Immunomodulating effect of B-glucans and mannan oligosaccharide on broiler chicks vaccinated with Newcastle disease virus

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This study was carried out to determine the immunomodulating effect of β-glucans and mannan oligosaccharide (MOS) on the immune response of chickens to Newcastle disease vaccine. The results showed that birds received β-glucans and MOS having higher average body weights values and significantly higher ND HI antibody titer than the other non medicated groups. Thymus, spleen and bursal indices of control negative showed significantly lower values than vaccinated medicated and non-medicated groups. Both total and differential leukocytic and lymphocytic counts showed significantly higher in medicated group than other groups. Liver function test showed lower AST and ALT in medicated group than other groups. Results of challenge test with NDV confirmed that MOS and B glucans immunostimulant improved protection rate by 15% in medicated than non-medicated ones. In conclusion MOS and B glucans can be given to chicken to improve both body weight and protection against VV NDV challenge that predominated in Egypt.

Commercial poultry flocks receive a lot number of vaccines to protect them from environmental pathogens; therefore, a great effort had been expanded to develop strategies to enhance chicken immune response, especially in facing immunosuppression caused by extraneous agents, infections, intoxication or by certain vaccine viruses. Immunomodulation could improve vaccinal immunity and possibly selectively promote responses that are critical for protection.

Immunomodulators usually classified according to their origin into biological and chemical products (Poli, 1984). This classification further broken down into physiological products, substances of microbial origin and synthesis compounds.

The mannan-oligosaccharide (MOS) is derived from the outer cell wall of yeast, and its evaluation in diets for breeders is of particular interest because it not only shifts gastrointestinal microflora balance toward beneficial organisms (Spring et al., 2000; Fairchild et al., 2001) but also has immunomodulatory properties (Cotter et al., 2002). The yeast cell wall has powerful antigenic stimulating properties, and it is well established that this property is a characteristic of the mannan chain (Ballou, 1970). This study

was carried out to determine the immumostimulant effects of commercial feed additive preparations containing a mannanoligosaccharide plus β-glucans on chicken, performance and immune response to ND vaccine. Body weight gain, HI and challenge With NDV that endemic in Egyptian poultery farms as well as bursal, thymic and spleen body weight ratio were taken as criteria for evaluation based.

Materials and methods

Immunostimulants (ALPHAMUNE®). It's a commercial feed additive product composed of (1-3, 1-6) β-glucans and (MOS) obtained from Alpharma Animal Health. USA (patch NO AG51242). It was used in ration at a rate of 500 gram/ ton of finished fed.

Chickes. A total number of 225 one day-old commercial (white HI-line rooster) chickens obtained from El-Wady Company were used in this study.

Newcastle disease (ND) vaccinal strains. 1-Hitchiner B₁ and La Sota strains, produced by Pfizer International Company, USA with each vial contain virus titre of 10⁹ EID₅₀ was used after titration for vaccination of experimental chicks via eye instillation route.

Clone 30. Vaccine nobilis clone 30 (Lot No: 06829AJ01, Intervet international B.V. Boxmeer – Holland) with virus titer of 10⁶ EID₅₀ was used for vaccination of experimental chicks via eye instillation

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Velogenic NDVs. A local velogenic viscerotropic Newcastle disease virus (vvNDV) isolate (Shible and Reda, 1976) was kindly supplied by Newcastle Diseases Department; Veterinary Serum & Vaccine Research Institute, Abbasia, Cairo. Egypt were used for challenge test.

Haemagglutination (HA) and haemagglutination inhibition (HI) tests. HA and HI test were carried out according to (Anon. 1971).

Haemtological studies. Total leukocytic count was counted according to the technique described by (Nutt and Herrick, 1952) while differential leukocytic count was done by the standard method of Battelnent described (Schalm, 1973).

Biochemical analysis. ALT and AST were done according to the method of (Reitman and Framkel, 1957). Serum uric acid was done according to the method descried by (Barham and Trinder, 1972) and serum creatinin was done according to the method described by (Houot, 1985).

