# EFFICACY OF SOME BIOAGENTS, CHITOSAN COMPOUNDS AND ORANGE PEELS EXTRACT IN CONTROLLING MELOIDOGYNE HAPLA ON STRAWBERRY IN EGYPT

#### Maisa L. Abd El- Moneim<sup>1</sup>, Samaa M. Shawky <sup>2</sup> and Manal M. Soliman<sup>2</sup>

- 1- Central Laboratory of Organic Agriculture, Agricultural Research Center, Giza, Egypt.
- 2- Nematode Research Department, Plant Pathology Research Institute, Agricultural Research Center, Giza, Egypt.

(Received: Apr. 11, 2010)

ABSTRACT: Five treatments (Bacillus subtilis, Trichoderma harzianum, chitocar product, soft guard and orange peels extract) with different concentrations were used to control root-knot nematode, Meloidogyne hapla under laboratory as well as greenhouse and field conditions on strawberry. Also, application number of the treatments were studied under both greenhouse and field conditions.

The most effective treatment in controlling root-knot nematode was soft guard, while Bacillus subtilis, Trichoderma harzianum, chitocar product occupied an intermediate position in the effectiveness whereas, the least effective one was orange peels extract under both laboratory and greenhouse conditions. So the treatment of orange peels extract was neglected under field conditions. The most effective treatment under field conditions in controlling Meloidogyne hapla was soft guard, whereas the least effective was Trichoderma harzianum.

Under laboratory conditions the five treatments were used to study their effect on percentage of juveniles mortality. All treatments led to high percentage of juvenile mortality especially at the highest concentration and after 72 hours exposure.

Under greenhouse conditions using soft guard and Bacillus subtilis were more effective in reducing numbers of developmental stages, females, galls, egg-masses and eggs/ egg-mass as well as number of 2 nd stage larvae in soil, whereas, the least effective was orange peels extract.

Adding the treatments as soil drench led to increase the fresh weights of both the root and shoot system of strawberry seedlings at all used concentrations especially at the highest concentration. Using three times of addition from each treatment achieved high decrease in nematodes in both roots and soil.

Also, all the treatments individually as soil drench decreased the number of nematodes in both roots and soil. In addition the fruit yield of strawberry

increased after adding the treatments at all three times and at high concentration under field conditions.

Key Words: Non chemical control, root-knot nematodes, Meloidogyne hapla, Bacillus subtilis, Trichoderma harzianum, chitocar product, soft guard, orange peels extract and strawberry.

#### INTRODUCTION

Strawberry (Fragaria × ananassa L.), is one of the most economic crops in Egypt. Several species of root- knot nematodes, Meloidogyne spp., have been recognized as a potentially serious problem to the crop productivity. (Annonymus, 1986) indicated that nematodes are a particular problem in strawberry crop in some countries, severe nematode infestations force grape growers to destroy strawberry plants and let the land lie fallow for many years before replanting.

In recent years, the awareness of the nematicides hazards to human and environment has directed the attention towards soil-borne antagonists as an alternative method to chemical control.

Biological control is gaining increasing role throughout the world for nematode suppression.

Bacillus subtilis is reported as a bio-control agent against root-knot nematodes. Nematode mortality was observed from 8 hours incubation. A concentration of at least 10 <sup>8</sup> cfu / ml were necessary to cause nematode mortality higher than 30%. Bacillus subtilis act through production of number of antibiotic as bacterocin (Ferreiro et al., 1991; Asaka and Shoda, 1996; Farahat, 1998; Khan et al., 2002 and Shawky & Abd El- Moneim, 2005).

Trichoderma harzianum act through different mechanisms including mycoparasitism, also through production of antibiotic substances (Abd El Moity & shatla, 1981and Banhamoud & Chet, 1993). Trichoderma harzianum also act through production of destructive enzymes i.e, chitenase ( Padares et al., 1992; Bolar et al., 2000; Sharon et. al., 2001; Faruk et al., 2002; Siddiqui and Shaukat., 2004; Shawky & Abd El- Moneim, 2005 and Sahebani & Hadavi, 2008). Abd El-Moity et al. (1998) said that application of Bacillus subtilis and Trichoderma harzianum to nematoue infested soil twice was always more effective compared with single treatment.

Loumedjiinon et al. (2007) used dried skin peels of orange to control root-knot nematode (RKN), Meloidogyne spp. in solanum macroatponl under field condition. They found significantly reduction in root galling and nematode soil population density and increasing in plant biomass as compared to control. Tsai (2008) stated that the extract of fresh orange peels showed significantly nematostatic effect against M. incognita second stage juveniles after 48 h of treatment. The nematicidal activity was very low in the extracts of fresh peels but was greatly enhanced in the extracts of stored extract peels with 93.5% mortality of nematodes. He explained that by the possibility that essential oils from the citrus peels might have released in the extract

during storage of the orange peels. He also mentioned that infection of *M. incognita* second stage juveniles on mungbean root was significantly inhibited by the extracts of the refrigerator-stored peels of orange.

(Mulawarman et al. 2001) said that chitosan compound is a natural product stimulate microbial activity in the soil and promote plant growth. They found positive effects of natural products in stimulating soil microbial activity and thereby the antagonistic potential in soils leading to a reduction in nematode infestation and improved plant growth.

(Vasyukova et al. 2001) stated that chitosan play an elicitor activity by inducing the local and systemic of resistance inside plant tissues against different pathogens and nematodes. They added that chitosan induced the accumulation of phytoalexins in tissues of host plants; decreased the total content; changed the composition of free sterols producing adverse effects on infesters; activated chitinases, -glucanases, and lipoxygenases; and stimulated the generation of reactive oxygen species.

(Hallmann et al 1999) said that the addition of chitin to soil at 1% (w/w) eliminated plant-parasitic nematodes in a first planting of cotton cv. 'Rowden' and significantly reduced Meloidogyne incognita infestation in a second planting, confirming long-term nematode suppressiveness induced by this organic amendment. They found that chitin amendment was associated with an increase in fungal and bacterial populations, especially those with chitinolytic activity.

The present work was carried out to study the efficacy of some bioagents, chitosan compound and extract of orange peels in controlling *Meloidogyne hapla* under laboratory, greenhouse and field conditions

#### MATERIALS AND METHODS

In this study five different treatments were used. These treatments included:-

- 1- Bacillus subtilis + Nematode (N).
- 2- Trichoderma harzianum + N.
- 3- Soft guard compound as a commercial product + N.
- 4- Chitocare compound as a commercial product + N.
- 5- Orange peels extract + N.
- 6- Check treatment with nematode only

All treatments were prepared in Central Laboratory of Organic Agriculture, Agricultural Research, Center, Giza.

1- Preparation of the treatments:-

#### A- Bacteria:-

Bacillus subtilis was grown on nutrant glucose broth (NG) for 48 hours. The bacterial suspension was adjusted to be containing  $(30 \times 10^6)$  cells /ml (Dowson, 1957)

#### B- Fungi:-

Trichoderma harzianum was grown in gliotoxin fermentation media (GFM) under complete darkness just to stimulate toxin production (Abd El- Moity and Shatla, 1981) for 9 days. The suspension of T. harzianum was prepared by adjusting the number of T. harzianum propagules, in the suspension to be (30 x  $10^6$ ) cfu /ml using sterilized distilled water.

#### C- Soft guard:-

It contains chitosan oligo Saccharin (molecular weight ≤ 3000) .Importer: Technogreen Group for agricultural production .Exporter: Leili; Agro Chemistry CO. LTD; China.Registration No. 4585. Application rates: for watering dilute 1000 times.

#### D- Chitocar product:-

It contains chitosan oligomers, and cheleated elements: Fe, Zn, Mn, Cu, B and NPK. Sentic Company. Application rates: for watering dilute 1000 times.

