

# Insecticidal Activity of Some Indole Derivatives Against *Spodoptera littoralis* (Boisd.)

Somaia E. Ali<sup>1</sup> and Ahmed S. Abdel-Aty<sup>2</sup>

<sup>1</sup>Department of Applied Entomology Faculty of Agriculture University; Alexandria, Egypt

<sup>2</sup>Department of Pesticide Chemistry and Technology University; Alexandria, Egypt

Received on: 23/2/2010

Accepted: 25/3/2010

## ABSTRACT

Eight indole derivatives were studied for their insecticidal activity on both larvae and eggs of *Spodoptera littoralis* (Boisd.). Insecticidal effects were a function of chemical structure, larval instar and concentration. All derivatives were more effective on the 4<sup>th</sup> larval instar after 5 days except compounds 3 and 7 with 612.8 and 437.7 µg/gm LC<sub>50</sub> on 6<sup>th</sup> instar, respectively. Indole-3-butyric acid (2) was the most effective with 70.9 and 39.7 µg/gm LC<sub>50</sub> on the 4<sup>th</sup> after 9 and 13 days, while 1-acetylindole-3-butyric acid (3) and 1-acetyl-2-phenylindole (7) were more effective with 151.4 and 80.6 µg/gm LC<sub>50</sub> values against sixth instar. While compounds 1, 2, 3 and 4 activated the larval weight of both instars, effect of the other derivatives was based on larval instar and concentration. Compound 3 was the most effective inhibiting pupation to 10% and blocking adult emergence to 25% after 21 and 45 days at 10 µg/gm in comparison to control. Malformations of intermediates and pupae as well as blocking adult emergence were higher in treated 4<sup>th</sup> larval instar based on the structural differences. Egg hatching was completely suppressed at 100 µg/gm of compounds 3 and 7. Dipping eggs in compound 2 solution inhibited hatching with IC<sub>50</sub> equaled 29.1 µg/ml and killed the produced larvae with LC<sub>50</sub> equaled 26.2 µg/ml. Transferring the immersed eggs to a poisoned medium enhanced the toxicity with IC<sub>50</sub> equaled 13.2 µg/gm and LC<sub>50</sub> equaled 15.2 µg/gm. Compound 7 multiplied the effect with IC<sub>50</sub> of 15.3 µg/gm and LC<sub>50</sub> of 7.5 µg/gm.

**Key words:** Indole ; *Spodoptera littoralis* ; larval mortality ; development ; egg hatching

## INTRODUCTION

The Egyptian cotton leaf-worm, *Spodoptera littoralis* (Boisd.) is an important polyphagous insect attacking cotton, several cultivated crops and ornamentals worldwide. Several synthetic insecticides that may not be environmentally safe are used for its control. Searching for other new insecticides is one of important concerns. Several plant extracts showed persuasive lethal and sublethal effects against it (Abbassy *et al.* 1998; Shonouda *et al.* 2000 & 2008). These activities were referred to plant originated heterocyclic alkaloids Ben Jannet *et al.*, (2000) & Tringali *et al.*, (2001).

Among heterocycles, the indole nucleus occupies a major importance as pharmaceuticals and antimicrobial agents. 5-Nitro-2-phenyl-1H-indole, 2-arylindole derivatives with ortho substitution on the phenyl ring and 5-methoxyindole-3-acetic acid exhibited potent antibacterial activity (Wang and Ng, 2002) & Samosorn *et al.*, (2005).

3-Acetyl-2,5,6-trichloro-1-(2-deoxy-beta-D-ribofuranosyl)-indole and some other derivatives induced potent antiviral activity Williams *et al.*, (2004). Indole-3-butyric acid controlled 23 bacteria and 15 fungi species Gulluce *et al.*, (2003). Combination of indole acetic acid at 100 µg/ml with *Cryptococcus laurentii* suppressed *Penicillium expansum* and *Botrytis cinerea* more than did *C. laurentii* alone (Yu and Zheng, 2007). Indole acetic acid, its 5-methoxy derivative, 1H-indole-4,7-diones and other indole alkaloids reduced spore germination, mycelial dry weight and protein

content of several fungi (Jingyong *et al.*, 2008, Kumar *et al.* (2007) and Ryu *et al.* (2007)). Antileishmanial activity of *Aspidosperma ramiflorum* is due to indole alkaloid content Tanaka *et al.*, (2007).

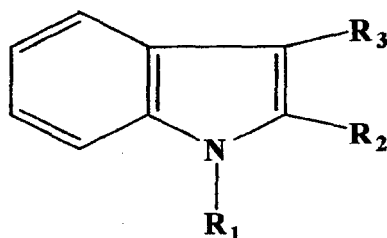
Regarding the insecticidal activity, sulfur-containing indole, camalexin plays an important role of defense in Brassicaceae, as toxic to pathogens and insects with antimicrobial and anti-fungal properties (Glazebrook, 2005). *S. littoralis* showed generally low sensitivity to feeding on different classes of alkaloids. Accumulation of indole glucosinolates reduced insect herbivory by *S. exigua* Gigolashvili *et al.*, (2007). Tryptanthrin, indole-3-acetonitrile showed insecticidal and anti-feeding activity against termites, *Reticulitermis santonensis* and larvae of the house longhorn beetle, *Hylotrupes bajulus* (Seifert and Unger, 1994).

Due to the mentioned biological activities and others of indole derivatives, this study aimed to examine eight indole derivatives for their effects against *S. littoralis*. These effects included larval lethality, reduction of larval weight, inhibition of pupation, blocking of adult emergence, developmental stages and malformations in the produced stages as well as egg hatching beside produced larval mortality.

## MATERIALS AND METHODS

### 1. Tested Compounds:

Both indol-3-acetic acid GRG, Batch No 971381, Code No L 17070 and indol-3-butyric acid, Sisco Research Laboratories PVT. LTD,



compound	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>
Indole-3-acetic acid (1)	H	H	-CH <sub>2</sub> -COOH
Indol-3-butyric acid (2)	H	H	-(CH <sub>2</sub> ) <sub>3</sub> -COOH
1-Acetylindole-3-butyric acid (3)	CH <sub>3</sub> CO	H	-(CH <sub>2</sub> ) <sub>3</sub> -COOH
1-Benzoylindole-3-acetic acid (4)	C <sub>6</sub> H <sub>5</sub> CO	H	-CH <sub>2</sub> -COOH
1-Benzoylindole-3-butyric acid (5)	C <sub>6</sub> H <sub>5</sub> CO	H	-(CH <sub>2</sub> ) <sub>3</sub> -COOH
2-Phenylindole (6)	H	C <sub>6</sub> H <sub>5</sub>	H
1-Acetyl-2-phenylindole (7)	CH <sub>3</sub> CO	C <sub>6</sub> H <sub>5</sub>	H
1-Benzoyl-2-phenylindole (8)	C <sub>6</sub> H <sub>5</sub> CO	C <sub>6</sub> H <sub>5</sub>	H

Mumbai, India, Batch No T828452 were purchased from El-Gomhouria Drug Company, Egypt. Other derivatives were prepared and identified (Abdel-Aty, 2009). These compounds are arranged as follows

