CLINICAL AND EPIDEMIOLOGICAL STUDIES ON RESPIRATORY MANIFESTATIONS IN SHEEP

A. A. Mahmoud, A. M. Khader and M. KH. Abd Elsalam Infectious Diseases department Fac. Of Vet. Med. Alex. Univ.

ABSTRACT

2390 Living sheep of different ages (1-12 months), sexes and breeds (Braky- Rhmany- Baladi) were examined clinically in the field for investigation of the animals which suffered from signs of respiratory manifestation [nasal discharge (serous - mucoid - mucopurulent), rapid breathing, sever dyspenea, pyrexia (rectal temp 40-41 c). congested mucous membranes, lacrimation, coughing and abnormal lung sound by auscultation. The study was carried out on (150) pneumonic lambs. The morbidity rate was 6.2 % (150/2390), mortality rate was 7.4 % and fatality rate was 19.3 % (29/150).

Bacteriological examination revealed that (75) samples were positive for bacterial growth with percentages (50%) from previously affected sites of samples. In total out of (127) bacterial isolates. Concerning Gram positive bacteria ; staphylococcus aureus was 10 (6.7 %), streptococcus 8 % staphylococcus (5.3) epidermidis 14(9.3 %).while the gram negative bacteria were as E,coli 30 (20%) identified Proteus sp 28 (13.3%) Enterobacter 9 (6%) klebseilla pneumoniae sp subsp pneumoniae 5 (3.4)%

klebseilla ozaenae 7 (4.7 %), citrobacter diversus 6 (4%), pasteurella multocida 2 (1.3 %), providencia 2 (1.3%) pseudomonas aerugenosa 14 (9.3 %) isolated from the different examined collected samples. Bacteriological examination for (14) lung samples revealed that staphylococcus aureus and Escherichia coli were isolated from two samples (14.2) % and other bacteria were mannheimia haemolytica pseudomonas aerugenosa, Enterobacter agglumerans, Haemophillus somuns every one of them were isolated from one sample (7.1%). Only actinomyces pyogenes was isolated from three samples (21.3%).

The bacterial examination of lung samples showed that ; all isolates were single such as actinomyces pyogens from two samples (14.2), mannheimia haemolytica haemophillus somnus Enterobacter agglomerans , Staph .aureus every one of them was isolated from one samples (7.1) % . Except two isolates were isolated from mixed cultures ; Escherichia coli + Actinomyces pyogenes and E.coli +Staph aureus .Every isolates was isolated from 1sample (7.1%). test revealed that Sensitivity

most isolates were highly sensitive to Enrofloxacin, Cephaloexine, and Ampicillin and were resistant to streptomycin

INTRODUCTION

Sheep play a vital economic role and support the survival of millions of people in our country, it is used as a source of meat, milk ,wool as well as quite and effective means of money disorder is still serious .respiratorv problem facing sheep rearing .(Hatem ;et.al2003.) the main causes of pneumonia are bacteria, fungi and viruses where poor hygienic measurement and climatic disorders are the most various predisposing factors to infection .(Radwan et al 2002). Communal bacteria were isolated with a percentage of 20% from clinically healthy sheep (Ibrahim and Mokhtar 2003). Also(Abdel.latif and El-dossoukv .,2006) isolated pasteurella multocida (5.71%), E.coli (2.86%) ,klebsiella pneumoniae and pseudomonas aerugenosa (5.71%) from clinically healthy living lambs .Moreover (Elyas .1993)recovered staphylococcus aureus (26%), E.coli (16%) and pasteurella multocida (3%) from clinical normal lambs .Mannheimia hemolytica were culturally isolated from the nasal secretion and lung of diseased pneumonic sheep (Quinn et al .,1994). Both mannheimia haemolytica and (pasteruella) pasteurella multocida were associated with pneumoniae in sheep and goat (Davies : 1985).moreover ,(Radwan et al..2002) reported that pasteurella multocida was the most important cause of pneumoniae in goat .also (Elyas .1993) isolated from pneumonic lungs of sheep and goat mannheimia haemolytica, E.coli and klebsiella pneumoniae. The clinical source of klebsiella pneumoniae infection is usually rapid with lung tissue necrosis and frequent formation of abscess (Reed .1973) pasteurella spps .klebsiella spps. And staphylococcus aureus are the most bacterial cause of sheep pneumoniae (Martin.1996).