#### E- Orange peels:-

Extract of orange peels was prepared by mixing 10 g of dry powder orange peels with 100 ml of water using electric blender for 5 minutes.

#### I- Laboratory experiment:-

Efficacy of some bioagents, chitosan compounds and orange peels extract treatments against active nematode juveniles under laboratory conditions:-

To test the efficacy of the five different treatments were used in inhibiting the activity of *M. hapla* juveniles in vitro, 1 ml of each (Bacillus subtilis, containing (30 x 10<sup>6</sup>) cells /ml., Trichoderma harzianum containing (30 x 10<sup>6</sup>) cfu /ml were added at three concentrations of (1:50, 1:100, 1:150) and I ml. of each (chitocar product, soft guard and orange peels extract) were added at three concentrations of (1:500, 1:800, 1:1000) to 1 ml of nematode suspension containing (200 active juveniles of *M. hapla*) in glass vials and compared with check treatment of 1 ml distilled water containing (200 active juveniles of *M. hapla*). The glass vials well examined after 24, 48 and 72 hours from exposure. The numbers of active and non active juveniles were counted microscopically. New blue R (0.05% aqueous solution), potassium permanganate (0.062-0.50% KMnO4 in aqueous solution) and chrysoidin have been used to stain dead nematodes (Barker et. al. 1985).

#### II- Greenhouse experiments:-

1- Evaluation the different concentrations of some bioagents, chitosan compounds and orange peels extract treatments to control *M. hapla* under greenhouse conditions:-

In all greenhouse experiments nematodes inoculation procedure were prepared <u>as</u>

#### Nematodes inoculation procedure:-

After strawberry seedlings (*Fragaria x ananassa L.*) Camarosa variety, were transplanted individually in 20 cm diameter clay pots each containing 2.5 Kg steam –sterilized soil. (1:2) loam and sand respectively, each pot was inoculated with 3000 newly hatched juveniles from pure culture of *M. hapla* by making 3 holes at different depths (2-3 cm) around the roots and immediately after inoculation the roots were covered with soil.

One week after inoculation different treatment which include *Bacillus* subtilis, *Trichoderma harzianum*, chitocar product, soft guard and orange peels extract were added. The five different treatments were prepared by diluting (1:50, 1:100, 1:150) for *Bacillus subtilis*, *Trichoderma harzianum* and orange peels extract and by diluted suspensions of (1:500, 1:800, 1:1000) for both chitocar product and soft guard. Pots were treated with nematode active larvae without the treatments served as control treatment.

All treatments received the same agricultural treatment such as amount of water, number of seedling /pot and amount of fertilizers. All pots were arranged in completely randomized design, and kept under greenhouse conditions.

2-Effect of soil drench application number of different treatments to control *M. hapla* under greenhouse conditions:-

This experiment was conducted to determine the effect of application numbers of the previous five treatments in controlling *M. hapla* under greenhouse conditions.

### All inoculated strawberry seedlings were divided into four groups.

- 1- The first group received one time of 100 ml/kg. soil from the highest concentrations of the different treatments.
- 2- The second group received two times of 100 ml/kg. soil from the highest concentrations of the different treatments.
- 3- The third group received three times of 100 ml/kg, soil from the highest concentrations of the different treatments.
- 4- The fourth group received nematode only as control.

In all greenhouse experiments, after 60 days, all plants were carefully uprooted. Root and shoot systems were weighted. Nematode populations in soil and roots were recorded according to (Franklin & Goodey, 1957). Eggs of *M. hapla* were extracted from galls of strawberry roots using sodium hypochoride (NaOCI) method as described by Husssey and Baker (1973).

#### III- Field experiments:-

Evaluation the different treatments in controlling *M. hapla* under field conditions:-

This experiment was conducted in sandy loamy naturally infested soil with M. hapla to determine the effect of numbers of application of four

treatments (*B. subtilis*, *T. harzianum*, chitocar product and soft guard) to control *M. hapla* under field conditions at the high concentration of (1:50) for *B. subtilis* and *T. harzianum*, and the concentration of (1:500) for both chitocar product and soft guard. At the end of the experiment nematodes in both soil and root were determined. Also, the totally fruit yield were determined. Each treatment include 10 replicates and every replicate contains 100 plants

#### Statistical Analysis:-

All obtained data were subjected to statistical analysis proposed by Gomez and Gomez (1984)

#### **RESULTS AND DISCUSSION**

#### I- Laboratory experiment:-

Efficacy of some treatments against active nematode juveniles under laboratory conditions:-

Data in Table (1) illustrated that the five tested treatments (*B. subtilis, T. harzianum*, chitocar product, soft guard and orange peels extract) had various degrees of effectiveness toward the nematode juveniles. Moreover, the percentage of mortality increased with increase of the treatments concentration and exposure period.

The highest concentration of (1:50) for *B. subtilis*, *T. harzianum* and orange peels extract and the concentration of (1:500) for both chitocar product and soft guard in all the tested treatments achieved the highest percentage of juvenile mortality during all exposure periods.

After 24 hours, the mortality caused by all used treatments (*B. subtilis, T. harzianum*, chitocar product, soft guard and orange peels extract ranged between 23.5 to 87.3% compared with control of mortality % (0.4 %). Also, after 72 hours at the concentration (1:50) the highest percentage of mortality % caused by soft guard was (95.4%) while the lowest percentage of mortality caused by orange peels extract was (57.2 %) compared with control of (1.5%). *B. subtilis* was occupied the second rank of effectiveness on mortality % by (92.5%). Chitocar product was occupied the third rank of effectiveness on mortality % by (87.2%) followed by *T. harzianum* treatment which achieved a percentage of mortality of (70.1%).

Bacillus subtilis acts through production of number of antibiotics (Ferreiro, 1991; Asaka and Shoda, 1996; Abd El Moity et. al., 1998 and Farahat., 1998). Bacillus subtilis can grow and multiplicate very fast under this circumstance.

Trichoderm harzianum act through different mechanisms including mycoparasitism, also through production of antibiotic substances (Abd El Moity & shatla, 1981 and Banhamoud & Chet, 1993). Trichoderma harzianum also act through production of destructive enzymes i.e., chitenase (Padares et al., 1992; Bolar et al., 2000).

Table (1): Effect of exposure period to some treatments on mortality percentage of *M. hapla* juveniles under laboratory conditions.

		Mortality % Exposure period (in hours)				
Treatments	Conc.					
		24	48	72		
	1:150	50.7	55.2	68.5		
Baçillus subtilis + N	1:100	71.9	75.6	81.4		
	1:50	83.5	88.3	92.5		
	1:150	35.2	38.5	46.8		
Trichoderma harzianum + N	1:100	48.9	55.7	64.2		
	1:50	55.5	62.8	70.1		
	1:1000	57.2	64.9	79.3		
Soft guard + N	1:800	76.9	82.5	85.8		
	1:500	87.3	91.7	95.4		
	1:1000	46.6	50.4	69.2		
Chitocar product + N	1:800	61.3	68.2	75.1		
	1:500	74.4	81.6	87.2		
Orange peels extract + N	1:150	23.5	26.9	35.6		
	1:100	28.7	33.8	38.2		
	1:50	41.3	46.7	57.2		
Nematodes in distilled water		0.4	1.1	1.5		
L.S.D. (5 %)		2.11	2.23	3.56		

Loumedjiinon et al (2007) used dried skin peels of orange to control root-knot nematode (RKN), Meloidogyne spp. in solanum macroatponl under field conditions. Data obtained showed that significantly reduction in number of galls and nematode population density in soil and increased plant biomass as compared to control. Tsai (2008) stated that the extract of fresh peels of orange showed significantly nematostatic effect against M. incognita second stage Juveniles after 48 h treatment. The nematicidal activity was very low in the extracts of fresh peels but was greatly enhanced in the extracts of stored orange peels with 93.5% mortality of nematodes. (Hallmann et al 1999) said that addition of chitin to soil at 1% (w/w) eliminated plant-parasitic nematodes in a first planting of cotton cv. 'Rowden' and significantly reduced M. incognita infestation in a second planting, confirming long-term nematode supressiveness induced by this organic amendment. The chitin amendment was associated with an increase in fungal and bacterial populations, especially those with chitinolytic activity.