## 2. Insecticidal effects:

### 2.1. Larval treatment:

The mother colony of *S. littoralis* was maintained under average laboratory conditions of  $27 \pm 3$  °C and  $60 \pm 5.0$  % RH. The insecticidal activity was tested on both the 4<sup>th</sup> and 6<sup>th</sup> larval instars. The tested larvae were reared on a semi artificial growing medium composed and modified from Shory (1963) by Hegazi *et al.* (1977). The tested compounds were dissolved in dimethylsulfoxide (DMSO) and well mixed with 2.0 gm of freshly prepared medium at concentrations of 10, 50, 100, 200, 500 and 1000 µg/gm. The control was concurrently kept under the same conditions. The dimethylsulfoxide concentration was as high as 1 %. After solidification of the used medium, fifteen larvae of the newly moulted 4<sup>th</sup> or 6<sup>th</sup> instar were placed on the treated medium in the plastic cups, covered with a cloth and secured with a rubber band as a replicate. Three replicates were used for each treatment. Each larval instar was individually treated. After feeding on the poisoned medium for 4 days, the treated larvae were transferred to an untreated medium. The alive larvae were daily counted. Mortality percents were calculated according to (Topps and Wain 1957) and its relation with exposure time was shown. Reduction in larval weight, larval developmental growth and malformations of intermediates, pupae and adults as well as percents of adult emergence were evaluated.

### 2.2. Egg treatment:

Some of the tested compounds that are persuasively active were studied for their effects on *S. littoralis* eggs. The compounds were prepared in 5, 10, 20, 50 and 100 µg/ml solutions. A consistent egg mass (0.030 gm) was dipped in the toxicant solutions for two minutes as a replicate. The contaminated egg masses were divided into two groups. The first group was transferred to 2.0 gm of non-poisoned medium. The other contaminated egg mass was transferred to a poisoned medium at the mentioned concentrations. Egg-hatching and the mortality of the produced larvae were evaluated and compared between the two groups as well as control. The concentration inhibited 50% of egg hatching (IC<sub>50</sub>) and the lethal concentration of 50% of the newly hatched larvae (LC<sub>50</sub>) were determined.

### 3- Statistical analysis:

Percentages of mortality and larval weight reduction were analyzed using the analysis of variance (ANOVA) and Student-Newman-Kules Test. LC<sub>50</sub>, IC<sub>50</sub> and EC<sub>50</sub> values were determined using probit analysis method (Finney, 1971).

## RESULTS AND DISCUSSION

### 1. Insect mortality:

Lethality results obtained as lethal concentration of 50% of the treated 4<sup>th</sup> and 6<sup>th</sup> larval instars are presented in Tables (1&2). All the tested compounds were more effective against the fourth larval instar than the sixth instar after 5 days except 1-acetylindole-3-butyric acid (3) and 1-acetyl-2-phenylindole (7). These compounds gave 612.8 and 437.7 µg/gm LC<sub>50</sub> values on the sixth larval instar, respectively comparing with >1000 and >1000 µg/gm on the fourth larval instar. The effect was increased after nine days in all cases. 1-Benzoyl-2-phenylindole (8) was less effective on the sixth and

its 1-acetyl derivative (7) were more effective on the sixth larval instar. The lethal effects were increased in all tested compounds against sixth larval instar except for compounds 2 and 5, which were more effective on the fourth instar. It was also found that substitution of compound 3 raised the toxicity on the sixth instar. The increase due to its acetylation appeared to be greater than benzoylation. Substitution of 2-phenyl moiety on the indole ring with removal of the side aliphatic carboxylic group increased the larval mortality in case of compound 6 more than in indole-3-acetic acid (1). Substitution with 1-acetyl on 2-phenylindole multiplied the lethality against the two tested larval instars, while substitution with 1-benzoyl in compound 8 enhanced the toxicity only against the fourth larval instar. The most effective compound was indole-3-butyric acid (2) with 70.9 and 39.7  $\mu\text{g/gm}$   $\text{LC}_{50}$  values on the fourth instar after 9 and 13 days, while 1-acetylindole-3-butyric acid (3) and 1-acetyl-2-phenylindole (7) were more effective with 151.4 and 80.6  $\mu\text{g/gm}$   $\text{LC}_{50}$  values against the sixth instar. So, compounds 2, 3 and 7 were chosen for egg treatment.

## 2. Sub-lethal effects:

### 2.1. Fresh body weight:

The larval weight of the 4<sup>th</sup> instar (after 7 days) was differently affected with the applied derivatives. Compound 1 and 2 raised larval weight at all concentrations in comparison to control. Benzoylation of indole-3-acetic acid in compound 4 affected the larval weight in non systematic arrangement with the tested concentrations. Acetylation of indole-3-butyric acid in compound 3 reduced the larval weight to 0.489 and 0.515 gm at 50 and 100  $\mu\text{g/gm}$ , followed by an increase at higher concentrations. On the contrary, its benzoyl derivative (compound 5) increased the larval weight when the lower concentration was used, followed by reduction at the two higher concentrations. Light reduction occurred at low concentrations, followed by gradual activation with increasing the concentration to 0.773 gm at 1000  $\mu\text{g/gm}$ , which was exhibited by compound 6. Substitution with 1-acetyl moiety in compound 7 increased the larval weight at low concentration with 16.0 and 2.3 %, followed by inhibition percents ranging from 3.2 to 18.5% of control at 100-1000  $\mu\text{g/gm}$ . Benzoylation of 2-phenylindole in compound 8 decreased the reduction effect more than compound 7.

Comparing with the untreated 6<sup>th</sup> larval weight (0.77 gm) after two days, all the tested compounds reduced the treated larval weight at all concentrations with different degrees and arrested their development to 7 days after treatment. Compounds 1 and 2 showed narrow differences among their concentrations and with less reducing effect, followed by compounds 8, 6, 5 and 7. Compounds 3 and 4 were the most active derivatives in weight reduction that may be due to

benzoylation. From these results, it could be realized that the hormonal effect was obviously clear through the activation of larval weight in most cases specially when applied earlier at the 4<sup>th</sup> instar more than at the 6<sup>th</sup> instar. These effects are shown in Figure (1).

### 2.2 Development:

The control of 4<sup>th</sup> instar larvae reached the pupal and adult stages after 6-7 and 9-10 days, respectively. Indole-3-acetic acid (1) at 10  $\mu\text{g/gm}$  delayed the development of the treated larvae to the pupae and adults to 29 and 45 days, respectively. However, the other compounds were less effective against pupation than compound 1 causing developing of 50, 10, 75, 92, 13, 63, and 83% of the treated larvae to pupae in case of compounds 2, 3, 4, 5, 6, 7 and 8, respectively after 21 days. While, compounds 5 and 7 caused complete transformation of the treated population to adults, compounds 2, 4, 6 and 8 caused developing of 75, 75, 55 and 67 % of population to adults. Compound 3 (1-acetylindole-3-butyric acid) was the most effective structure blocking adult emergence to 25% of the treated population after 45 days.

Regarding the 6<sup>th</sup> larval instar, its control larvae completely reached the pupal and adult stages after 2-3 and 7-8 days, respectively. All compounds arrested the larval development except compounds 3 and 5, which caused 25 and 18% pupation after 13 days. Compound 1 was the most effective inhibiting the adult emergence, followed by its 1-benzoyl derivative (4), 1-benzoylindole-3-butyric acid (5), indole-3-butyric acid (2), 2-phenylindole (6), 1-acetylindole-3-butyric acid (3), 1-benzoyl-2-phenylindole (8) and 1-acetyl-2-phenylindole (7). They blocked the adult emergence to 7, 14, 31, 33, 39, 46, 48 and 50% of the treated population after 35 days.