This study was directed mainly to determine:

- 1. Epidemiological studies of ovine pneumonia.
- 2. Clinical finding of ovine pneumonia.
- 3.Bacterial causes of ovine pneumonia.
- 4. The sensitivity of the isolated bacteria to different drugs to aid in the choice of drug.

MATERIAL & METHODS

I- Material

1-Samples

1- Nasal swabs: (2390) Living sheep of different ages (1-12 months), sexes and breeds (Braky- Rhmany- Baladi) were examined clinically in the field for investigation of the animals which suffered from signs of respiratory manifestation Nasal swabs were taken under septic condition from (150) pneumonic lambs of both sexes' aged from 1-12 months suffering from respiratory disorder includes [nasal (serous discharge – mucoid mucopurulent), rapid breathing, sever dyspenea, pyrexia (rectal temp 40-41 c). congested mucous membranes, lacrimation, coughing and abnormal

lung sound by auscultation. These lambs belonged to different herds at Alexandria governorate.

2- **Lung samples** (14) lung samples were obtained from slaughtered pneumonic lambs at abattoirs. And sent to laboratory directly for bacteriological directly examination

<u>2-Media</u>

The following media were used according to *(Finegold and martin 1982)* Nutrient agar, Blood agar, Tryptose soya agar. MacCkonkey agar, mannitol salt agar, dextrose starch agar. Nutrient broth, Brain heart infusion broth Muller Hinton agar, Muller Hinton broth and were obtained from (oxoid Lt d)

Antibiotic sensitivity test ambicillin 10 ug , ampicillin 10 ug , cephaloxin 10mg , Enrofloxacin 10 mg , Erythromycin 10 mg ,Gentamycin 10 ug , oxyteracycline 30 ug penicillin 10 ug were used according to the instruction of the manufactures on random isolates from each species and obtained from (oxoid Ltd)

<u>3-Stains</u>; Gram stain and Giemsa stain were used according to *(Finegold and martin .1982)*

II- Methods

1- Collection of samples

Nasal swabs collected from clinical cases. The cotton swabs were pushed as possible into one nostril then transferred to sterile nutrient broth in tubes. (Woldehiwt.et al 1990).

Lung samples: the sterilization of lung tissue was done by flamed spatula and cut, take swabs or apiece of lung to cultures into peptone water. The samples were taken under aseptic condition and sent without delay to laboratory. All media previously melted and cooled till 40c inculated and control plates were incubated at 37c for 24-48 hours (*Cruickshank et al* 1975)

2- Isolation and identification of the isolates according to (Quinn et al, 1994)

All previously collected samples were inculated into the following solid media ; nutrient agar . MacCconkey agar .mannitol salt agar 7.5 % . tryptose soya agar and dextrose starch agar . Incubated aerobically at 37c for 24-48 hours, the isolated colonies were streaked on new sterile pure culture of Nutrient agar and subcultured on slopes agar for further identification by study the culture characters pigment production. Staining reaction and cell morphology as well as aggregation. The biochemical activities of pure isolates were carried out according to (Finegold and martin.1982)

3- Sensitivity of microorganism to antibiotics was determined by using Muller Hinton agar plates and standard disc technique according to *(Quinn et al 1994)*

RESULTS AND DISCUSSION

((2390) Living sheep of different ages (1-12 months), sexes and breeds (Braky- Rhmany- Baladi) were examined clinically in the field for investigation of the animals which suffered from signs of respiratory manifestation Nasal swabs were taken under septic condition from (150) pneumonic lambs of both sexes' aged

from 1-12 months suffering from respiratory disorder includes [nasal discharge (serous – mucoid mucopurulent), rapid breathing, sever dyspenea, pyrexia (rectal temp 40-41 c). congested mucous membranes, lacrimation, coughing and abnormal lung sound by auscultation. our results agree with **Radostits at al. (1995).**

The study was carried out on (150) pneumonic lambs. The morbidity rate was 6.2 % (150/2390), mortality rate was 7.4 % and fatality rate was 19.3 % (29/150) as shown in table (1).