Bursa body weight index. It was calculated according to (Ying et al., 2003) as following: Bursa: body weight ratio = bursa weight/ body weight. Bursal index = Bursa: body weight ratio X 1000.

Challenge test. Chickens were challenged via intramuscular route. Each chicken received a dose of 0.5ml / bird containing 10⁶ EID_{50 VV NDV} according to (Afify, 1990). Birds with persisted symptoms till the end of the observation period were considered as if dead.

Statistical analysis. Statistical analysis of variance (ANOVA) test was used to estimate differences among treatments according to (Steel and Torrie, 1960).

Experimental design. The used chicks (225) were floor reared and fed on balanced commercial ration free from antimicrobial agents. At the 1st day of life, 5 chicks were sacrificed for organ body weight ratio and serum while the rest of obtained chicks were divided into 4 groups (1-4). Groups (1 and 3) containing 60 chicks each, while groups 2 and 4 containing 50 chicks each. Each group was kept in a separate clean disinfected room.

Chicks of groups 1 and 3 were feed on ration without additives, while those of groups 2 and 4 were feed on ration supplemented with the immunostimulant (Alphamune®), in dose of 0.5 gm/ kg. At the 5th day of age all chicks were S/C

vaccinated with inactivated avian influenza (H5N1) (0.3 ml/ bird). At the 7th day of age, chicks of group (1 and 2) were kept ND nonvaccinated control while birds of groups (3 and 4) were vaccinated each with 10⁶ EID₅₀ Hitchiner B1 via eye instillation revaccinated at the 18th day of age as each bird was given 10⁶ EID₅₀ La Sota via eye instillation route. At 35 day of age 20 birds from groups were separated and challenged with 0.5 ml containing 106 VVND. Challenged chickens were kept under daily observation for 21 days with daily record of symptoms, deaths and post - mortem lesions. Ten birds from group 1 and 3 were left without challenge to be control.

Experimental chicken groups were weekly subjected to the following: Life body weight of random 5 birds / group as well as weight of bursal, thymus and spleen of each bird was recorded to calculate organ body weight ratio. Random 5 non-coagulated blood samples on EDTA were collected for total anddifferential leukocytic count. Random coagulated 5 blood samples from wing vein were collected for serum collection. The collected serum samples were divided into two equal quantities, labeled and stored at-20°C until use. The collected sera were tested for detection of NDV HI as well as liver and kidney function test.

Results and Discussion

numbers There are large of immunostimulatory components were reported to be used for stimulating the chicken immune response to face the problem of vaccination failure, which constitute a challenge to poultry industry all over the world. The application of immunostimulant is not only to raise resistance of birds but also to improve their immune response to vaccination (Afify, 1990; Awaad et al., 2000). The work was designed to evaluate the effect of Alphamune as immunomodulator in chickens, where data presented in (Table 1) showed that administration of Alphamune was significantly increased body weight at 7 days old 80.30 ± 1.88 gm verses 72.90 ± 1.58 for control group. While from 14 to 35 days there is no significant difference could be detected values of different groups as well as that of control group. This result agrees with those of Solis de los (Santos et al., 2007) where a significant weight difference at 7 days only between treated and non treated poults was found while no difference at 3 weeks old.

Table (1): Effect of immunostimulant on average body weight of ND vaccinated and non-vaccinated chickens.

Group	Tre	atment		Age / week								
No.	I.S	Vace	0	1	2	3	4	5				
1	-	. •	36.39±1.2	72.90±1.58	130.00±2.31	221.80±8.40	319.30±4.29	428.40±9.93				
2	+	•		80.30±1.88*	133.00±3.21	228.50±5.5	332.00±5.09	446.25±17.95				
3	-	. +			135.40±2.85	225.00±4.18	325.60±5.26	428.60±15.93				
4	+	+			137.60±2.35	229.60±5.28	343.50±12.04	467.50±14.16				

Each value represents mean ±S.E.

Table (2): The effect of immunostimulant on mean ND HI antibody titer in vaccinated and non-vaccinated chickens.