#### II- Greenhouse experiments:-

- 1- Evaluation the different concentrations of some bioagents, chitosan compounds and orange peels extract treatments to control *M. hapla* under greenhouse conditions:-
- 1- Effect on nematode population:-
- a- Number of root galls:-

However all the treatments showed remarkable decrease of the number of root galls Fig.(1). Soft guard performed the highest reduction of galls

number compared with the other treatments. *B. subtilis* ranked on the second place, whereas orange peels extract resulted in the lowest reduction of root galling. *T. harzianum* and chitocar product occupied an intermediate position in decreasing the root galling. Increase the concentration of soft guard from (1:1000) to (1:500) resulted in decrease the root galling from 46 to 29 galls/root system. Similar decrease was found in the case of all the other treatments.

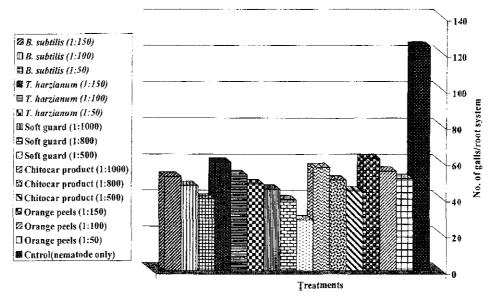


Fig. (1): Effect of some bioagents, chitosan compounds and orange peels extract treatments on number of galls /root of strawberry plants infected by *M. hapla* under greenhouse conditions.

b- Number of juveniles in soil, developmental stages, females, egg masses, eggs numbers and rate of build up of *M. hapla*:-

Data in Table (2) illustrated that all tested treatments were effective in decrease the final nematode population and rate of build up of root knot nematode in both soil and roots especially at the highest concentrations. Soft guard was the most effective treatment whereas the least effective treatment one was orange peels extract. B. subtilis, T. harzianum, and chitocar product occupied an intermediate position. Also, data showed that positive correlation between efficacy of the treatments and concentrations.

The same trend obtained with the effect on juveniles in soil, and developmental stages, females, egg -masses and eggs numbers in roots.

Table (2): Efficacy of some bioagents, chitosan compounds and orange peels extract treatments on reproduction of *M. hapla* infecting strawberry plants under greenhouse conditions.

	i		Nematode	e populatio	n in			
Treatments Conc.		No. of		*Final	Rate of			
	juveniles in 250g soil	No. of development al stages	No. of females	No. of egg- masses	No. of eggs/ egg- mass	nematode population (PF)	build- up (PF/PI)	
	1:150	180	63	52	48	341	16663	5.55
Bacillus	1:100	120	57	47	41	335	13959	4.65
subtilis + N	1:50	100	50	40	37	317	11919	3.97
	1:150	340	71	60	55	360	20271	6.75
Trichoderma	1:100	300	64	53	50	345	17667	5.88
harzianum +N	1:50	220	58	48	42	333	14312	4.77
	1:1000	140	58	45	40	320	13043	4.34
Soft	1:800	100	51	39	35	308	10970	3.65
Guard + N	1:500	80	45	28	26	295	7823	2.60
	1:1000	260	66	57	54	352	19391	6.46
Chitocar	1:800	200	60	50	47	340	16290	5.43
Product + N	1:500	140	52	44	40	331	13476	4.49
Orange	1:150	400	73	62	60	380	23335	7.78
Peels + N	1:100	320	67	55	52	351	18684	6.23
extract	1:50	280	62	51	49	343	17200	5.73
Control (nematodes only)	(N)	2400	112	124	119	430	53806	17.93
L.S.D. (5 %)		18.36	1.23	1.54	1.12	11.87	465.61	0.23

Each value presented the mean of four replicates.

<sup>\*</sup>Final nematode population(PF)=(No. of egg-masses x No. of eggs/egg-mass)+ No. of females +No. of developmental stages+ No. of juveniles in soil/pot.

Rate of build-up =	Final nematode population (PF)
	Initial nematode population (PI)

All data are in agreement with those obtained by Hanna et. al. (1999) who mentioned that B. subtilis was effective against root-knot nematode M. incognita on tomato plants. Also agree with Abd El-Moity, et. al. (1998) who mentioned that the most effective in controlling the root-knot nematode was Bacillus spp., whereas the least effective was T. harzianum. Also, Khan et. al.(2002) said that treatment with B. subtilis or Beijerinckia indica reduced galling by 33-34 % and increased the dry weight of shoots by 22-24 %, respectively. Trichoderma spp. can produce various toxin metabolites and different enzymes that improve photolytic activity of the antagonist and control of nematodes. ((Sharon et. al., 2001; Faruk et. al., 2002; Siddiqui and Shawkat, 2004 and Shawky & Abd El-Moneim, 2005).

#### 2 - Effect on the vegetative growth of the treated seedlings:-

The effect of the treatments (B. subtilis, T. harzianum, chitocar product, soft guard and orange peels extract) on fresh weights of the treated plants were illustrated in Fig.(2).

All the treatments enhanced increase in fresh weights of the whole plants compared with control plants. The maximum increasing % at the highest concentration was recorded in treatment with soft guard (223%) followed by *B. subtilis* with (199%), chitocar product (171%) followed by *T. harzianum* with (152%). The treatment of orange peels showed the lowest shoot weight increasing %, which reached to (83 %) compared with the control.

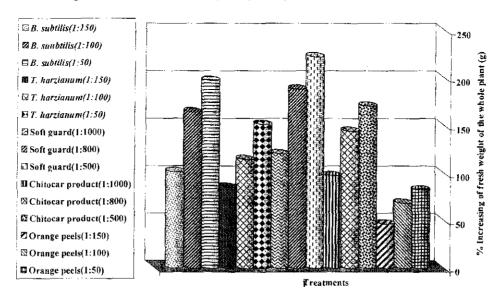


Fig. (2): Effect of some bioagents, chitosan compounds and orange peels extract treatments on increasing % of fresh weight of the whole plant of strawberry infected by *M. hapla* under greenhouse conditions.

- 2- Effect of application number of treatments at different preparation to control *M. hapla* on strawberry us der greenhouse conditions:-
- A -Evaluation the application number on the nematode infecting strawberry plants under greenhouse conditions:-
- 1- Effect on nematode population:-

#### a- Number of root galls:-

However all the treatments showed remarkable decrease of the number of root galls Fig. (3), soft guard performed the lowest galls numbers compared with the other treatments especially after three times of applications at the highest concentration (1:500). B. subtilis ranked on the second place, whereas orange peels extract resulted in the lowest decrease in the number

of root galls. Both Chitocar product and *T. harzianum* were occupied an intermediate position in decrease number of root galling. The decrease in root galls was affected greatly with the number of applications of different treatments.