Nasal swabs were collected under septic condition from (150) clinically diseased sheep suffering from respiratory disorders and examined bacteriology.

Table (2), showing (75) Samples were
bacteriologically positive (50%), 50
from them showed single cultures
(33.3 %) and 25 mixed cultures (16.7
%).

Table (3), revealed The bacteriological examination of (150) nasal samples collected from lambs showing respiratory manifestation revealed that Escherichia coli, pseudomonas aerogenosa and staphylococcus the epidermidis were most predominant bacteria. As Escherichia coli was isolated from 30 samples (20%), pseudomonas aerogenosa and staphylococcus epidermidis as each of them were isolated from 14 (9.3%) followed by proteus vulgaris and staphylococcu aereus as each them were isolated from 10 (6.7%). proteus mirabilis and streptococcus sp bacteria as each of them were 8 (5.3%), klebsiella isolated from subsp ozaenae was isolated from 7 (4.7%), klebsiella pneumoniae subsp pneumoniae and enterobacter

agglomerans as each of them were isolated from 5 (3.4%), citrobacter diversus from 6 (4%), enterobacter cloaca was from 4 (2.6%) and proteus myxofaciens, providencia and pasteurella multocia as each of them were isolated from 2 (1.3%).

Table (4), showed that E.coli were the most isolate as a single culture were isolated from 17 samples (11.3 followed by staphylococcus %) epidermidis from 11 samples (7.3%), pseudomonas aerugenosa from 10 samples (6.7 %), streptococcus sp from 4 samples (2.7%), staph. aureus from 3 (2%), klebsiella pneumoniae subsp pneumoniae and pasteurella multocida as each of them were isolated from 2 (1.3%) and Enterobacter agglumerans from one samples (0.7). The total isolates 50 isolates (33.3 %).

Table (5), showed that mixed cultures of total number of isolation from pneumonic lambs was 25 isolates (16.75%)E.coli, Proteus were Staphylococcus vulgaris and aurous were isolated from 7 samples (4.7 %), Proteus vulgaris Klebsiella ozaenae and were 3 (2%), isolated from Proteus mirabilis. Citrobacter ,Enterobacter from (2%), E.coli, cloaca 3 Enterobacter agglumerans and ozaenae were isolated Klebsiella from 3 (2%), Proteus mirabilis, Pseudomonas Citrobacter aerugenosa and Staph . epidermidis were isolated from 3 (2 %). E.coli, Previdencia and Streptococcus sp were isolated from 2 (1.3%). Klebsiella Proteus mvxofaciens. pneumoniae pneumoniae subsp and streptococcus were sp isolated from 2 (1.3%). proteus mirabilis, pseudomonas aerugenosa,

enterobacter agglumerans and klebsiella pneumoniae subsp pneumoniae were isolated from 1 proteus mirabilis E.coli (0.7%). Klebsiella ozaenaeand and Enterobacter cloaca were isolated from one samples (0.75%)

These finding in general are somewhat similar to those obtained by El-Sherif and Abd-El-Ghani (1974) who found that from 157 sheep with respiratory disease, 135 contained specific bacteria. pasteurella multocida was recovered at percent of (3.9%). Esherichia coli representing 12%. Pseudomonas aeroginosa were also recovered at percent of (7.352%). Staphylococcus epidermidis have been isolated with staphylococcus alpha-haemolytic aureus and streptococci from diseased sheep. The percent of last two bacteria were 6.2% 8.5%, and respectively. Corynebacterium pyogenes have been isolated with unclassified Corynbacterium at percent 7% and 6.2% respectively. Gameel et al. (1991) isolated Klebsiella pneumoniae from sheep suffering from pneumonia. Elyas (1993) isolated Staphylococcus aureus, staphylococcus epidermidis, and Streptococcus pneumoniae from 60 affected sheep at percent 6%, 5%, 8%, and respectively. Corynebacterium pseudotuberculosis also have been isolated at percent of 8%. On the other hand Escherichia coli. Klebsiella pneumoniae, and Pseudomonas aeruginosa were also isolated at a percentage of 3%, 7%, and 7%, respectively.

