- 20.Singh K V and Baz Thanaa 1 (1966): Isolation of Para Influnza-3 virus from water buffaloes in Egypt. Nature Land 210 (50): 616-652.
- 21.El-Sabbagh M M, Samira S T, Ghally H M and Saad M S (1995): Binary Ethylene imine as an inactivant for Infectious Bovine Rhinotracheitis (IBR), Bovine Viral Diarrhea (BVD) and Para Influenza-3 (PI-3) viruses and its application for vaccine production. Beni-Suef Vet. Med. Res. 5 (2): 29-57.
- 22.Zeidan S M, Samira S T, El-Sabbagh M M and El-Kholly A A (2005): Trials for preparation of combined inactivated respiratory virus's vaccine containing BVD genotypes I and II, IBR, PI-3 and BRSV viruses. 4th Inter. Sci. Conf. Mansoura University, 433-446.
- 23. Samira S T, El-Sabbagh M M and Ghally H M (2001): Preparation of combined inactivated BVD, IBR, PI-3 and respiratory Syncytia virus (BRSV). Egypt. Vet. Med. Assoc. 61 (4): 251-263.
- 24.Mackie T J and McCartney J E (1989):
 Practical Medical Microbiology.
 Thirteenth Edition, volume 2, Edinburgh
 London Melbourne and New York.
- 25.Hannan I M (2007): Immunological studies on salmonella and bovine viral diarrhea vaccine in cattle. Ph. D. Thesis, Beni-Suef University, Faculty of Vet. Medicine.
- 26.Collins FM (1972): Effect of adjuvant on immunogenicity of a heat killed salmonella vaccine. J. Infect. Dis., 126 (1): 69-76.
- 27. USA code of Federal Regulation (1987):
 Animal products No. 9 part 1 to 199
 published by the office of Federal
 Register National Archives and Records
 Administration.
- 28.Rossi C R and Keiesel G K (1971): Microtitre tests for detecting antibodies in

- bovine serum to Para Influenza-3 virus, Infectious Bovine Rhinotracheitis virus and Bovine viral diarrhea virus. Microbiology, 22: 32-36.
- 29.Dannacher G and Martel J L (1978):
 Titration of antibody to bovine viral diarrhea virus by micro-method of serum neutralization. Reuceil de Medicine Veterinire, 154 (1): 31-37.
- 30.Reed L J and Munech H (1938): A simple method for estimating fifty percent end point. Amer. J. Hygiene, 27: 493-497.
- 31.Barrow, P. A. (2000): Serological diagnosis of salmonella by ELIZA and other tests. Salmonella in domestic animals, 407-427.
- 32. Wray C, Morris J A and Frinely W J (1977): The immunization of mice and calves with gel E. mutant of S. typhimurium. J. Hyg. Comb. 79: 17-24.
- 33.Sojka W J and Wray C (1978): Experimental S. typhimurium infection in calves. Res. Vet. Sc., 25 (2): 139-143.
- 34. Wray C and Davies R H (2000): Salmonella infection in cattle. Salmonella Domestic Animals 169-196.
- 35.Reggiardo C and Koeberle M L (1981):
 Detection of bacteraemia in cattle inoculated with bovine viral diarrhea virus. Am. J. Vet. Res., 42 (2): 218.
- 36. Vanoirschot J T (1999): Bovine viral vaccine, diagnostic and eradication in veterinary vaccines and diagnostics. Sehaltz, R. O. ed. Academic press, London, U K: 197-213.
- 37.Brysan D (1996): Infectious bovine respiratory disease emerging issues and progress towards control BCVA. Edinburgh, pp. 1-8.
- 38.Code of American Federal Regulation (1987): Animal products No. 9 part 1:

- 199. Published by the office of Federal Register National Achieves and Record Administration.
- 39. Cameron C M and Fuls W J (1976): Immunization of calves against S. dublin with attenuated live and inactivated vaccine Ondersteport, J. Vet. Res., 43 (2): 38.
- 40.Morein B, Eriksson MV, Sjolander A and Bengtsson K L (1996): Novel adjuvant and vaccine delivery systems. Vet. Immunol-Immunopath. 54: 373-384.
- 41. Khristov S, Karadzov I, Ignatov G and Khristova V (1976): Immunogenic properties of polyvalent inactivated vaccine prepared from respiratory viruses of cattle. Veterinarmonn, Editisimisk 13 (4): 36-44.
- 42.Fulton RW, Confer A M, Burge L T, Perino L J and Offay J M (1995): Antibody responses by cattle after vaccination with commercial viral vaccine containing BHV-1, BVDR, PI-3 and BRSV, immunogens and subsequent revaccination at day 140. Vaccine, 13: 725-733.
- 43.Bittle J L (1968): Vaccination for bovine viral diarrhea-mucosal disease. J. Am. Vet. Med. Assoc., 152 (6): 861-865.
- 44. Mihaylovic B, Cvetovic A, Kazmanovic M, Vuhobrat D, Asuin R and Lazurevic T (1979): Comparison of vaccine against Respiratory viral disease of cattle and Infectious bovine Rhinotracheitis (IBR) and Para Influenza-3 (PI-3) viruses. Veterinarski, Glasrik, 33 (1): 33-39.
- 45.Zuffa A and Fekeotova N (1980):
 Protective action of inactivated adjuvanted IBR vaccine against experimental infection. Veterinaria, Med. 25 (1): 51-61.

الملخص العربي

محاولة تحضير و معايرة لقاح جامع مثبط يحتوى على فيروسات مرض الأسهال البقرى الفيروسى ومرض ألتهاب القصبة الهوائية الرغامى المعدى و فيروس البارا أنفلونزا-٣ و بكتيريا السالمونيلا ومرض ألتهاب القصبة تيفيميوريوم و السالمونيلا دبلن في العجول

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تهدف هذة الدراسة إلى تحضير و معايرة لفاح جامع مثبط بالبينارى يحتوى على فيروسات مرض الأسهال البقرى الفيروسي (بي في دي)

و فيروس مرض التهاب القصبة الهوائية الرغامي المعدى (أى بى أر) و فيروس البيار ا إنفلونزا-٣ (بى أى-٣) و بكتيريا السالمونيلا تيفيميوريوم و السالمونيلا دبلن و محمل بهيدروكسيد الألمونيوم الجيل كعامل مساعد لتفعيل اللقاح.

تمت معايرة اللقاح الجامع المخمد المحضر عن طريق أختبارات النقاوة و الأمان و الكفاءة المناعية . أثبتت الأختبارات أن اللقياح المحضر محليا أنه نقى و أمن تماما للاستحدام فى العجول بدون ظهور أى أعراض أو رد فعل غير مستحب سيواء عضوى أو موضعى فى مكان الحقن ولتقييم كفاءة اللقاح مناعيا تم إستخدام عدد ١٢ عجل بقرى بالغ سليم ظاهريا تم تقسيمهم إلى ٤ مجموعات تحتوى كل مجموعة على ٣ عجول. أستخدمت فى تقييم الكفاءة المناعية للعجول (أختبارات المصل التعادلي التلازن الدقيق و أختبار الحماية للفنران). أتضح مسن النتائج أنه لا يوجد أى تفاعل أو تضاد بين أى من مكونات اللقاح المحضر. كذلك أستمرت الأستجابة المناعية للعجول بمعدل حمسياية مسرتفع حتى نهاية التجربة (الأسبوع ٢٤ بعد التحصين).

أستنتجت من النتائج السابقة أن اللقاح المحضر محليا هو لقاح نقى و أمن و ذو كفاءة مناعية عالية لذلك ينصح باستخدامة في الحقل البيطري لتحصين العجول لحمايتها من الأسهال و الأمراض التنفسية.