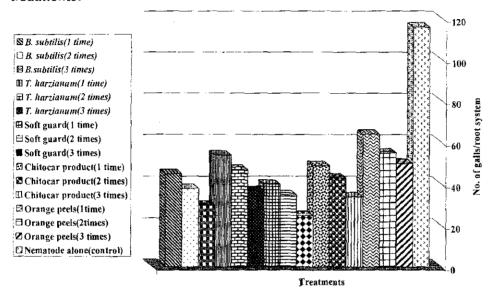


Fig. (3): Effect of application number of some bioagents, chitosan compounds and orange peels extract treatments on number of galls /root of strawberry plants infected by *M. hapla* under greenhouse conditions.

b- Number of juveniles in soil, developmental stages, females, egg masses, eggs numbers/ root and rate of build up of *M. hapla*:-

Data in Table (3) showed that all tested treatments were effective in decrease the final nematode population and rate of build up of root knot nematode in both soil and roots especially after three times of application. Soft guard was the most effective treatment, whereas the least effective one was orange peels extract. B. subtilis, T. harzianum and chitocar product occupied an intermediate position. Also, data showed that positive correlation between efficacy of the treatments and applications number of the treatments.

The same trend obtained with the effect on juveniles in soil, and developmental stages, females, egg -masses and eggs numbers in roots.

Table (3): Effect of application number of some bioagents, chitosan compounds and orange peels extract treatments on reproduction of M. hapla infecting strawberry plants under greenhouse conditions

Treatments		Nematode population in						
	Number	Root					#F11	
	application juv	No. of juveniles in 250g soil	No. of developme ntal stages	No. of females	No. of egg- masses	No. of eggs/ egg- mass	*Final nematode population (PF)	build- up (PF/PI)
	One time	160	56	45	41	360	15021	5.00
Bacillus	Two times	120	51	38	35	340	12109	4.03
subtilis + N	Three times	80	43	30	28	310	8833	2.94
	One time	280	65	54	50	385	19649	6.55
Trichoderma	Two times	240	59	47	45	375	17221	5.72
harzianum + N	Three times	180	50	37	34	354	12303	4.10
	One time	120	52	40	38	337	13018	4.34
Soft	Two times	100	48	35	32	315	10263	3.42
guard + N	Three times	60	40	25	22	280	6285	2.09
	One time	200	62	49	46	378	17699	5.90
Chitocar	Two times	180	57	43	41	366	15286	5.09
product + N	Three times	140	46	34	30	337	10330	3.44
Orange peels + N extract	One time	360	72	64	61	405	25201	8.40
	Two times	300	65	55	51	389	20259	6.75
	Three times	240	54	50	47	373	17875	5.95
Control (nematode	s only) (N)	2600	112	116	111	430	50558	16.85
L.S.D. (5 %)		15.86	1,23	0.89	0.81	10.12	415.16	0.06

Each value presented the mean of four replicates.

D. C. of building	Final nematode population (PF)
Rate of build-up =	
	Initial nematode population (PI)

#### 2 - Effect on the vegetative growth of the treated seedlings:-

The effect of number of applications of soil drench of the treatments (Bacillus subtilis, Trichoderma harzainum, chitocar product, soft guard and orange peels extract) on fresh weight of the reated plants were studied in Fig.(4).

All the treatments enhanced the increase in fresh weight of the whole plants compared with the control. The maximum increasing % after three times of applications was recorded with the seedlings treated with soft guard (255%) followed by *B. subtilis* with (232%), chitocar product (221%) followed by *T. harzianum* with (198%). The treatment of orange peels showed the lowest shoot weight increasing%, which reached (112 %) compared with control.

<sup>\*</sup>Final nematode population(PF)=(No. of egg-masses x No. of eggs/egg-mass)+ No. of females +No. of developmental stages+ No. of juveniles in soil/pot.

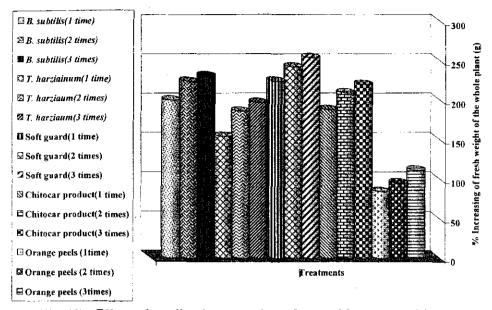


Fig. (4): Effect of applications number of some bioagents, chitosan compounds and orange peels extract treatments on increasing of fresh weight of the whole plant of strawberry infected by *M. hapla* under greenhouse conditions.

#### III- Field experiments:-

Effect of some bioagents and chitosan compounds treatments to controlling *M. hapla* in naturally infecting strawberry plants under field conditions:-

#### 1- Number of root galls:-

However all the treatments showed remarkable reduction of the number of root galls as shown in Table (4), the soft guard performed the lowest galls numbers as reached to 24 galls/root compared with the other treatments at (1:500) with three times of application. *B. subtilus* ranked on the second place resulting in reduce galls to 28 galls/root, whereas *T. harzianum* resulted in the lowest reduction in the number of root galls reached to 35 galls/root. Chitocar product occupied an intermediate position in decrease number of galls/ root reached to 31 galls/root in compared with the control 113 galls/root.

2- Number of juveniles in soil, developmental stages, females, egg masses and eggs numbers in roots:-

#### Estimates of combining ability for grain yield and other.....

Table 3: Estimates of GCA effects for grain yield and other attributes, data of the combined over the three locations.

Traits Lines	Days to	Plant height	Ear height	Resistance to late wilt	Grain yield
L1	silking 0.729**	0.783	1,002	0.626	2.237**
L2	0.479*	16.050**	6.414**	0.318	0.612
L3	1.284**	10.550**	3.391*	1.997*	0.178
L4	0.035	9.338**	7.252**	0.452	0.244
L5	0.437*	6.106**	1.386	0.152	1.836**
L6	0.687**	21.004**	10.057**	3.520**	1.598**
L7	0.687**	4,449**	4.692*	4.208	0.796
L8	0.395*	3.467*	4.469*	1.701	0.216
L9	1.396**	4.004**	0.331	3.375**	0.594
L10	0.576**	0.865	0.613	1.726	0.346
L11	0.187	6.643**	2.752	3.051**	0.533
L12	1.743**	10.912**	7.775**	3.074**	2.060**
Tester (T)					
GZ638	0.118	5,102**	6.516**	0.160	0.954**
GZ650	0.076	5.594**	2.377*	1.171**	0.896**
Gm1021	0.042	10.696**	8.893**	1.331**	1.850**
SE gi L SE gi T	0.189 0.094	1.460 0.730	1.698 0.849	0.691 0.345	0.427 0.213

<sup>\*, \*\*</sup> Significant at 0.05 and 0.01 level of probability, respectively.

#### M.E.M. Abd El Azeem, M.A. Abd El Moula and A.E.M.K. El-Galfy

Table 4: Estimates of SCA effects for grain yield and other attributes, data of the combined over the three locations.