On other hand these findings were contrast with **Alley**, (1975) who recorded that The percentages of mannheimia haemolytica (pasteruella) isolated from 184 normal sheep and 246 sheep with subacute and chronic forms of pneumonia were 73% from the nasal cavities, 5% from trachea and 5% from lung of normal sheep, and 78% from the nasal cavities, 54% from trachea and 59% from lung of pneumonic sheep. Also Escherichia coli have been recovered from normal and diseased sheep suffering from pneumonia. Kaya and Erganis (1991) isolated pathogenic bacteria from 57 out of the 102 affected adult sheep and 41 out of 50 affected lambs with pneumonia. The commonest bacteria isolated were Pasteurella haemolytica (16) isolates and Pasteurella multocida (7) isolates. Staphylococcus aureus was recovered alone or in combination with other organism from 23 cases. Also Klebsiella pneumoniae have been isolated from 15 out of 41 pneumonic lambs. Blanco-viera, et al. (1995) reported that about 112 strains of pasteurella spp. were isolated from cattle and sheep. Forty isolates were identified as mannheimia haemolvtica (pasteruella) and 72 isolates as pasteurella multocida. 100% of mannheimia haemolytica (pasteruella) belonged to biotype A. But 61% of multocida pasteurella isolates belonged to type A, 25% to type D and 14% were untypified. Bouljihad and Leipold (1995) reported that about 64 sheep suffering from respiratory disorders,were examined bacteriologically the bacteriological examination resulted in isolation of pasteurella multocida from 55 sheep mannheimia haemolytica and (pasteruella) from 9 sheep. Mohamed (2002) and Shaker isolated mannheimia haemolytica (pasteruella) from apparently healthy and diseased sheep at percent of 56% and 75%,

respectively. And 66% and 90% from emergency slaughtered and dead sheep, respectively. Those studies showed that the pasteurella multocia and mannheimia haemolytica (pasteruella) were the predominant bacteria contrast to our study may be due to pasteuralla sp be located at intervals in nasal cavities several studies indicated the high incidence of Escherichia coli isolate and pseudomonas aerugenosa from nasal swabs of pneumonic lungs. (Sayed 1996) and (Quinn et al 1994) cited that Pseudomonas aerugenosa Bordettella pyogens parapertusis actinomysis and klebsiella sp were implication as causes of pneumonia among and lambs adult sheep concentration of her member of Enterobacteriaceae and staphylococcus epidermidis, their role in the pathogenesis of lambs pneumonia is unkown But may indicated poor hygienic practice or factores other such as overcrowding bad climates or method husbandry regarding of single and mixed cultures. Escherichia coli and pseudomonas aerugenosa the were most frequent bacteria isolated as а single and mixed pathogen as shown in table (2,3,4).

Table (6), showed bacteriologicalexamination for (14) lung samplesrevealed that staphylococcus aureusand Escherichia coli were isolatedfrom two samples (14.2) % and otherbacteria;- mannheimia haemolytica(pasteruella),aerugenosa,Enterobacteragglumerans, Haemophillus somunsevery one of them were isolatedfrom one sample (7.1%).

actinomyces pyogenes was isolated from three samples (21.3%).

Table (7), showed that the bacterial isolates from lung samples were single such as actinomyces pyogens from two samples (14.2), mannheimia haemolytica (pasteruella), haemophillus somnus Enterobacter agglomerans, Staph .aureus every one of them was isolated from one samples (7.1%). Except two isolates were isolated from mixed cultures Escherichia coli + Actinomyces pyogenes and E.coli +Staph aureus, every isolates was isolated from one sample (7.1%).