From the obtained results, the duration of *S. littoralis* larval stage was significantly affected by the tested compounds. It required long time to reach next stadium differing from control and differently among the studied derivatives (Figures 2a & 2b).

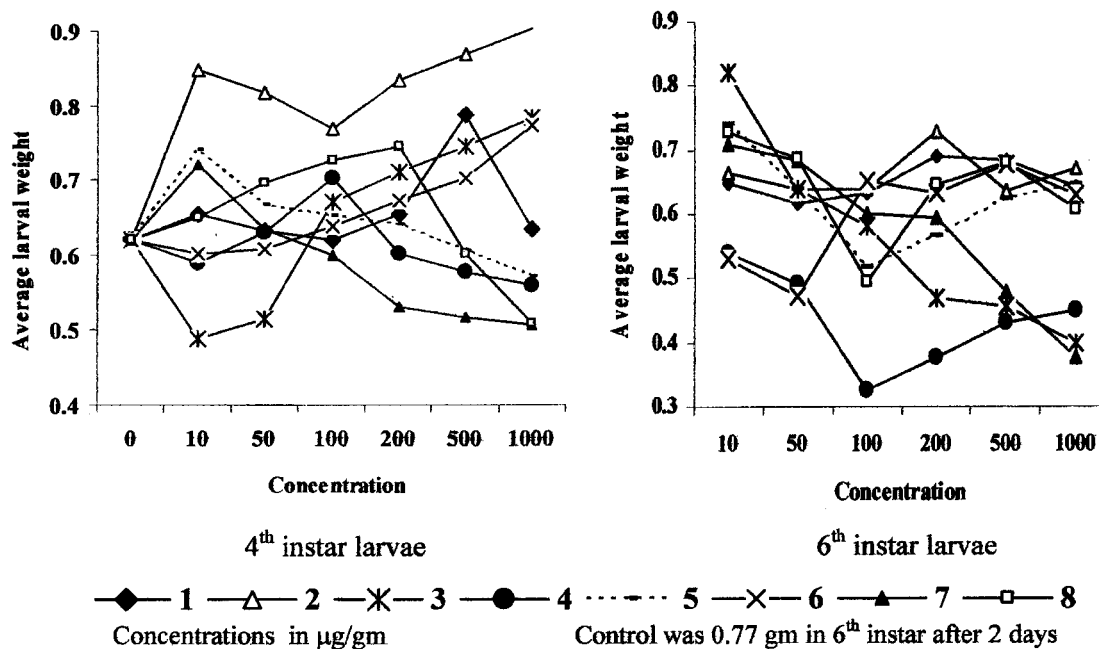
### 2.3 Malformations:

Several morphogenic effects in each stage at 10-200  $\mu\text{g/gm}$  concentration are recorded in Table (3). Comparing with the untreated larvae, both compounds 1 and 2 exhibited 14.6% and 16.7% malformation in the intermediates of the treated 4<sup>th</sup> larval instar at 200  $\mu\text{g/ml}$ , with no effect on that of the 6<sup>th</sup> larvae. Acetylation of indole-3-butyric acid in compound 3 affected the intermediates at lower concentrations in the 6<sup>th</sup> larval instar, while its benzoylation increased this effect against the 4<sup>th</sup> instar only. Acetylation of 2-phenylindole caused 32.6 and 61.1% malformation at 100 and 200  $\mu\text{g/ml}$  in intermediates of the treated 4<sup>th</sup> instar larvae with 1-benzoyl-2-phenylindole affected 4<sup>th</sup> larvae at 10  $\mu\text{g/ml}$  with 7.6% malformation. However, its effect was as high as 10.1% at the higher concentrations against 6<sup>th</sup> instar intermediates

**Table 1: Mortality effects of the tested compounds against the 4<sup>th</sup> instar of *S. littoralis* larvae**

Compound	LC <sub>50</sub> (µg/gm) after different times								
	5 Days			9 Days			13 Days		
	LC <sub>50</sub> 95%CL	Slope ± SE	χ <sup>2</sup>	LC <sub>50</sub> 95%CL	Slope ± SE	χ <sup>2</sup>	LC <sub>50</sub> 95%CL	Slope ± SE	χ <sup>2</sup>
Indole-3-acetic acid (1)	> 1000			804.7 503-1300	0.79± 9.0	1.94	543.2 373-795	0.86± 9.2	1.92
Indol-3-butyric acid (2)	117.8 69-201	0.44± 6.4	0.42	70.9 37.5-132	0.40± 6.4	1.52	39.7 21.9- 71	0.51± 6.6	1.10
1-Acetylindole-3-butyric acid (3)	> 1000			459.3 263-815	0.54± 7.1	0.75	243.2 159 - 375	0.59± 6.9	4.06
1-Benzoylindole-3-acetic acid (4)	828 535- 1293	0.87± 10.3	0.57	637.5 445-919	0.96± 10.7	0.84	376 287-494	1.07± 10.4	1.47
1-Benzoylindole-3-butyric acid (5)	424.4 297-609	0.83± 8.6	3.93	248.1 181-341	0.81± 7.9	2.17	116.7 86-159	0.79± 7.4	1.25
2-Phenylindole (6)	> 1000			779 441-1398	0.64± 8.0	5.37	352 235-532	0.69± 7.5	7.44
1-Acetyl-2-phenylindole (7)	> 1000			561.4 357-891	0.72± 8.2	6.83	223.5 157-319	0.71± 7.4	4.78
1-Benzoyl-2-phenylindole (8)	> 1000			514 316-845	0.65± 7.6	0.87	207 147-293	0.72± 7.3	1.55

Degree of freedom= 4      SE, Standard error × 10<sup>1</sup>



**Fig. 1: Effect of the tested compounds on fresh larval weight of *S. littoralis*; shown as average weight (gm) after 7 days of treatment**

**Table 2: Mortality effects against the 6<sup>th</sup> instar of *S. littoralis* larvae treated with different compounds**

Tested compound	LC <sub>50</sub> after different times					
	5 Days			9 Days		
	LC <sub>50</sub> 95%CL	Slope± SE	χ <sup>2</sup>	LC <sub>50</sub> 95%CL	Slope ± SE	χ <sup>2</sup>
Indole-3-acetic acid (1)	> 1000			680.4 428.9 – 1090.6	0.76 ± 8.72	3.93
Indol-3-butyric acid (2)	> 1000			794.9 473.7 – 1351.7	0.71 ± 8.60	1.0
1-Acetylindole-3-butyric acid (3)	612.8 395.6 – 958.4	0.77 ± 8.7	1.3	151.4 113.5 – 201.9	0.85 ± 7.7	3.93
1-Benzoylindole-3-acetic acid (4)	> 1000			209.1 146.9 – 298.6	0.76 ± 7.3	1.0
1-Benzoylindole-3-butyric acid (5)	> 1000			397.8 295.3 – 537.6	0.98 ± 9.7	1.01
2-Phenylindole (6)	> 1000			307.6 216.5 – 439.1	0.77 ± 7.84	1.92
1-Acetyl-2-phenyl indole (7)	437.7 256.3 – 758.6	0.55 ± 7.1	4.5	80.6 59.6 – 108.6	0.86 ± 7.61	5.8
1-Benzoyl-2-phenyl indole (8)	> 1000			> 1000		

Degree of freedom= 4 SE, Standard error × 10<sup>4</sup>

These malformation symptoms appeared as larval-pupal intermediates in which the posterior portion of the body only exhibited the pupal shape, while the anterior portion had larval head capsule and thoracic legs.