Crosses	Days to 50% silking	Plant height (cm)	Ear height (cm)	Resistance to late wilt	Grain yield
L1 x GZ638	0.118	11.981**	5.594*	1.227	1.774*
L2x GZ638	0.118	17.685**	12.738**	1.722	3.174**
L3 x GZ638	0.479	4.815	5.206*	1.358	0.057
L4 x GZ638	0.479	7.426**	4.178*	0.436	0.052
L5 x GZ638	0.548	6.796*	7.099**	2.559*	2.240**
L6 x GZ638	0.382	2.342	0.456	6.585**	0.233
L7 x GZ638	0.298	5.231*	6.372**	0.125	2.218**
L8 x GZ638	0.548	5.185*	0.349	0.854	0.273
L9 x GZ638	0.285	2.324	1.571	0.427	0.637
L10 x GZ638	0.090	3.101	0.567	0.988	0.187
L11 x GZ638	0.034	10.954**	6.761**	2.037	1.400
L12 x GZ638	0.173	7.657**	7.377**	1.756	0.871
L1 x GZ650	0.493	1.678	1.067	1.576	1.672*
L2 x GZ650	0.340	3.571	2.817	0.8433	1.312
L3 x GZ650	0.368	3.511	2.155	0.116	3.316**
L4 x GZ650	0.049	4.317	2.182	2.450*	0.506
L5 x GZ650	0.243	7.627**	2.956	1.359	1.601*
L6 x GZ650	0.090	0.984	2.488	3.399**	1.105
L7 x GZ650	0.674*	5.155*	1.988	0.768	1.148
L8 x GZ650	0.076	5.322*	0.373	0.635	1.434
L9 x GZ650	0.659*	3.099	3.400	0.389	0.776
L10 x GZ650	0.021	.1.488	0.28 <del>9</del>	0.025	0.892
L11 x GZ650	0.076	18.206**	8.432**	2.790*	2.934**
L12 x GZ650	0.145	2.432	6.344**	0.056	0.110
L1 x Gm1021	0.375	10.303**	6.662**	0.349	0.102
L2 x Gm1021	0.458	14.113**	9.921**	2.565*	1.862*
L3 x Gm1021	0.8 47*	1.303	3.050	1.474	3.373**
L4 x Gm1021	0.430	<b>3.10</b> 0	1.995	2.886*	0.559
L5 x Gm1021	0.791*	0.831	4.143	1.199	0.639
L6 x Gm1021	0.291	1.358	2.072	3.186**	0.872
L7x Gm1021	0.375	10.386**	4.384*	0.894	1.069
L8 x Gm1021	0.625	0.136	0.023	1.490	1.708*
L9 x Gm1021	0.375	0.775	1.828	0.037	1.413
L10 x Gm1021	0.069	1.613	0.856	0.962	1.079
L12 x Gm1021	0.319	5.224*	1.032	1.812	0.982
L11 x Gm1021	0.041	7.252**	1.671	0.753	1.534*
SE (sij)	0.328	2.530	2.080	1.198	0.739

<sup>\*, \*\*</sup> Significant at 0.05 and 0.01 level of probability, respectively.

#### Variance components

Estimates of the variance components of lines and testers ( $\sigma_{GCA}^2$ ) and of crosses (o<sup>2</sup><sub>SCA</sub>) for grain yield and other agronomic traits across locations are presented in Table (5). Results revealed that values of  $\sigma^2_{GCA}$  for lines (L) were higher than those of  $\sigma^2_{GCA}$  for testers (T) for number of days to 50% silking and resistance to late wilt. These results indicate that most of the total variance was due to GCA of the lines. While, values of  $\sigma^2_{GCA}$  for testers (T) were higher than those of  $\sigma^2_{GCA}$  for lines (L) for plant and ear height and grain yield. These results indicate that most of the total variance was due to GCA of the testers of these traits. The variance interaction of  $\sigma^2_{GCA}$  testers x Loc. was larger than  $\sigma^2_{GCA}$  lines x Loc. for ear height and grain yield, indicating that  $\sigma^2_{GCA}$  for testers was more affected by environmental conditions than that for lines. On the other hand the magnitude of variance interaction of  $\sigma^2_{GCA}$  lines x Loc. was larger than that of  $\sigma^2_{GCA}$  testers x Loc for number of days to 50% silking, plant height and resistance to late wilt. The  $\sigma^2_{SCA}$ variances were larger than that of  $\sigma^2_{GCA}$  for days to 50%silking, plant height and late wilt resistance. These results indicate that the non-additive gene effects were more important than additive gene effects in the inheritance of these traits. While the  $\sigma^2_{GCA}$  variances was larger than those of  $\sigma^2_{SCA}$  for ear height and grain yield. Furthermore, the magnitude of  $\sigma^2_{SCA}$  x Loc. interaction was greater than  $\sigma^2_{GCA}$  x Loc interaction for all the traits studied except, for number of days to 50% silking, indicating that the non-additive gene action interacted more with the environmental conditions than the additive component for these trait. These results are in agreement with the findings of several investigators who reported that specific combining ability variance was more sensitive to environmental changes than general combining ability variance (Gilbert, 1958). Also, Shehata and Dhawan (1975) and Sadek et al (2000 and 2002) also found that the non-additive genetic effects interacted more with the environment than the additive component. On the other hand, El-Itriby et al (1990), and Soliman et al (2001) reported that the additive types of gene action were more affected by the environment than non-additive ones.

#### M.E.M. Abd El Azeem, M.A. Abd El Moula and A.E.M.K. El-Galfy

Table 5. Estimates of general  $(\sigma^2_{GCA})$  and specific  $(\sigma^2_{SCA})$  combining ability variances for grain yield and other plant traits combined over three locations.

	Traits								
Parameters	Days to 50% silking	Plant height (cm)	Ear height (cm)	Resistance to late wilt	Grain	yield			
σ <sup>2</sup> <sub>gca</sub> (lines)	0.737	76.183	16.993	4.413	0.2	45			
σ <sup>2</sup> <sub>gca</sub> (testers)	0.10	78.852	60.582	1.078	2.242				
σ <sup>2</sup> <sub>gca</sub> x Loc (lines)	0.281	17.326	4.732	5.786	0.090				
σ <sup>2</sup> <sub>gca</sub> x Loc (testers)	0.002	10.013	2.614	0.316	1.2	14			
$\sigma^2_{gca}$	0.125	77.062	53.258	2.011	7.3	12			
σ <sup>2</sup> <sub>gca</sub> x Loc	0.037	0.789	0.623	0.623	0.6	41			
$\sigma^2_{sca}$	0.150	77.883	31.92 <del>1</del>	4.642	3.3	75			
σ² <sub>sca</sub> x Loc	0.008	21.548	15.444	6.553	3.7	63			

#### REFERENCES

- Abd El Moula, M.A., A.A.Barakat and A.A. Ahmed (2004). Combining ability and type of gene action for grain yield and other attributes in maize (Zea mays L.) Assiut J. of Agric. Sci. 35(3):129142
- Abd El Moula, M.A. and M.E.M. Abd ElAzeem (2008). Selection among S<sub>3</sub> yellow maize using combining ability. Assiut J.of Agric.Sci. 39(4):117
- Ali, M.L. and N.M. Tepora (1986). Comparative performance of four types of testers for evaluating corn inbred lines from two populations. Crop Sci. 11 (3): 175179.
- Amer, E.A and A.A. ElShenawy (2007). Combining ability for new twenty one yellow maize inbred lines. J. of Agric. Sci. Mansoura Univ., 32(9):70537062.
- Davis, R.L. (1927). Report of the plant breeding Ann. Rep. Pureto Rico Agric. Exp., P:1415.
- Elltriby, A., H.Y. ElSherbieny, M.M. Ragheb and M.A.K. Shalaby (1990). Estimation of combining ability of maize inbred lines in top crosses and its interaction with environments. Laypt. J. Appl. Sci. 5(8):354370.
- ElMorshidy, M.A. and E.A. Hassaballa (1982). Relative values of five testers in evaluating combining ability of maize inbred lines. Assiut J. of Agric. Sci.13 (1):95102.
- ElMorshidy, M.A., E.A. Hassaballa, Sh.F. AbouElsaad and M.A. Abd ElMoula (2003). Combining ability and type of gene action in maize under favorable and water stress environments. Proceed. Pl. Breed. Con. April 26, 2003: 5557.
- ElZeir, F.A., E.A. Amer, A.A. Abdel Aziz and A.A. Mahmoud (2000). Combining ability of new maize inbred lines and type of gene action using top crosses of maize. Egypt. J. Appl. Sci. 15(2): 116128.