Group	Treatment		Age / week							
No.	I.S	Vacc	. 0	1	2	3	- 4	5		
-1		-	7.8±86	6 ± 0.32	#4±0.55 b	# 2.4±0.51b	# 1.6±0.51b	# 1.6±0.51b		
2	+			6.2 ± 0.37	4.4± 0.51b	2.8 ± 0.37 b	$1.8 \pm 0.37 b$	$1.8 \pm 0.55 b$		
3	•	+		6 ± 0.32	6.4± 0.51a	$7.2 \pm 0.58a$	6.6± 0.69a	$5.6 \pm 04a$		
4	+	+		6.2±0.37	7.2±0.37a	7.8± 0.58a	7.2± 0.37a	6.2± 0.49a		

Each value represents mean ±S.E.

#: Significant variation between groups by ANOVA test at P≤0.05.

Different superscript letters a and b denote significant variation respectively by LSD at P≤0.05.

Table (3): Effect of immunostimulant on thymus index of vaccinated and non-vaccinated chickens with NDV live vaccine.

Group	Treatment		Age / week							
No.	I.S	Vacc.	0	1	2	3	4	5		
1	-	· · ·	4.60±0.12	5.60±0.06	#6.00±0.10b	#6.10±0.08b	#6.70±0.12c	#5.60±0.18c		
2	+	. •		5.9±0.07*	6.60±0.09a	7.21±0.15a	7.65±0.23b	5.80±0.20b		
3	-	. +			6.63±.0.11a	7.25±0.20a	7.91±0.22ab	6,33±0.21ab		
4	+	+			6.71±0.10a	7.50±0.2a	8.35±0.30a	6.50±0.25a		

Each value represents mean ±S.E.

Different superscript letters a, b and c denote significant variation respectively by LSD at P≤0.05.

Table (4): Effect of immunostimulant on mean spleen index of vaccinated and non-vaccinated chickens with NDV live vaccine.

Group	Trea	tment		Age / week						
No	_I.S	Vacc.	0	1	2	3	4	5		
	-		0.50±0.01	1.00±0.05	#1.50±0.08c	#1.90±0.10c	#2.05±0.08c	2.30±0.05		
2	. +		-	1.35±0.07*	1.70±0.07c	2.10±0.10bc	2.10±0.15c	2.34±0.06		
3	-	+			2.30±0.11b	$2.46\pm0.12b$	2.50±0.11b	2.40 ± 0.09		
4	+	+	. ·	· ·	2.63±0.15a	2.87±0.17a	2.90±0.18a	2.44±0.15		

Each value represents mean ±S.E.

Different superscript letters a, b and c denote significant variation respectively by LSD at P≤0.05.

Table (5): Effect of immunostimulant on mean bursal index of vaccinated and non-vaccinated chickens with NDV live vaccine.

Group	Treat	ment		Age / week						
No.	I.S	Vacc.	. 0	1	2	3	4	5		
1	-	-	1.80±0.08	2.50±0.10	#2.80±0.15c	#3.20±0.12b	#2.41±0.15c	#1.55±0.11b		
2	+ .	-		2.85±0.12*	2.90±0.14c	3.40±0.12b	2.60±0.13bc	$1.85\pm0.12b$		
. 3		+	e produce in the		3.20±0.13b	3.80±0.14ab	3.00±0.19b	2,50±0.15a		
4	+	. +			$3.43\pm0.13a$	4.10±0.18a	3.55±0.21a	2.75±0.20a		

Each value represents mean ±S.E.

Different superscript letters a, b and c denote significant variation respectively by LSD at P≤0.05.

^{*} Significant difference between groups by t-student test at P≤0.05.

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^{#:} Significant variation between groups by ANOVA test at P≤0.05.

Table (6): Effect of immunostimulant on mean tot	al leucocytic count X 103 of vaccinated and non	-
vaccinated chickens with NDV live vaccine.		

Group	Trea	tment	Age / week									
No.	I.S	Vacc.	0_	1	2	3	4	5				
1	•	•	16.1±1.30	14.3±1.10	#11.6±0.50b	#12.1±0.74b	#13.1±0.51b	#13.2±0.9b				
2	+	-		22.5±1.30**	17.3±1.30ab	15.6±0.75ab	14.8±0.8ab	13.4±0.95b				
3	-	, +			20.6±1.15a	21.5±1.20a	22.2±1.23a	21.1±1.25b				
4	+	+			26.2±1.20a	28.5±1.9a	32.3±1.7a	30.6±2.3a				

Each value represents mean ±S.E.