Malformation of the produced pupa (forming abnormal pupa without wings or that failed to shed of the larval cuticle) resulted from the 4<sup>th</sup> larval instar, which was more sensitive than that from 6<sup>th</sup> larval instar to treatment with compounds 1-3, 2-

phenylindole (6) and 1-benzoyl-2-phenylindole (8). The effects of compounds 4 and 5 depended on the applied concentration. Benzoylation of 2-phenylindole increased the pupae malformation. Adult malformation (adult failed to shed the pupal cuticle or adult with dwarf wings) was affected with the tested compounds, concentration and treated larval instar.

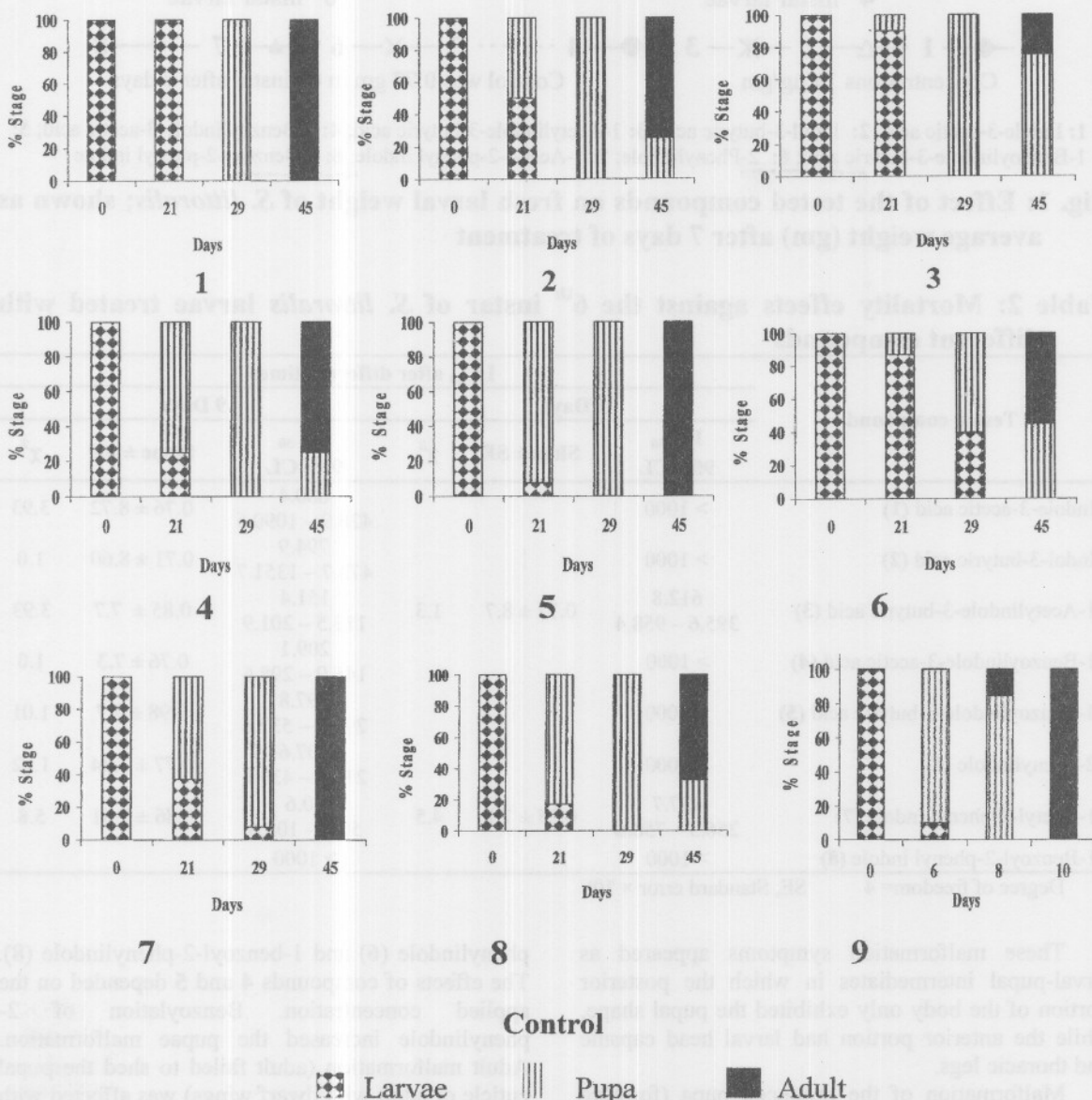
Adult emergence from the both treated larval instars was also affected. Compounds 1, 2, 3 and 5

blocked the adult emergence to 10.3 - 47.4%, 16.7 - 50%, 20.2 - 50.6% and 10.6 - 55.7% in systematic arrangement, respectively from 4<sup>th</sup> larval populations comparing with 100% of control. The blocking effect was reduced with increasing the concentration. They blocked adult emergence to 25.9-43.7%, 36.8-57.0%, 31.9-40.9% and 32.5-66.9%, respectively in non systematic arrangement in case of the 6<sup>th</sup> larval population. Compound 4 caused 9.5-20.8% and 22.9-69.5% adult emergence in case of the treated 4<sup>th</sup> and 6<sup>th</sup> larval instars, respectively. Although 2-phenylindole and its 1-acetyl derivative affected the adult emergence from both treated larval instars in non systematic arrangement, its 1-benzoyl derivative blocked the adult emergence with increasing the concentration. Adult emergence was more inhibited from 4<sup>th</sup> larval

instar treatment indicating that treatment of the lower larval instars gave good result of control. 1-Benzoylindole-3-acetic acid (4) was found more effective than indole-3-butyric acid (2) than indole-3-acetic acid (1) in their effect (Figure 3).