- Gado, H.E., M.S.M. Soliman and M.A.K. Shalaby (2000). Combining ability analysis of white maize (*Zea mays L.*) inbred lines. J. Agric. Sci. Mansoura Univ. 25:37193729
- Gilbert, N.E.G. (1958). Diallel cross in plant breeding. Heredity.12:477492
- Hallauer, A.R. and J.E. Miranda (1981). Quantitative genetics in maize breeding. 2<sup>nd</sup> Ed., Iowa State Univ. press, Ames, USA.
- Hassaballa, E.S., M.A. ElMorshidy, M. Khalifa and E.M. Shaiaby (1980). Combining ability analysis in maize. 1 Flowering. Res. Bull. Fac. of Agric., Ain Shams Univ., 1291,8pp.
- Hede, A.R., G. Srinvasan, G. Stolen and S.K. Vasal (1999). Identification of heterotic pattern in tropical inbred maize lines using broadbase synthetic testers. Maydica 44(4):325331.
- Kempthorne, O. (1957). An Introduction to Genetic Statistics .John Wiley and Sons Inc., NY,USA.
- Konak,G., A. Unay, E. Serter and H. Basal (1999). Estimation of combining ability effects, heterosis and heterobeltiosis by line x tester method in maize. Turkish J. of Field Crops. 4(1): 19 [C.F.Pl.Br.Abst. 69(11):10711].
- Lonnquist, J.H. and M.F. Lindsey (1964). Top cross versus S1 lines performance in corn (Zea mays L). Crop.Sci, 4:580584.
- Mahmoud, A.A. (1996). Evaluation of combining ability of newdeveloped inbred lines of maize. Ph.D. Thesis, Fac. Agric., Cairo University Egypt.
- Mathur, R. K., Chunilal, S.K. Bhatnagar and V. Singh (1998). Combining ability for yield, phenological and ear characters in white seeded maize. Indian J. of Genet. and Pl.Breed.58(2):177182.
- Matzinger, D.F. (1953). Comparison of three types of testers for the evaluation of inbreed lines of corn. Agron. J., 45:493495.
- Nass, L.L., M. Lima, R.Vencovesky and P.B. Gallo (2000). Combining ability of maize inbred lines evaluated in three environments in Brazil. Scientia Agricola, 57(1): 129134.
- Russell, W.A, S.A. Eberhart and U.A. Vega (1973). Recurrent selection for specific combining ability in two maize populations. Crop. Sci., 13:257-261.
- Sadek, E.S., H.E. Gado and M.S.M. Soliman (2000). Combining ability and type of gene action for maize grain yield and other attributes. J. Agric. Sci. Mansoura Univ. 25(5): 24912502.
- Sadek, S.E., M.S.M. Soliman, A.A. Barakat and K.I. Khalifa (2002). Topcrosses analysis for selecting maize lines in the early self generations. Minufiya J. Agric. Res., 27:197213.
- Shehata, A.M., F.A. ElZeir and E.A. Amer (1997). Influence of tester lines on evaluating combining ability of some new maize inbred lines. J. Agric. Sci. Mansoura Univ. 25(5):2491 2502.
- Shehata, A.H. and N.L. Dhawan (1975). Genetic analysis of grain yield in maize as manifested in genetically diverse varietal populations and their crosses. Egypt. J. Genet. Cytol. 4:90116.

#### M.E.M. Abd El Azeem, M.A. Abd El Moula and A.E.M.K. El-Galfy

- Singh, I.S. and D.B. Chaudhary (1979). Biometrical Methods in Quantitative Genetic Analysis. Kalyani Publishers. New Delhi, 3<sup>rd</sup> Ed., P.3968.
- Soliman, F.H.S. (2000). Comparative combining ability of newly developed inbred lines of yellow maize (*Zea mays* L.). Egypt. J. Appl. Sci. 15:87102.
- Soliman, F.H.S. and S.E. Sadek (1999). Combining ability of new maize inbred lines and its utilization in the Egyptian hybrids program. Bull. Fac. Agric., Cairo Univ. 50(1):120.
- Soliman, M.S.M., A.A. Mahmoud, F.A. ElZeir, Afaf A.I. Gaber and F.H.Soliman (2001). Utilization of narrow base tester for evaluating combining ability of newly developed maize inbred lines (*Zea mays* L.). Egypt. J. Plant Breed. 5:6176.
- Soliman, M.S.M. and M.M.A. Osman (2006). Type of gene action for grain yield using testcross analysis in new developed maize inbred lines. J. Agric. Sci. Mansoura Univ., 31(5):2615-2630.
- Sprague, G.F. and L.A. Tatum (1942). General vs. specific combining ability in single crosses of corn. J. Am., Agron, 34: 923-932.
- Steel, R.G. and J.H. Torrie (1980). Principles and Procedures of Statistics. Mc Grow Hill Book Inc., New York, USA.
- Waljko, R.N and W.A. Russell (1977). Evaluation of recurrent selection for specific combining ability in two open pollinated varieties. Crop Sci., 13:647-651.
- Zelleke, H. (2000). Combining ability for grain yield and other agronomic characters in inbred lines of maize (Zea mays L.). Indian J. of Genet. And Pl. Breed. 60(1): 6370.

## تقدير القدرة على التآلف لمحصول الحبوب وبعض الصفات الأخرى في الذرة الشامية

محمد المهدي محمد عبد العظيم ، مجدي احمد عبد المولى ، علاء الدين محمود خليل الجلفي

قسم بحوث الذرة الشامية - معهد بحوث المحاصيل الحقلية - مركز البحوث الزراعية

#### الملخص العربي

هجنت ١٢ سلالة في الجيل الاختصابي النذاتي الثالث(S<sub>3</sub>) من النذرة التشامية الصفراء مع ٣ كشافات (سلالات تربية داخلية) مختلفة هي : جيزة ٣٨٨ وجيزة ٢٥٠ وجميزة ١٠٢١ وذلك بمحطة البحوث الزراعية بسندس في موسيم ٢٠٠٧ . وفيي موسم ۲۰۰۸ تم تقییم ۳۳ هجین قمی مسع هجین للمقارنسة (هـــــ. ف. ۱۵۵ ، هـــــ. ف. ١٦٢ في كل من محطة البحوث الزراعية بسخا والجمييزة ، وسيدس وذليك ليصفات عد الأيام من الزراعية حتى ظهور ٥٠ % من الحرايس ، ارتفاع كل من النبات والكوز، المقاومة لمسرض السذبول المتسأخر ، محسصول الحبسوب بسالأردب للفسدان. وقسد وجدت اختلافات معوية بين الهجين القمية ، السلالات ، الكشافات لكل المصفات المدروسة ما عدا صفة عدد الأيسام من الزراعة حتى ظهور ٥٠ % من الحرايس للكشافات و صفة ارتفاع الكوز للسلالات ومعنوية لتباين تفاعل السلالات × الكشافات لكل الصفات موضع الدراسة . كذلك وجدت اختلافات معنوية بين المواقع لجميع الصفات المدروسة مما يدل عل إن المواقع مختلفة قيى الظروف البيئية. وكان تباين النفاعل بين المواقع والهجن القمية والسملالات والكشافات معنويا لجميع الصفات المدروسة ما عدا صفة ارتفاع الكوز للسلالات و صفة عدد الأيام من الزراعة حتى ظهور ٥٠ % من الحرايس للكشافات .. كمنا كنان تبناين التفاعيل بنين السلالات × الكشافات × المواقع معنوى لصفات ارتفاع النبات والكوز ومحصول الحبوب. أظهرت السلالات رقم ١و ١٢ أحسن قدرة عامة (مرغوية) لسصفة المحصول وقد اظهرت ٣

#### M.E.M. Abd El Azeem, M.A. Abd El Moula and A.E.M.K. El-Galfy

هجن قمية هي and L11 x GZ650, L2 x GZ638, L3 x GM1021, L5 x GZ638 أحصية هجن قمية المحصول. على التالف للصفة المحصول. على تباين القدرة العامة على التآلف عاليا للصفات التزهير والمحصول. وكان التباين الراجع لتفاعل القدرة الخاصة على التآلف مع المواقع أعلى من تباين تفاعل القدرة العامة على الاتتلاف مع المواقع لجميع الصفات المدروسية عدا صفة عدد الايام حتى ظهور ٥٠% من الحراير، مما يدل على ان القعل الجيني غير المحضيف لتلك المصفات الكثر تاثرا بالمواقع عن الفعل المضيف.