** Significant difference between groups by t-student test at P≤ 0.01.

#: Significant variation between groups by ANOVA test at $P \le 0.05$.

Different superscript letters a, b and c denote significant variation respectively by LSD at P≤0.05.

Regarding thymus index, spleen index and bursal index of chicken fed on Alphamune supplemented ration and vaccinated with NDV revaled significant increase in values than results of other groups where it give 6.50 ± 0.25 , $2.44 \pm$ 0.15 and 2.75 ± 0.20 respectively at 35 days of age (Table 3-5). These results come in agreement with the finding of Ying et al., (2003) where mean percentage of organ body weight ratios of liver, spleen, Kidney, thymus and bursa of Fabricius exhibited a significant (P<0.05) increase in MOS as compared to those of control group. Results of total and differential leucocytic count (Table 6, 7) were significantly higher TLC on group 4 at 35 days of age $(30.6 \pm$ 2.3) in comparison to $(13.2 \pm 0.9, 13.4 \pm 0.95,$ 21.1 ± 1.5) for the other groups. It was observed that the source of increased in TLC is the significantly increased lymphocyte counts due to use of Alphamune®, Increased TLC in group 4 can be attributed to immunostimulation effect of compounds of Alphamune. This result was previously observed by (Fleischer et al., 2000; Acevedo et al., 2001) who recoded increased TLC with administration of MOS and b-glucan respectively. Chicken group 4 that fed on Alphamune® supplemented ration and vaccinated with NDV vaccine showed significant lower AST and ALT levels at 35 days of age (Table 8, 9). Where the results is $172.17 \pm$ 7.15 and 9.85 ± 0.20 respectively verses 201.56 \pm 7.53 and 14.3 \pm 0.25 in untreated vaccinated group . This result was observed by (Santhosh et al., 2003) in treated group with MOS all over the breeding period.

Statistical analysis of uric acid and creatinine values (Table 10, 11) resulted in non significant difference between different groups up to 35 days of age. The instability in creatinine value from week to week may be related to change in feed and protein concentration. Birds of group (4) showed significant HI titers to NDV at 35

days of age than other groups (Table 2); this higher HI titers resulted in 95% protection in this group 4 compared to 85% protection in group 3, 20% protection in group 2 and 0% protection in group 1 (Table 13). Our results clearly showed the specific immune stimulation and protection against challenge in group 4 were attributed to B-glucan compound of Alphamune due to increasing functional activity of macrophage and neutrophils. Yun et al., (2003); Sakurai et al., (1992) reported that orally B-glucan indirectly stimulate the immunity in the respiratory system of mice by activating macrophage in the payer's patches of the gut.

From the above discussed data we could conclude that Alphamune® could increase body weight gain, improve immunity of the birds and decrease susceptibility to NDV challenge.

References

Acevedo, A. M.; Pedroso, M. and Miranda, I. (2001): Effect of treatment with lineal particulate beta1-3 glucan by oral route on humoral response to Newcastle vaccine in chickens. Revista-Cubana-de-Ciencia-Avicola. 25(2):107-112;

Afify, M. A. (1990): Studies on the role of some immunostimulants in using poultry vaccines. Ph. D. Thesis, Fac. Vet. Med., Cairo Univ., Egypt.

Afify, M. A.; Elshazly, O. A.; Zouelfakar, S. A.; Shaheed, I.B. and Awaad, M. H. H. (2003): Immunopotentiation properties of Propionibacterium granulosum versus vitamin E in cyclophosphamide immunosuppressed chickens. Vet. Med. J. Giza, 51(2): 257-272.

Anon, (1971): Methods for examining poultry biologics and for identifying avian pathogens. Nat. Acad. Sci., Washington, D.C.