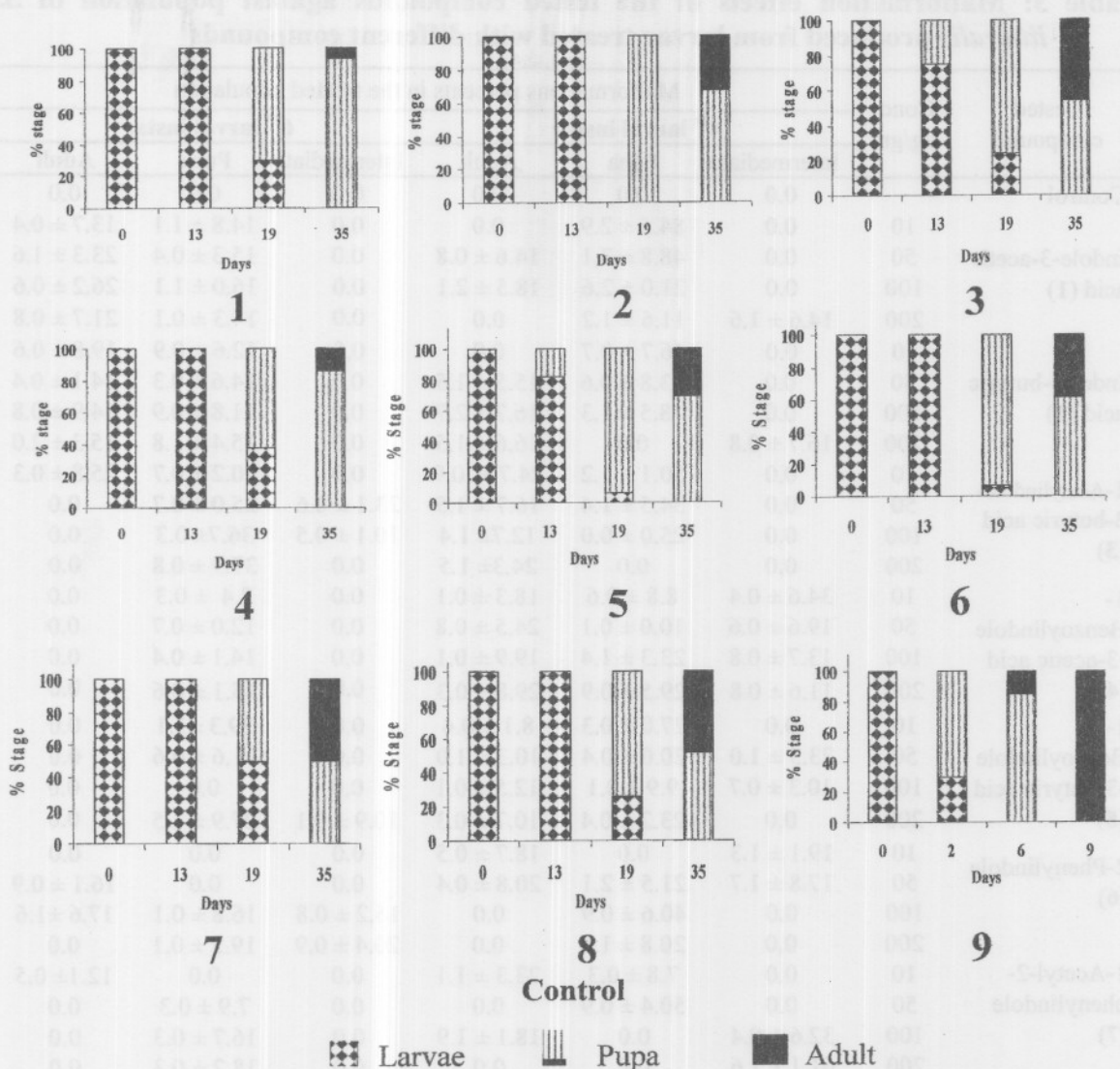
**3. Effect on eggs:**

Inhibition of egg hatchability was increased in systematic arrangement with the tested concentrations. Both 1-acetylindole-3-butyric acid (3) and 1-acetyl-2-phenylindole (7) completely stopped hatching when mixed at 100 µg/gm with the used medium. As the untreated egg mass hatched completely within 24 hours, treated eggs took 48-96 hours and 6-7 days at high concentrations of compound 2 and both of compounds 3 and 7, respectively.



1: Indole-3-acetic acid; 2: Indol-3-butyric acid; 3: 1-Acetylindole-3-butyric acid; 4: 1-Benzoylindole-3-acetic acid; 5: 1-Benzoylindole-3-butyric acid; 6: 2-Phenylindole; 7: 1-Acetyl-2-phenyl indole; 8: 1-Benzoyl-2-phenyl indole

**Fig. 2a : Effect of the tested compounds on *S. littoralis* 4<sup>th</sup> larval development at 10 µg/gm**



1: Indole-3-acetic acid; 2: Indol-3-butyric acid; 3: 1-Acetylindole-3-butyric acid; 4: 1-Benzoylindole-3-acetic acid; 5: 1-Benzoylindole-3-butyric acid; 6: 2-Phenylindole; 7: 1-Acetyl-2-phenyl indole; 8: 1-Benzoyl-2-phenyl indole

**Fig. 2b: Effect of the tested compounds on *S. littoralis* 6<sup>th</sup> larval development at 10  $\mu\text{g}/\text{gm}$**

Comparing the total number of hatched eggs and lethal effects on the produced larvae after 48 hours, only dipping the egg masses in solutions of compound 2 inhibited hatching with  $\text{IC}_{50}$  value equaled 29.1  $\mu\text{g}/\text{ml}$  and killed the produced larvae with  $\text{LC}_{50}$  value equaled 26.2  $\mu\text{g}/\text{ml}$ . Transferring treated eggs to the poisoned medium enhanced the toxicity on both egg-hatching with  $\text{IC}_{50}$  equaled 13.2  $\mu\text{g}/\text{gm}$  and on larval mortality with  $\text{LC}_{50}$  equaled 15.2  $\mu\text{g}/\text{gm}$ . Although acetylation in compound 3 decreased larval mortality in dipping technique with or without transferring the eggs to the poisoned medium, it enhanced egg-hatching inhibition when dipped only in the toxic solutions. Although compound 7 was less effective when egg masses

were dipped in it, its mixing with the used medium greatly enhanced the effect with  $\text{IC}_{50}$  value equaled 15.3  $\mu\text{g}/\text{gm}$  on egg-hatching and  $\text{LC}_{50}$  value equaled 7.5  $\mu\text{g}/\text{gm}$  on larval mortality. The results are shown in Table (4).

It could be concluded that mortality of the 4<sup>th</sup> instar larvae was increased with increasing the aliphatic side chain. Substitution of N-H of 2-phenylindole raised the toxicity, vice versa in case of indole-3-butyric acid against the same instar. The tested compounds affected the larval weight, pupation and adult emergence. Several malformations in intermediates, pupae and adults were noticed with A type of juvenilization as larval-pupal intermediates were observed where the

**Table 3: Malformation effects of the tested compounds against population of *S. littoralis* produced from larvae treated with different compounds**