### FACTOR ANALYSIS AND ITS RELATIONSHIP WITH GENETIC DIVERSITY IN COTTON.

M.E. Abdelsalam, M.A. AL-Ameer and Y.M. EL-Mansy Cotton Research Institute, Agricultural Research Center, Egypt.

(Received: Apr. 4, 2010)

ABSTRACT: The relationship between the biometrical methods depended on single trait and multivariate analysis of genotypes in breeding programs are very important. Therefore, this study was carried out to access factor analysis and diversity among 13 parents and 36 F<sub>1</sub> hybrids performance to evaluate 12 variables into separate groups at the Agricultural Research Station, Sakha, Kafr El-sheikh governorate, Egypt during 2008 and 2009 seasons. The analysis of variance revealed that highly significant genotypic differences for the most traits among parents and hybrids. Multivariate analysis reported that, the first factors which accounted for 70 % of the total variance are important. Factor 1, which accounted for about 25.3 % and it was associated with micronaire reading (mic), lint index (L.I.), lint percentage (L.P. %) and degree of yellowness (+ b). Factor 2, which accounted 17.3 % and it was associated with lint quality traits i.e., fiber length (F.L.), uniformity ratio (U.R. %) and lint color (+ b).

The male parents Kar.2, Seuvin, G.75 and G.76 were grouped into 4 separate groups, these parents varied in general combining ability for the most traits. The female parents were also grouped into 4 different groups. Some of these were grouped with male parents in the same cluster showing nearly related and the other grouped in the same cluster.

Specific combining ability (S.C.A.) effect revealed that most of the combinations having high of (S.C.A.) effect were found between genetically diverse parents. The cross combination Kar.2 x (Pima S 6 x G.89) surpassed all crosses for earliness index and the common parents were distantly related .Also, not only the genetic divergence might be used choose parents for crossing, but also their performance of parents and the  $F_1$ .However (G.C.A.) and (S.C.A.) effects are more informative than performance values.

Generally, the breeder can use the parents according to divergence with performance. Also, breeder might be evaluates characters to know the relative importance of such characters in genetic variability and divergence.

Kay Words: Factor analysis, Combining ability, Genetic diversity, Cotton.

#### INTRODUCTION

Genetic relationships among various genotypes can be measured by similarity of any number of quantitative characters, where characters are agronomic parameters of plant. In determining the potential of genetically different lines and cultivars, breeders have to observe among, many different

characters that influence yield. Accurate evaluation of those characters is made more difficult by genotypes x environment interaction. Thus multivariate technique which using factor analysis have analogous efficacy to determine the most suitable combination of characters Suinaga et al (2005). Few research workers were studied factor analysis such as Walton (1972) who suggested factor analysis as a new technique to identify growth and plant characters as related to yield in spring wheat. Seyam et al (1984) used factor analysis in determining traits that could be selected for high yield in cotton program. Tadess and Bekele (2001) indicated the selection of variable in factor 1 could enable breeder to better realize the desired increment in seed yield of grass pea.

Multivariate analysis of quantitative characters has been used previously to measure genetic relationships within cotton genotypes. Categorizing genotypes accession into morphological similar groups is most useful for analysis of cultivar variability (Cox et al, 1986), selection parents for hybrids (El-Lawendey et al, 2008, Abo El-Yazeid et al, 2009 and El-Mansy et al, 2010) and for predication of variances for some characters in the  $F_2$  and inbred generations (Cowen and Frey 1987). However Hemada et al, 2006 and El-Mansy et al, 2008 used canonical analysis and principal components analysis respectively to create the genetic variability in some Egyptian cottons and estimate the relative importance of each character on total variability.

This study was undertaken in order to determine the dependence relationship between morphological characters of thirteen cotton parents and 36 hybrids using factors analysis. The study was extension to determine genetic divergence among cotton genotypes as related to general and specific combining ability to select the most suitable combinations and parents.

#### MATERIALS AND METHODS

Nine female parents comprising a broad range of Egyptian cottons and characters viz ( Giza 77, Giza 80, Giza 81, Giza 85, Giza 86, Giza 88, Giza 89, promising cross (Giza 89 x Pima S 6) and (Giza 86 x Giza 89 ) and four genetically diverse male parents. These male parents have earliness, high seed cotton yield and high lint quality characters i.e., Karshenky 2 as a Russian genotype, Seuvin as Indian genotype, Giza 75 and Giza 76 as Egyptian genotypes were crossed during 2008 growing season to generate a total of 36 hybrids.

These 36 hybrids along with 13 parents were grown in randomized block design with three replications at Agriculture Research Station, Sakha during 2009 season having 4.0 m plot length with spacing of 70 x 30 cm. Five competitive random plants were chosen from each replicate of each genotypes to record data on earliness index (E.I.), Lint yield per plant (L.Y./p.), boll weight (B.W.), lint percentage (L.P. %), lint index (L.I.), seed

index (S.I.), span length (F.L.), uniformity ratio (U.R.), fiber strength (Press.), micronaire reading (Mic.), lint reflectness (R.D. %) and yellowness (+b).

Data analysis followed three steps (i) fisher's analysis of variance. combining ability effects were computed following Singh and Chaudhry (1977). Heterozygous superiority was determined as the mean average of heterosis of F<sub>1</sub> over the mean average of their homozygous parents for each characters (ii) factor analysis as a multivariate analysis methods which aims to explain the correlation between a large set of variable in terms of a small number of underlying independent factors. It is assumed that each of the variables measures depends upon the underlying factors. The principal factor analysis method explained by Harman (1976) was followed in the extraction of the factor loading. The array of communality, the amount of the variance of a variable accounted by the common factor together, was estimated by the highest correlation coefficient in each array as suggested by Seiller and Stafford (1985). The factor loadings of the rotated matrix, the percentage variability explained by each factor and the communalities for each variable were determined. The third step (iii) clustering genotypes 13 parents and 36 hybrids into similarity groups using principal components coefficient according to principal component axis. All these computation were performed by using SPSS evaluation version 10.0 production mode facility and Minitab computer programs.

#### RESULTS AND DISCUSSION

The analysis of variance for 12 characters studied was presented in Table 1. It's revealed highly significant genotypic differences for all characters. Factorial analysis of population indicated significant differences among parents for all characters except for boll weight, seed index and Presley index. The female parents showed differences for majority of the characters.. The interaction between female and male parents was found to be significant for the characters *viz* lint yield per plant (L.P./P.), (F. L.), (U.R.), (mic.), (press.) and (+ b). Hybrids showed significant differences for most characters.

The previous results indicating that the experimental material possessed considerable amount of variability, while general and specific combining ability were involved in the genetic expression of these characters.

The magnitude of (S.C.A.) variances were greater than (G.C.A.) for the traits i.e., L.Y.,B.W, E.I,F.L,U.R and PRES, indicating that non additive type of interactions were higher among hybrids which could be exploited by heterosis breeding (Tuteja and Kumer, 2003 and Abo El-Yazied et al, 2009).

Multivariate analysis which used factor analysis was performed on 12 agronomic and fiber characters to determine which factor more effect on total variability than other, also to extract important component of variation in agronomic attributes and to obtain the initial factor solution using eigen value.

