Histomorphometric Studies of the Urinary Tubules of the African Grasscutter (*Thryonomys swinderianus*)

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With 2 figures

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Abstract

The study was carried out to elucidate diameters the luminal of some segments of the urinary tubules in the grasscutter (Thryonomys swinderianus). Seven adult, apparently, healthy grasscutters were used in this study. They were anaesthesized with chloroform, dissected and the kidneys removed. The kidneys were fixed and processed by routine histological procedures. The various segments of the nephron were identified by their cell and their diameters types were measured. The data obtained were subjected to Pearson's correlation analysis. The diameters of the Bowman's capsule and glomerulus were 11.24 ± 0.460 and 9.21 ± 0.347 um, respectively. There was a very strong positive correlation between the diameters of the Bowman's capsule and those of the glomerulus (r = 0.975, P < 0.01). A significant positive correlation was also observed between the distal convoluted tubule and the thin segment of the loop of Henle (r = 0.786. P < 0.05), the collecting duct and the proximal convoluted tubule (r = 0.781, P < 0.05), and between the collecting duct and the thin segment of the loop of Henle (r = 0.821, P < 0.05). This study has attempted to present base-line data of the diameters of the various segments of the nephron.

Keywords

Histomorphometry, kidney, Urinary tubules, grasscutter

Introduction

The African grasscutter (Thryonomys swinderianus), also known as the greater cane rat and cutting grass, is the second largest rodent after the porcupine. It is found only in Africa (Adoun 1993) and it is a very good source of high-quality animal protein. Unfortunately, the animal has been aggressively hunted over the years, resulting in a depletion of the species. Commercial production of grasscutters as micro-livestock (National Research Council. 1991) and as laboratory animals (Asibey and Addo 2000) may be beneficial in conserving the species in countries where it is irrationally overhunted. The domestication of the rat has been with varying degrees of success (Jori et al., 1995). Various investigations have been carried out on the grasscutter all in an attempt to exhaustively understand the biology of the animal. There is no doubt that it is only after extensive research on the

special features of the biology of the rat that explanations to questions that may arise in the course of domestication and breeding of the grasscutter may be provided.

The primary function of the kidneys is to maintain proper balance of water and minerals (including electrolytes) in the body (Charmi et al., 2009). Additional functions include filtration and excretion of waste products from the processing of food, drugs, and harmful substances (toxins); regulation of blood pressure; and secretion of certain hormones (Cutler, 2006; Onveanusi et al., 2007). these functions All of contribute immensely to the maintenance of homeostasis. Certain features in the renal anatomy of different mammals and variations with the aridity of their habitat have been reported (Sperber, 1944). Studies have shown that animals from dry arid areas have a relatively well-developed renal medulla, which reflects their high capacity for reducing excretory water loss (Schmidt and O'Dell, 1961) as the length of the kidney medulla is suggestive of the length of the loop of Henle. Furthermore, Warui (1989) reported that the quantitative structural characteristics of the kidneys of mammals and birds may reflect the influence of environmental as well as many other factors on renal function. Although, the urinary tubules are the fundamental units of the organ structure. little is known about the mechanisms governing the size of the tubules (Germino, 2005). However, a good knowledge of their sizes is important as the morphology and function of the kidney have been modified through evolution to fulfil physiological different requirements (Charmi et al., 2009).

Baylis and Corman, (1998) reported that the rat provides a useful

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experimental model for the study of the mechanisms of renal ageing. Again, the first step to this, is studying the normal structures in their peculiarities and dimensions. There is a dearth of information on the microscopic quantitative characteristics of the renal structure of the African grasscutter.

The specific aim of this study was to obtain quantitative data on some metric proportions of the main structural component of the kidney of the African grasscutter (*Thryonimys swinderianus*).

Materials and Methods

Animal and tissue preservation

Seven adult grasscutters (Thryonomys swinderianus) were obtained from local hunters in Otukpo (6° 49' North, 8° 40' East) Benue State, Nigeria. They had a weight range of 1.5 to 3.5 kg. The rats were transported in a motor car to the Anatomy Research Laboratory of the Department of Veterinary Anatomy, Ahmadu Bello University, Zaria, using standard laboratory grasscutter cages. They were given access to fresh elephant grass (Pennisetum purpureum) and water ad libitum. The animals were anaesthesized with chloroform in closed aaseous а container, after which a simple exploratory laparatomy procedure was done to remove the kidneys. The whole kidneys were preserved in 10 % buffered neutral-formalin in the ratio of one part of tissue to ten parts of formalin to achieve adequate fixation. The kidneys were left in the fixative for 24 hours.