The important of actinomyces and Escherichia coli. pasteurella. haemophillus influenziae as causative agents of pneumonia among sheep was fullv described by Shurma woldehivet (1990), and Elyas (1993). Who compare the bacterial groups which were isolated from nasal swabs of lambs and those from pneumonic isolated lungs. He noticed that prevalence of Escherichia coli and pseudompnas areugenosa, staphylococcus epidermidis following proteus vulgaris and staphylococcus aureus were high in nasal swab samples of affected lambs in contrast to lung samples where it was not found also Haemophillus somnos was not isolated from swabs samples.

Table (8) showed that, Sensitivity test revealed that most isolates were highly sensitive to Enrofloxacin, Cephaloexine, and Ampicillin and were resistant to streptomycin

REFERENCES

Alley, M. R. (1975): The bacterial flora of the respiratory tract of normal and pneumonic sheep. New-Zealand Vet. J. 23, 113-118.

Blanco-viera, F. J., Trigo, F. J., Jaramillo-Meza, L. and Aguilar-Romero, F. (1995): Serotypes of pasteurella multocida and Pasteurella haemolytica from pneumonic lesions in cattle and sheep from Mexico. Rev Lantinoam Microbial. 37(2), 121-126.

Bouljihad, *M.* and Leipold, *H.W.* (1995): Preliminary pathological observations of sheep with chronic progressive pneumonia. Agri-Practice 16(2), 25-27.

Cruickshank ,R.; Duguid,J.P; and swain ,R.H.A.(1975): Medical microbiology . vol.2,the practice of medical microbiology .12thEd .churchill livingstone ,Edinburgh .london .

Davies D.H (1995) pasteurellosis of sheep .in progress in vet .microb Imm.vol .229 Basel;karger .

El-Sherif, M.T. and Abd El-Ghani, M. (1974): Some studies on the respiratory affections of lambs. Assiut Vet. Med. J. 1(1-2), 199-211.

Elyas , A.H(1993): some studies on sheep pneumoniae of bacterial and fungal origin A ssuit .Vet. Med. J.(29),89-95.

Finegold, E.M and Martin, W.J. (1982): Baily and Scott diagnostic microbiology 6th the C.V; mosby C, Torento London.

Gameel, A. A., EL-Sanousi, S. M.; Al-Nawawi, F. and Al-Shazly, M. O. (1991): Association of Klebsiella organisms with pulmonary lesions in sheep.Revue d'Elevage et de Med. Vet. Des pays Tropicaux. 44(2), 161-164.

Hatem M, E, Mona, S. Zaki Osman A, H. and Mona El Shabrawy (2003): bacteriological, histopathologycial and clinical pathological studies on respirotary affection in sheep and goats in Egypt. J. Egypt Vet. Med. Assoc, 63(1): 97-109.

Ibrahim E.mohamed and mokhtar A ;(2003) pathology and bacteriology studies on out break of pneumonic pasteurellosis in sheep at sharkia proviance ; Egypt J.Agric Res .81 (1):32-49.

Kaya, O. and Erganis, O. (1991): A etiological survey of pneumonia in sheep and lambs.Veterinarium 2(3-4), 27-29.

Martin ,W.B.(1996): Respiratory infection of sheep comp. immune microb infec.Dis.19.

Mohamed, S.R. and Shaker, M.H. (2002): Immuno-etiological studies on Pasteurella haemolytica in sheep. Vet. Med. J. Giza. 50(4), 695-705.

Quinn,p.j;catrter,M.F;Markey,Bandcarter,G.R.(1994):clinicalveterinarymicrobiology16^stEd.mosby.yearBook.Europelimited

Radwan 1.A;Abd El-twab A,A. and Mona A. El shabrawy(2002):Atreatise on the Bacterial causes of pneumoniae in goat ;J.Egypt Vet ,Med Ass-c,63 (6):179-187

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Reed, W.P.(1973): Indolent pulmonary abscess associated with klebsiella and Enterobacter . AM Rev Res ; 107: 1055-1059.

Radostits OM, Blood DC and Gay CC (1995): Veterinary Medicine. A textbook of the Diseases of Cattle, Sheep, Pigs, Goat and Horses. Bailliere Tindall, London.