Awaad, M. H. H.; Zouelfakar, S. A.; EL-Shazly, O. A.; Afify, M. A. and Shaheed, I. B. (2000): Immunomodulatory properties of inactivated probacterium granulosum (IMR). I.In non-immunosuppressed chickens. J. Egypt. Med. Assoc., 66(7):137-148.

Ballou, C. E. (1970): A study of the

Table (7): Mean differential leucocytic count in immunomodulator medicated vaccinated or non-vaccinated chickens with NDV live vaccine.

Age								Group No.	and treatme	nt						
1	Group 1: Control Group 2: I.S					Group 3: vaccinated				Group 4: I.s +vaccination						
week	H	L	M	E	Н	L	M	E	Н	L	M	E	н	L	M	E
0	20.5	74.7	2.9	1.9	20.5	74.7	2.9	1.9	20.5	74.7	2.9	1.9	20.5	74.7	2.9	1.9
1	25.8*±1.08	67.8±1.07	3.6±0,24	2.8±0.2	17.2±0.58	77.8*±0.37	3±0.32	2±0.32	25.8*±1.08	67.8±1.07	3.6±0.24	2.8 ± 0.2	17.2±0.58	77.8*±0.37	3±0.32	2±0.32
2	23.8±1.02	73.4±0.75	2.0±0.32	1.2±0.37	20.4±0.51	75.6°±0.4	2.2±0.37	1.8±0.2	45.8*±0.37	48.0±0.89	3.4*±0.24	2.8*±0.37	41.6*±0.51	52.1 ± 0.32	2.7±0.37	3.6°±0.4
3	20.4±0.81	76.4*±0.51	2.6±0.24	1.2±0.58	18.2±0.37	77.2*±0.37	3.4±0.6	0.6 ± 0.24	49.8*±0.37	43.6±0.24	2.8±0.37	4.0*±0.2	44.8*±0.37	49.6±0.24	2.2±0.37	3.4*±0.24
4	17.6 ± 0.51	76.8*±0.37	2.8±0.51	2.8±0.37	18.4 ± 0.51	77.6*±0.51	1.8±0.2	2.2±0.37	46.6*±0.75	48. 6± 0.51	2.2 ± 0.37	2.6±0.24	34.1°±0.32	59.5*±0.52	2.2±0.37	2.6±0.24
5	20.4±0.93	74.4±0.93	1.8±0.2	2.4±0.24	17.2±0.86	77.6*±1.12	2.4±0.37	2.8±0.37	42.0*±0.71	5 <u>2.2</u> ±0.58	1.8±0.2	3.6±0.37	33.6*±0.57	61.0*±0.71	1.8±0.2	3.4±0.4

^{*}Significant difference at P≤ 0.05 between treated and non treated groups.

Table (8): Effect of immunostimulant onmean of AST in sera of vaccinated and non-vaccinated chickens with NDV live vaccine.

C N-	Treatment			•	A			
Group No.	I.S	Vacc.	0	1	2	. 3	4	5
1	-	-	101.30±4.15	131.30±6.21	#155.00±5.90 a	#162.30±5.35 a	#174.55±5.84 b	#190.10±6.82 a
2	+	-	•	125.32±5.85	130.10±6.15 b	142.60±5.60 b	159.21±6.17 b	168.32±6.25 b
. 3		+ .			160.30±7.51 a	171.25±5.85 a	194.80±8.13 a	201.56±7.53 a
4	+ .	_ +			136.50±6.22 b	139.50±5.51 b	162.23±6.54 b	172.17±7.15 ab

Each value represents mean ±S.E.

Different superscript letters a,b and c denote significant variation respectively by LSD at P≤0.05.

Table (9): Effect of immunostimulant on mean of ALT in vaccinated and non-vaccinated chickens with NDV live vaccine.