Tissue processing and staining

Each kidney was cut in the saggital plane. Tissue processing was done according to the method described by Gordon (1990), as outlined below:

The fixed tissues were dehydrated in an ascending series of graded concentrations of alcohol (70 %, 80 %, 95 % and 100 %) with a time interval of 1 hour for each stage of dehydration. The tissues were then cleared with xylene for about 2 hours, infiltrated with molten paraffin wax at 50 °C for 2 hours and embedded in paraffin blocks. Sections of 5 µm thick were then made using a 42339. microtome (Model Berlin, Germany), mounted on glass slides in the presence of egg albumin, deparafinized and dried at room temperature (25 °C). They were stained with hematoxylin and eosin (H & E), dehydrated, and coverslipped using Permount as the mounting medium and viewed under a light microscope (Olympus® CH20) at the magnification of x 400.

Measurements

The diameters of each kidney segments considered were measured using an ocular micrometer (Leitz Wetzlar®, Germany) following a calibration of the microscope with a stage micrometer (Graticules Ltd., London) as described by Reid (1968). The various histologic parts of the kidney were recognised based on functional histology (Young et al., 2006). Bowman's capsules that were considered for measurement were those with conspicuous vascular poles. This was done to reduce the risk of measuring sections that were not in the middle of the Bowman's capsule, and consequently, improving the accuracy of the results. For each animal, the Bowman's capsules, Bowman's spaces, glomeruli, proximal convoluted tubules (PCT), thin segment of the loop of Henle (LOH), distal convoluted tubules (DCT) and collecting ducts (CD) were observed.

Results and Discussion:

The mean (\pm SEM) body weight and kidney weights for the animals are shown in Table 1. The weights of the rats ranged between 1.5 – 3.5 kg, and the weights of the kidneys ranged between 3.41 – 6.34 g.

The PCT was lined with simple low cuboidal epithelim that possessed conspicuous 'brush borders' (Figure 1). The thin segment of the LOH was lined by simple squamous epithelium (Figure 2) which was obvious under the microscope. The DCT was also lined by simple cuboidal epithelium (Figure 1), although, the 'brush borders' of the latter were absent and their lumina were more clearly defined.

The lumina of the DCT were larger than those of the PCT. The ranges for the diameters of the Bowman's capsule and glomerulus were 9.12 - 13.02 and 7.71 - 10.64 µm, respectively, while those of the PCT, LOH, DCT and CD were 2.71 - 3.47; 1.79 - 2.26; 3.09 - 3.85 and 3.85 - 5.16 µm, respectively. The individual mean (± SEM) values are shown in Table 2. There was a very strong positive correlation between the diameters of the Bowman's capsule and those of glomerulus (r = 0.975, P < 0.01). Also, significant relationships were observed between the diameters of the CD and the PCT (r = 0.781, P < 0.05); the CD and the thin segment of the LOH (r = 0.821, P < 0.05).

From a physiological perspective, Du *et al.* (2004) reported that measurements of the luminal diameter are critical to assessing changes in hydrodynamic forces and torques. Therefore, a knowledge of the diameters of the tubules is useful in determining the rate of fluid flow through the urinary tubules. The larger sizes in the diameters of the

DCT and CD suggests that urine flows more slowly in the larger tubules (Endo and Kimura, 2005). However, the finding in this study is contrary to the observation of Bentley et al. (2007), who observed that the luminal diameter of the PCT was greater than those of the DCT and the CD in rats. The majority of clinical references concentrate on the functional evaluation of kidneys (Lauschova et al., 2004). This may, however, be inadequate in thorough studies as functional aberrations are often accompanied by structural changes. Also, Damme and Koudstaal (1976) reported that in some kidney diseases it is important to know the diameter of the glomerulus. This is further buttressed by the work of Roman et al. (2004), who noted a significant increase in the diameters of the proximal tubules, distal tubules and collecting ducts of rats treated with cadmium. Normal kidneys are nearly always recognisable in survey radiography. However, an assessment of the histology of the renal pelvis and the ureter is not possible by radiography (SevrekIntas and Kramer. 2008). Hence, the relevance of histological studies.

The method of using the ocular micrometer and a formalin fixed tissue section may hamper the accuracy of the measurements due to some level of shrinkage that occur during tissue processing with formalin (Baker, 1960). However, bearing in mind that the level of shrinkage in formalin fixed and paraffin embedded tissues is about 33.3 % (Baker, 1960), a working estimate may be deduced easily to reflect this tissue alteration.