Tested compound	Conc. $\mu\text{g/gm}$	Malformations percents in the treated population					
		4 <sup>th</sup> larval instar			6 <sup>th</sup> larval instar		
		Intermediate	Pupa	Adult	Intermediate	Pupa	Adult
Control		0.0	0.0	0.0	0.0	0.0	0.0
	10	0.0	84.5 $\pm$ 2.9	0.0	0.0	14.8 $\pm$ 1.1	13.7 $\pm$ 0.4
Indole-3-acetic acid (1)	50	0.0	48.8 $\pm$ 3.1	14.6 $\pm$ 0.8	0.0	15.3 $\pm$ 0.4	23.3 $\pm$ 1.6
	100	0.0	21.0 $\pm$ 2.6	18.5 $\pm$ 2.1	0.0	16.0 $\pm$ 1.1	26.2 $\pm$ 0.6
	200	14.6 $\pm$ 1.6	11.6 $\pm$ 1.2	0.0	0.0	14.3 $\pm$ 0.1	21.7 $\pm$ 0.8
	10	0.0	46.7 $\pm$ 0.7	0.0	0.0	12.6 $\pm$ 0.9	19.8 $\pm$ 0.6
Indol-3-butyric acid (2)	50	0.0	33.8 $\pm$ 0.6	15.5 $\pm$ 1.5	0.0	14.6 $\pm$ 0.3	14.1 $\pm$ 0.4
	100	0.0	18.5 $\pm$ 1.3	16.7 $\pm$ 2.7	0.0	11.8 $\pm$ 0.9	14.9 $\pm$ 0.8
	200	16.7 $\pm$ 0.8	0.0	16.6 $\pm$ 1.5	0.0	15.4 $\pm$ 1.8	15.3 $\pm$ 2.0
	10	0.0	50.1 $\pm$ 1.2	24.7 $\pm$ 0.9	0.0	20.2 $\pm$ 0.7	25.8 $\pm$ 0.3
1-Acetylindole-3-butyric acid (3)	50	0.0	34.5 $\pm$ 1.4	16.7 $\pm$ 1.5	23.1 $\pm$ 0.6	25.0 $\pm$ 0.7	0.0
	100	0.0	25.0 $\pm$ 0.0	12.7 $\pm$ 1.4	10.1 $\pm$ 0.5	36.7 $\pm$ 0.3	0.0
	200	0.0	0.0	24.3 $\pm$ 1.5	0.0	37.9 $\pm$ 0.8	0.0
	10	34.6 $\pm$ 0.4	8.8 $\pm$ 0.6	18.3 $\pm$ 0.1	0.0	7.4 $\pm$ 0.3	0.0
1-Benzoylindole-3-acetic acid (4)	50	19.6 $\pm$ 0.6	10.0 $\pm$ 0.1	24.5 $\pm$ 0.8	0.0	12.0 $\pm$ 0.7	0.0
	100	13.7 $\pm$ 0.8	23.3 $\pm$ 1.4	19.9 $\pm$ 0.1	0.0	14.1 $\pm$ 0.4	0.0
	200	11.6 $\pm$ 0.8	29.5 $\pm$ 0.9	29.8 $\pm$ 0.3	0.0	15.1 $\pm$ 0.6	0.0
	10	0.0	27.0 $\pm$ 0.3	8.1 $\pm$ 0.4	0.0	29.3 $\pm$ 1.1	0.0
1-Benzoylindole-3-butyric acid (5)	50	23.3 $\pm$ 1.0	20.0 $\pm$ 0.4	10.3 $\pm$ 1.0	0.0	11.6 $\pm$ 0.6	0.0
	100	10.3 $\pm$ 0.7	9.9 $\pm$ 0.1	12.5 $\pm$ 0.1	0.0	0.0	0.0
	200	0.0	23.2 $\pm$ 0.4	10.7 $\pm$ 0.3	10.9 $\pm$ 0.1	27.9 $\pm$ 1.5	0.0
	10	19.1 $\pm$ 1.3	0.0	18.7 $\pm$ 0.5	0.0	0.0	0.0
2-Phenylindole (6)	50	17.8 $\pm$ 1.7	21.5 $\pm$ 2.1	20.8 $\pm$ 0.4	0.0	0.0	16.1 $\pm$ 0.9
	100	0.0	40.6 $\pm$ 0.9	0.0	16.2 $\pm$ 0.8	16.8 $\pm$ 0.1	17.6 $\pm$ 1.6
	200	0.0	20.8 $\pm$ 1.1	0.0	20.4 $\pm$ 0.9	19.3 $\pm$ 0.1	0.0
	10	0.0	7.8 $\pm$ 0.3	23.3 $\pm$ 1.1	0.0	0.0	12.1 $\pm$ 0.5
1-Acetyl-2-phenylindole (7)	50	0.0	50.4 $\pm$ 0.9	0.0	0.0	7.9 $\pm$ 0.3	0.0
	100	32.6 $\pm$ 0.4	0.0	18.1 $\pm$ 1.9	0.0	16.7 $\pm$ 0.3	0.0
	200	61.1 $\pm$ 1.6	0.0	0.0	0.0	18.2 $\pm$ 0.3	0.0
	10	17.6 $\pm$ 1.3	17.3 $\pm$ 0.9	15.1 $\pm$ 0.6	5.8 $\pm$ 0.6	11.9 $\pm$ 1.1	18.2 $\pm$ 0.6
1-Benzoyl-2-phenylindole (8)	50	0.0	21.1 $\pm$ 1.6	19.4 $\pm$ 0.8	5.7 $\pm$ 1.1	10.8 $\pm$ 0.3	10.8 $\pm$ 1.7
	100	0.0	35.6 $\pm$ 0.6	13.5 $\pm$ 1.5	6.6 $\pm$ 1.6	5.1 $\pm$ 0.9	0.0
	200	0.0	32.3 $\pm$ 0.6	19.9 $\pm$ 0.8	10.1 $\pm$ 1.5	0.0	0.0