Sayed, **A.M**. (1996):Some bacteriological and mycological studies on sheep pneumonia at Assiut governorate .Assiut Vet .Med ., 36(71):68-73. Sharma ,R. and Woldehiwet

,z.(1990) : Increased susceptibility to pasteurella haemolytica in lambs infected with bovine respiratory syncytial virus

.j.comp.pathol.,103:411-420.

Woldehiwt, Z., Hamache B. and Rowan T.G. (1990): the effect of age, environmental temperature and relative humidity on the bacterial flora of the upper respiratory tract in calves. Br. Vet. J. 146:211-215.

| Table (1). Marbidity | | fotoliti i rotoo a | |
|-----------------------|-------------------|--------------------|-------------------|
| Table (T). Morbially, | monality and case | e ratality rates c | prieumonic lamos. |

| No .of sheep | pneumonic sheep no. | morbidity rate | | Mortality rate | | Fatality rate | |
|-----------------|------------------------|----------------|-----|----------------|------|---------------|------|
| | No. | % | No. | % | No. | % | |
| 2390 | 150 | 150 | 6.2 | 179 | 7.4% | 29 | 19.3 |

Table (2): No of positive bacterial isolates from nasal swabs collected from pneumonic lambs.

| Total no | Negative samples | | positive samples | | | |
|--------------------|------------------|-------|------------------|----|------|-------------|
| of Samples | No. | No. % | | | | % |
| 150 nasal swabs | 75 | 50 | | 75 | 50 | |
| | | | Mixed cultures | | Sing | le cultures |
| | | | NO. % | | NO. | % |
| | | | 25 16.7 | | 50 | 33.3 |

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| Bacterial | No. of | No. of isolates | % |
|------------------|---------|-----------------|-----|
| Isolates | samples | | |
| Escherichia coli | 150 | 30 | 20 |
| Proteus | | 10 | 6.7 |
| vulgaris | | | |
| Proteus | | 8 | 5.3 |
| mirabilis | | | |
| Proteus | | 2 | 1.3 |
| myxofaciens | | | |
| Enterobacter | | 4 | 2.6 |
| cloaca | | | |
| Enterobacter | | 5 | 3.4 |
| agglumerans | | | |
| Klebsiella | | 4 | 3.4 |
| pneumoniae | | | |
| subsp | | | |
| pneumoniae | | | |
| Klebsiella | | 7 | 4.7 |
| subsp | | | |
| ozaenae | | | |
| Staphylococcus | | 10 | 6.7 |
| aereus | | _ | - |
| Staphylococcus | | 14 | 9.3 |
| epidermidis | | | |
| Streptococcus | | 8 | 5.3 |
| SD | | _ | |
| Citrobacter | | 6 | 4 |
| diversus | | - | - |
| Pasteurella | | 2 | 13 |
| multocida | | _ | |
| Providencia | | 2 | 1.3 |
| Pseudomonas | | 14 | 9.3 |
| aerogenosa | | | 0.0 |
| Total isolates | | 126 | 84 |
| | | .20 | |
| | | | |

Table (3) bacterial isolatesfrom nasal swabscollected from pneumonic lambs.

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Table (4): Single cultures of bacteriological examination of nasal swabs collected from pneumonic lambs.

| Bacterial isolates | No. of samples | No. of isolates | % |
|---|----------------|--------------------|------|
| Escherichia coli | 150 | 17 | 11.3 |
| Pseudomonas aerogenosa | | 10 | 6.7 |
| Staphylococcus epiderimidis | | 11 | 7.3 |
| Staphylococcus aereus | | 3 | 2 |
| Klebsiella pneumoniae subsp pneumoniae | | 2 | 1.3 |
| Enterobacter agglumerans | | 1 | 0.7 |
| Pasteurella multicida | | 2 | 1.3 |
| Streptococcus sp | | 4 | 2.7 |
| Total no. of isolates | | 50 | 33.3 |

Table (5) Mixed cultures ofbacteriological examination of nasal swabscollectedfrom pneumonic lambs.