The results have for the first time provided data on the tubular diameters of the various segments of the urinary tubule of the grasscutter. Ajayi et al.

Conclusion

In conclusion, this study has attempted to generate base-line data on the diameters of the various parts of the nephron. This may be useful as a relative control when studying the effect of certain drugs or toxins on the sizes of the various segments of the urinary tubules. However, further anatomic studies involving the use of more advanced equipment and methods like electron microscopy, radiology, ultrasonography, and, if necessary, computer tomography (CT) or magnetic resonance imaging are required for comprehensive elucidation of the kidneys in the arasscutter.

 Table 1: Mean body weight and kidney weight (n = 7).

Variables (g)	Mean ± SEM
Body weight	2571.4 ± 0.283
Kidney weight	4.67 ± 0.400

Table 2 : Mean diameters of the urinary	
tubule segments measured $(n = 7)$.	

Variables (µm)	Mean ± SEM	Minimum	Maximum
Bowman's	11.244	9.12	13.02
capsule	±		
	0.460		
Glomerulus	9.213	7.71	10.64
	±		
	0.347		
PCT	3.119	2.71	3.47
	±		
	0.106		
LOH	1.921	1.79	2.26
	±		
	0.058		
DCT	3.459	3.09	3.85
	±0.090		
CD	4.356	3.85	5.16
	±		
	0.171		

PCT- Proximal convoluted tubule, LOH-Loop of Henle, DCT- Distal convoluted tubule and CD- Collecting duct.

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References

- Adoun, C. (1993). Place de l'aulacode (*Thryonomys swinderianus*) dans le règne animal et sa répartition géographique In: Schrage R and Yewadan L T (editors) 1ère Conférence Internationale sur l'Aulacodiculture: *Acquis et Perspectives*, 35-40.
- Asibey, E. O. A. and Addo, P. G. (2000). The grasscutter, a promising animal for meat production. In: Turnham D, (Ed) *African Perspectives. Practices and Policies Supporting Sustainable Development.* Scandinavian Seminar College, Denmark, in association with Weaver Press, Harare, Zimbabwe). P. 120.
- Baker, J. R. (1960). Principles of biological microtechniques. *In: Theory and Practice of Histological Techniques*. Third Edition. Churchill livingstone, Edinburgh, 2: 29
- Baylis, C. and Corman, B. (1998). The aging kidney: insights from experimental studies. *Journal of the American Society of Nephrology*, 9: 699–709.
- Bentley, M. D., Jorgensen, S. M., Lerman, L. O., Ritman, E. L., Romero, J. C. (2007). Visualization of three-dimensional nephron structure with microcomputed tomography. *The Anatomical Record*, 290(3): 277-283.
- Charmi, A., Bahmani, M., Sajjadi, M.M. and Kazemi, R. (2009). Morpho-histological study of kidney in farmed juvenile Beluga, Huso huso (Linnaeus, 1758). Pakistan Journal of Biological Sciences, 12(1): 11-18.

- Cutler, R. E. (2006). Kidney: Biology of the Kidneys and Urinary Tract. *In: The Merck Manual* Home Edition. Retrieved From <u>http://www.merck.com/mmhe/s</u> <u>ec11/ ch141/ ch141b.html on</u> <u>29/12/09 at 13</u>: 23 GMT +1 West Africa.
- Damme, B. and Koudstaal, J. (1976). Measuring glomerular diameters in tissue sections. *Virchows Arch. A Path. Anat. and Histol.*, 369: 283-291.
- Du, Z., Duan, Y.,Yan, Q., Weinstein, A. M., Weinbaum, S. and Wang, T. (2004). Mechanosensory function of microvilli of the kidney proximal tubule. *Proceedings of the National Academy of Sciences of the United States of America*, 101(35): 13060-13073.
- Endo, M. and Kimura, M. (2005). Histological and enzyme histochemical studies on the neprons of the fresh water fishes, *Cyprims carpio* and *Carassius auratus. Journal of Morphology*, 173(1): 29-33.
- Germimo, G. G. (2005). Linking cilia to Wnts. *Nature Genetics*, 37(5): 455-457.
- Gordon, K. C. (1990). Tissue processing. *In: Theory and Practice of Histological Techniques* (Ed. Bancroft, J. D. and Stevens, A.). Churchill Livingstone. Edinburgh. p.45.
- Jori, F., Mensah G. A. and Adjanohoun, E. (1995). Grasscutter production: an example of rational exploitation of wildlife. *Biodiversity and Conservation*, 4: 257-265.
- Lauschova, I., Krejcirova, L., Hork, D., Doubek, M., Mayer, J. and Doubek, J. (2004). Ultrastructural morphometry of renal tubule epithelium in rats treated

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with conventional amphotericin B deoxycholate or amphotericin B colloidal dispersion. *Acta Veterinaria Brno,* 73: 165-169.