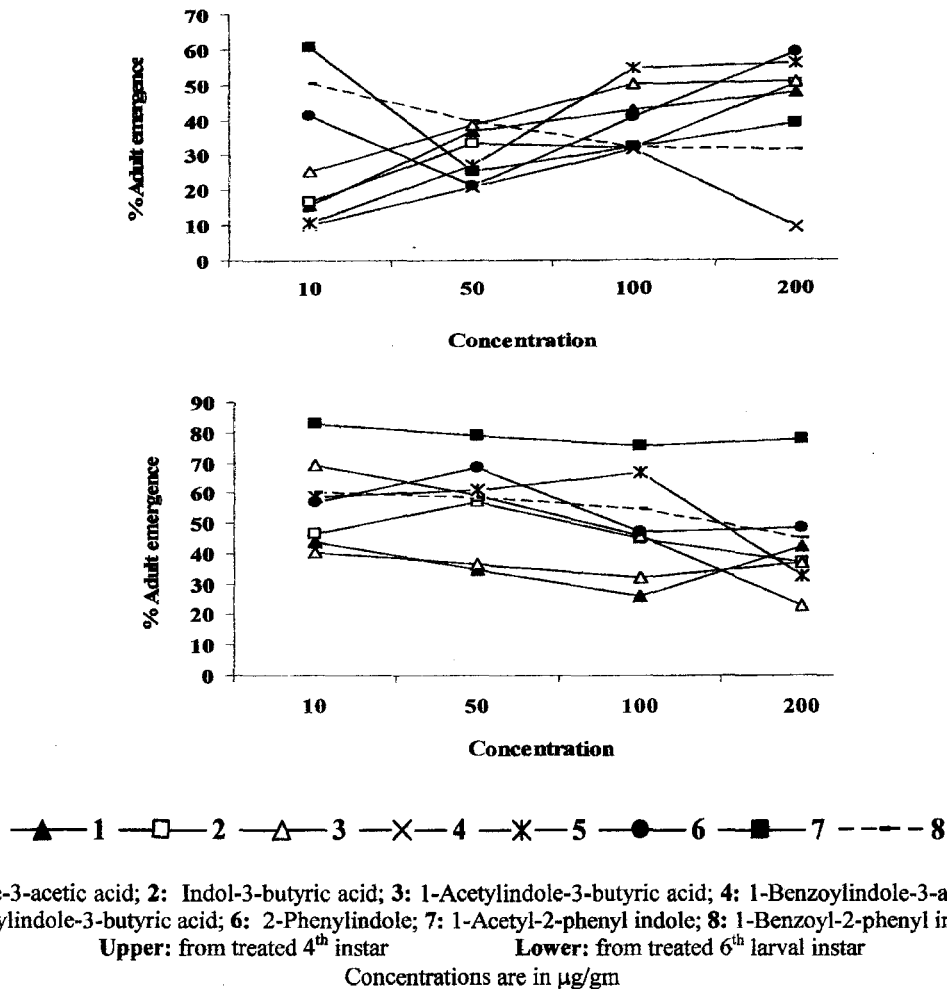
Data are shown in percentages

**Table 4: Effect of the active derivatives on *S. littoralis* eggs**

Tested compound	Used technique	Inhibition of egg-hatching			Lethality on the hatched larvae*		
		IC <sub>50</sub>	Slope $\pm$ SE	Chi <sup>2</sup>	LC <sub>50</sub>	Slope $\pm$ SE	Chi <sup>2</sup>
		95% CL			95% CL		
Indol-3-butyric acid (2)	Dipping	29.1 25.0-33.9	1.96 $\pm$ 0.024	0.34	26.2 20.7-33.4	1.13 $\pm$ 0.017	4.52
	Dipping + Mixing	13.2 11.5-15.1	2.27 $\pm$ 0.032	5.81	15.2 13.5-17.2	2.7 $\pm$ 0.041	1.9
1-Acetylindole-3-butyric acid (3)	Dipping	19.3 16.8-22.1	2.2 $\pm$ 0.027	4.8	35.1 27.6-44.9	1.18 $\pm$ 0.018	1.97
	Dipping + Mixing	16.6 14.3-19.2	2.0 $\pm$ 0.025	2.47	31.6 21.5-46.9	0.7 $\pm$ 0.015	3.17
1-Acetyl-2-phenylindole (7)	Dipping	28.1 23.6-33.6	1.62 $\pm$ 0.02	3.30	> 100		
	Dipping + Mixing	15.3 13.1-17.9	1.88 $\pm$ 0.024	3.04	7.5 6.2-9.1	2.0 $\pm$ 0.05	5.03

\* Lethality on the hatched larvae were evaluated after two days





**Fig. 3: Emergence percents of *S. littoralis* adults produced from treated larvae**

posterior of the body only showed the pupal shape while, the anterior portion had the larval head capsule and thoracic legs. This further indicates that treatment induced an effect typical to juvenile hormone excess. Juvenalized adults failed to emerge from the pupal cuticle, *i.e.* produced imperfect alive adults. This effect varied according to the tested compound.

These delayed effects are expressed as developmental abnormalities in the adult stage *e.g.* imperfect adults, adults with eclosion problems and dead adults before emergence. These effects may be due to oxidative decarboxylation to cytotoxic species. It is thought to be due to the formation of 3-methylene-2-oxindole, which may conjugate with DNA bases and protein thiols (Folkes and Wardman, 2001). It may be also due to inhibition of cholinesterase as indole-3-butyric acid inhibited the butyrylcholinesterase (BuChE) from purified human serum and horse serum (Bodur and Cokugras, 2005). Its effect is associated with cell peroxidase activity (De Melo *et al.*, 2004). 2-phenylindole and 1-acetylindole-3-butyric acid affected fungal polyphenoloxidase (PPO), peroxidase (PO), DNA and RNA contents (Abdel-Aty, 2009). At the same time, phenoloxidase (PO) is believed to be a key

mediator of immune function in insects where in larvae of the Egyptian cotton leafworm, *S. littoralis*, haemolymph PO activity varied markedly between individuals. The heritability estimate of haemolymph PO activity was high, while PO activity in the haemolymph was strongly correlated with PO activity in both the cuticle and midgut (Cotter and Wilson, 2002). This notice may clarify the effect of the tested compounds on adult emergence and pupation. N-H and N-substituted indole-2- and 3-carboxamide showed a strong inhibitory (95-100%) effect on superoxide anion (SOD). Some showed similar potency for the inhibition of lipid peroxidation (81-94%) which revealed their highly potent antioxidant properties. Substitution in the 1-position of the indole ring caused significant differences between the activity results regarding lipid peroxidation inhibition Olgen *et al.*, (2007) emphasizing the differences in effects due to the derivative structure.

## REFERENCES

- Abdel-Aty, A. S. 2009. Fungitoxic effect of certain indole derivatives against some plant pathogenic fungi. *Alex. J. Agric. Res.*, 54 (2): 113-125.

- Abbassy, M.A., O. A. el-Gougary, S. el-Hamady and M. A. Sholo (1998). Insecticidal, acaricidal and synergistic effects of soosan, *Pancreaticum maritimum* extracts and constituents. *J Egypt Society Parasitology*, **28** (1): 197–205.
- Ben Jannet, H., F. Harzallah-Skhiri, Z. Mighri, M. S. Simmonds and W. M. Blaney 2000. Responses of *Spodoptera littoralis* larvae to Tunisian plant extracts and to neo-clerodane diterpenoids isolated from *Ajuga pseudoiva* leaves. *Fitoterapia*, **71** (2): 105–112.
- Bodur, E.; A. N. Cokugras 2005. The effects of indole-3-acetic acid on human and horse serum butyryl cholinesterase. *Extended Abstracts/Chemico-Biological Interactions*, **157-158**: 373–377.
- Cotter, S. C. and K. Wilson 2002 Heritability of immune function in the caterpillar *Spodoptera littoralis*. *Heredity*, **88** (4): 229–234.
- De Melo, M. P.; T. C. Pithon-Curi and R. Curi 2004. Indole-3-acetic acid increases glutamine utilization by high peroxidase activity-presenting leukocytes. *Life Sci.*, **75** (14): 1713–1725.
- Finney, D. J. 1971. Probit analysis. 3<sup>rd</sup> edition Cambridge University Press, London; page: 138.
- Folkes, L. K. and P. wardman 2001. Oxidative activation of indole-3-acetic acid to cytotoxic species a potential new role for plant auxins in cancer therapy. *Biochemical pharmacology*, **61**: 129–136.
- Gigolashvili, T.; B. Berger; H. P. Mock; C. Muller; B. Weisshaar and U. I. Flugge 2007. The transcription factor HIG1/MYB51 regulates indolic glucosinolate biosynthesis in *Arabidopsis thaliana*. *Plant J.* **50**, 886–901.
- Glazebrook, J. 2005. Contrasting mechanisms of defense against biotrophic and necrotrophic pathogens. *Annu. Rev. Phytopathol.* **43**, 205–227.
- Gulluce, M; M. Sokmen; D. Daferera; G. Agar; H. Ozkan; N. Kartal; M. Polissiou; A. Sokmen and F. Sahin 2003. *In vitro* antibacterial, antifungal, and antioxidant activities of the essential oil and methanol extracts of herbal parts and callus cultures of *Satureja hortensis* L. *J Agric Food Chem.*, **51** (14): 3958–3965.
- Hegazi, E. M.; A. M. El-Menshawy and S. M. Hammad 1977. Mass rearing of the Egyptian cotton leafworm, *Spodoptera littoralis* (Boisd.) on semi-artificial diet. *Proc. 2<sup>nd</sup> Arab Pesticide Conf., Tanta Univ.* pp. 61–70.
- Jingyong S.; L. Hongxiang; D. Shengjun; H. Xu; F. Zhao and K. Liu 2008. Indole alkaloids from *Nuclea officinalis* with weak antimalarial activity. *The Plant Journal* **55** (5): 774–786.
- Kumar, V.; A. Kumar and R. N. Kharwar 2007. Antagonistic potential of fluorescent pseudomonads and control of charcoal rot of chickpea caused by *Macrophomina phaseolina*. *J Environ Biol.* **28** (1): 15–20.
- Olgen, S.; Z. Kiliç; A. O. Ada and T. Coban 2007 Synthesis and evaluation of novel N-H and N-substituted indole-2- and 3-carboxamide derivatives as antioxidants agents. *J Enzyme Inhib Med Chem.*, **22** (4): 457–462.
- Ryu, C. K. ; J. Y. Lee; R. E. Park; M.Y. Ma and J. H. Nho 2007. Synthesis and antifungal activity of 1H-indole-4,7-diones. *Bioorg Med Chem Lett.* **17** (1): 127–131.
- Samosorn, S.; J. B. Bremner; A. Ball and K. Lewis 2005. Synthesis of functionalized 2-aryl-5-nitro-1H- indoles and their activity as bacterial N- or A- efflux pump inhibitor. *Bioorganoc & medicinal Chemistry*, **14**: 857–865.
- Seifert, K and W. Unger 1994. Insecticidal and fungicidal compounds from *Isatis tinctoria*. *Z: Naturforsch [C]*, **49** (1-2): 44–48.
- Shonouda, M. L, R. M. Farrag and O. M. Salama 2000. Efficacy of the botanical extract (myrrh), chemical insecticides and their combinations on the cotton leaf worm, *Spodoptera littoralis* (Boisd.) (Lepidoptera : Noctuidae). *J Environmental Science Health B.* **35** (3): 347–356.
- Shonouda, M. L.; S. Osman; O. Salama and A. Ayoub 2008. Toxic effect of *Peganum harmala* L. leaves on the cotton leaf worm, *Spodoptera littoralis* (Boisd.) and its parasitoid *Microplitis rufiventris* Kok. *Pak J Biological Science*, **11** (4): 546–452.
- Shory 1963. A simple artificial rearing medium for the cabbage looper. *J. Econ Entomol.*, **56**: 536–537.
- Tanaka, J. C.; C. C. da Silva; I. C. Ferreira; G. M. Machado; L. L. Leon and A. J. de Oliveira 2007. Antileishmanial activity of indole alkaloids from *Aspidosperma ramiflorum*. *Phytomedicine*, **14** (6): 377–380.
- Topps, J. H. and R. L. Wain 1957. Investigation on fungicides. III. The fungitoxicity of and 5-alkyl salicylanilide and p-chloroanilines. *Ann. Appl. Biol.*, **45** (3): 506–511.
- Tringali, C.; C. Spatafora; V. Cali and M. S. Simmonds 2001. Antifeedant constituents from *Fagara macrophylla*. *Fitoterapia*, **72** (5): 538–543.
- Wang, H. X. and T. B. Ng 2002. Demonstration of antifungal and anti-human immunodeficiency virus reverse transcriptase activities of 6-methoxy-2-benzoxazolinone and antibacterial activity of the pineal indole 5-methoxyindole-3-acetic acid. *Comp Biochem Physiol C Toxicol Pharmacol.* **132** (2): 261–268.
- Williams, J. D.; R. G. Ptak and J. C. Drach 2004. Synthesis, antiviral activity, and mode of action of Some 3-Substituted 2,5,6-