Group No.	Trea	tment						
	I.S	Vacc.	0	1	2	3	4	5
1	-	-	4.92±0.15	7.90±0.20	#10.17±0.20 a	#12.77±0.22 a	#13.12±0.25 b	# 13.50±0.26 b
2	+	-		7.76±0.21	8.65±0.22 b	11.20±0.25 b	10.57±0.22 c	12.30±0.26 c
3	-	+			10.50±0.21a	12.89±0.26 a	14.05±0.27 a	14.30±0.25 a
4	+	+	<u></u>		8.10±0.18 b	9.60±0.20 c	9.70±0.21 d	9.85±0.20 d

^{*}H = Heterophils; L = Lymphocytes; M = Monocytes; E = Eosinophils.

^{#:} Significant variation between groups by ANOVA test at P≤0.05.

Table (10): Effect of immunostimulant on uric acid in sera of vaccinated and non-vaccinated chickens with NDV live vaccine.

Group No.	Tre	tment	. 14	Age / week						
	I.S	Vacc.	. 0	1	2	3	4	5		
1	-	-	5.30±0.13	6.10±0.15	6.30±0.12	6.42±0.20	6.20±0.21	5.90±0.20		
2	+	-		6.21±0.14	6.20±0.13	6.40±0.18	6.18±0.25	6.00±0.22		
3	-	+			6.50±0.17	6.48±0.19	6.51±0.23	6.25±0.20		
4	+	_+	_		6.40±0.16	6.41±0.20	6.48±0.25	6.20±1.54		

Table (11): Effect of immunostimulant on creatinine level in sera of vaccinated and non-vaccinated chickens with NDV live vaccine.

Gre	Group No.		itment		Age / week								
2000		I.S	Vacc.	0	1	2	3	4	5				
7	1 -	-	-	1.12±0.10	1.05±0.03	1.15±0.03	1.25±0.04	1.30±0.06	1.28±0.04				
r-=-	2	+	-	•	1.10±0.04	1.10±0.04	1.20±0.04	1.26±0.03	1.30 ± 0.04				
٠.	3	-	+			1.21±0.05	1.25±0.05	1.29±0.05	1.40±0.05				
	4	+	+			1.20±0.03	1.23±0.05	1.31±0.06	1.34±0.06				

Table (12): Daily distribution of morbidity and mortality in challenged chickens.

Group No.	Treatment			Days post-challenge								Total	%			
	I.S	Vacc.	Observation	1	2	3	4	5	6	7	8	9	10	11- 21		
1	-	-	Diseased No				5	7	4	2	1	-	-	-	19	95
			Died No.			3	4	5	5	2	1	-	-	- '	20	100
2			Diseased No				2	3	5.	3	2	2	1	-	18	90
2	т		Died No.				1	3	4	5	2	1	1	•	17	85
3	-	+	Diseased No				1	-1	2	1	1	-	-	-	6	30
			Died No.					1	2	-	-	-	-	-	3	15
4	+	-+ :	Diseased No					1	ı	1	1	-	-	-	4	20
4			Died No.						1	-	-	-	-	- '	1	5

Table (13): Results of VVND challenge test in immunostimulant medicated on vaccinated and non-vaccinated chickens.

Crown No.	Trea	atment	Total No achinda	No of dead binds	No. of assumband blands	Protection %	
Group No	I.S	Vacc.	- Lotal No of Diras	No of dead birds	NO 01 SURVIVED DIFUS		
1	-		20	20	0	0	
2	. +		20	. 17	3	15	
3	-	+	20	' 3	17	85	
4	+	+ '	20	1	19	95	

immunochemistry of three yeast mannans. J. Biol. Chem., 245: 1197-1203.

Barham, D. and Trinder, P. (1972): Enzymatic determination of uric acid analyst, 97:142-145.

Cotter, P. F.; Sefton, A. E. and Lilburn, M. S. (2002): Manipulating the immune system of layers and breeders: Novel applications for mannan oligosaccharides. Pages 21–28 in Nutritional Biotechnology in the Feed and food Industries. T. P. Lyons and K. A. Jacques, ed. Nottingham Univ., Press, Nottingham.

Fairchild, A. S.; Grimes, J. L.; Jones, F. T.; Wineland, M. J.; Edens, F. W.; and Sefton, A. E. (2001): Effects of Hen Age, Bio-Mos, and Flavomycin on Poult Susceptibility to Oral Escherichia coli Challenge. Poult. Sci., 80:562-571.