| Bacterial isolates | No. of samples | No. of isolate s | % |
|--|----------------|------------------------|-----|
| Proteus vulgaris +Escherichia coli | 150 | 7 | 4.7 |
| Staph. aureus+ | | | |
| Proteus vulgaris + klebsiella ozaenae | | 3 | 2 |
| Proteus mirabilis + citrobacter +entero- bacter | | 3 | 2 |
| cloaca- | | | |
| Escherichia coli +enterobacter agglomerans + | | 3 | 2 |
| Klebsiella ozaenae | | | |
| Proteus mirabilis +citrobacter +pseudomonas | | 3 | 2 |
| aerugenosa + staphylococcus epidermidis | | | |
| Escherichia coli+ providencia + streptococcus sp | | 2 | 1.3 |
| Proteus myxofaciens + klebsiella pneumoniae + | | 2 | 1.3 |
| Streptococcus sp | | | |
| Proteus mirabilis + pseudomonas aerogenosa + | | 1 | 0.6 |
| Enterobacter agglomerans +klebsiella pneumoniae | | | 6 |
| Proteus mirabilis + Escherichia coli+ klebsiella | | 1 | 0.6 |
| Ozaenae +enterobacter cloaca | | | 6 |
| Total no. of isolates | | 25 | |

| Bacterial isolates | No. of samples | No. of isolates | % |
|--------------------------|----------------|-----------------|------|
| Pasteurella haemolytica | 14 | 1 | 7.1 |
| Staphylococcus aereus | | 2 | 14.2 |
| Escherichia coli | | 2 | 14.2 |
| Enterobacter agglomerans | | 1 | 7.1 |
| Actinomyces pyogenes | | 3 | 21.3 |
| Pseudomonas aerugenosa | | 1 | 7.1 |
| Haemophillus somnus | | 1 | 7.1 |
| Total of isolated | | 11 | |

Table (6): Bacterial isolates of lung samples collected from pneumonic lambs.

Table (7): Single and mixed cultures of bacteriological examination of lung samples collected from pneumonic lambs.

| Bacterial isolates | No. of samples | No. of isolates | % |
|--|----------------|-----------------|------|
| Actinomyces pyogenes | | 2 | 14.2 |
| Pasteurella haemolytica | | 1 | 7.1 |
| Pseudomonas aerugenosa | | 1 | 7.1 |
| Haemophillus somnus | | 1 | 7.1 |
| Enterobacter agglomerans | | 1 | 7.1 |
| Staphylococcus auereus | | 1 | 7.1 |
| Escherichia coli + actinomyces pyogenes | | 1 | 7.1 |
| Escherichia coli + actinomyces pyogenes | | 1 | 7.1 |
| Total of isolated | | 9 | 64.3 |

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| Bacterial isolates /antibiotics | amoxy cillin | ampi cillin | cepha loxin | Enrof Ioxin | erythro mycin | genta mycin | oxytetra cycline | pene cillin | strepto mycin |
|---------------------------------------|-----------------|----------------|----------------|----------------|------------------|----------------|---------------------|----------------|------------------|
| Staph .aureus | S | SS | SSS | S | SSS | R | S | R | SS |
| Staph .epidermidis | S | SS | SSS | S | SSS | R | S | R | S |
| Strepto. Sp | S | SS | SSS | SS | SSS | R | S | R | R |
| E ,coli | R | SS | SSS | R | SSS | S | S | R | R |
| Klebsielliae pneumoniae | R | S | SS | S | SS | S | R | R | R |
| Proteus vulgaris | R | SSS | SSS | S | SSS | S | S | SS | SS |
| P . haemolytica | R | S | SSS | R | SSS | R | R | R | R |
| P . multocida | R | S | SSS | S | SSS | S | SS | S | S |
| Citrobacter sp | S | SS | SSS | S | SSS | R | SS | R | SS |
| Enterobacte r aerogenes | R | SSS | SSS | S | SS | R | SS | S | S |
| Pseudomon as aeruginosa | R | SS | SSS | R | SS | R | R | R | R |
| R- resistant | | | S- ellecor | vriblity | | | | | |

Table (8): sentivity of bacterial isolates to antibiotics.

Staph- Saphylococcus

Srept= Streptococcus

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