Trichloroindole 2'- and 5'-Deoxyribonucleosides. *J Med. Chem.*, 47 (23): 5773-5782.  
Yu, T. and X. D. Zheng 2007. Indole-3-acetic acid enhances the biocontrol of *Penicillium*

*expansum* and *Botrytis cinerea* on pear fruit by *Cryptococcus laurentii*. *FEMS Yeast Res.*, 7 (3):459-464.

### الملخص العربي

## النشاط الإبادي لبعض مشتقات الأندول على دودة ورق القطن

سمية السيد على<sup>١</sup>، أحمد صبرى عبد العاطى<sup>٢</sup>

قسم علم الحشرات التطبيقي<sup>١</sup> قسم كيمياء و تقنية المبيدات<sup>٢</sup>

كلية الزراعة - جامعة الإسكندرية - الشاطبي - الإسكندرية - مصر

تم تقدير النشاط الإبادي الحشري لثمانية من مشتقات الأندول ضد بيض و يرقات دودة ورق القطن (العمرين الرابع و السادس).

ظهر التأثير الإبادي لهذه المركبات كدالة في كل من العمر اليرقي و المركب المختبر و كذلك تركيزه المستخدم . أظهرت كل المشتقات المختبرة سمية أعلى على العمر اليرقي الرابع بعد خمسة أيام من المعاملة عدا مركبي ٣ و ٧ اللذان سببا تركيزا قاتلا لـ ٥٠% من عشيرة يرقات العمر السادس قدره ٦١٢,٨ و ٤٣٧,٧ ميكروجرام/جم بيئة مسممة.

كما ظهر مركب الـ إندول-٣- بيوتيريك أسيد (٢) أعلى نشاطا بتركيز قاتل لـ ٥٠% من عشيرة يرقات العمر الرابع قدره ٧٠,٩ و ٣٩,٧ ميكروجرام/جم بعد ٩ و ١٣ يوم على الترتيب من بداية المعاملة بينما ظهر مركبي الـ ١-أسيتيل اندول-٣-بيوتيريك أسيد (٣) و الـ ١-أسيتيل-٢-فينيل اندول (٧) أكثر نشاطا على عشيرة يرقات العمر السادس بتركيز قاتل لـ ٥٠% قدره ١٥١,٤ و ٨٠,٦ ميكروجرام/جم. في حين أزادت مركبات ١ و ٢ و ٣ و ٤ وزن اليرقات بينما إختلف تأثير باقى المركبات حسب التركيز و العمر اليرقي.

ظهر مركب الـ ١-أسيتيل اندول-٣-بيوتيريك أسيد (٣) الأكثر نشاطا حيث ثبت تعذير اليرقات المعاملة إلى ١٠% من العشيرة بعد ٢١ يوم كما أخدم الإنسلاخ إلى الطور البالغ إلى ٢٥% من العشيرة بعد ٤٥ يوم على تركيز ١٠ ميكروجرام/جم مقارنة بالكنترول الذى أعطى ١٠٠% تعذير و ١٠٠% تحول إلى الطور الكامل بعد ٦-٧ و ٩-١٠ أيام على الترتيب. أثبتت النتائج حدوث تشوهات فى كل الأطوار الناتجة بصورة أكبر فى حالة المعاملة فى الطور اليرقي الرابع عنها حين عومل الطور اليرقي السادس.

تم إخماد فقس البيض تماما بمعاملته بتركيز ١٠٠ ميكروجرام/جم من مركبي الـ ١-أسيتيل اندول-٣-بيوتيريك أسيد (٣) و الـ ١-أسيتيل-٢-فينيل اندول (٧).

غمس البيض فى محلول مركب الـ إندول-٣-بيوتيريك أسيد (٢) ثبط فقسه بتركيز لازم لتثبيط ٥٠% قدره ٢٩,١ ميكروجرام/مل كما سبب موت اليرقات الناتجة بتركيز لازم لقتل ٥٠% قدره ٢٦,٢ ميكروجرام/مل كما أن وضع البيض على بيئة مسممة بعد الغمس أدى إلى زيادة السمية بتركيز لازم لتثبيط فقس ٥٠% قدره ميكروجرام/جم ١٣,٢ و بتركيز لازم لقتل ٥٠% قدره ١٥,٢ ميكروجرام/جم. ضاعف مركب الـ ١-أسيتيل-٢-فينيل اندول (٧) السمية بتركيز لازم لتثبيط فقس ٥٠% قدره ميكروجرام/جم ١٥,٣ و الناتجة بتركيز لازم لقتل ٥٠% من اليرقات قدره ٧,٥ ميكروجرام/جم.