Fleischer, L. G.; Gerber, G.; Liezenga, R. W.;

Lippert, E.; Scholl, M. A. and Westphal, G. (2000): Blood cells and plasma proteins of chickens fed a diet supplemented with beta-D-glucan. Arch. of Animal Nutr., 53(1): 59-73.

Houot, O. (1985): interpretation of clinical laboratory test. cited By siest, G., Henny, J.; schiele, F. and Young, D. s. in panphlete of Bio Merieux, France.

Nutt, M. P. and Herrick, C. A. (1952): A new blood diluent for counting the leucocytes of the chickens. Poult. Sci., 31:735-738.

Poli, G. (1984): Immunomodulators. In Adjuvents, Interferon and Non-Specific Immunity, Eds Cancellotti, F.M. and Galassi, D., pp. 111-126. EEC, Luxembourg.

Reitmen, S. and Frankel, S. (1957): A calorimetric method for determination of serum glutamic-oxalacetic and glutamic-pyruvic transaminases.

Amer. J. of clinic. Pathol., 28, 56-63.

Sakurai, T.; Hashimoto, K.; Suzuki, I.; Ohno, N.; Oikawa, S.; Masuda, A. and Yadomae, T. (1992): Enhancement of murine alveolar macrophage functions by orally administered β-glucan. Int. J. Immunopharmacol., 14:821–830.

Schalm, A. (1973): Veterinary haematology 3rd Ed., Lea and Fabiger, Philadelphia.

Shible, A. and Reda, I. M. (1976): Cited by Khaphgy, A.U. (1977) thesis MVSc., Fac. Vet. Med., Cairo Univ., Egypt.

Solis de los Santos, F.; Donoghue, A. M.; Farnell, M. B.; Huff, G. R.; Huff, W. E.; and Donoghue, D. J. (2007): Gastrointestinal Maturation is accelerated in Turkey Poults Supplemented with a Mannan-Oligosaccharide Yeast Extract (Alphamune) Poult.

Sci., 86(5): 921-930.

Spring, P. C.; Wenk, K.; Dawson, A. and Newman. K. E. (2000): The effects of dietary mannanoligosaccharides on cecal parameters and the concentrations of enteric bacteria in the ceca of Salmonella-challenged broiler chicks. Poult. Sci., 79:205-211.

Ying, L.; YuMing, G.; JianMin, Y. and Wei, N. (2003): Effects of beta -1, 3/1, 6-glucan on performance and immune response of broilers. J. of China Agri. Univ., 8 (1): 91-94.

Yun, C. H.; Estrada, A.; VanKessel, A.; Park, B. C. and Laarveld, B. (2003): Beta-glucan, extracted from oat, enhances disease resistance against bacterial and parasitic infections. FEMS Immunol. Med. Microbiol., 35:67-75.

التأثير المناعي لمادة البيتا جلوكاتز والمانان اوليجو سكاريدز على الكتاكيت المحصنة بلقاح النيوكاسل

تم دراسة التأثير المتاعي لمادة البيتا جلوكاتر والماتان اوليجو سكارينزفي الكتاكيت المحصنة يلقاح النيوكاسل وأظهرت النتاتج أن الكتاكيت التي تم معاملتها بهذه المعاملات أعطت معدل أعلى في أوزان الجسم والفدة التيموسية والطحال و عَدة فابريشيس كما أظهرت المجموعة المعاملة بهذه المواد ممنوى أعلى في الكم والنوع في كرات الدم البيضاء وأظهرت القياسات للمجموعة المعاملة مستوى اقل من المجموعات الاخرى في إنزيمات الكيد كما أظهرت النتائج ارتفاع مستوى الأجسام المتاعية للقاح النيوكاسل في هذه المجموعة مما أعطى معدل حماية اعلى في اختبارا لتحدى. من النتائج بمكتنا أن نستنتج أن استخدام البيتا جلوكاتر والماتان يودى إلى تحسن في الأوزان وكذا معدلات الصد ضد عوى التحدي يقيرومن النيوكاسل شديد الضراوة والمنتشر في مصر