- National Research Council, NRC (1991). Microlivestock: Little known small animals with promising economic future. *In: Veit Meyer Noel* (Ed.). National Academy Press, Washington, D. C., p. 183-185.
- Onyeanusi, B. I., Adeniyi, A. A., Ayo, J. O. and Nzalak, J. O. (2007). Morphometric studies on the kidneys of the African giant rat (*Cricetomys gambianus* Waterhouse). *Journal of Animal and Veterinary Advances*, 6(11): 1273-1276.
- Reid, W. M. (1968). A diagnostic chart for nine species of fowl coccidian. Department of Poultry Science College Station Athens, Georgia, USA. Technical Bulletin, 39: 17-18.
- Roman, T. R. N., de Lima, E. G., Azoubel, R. and Batigália. F. (2004). Renal morphometry of fetuses' rats treated with cadmium. *International Journal* of *Morphology*, 22(3):231-236.
- Schmidt-Nielsen, B. and O'Dell, R. (1961). Structure and concen-

trating mechanism of the mammalian kidney. *American Journal of Physiology*, 200: 1119-1124.

- Seyrek-Intas, D. and Kramer, M. (2008). Renal imaging in cats. *Veterinary Focus*, 18(2): 23.
- Sperber, J. (1944). Studies on the mammalian kidney. *Zoological Bulletin*, 22: 249-432.
- Warui, C. N. (1989). Light microscopic morphometry of the kidneys of fourteen avian species. *Journal of Anatomy*, 162: 19-31.
- Young, B., Lowe, J. S., Stevens, A. and Heath, J. W. (2006). *Wheaters Functional Histology. A Text and Colour Atlas.* Fifth Edition. Churchill livingstone, Elsevier, Pp. 302-325.

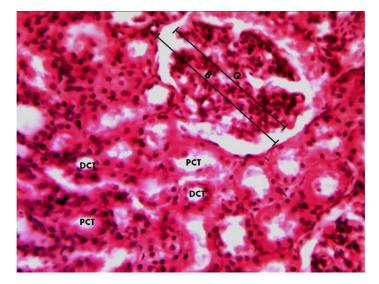


Fig (1): A section through the cortex of the kidney of the grasscutter (*Thryonomys swinderianus*) showing the glomerulus (G), the Bowman's capsule (B), the proximal convoluted tubule (PCT) and the distal convoluted tubule (DCT). (H & E, x 400)

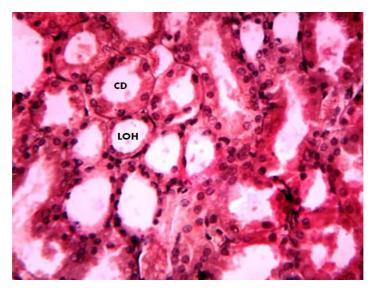


Fig (2): A section through the medulla of the kidney showing the collecting ducts (CD) and the thin segments of the loop of Henle (LOH). (H & E, x 400)

Animal species of this issue

The Greater Cane Rat or the Grass cutter (*Thryonomys swinderianus*)



Kingdom: Animalia & Phylum: Chordata & Class: Mammalia & Order: Rodentia & Family: Thryonomyidae & Genus: *Thryonomys* & Species: *T.swinderianus*

The **Greater Cane Rat** (*Thryonomys swinderianus*) is one of two species of cane rats, a small family of African hystricognath rodents. It inhabits Africa, south of the Saharan Desert. The cane rat lives by reedbeds and riverbanks.

Cane rats can grow about two feet long in the longest individuals and weighs a little less than 19 lb (8.6 kg). It has rounded ears, short nose, and coarse bristly hair. Its forefeet are smaller than its hindfeet and supports its weight on only three toes. Cane rats live in small groups led by a single male. They are nocturnal and make nests from grasses or burrow underground. The oldest cane rat can live more than four years.

In the country of Ghana and other regions of West Africa, the Greater Cane Rat is usually called a **grasscutter** or **cutting grass**. In both West Africa and Southern Africa, it is considered a delicacy. As a consequence, "grasscutters" are beginning to be raised in cages for sale, and so are sometimes referred to as micro livestock.