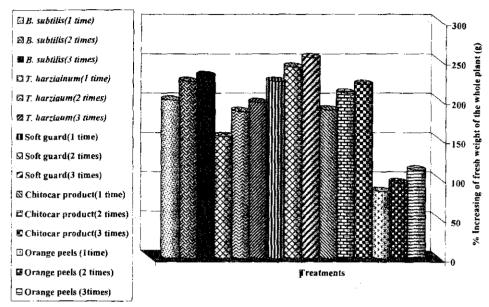


Fig. (4): Effect of applications number of some bioagents, chitosan compounds and orange peels extract treatments on increasing of fresh weight of the whole plant of strawberry infected by *M. hapla* under greenhouse conditions.

#### III- Field experiments:-

Effect of some bioagents and chitosan compounds treatments to controlling *M. hapla* in naturally infecting strawberry plants under field conditions:-

#### 1- Number of root galls:-

However all the treatments showed remarkable reduction of the number of root galls as shown in Table (4), the soft guard performed the lowest galls numbers as reached to 24 galls/root compared with the other treatments at (1:500) with three times of application. *B. subtilus* ranked on the second place resulting in reduce galls to 28 galls/root, whereas *T. harzianum* resulted in the lowest reduction in the number of root galls reached to 35 galls/root. Chitocar product occupied an intermediate position in decrease number of galls/ root reached to 31 galls/root in compared with the control 113 galls/root.

2- Number of juveniles in soil, developmental stages, females, egg masses and eggs numbers in roots:-

The same trend on number of root galls also was obtained with the effect on juveniles in soil, developmental stages, females, egg -masses and eggs numbers.

Table (4): Efficacy of some bioagents and chitosan compounds treatments on reproduction *M. hapla* in naturally infecting strawberry plants under field conditions.

nea cond								
Treatments	Nematode population in							
	No. of			Root				
ļ	in 250g No. of soil galls	No. of galls	No. of developmen tal stages	No. of females	No. of egg- masses	No. of eggs/ egg- mass		
Bacillus subtilís + N	100	28	45	31	29	330		
Trichoderma Harzianum + N	200	35	53	38	36	372		
Soft Guard +N	80	24	42	26	24	310		
Chitocar product + N	160	31	48	35	30	356		
Control (nematodes only) (N)	2800	113	117	118	117	460		
L.S.D. (5 %)	18.91	3.22	2.33	2.75	1.71	18.32		

Data in Fig.(5) illustrated the effect of the treatments on fruit yield weights of strawberry after treatments under field conditions. All the treatments showed remarkable increasing in yield weights. Using soft guard was the highest effective treatment in the increasing in fruit yield weights as reached to 8640 Kg/F. While, *T. harzianum* was the lowest effective one to increase fruit yield as reached to

7920 Kg/F.

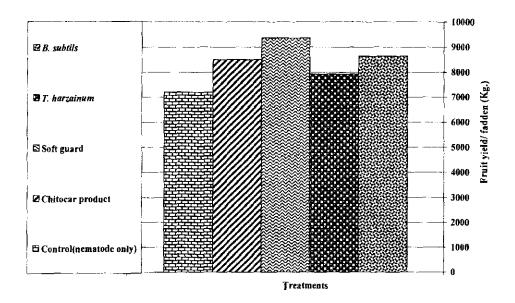


Fig. (5): Effect of some bioagents and chitosan compounds on fruit yield weight of strawberry infected naturally by *M. hapla* under field conditions.

#### Conclusion:-

From the above mentioned results, it can be concluded that soft guard compound was the most effective treatment; whereas *T. harzianum* was the lowest effective one three times soil drenching in comparing with control to control *M. hapla* under both greenhouse and field conditions and improved fruit yield of strawberry under field conditions.

#### REFERANCES

Abd El-Moity, T. H., M. H. El-Hamawi, T. El-Sharkawy and K. Tiilikkala (1998). Effect of some biologica agents on reproduction of *Meloidogyne incognita* on tomato plants. Egyptian journal of Agricultural Research. 76(1): 51-62.

Abd El- Moity, T. H. and M. N. Shatla (1981). Biological control of white disease of onion *Sclerotium cepivorum by Trichodermn harzianum*. Phytopathology (4):1187-1194.

Anonymous (1986). Manual for field trail in plant protection. Edited and Published By Werner Puntener, Agricultural Divsion, Ciba-Gaigy Limited, Basle, Switzerland.

Asaka, O. and M. Shoda (1996). Biocontrol of *Rhizoctonia solani* damping off of tomato with *Bacillus subtilis* Rb14. Applied and Environmental Microbiology, 62 (11): 4081-4085.

- Barker, K. R., C. C. Carter and J. N. Sasser (1985). An advanced treatise on *Meloidogyne* Vol.2 Methodology pag. 41
- Banhamoud, W. and I. Chet (1993). Hyphal interaction between *Trichoderma* harzianum and *Rhizoctonia solani* ultrastructure and chemistry of mycoparasitic process. Phytopathology, 83: 1026-1071.
- Bolar, J. P., J. L. Norelli, K.W. Wong, C. K. Hayes, Q. E. Harman and H. S. Aldwinckle (2000). Expression of endochitinase from *Trichodermn harziianum* intransgenic apple increase resistance to apple scab and reduces vigour. Phytopathology, 90: 72-77.
- Dowson, W. G. (1957). Plant diseases due to bacteria. Second Ed., Cambridge University, Press., London, pp. 231.
- Farahat, A. (1998). Biological control of some potato bacterial diseases. PHD. Thesis, Fac. Agric. Monofia Univ. PP118.
- Faruk, M. I., M. L. Rahman and M. A. Bari (2002). Mangement of root-knot nematodes of tomato using *Trichoderma harzianum* and organic soil amendment. Bangladesh Journal of Plant Pathology, 18: (1), 33-37,
- Ferreiro, J. H. S., F.N. Matthee and A. C. Thomas (1991). Biological control of Eutypalota on grapevine by an antagonistic strain of *Bacillus subtilis*. Phytopathology, 81: 283-287.
- Franklin, M. T. and J. B. Goodey (1957). A cotton-blue lactophenol technique for mounting plant parasitic nematodes. J. Helminthological Abstracts; 23:175-178.
- Gomez, K.W. and A. A. Gomez (1984). Statistical procedures for Agricultural research and ED. John Wiley and sons ltd , New York 680P.
- Hallmann, J., R. Rodriguez-Kabana and J.W. Kloepper (1999). Chitin-mediated changes in bacterial communities of the soil, rhizosphere and within roots of cotton in relation to nematode control. Soil Biology and Biochemistry volume 31(4): 551-560.
- Hanna, A. I., F. W. Riad and A. E. Tawfik (1999). Efficacy of antagonistic rhizobacteria on the control of root-knot nematode, *Meloidogyne incognita* in tomato plants. Egyptian Journal of Agricultural Research. 77(4): 1467-1476.
- Hussey, R. S. and K. R. Barker (1973). A comparison on methods of collecting inocula of *Meloidogyne spp.* including a new technique. Plant Dis. Reptr., 57:1925-1928.
- Khan, M. R., K. Kounsar and A. Hamid (2002). Effect of certain rhizobacteria and antagonistic fungi on root- knot. Nodulation and root-knot nematode disease of green gram. Nematologia Mediterranea, 30(1): 85-89.
- Lournedjiinon, S., I. Godonou, C. Atcha-Ahowe, B. James, H. Baimey, D. Coyne and A. Ahanchede (2007). Mangement of Root-Knot nematodes on solanum macrocarpon using Botanicals in Benin.ISHS Acta Horticulturae 752:1 International conference on indigenous vegetables and legumes. Prospects for fighting porerty trunger and malnutrition.

- Mulawarman, J., D. Hallmann, B. Bell, Kopp-Holtwiesche and R. A. Sikora (2001). Effect of natural products on soil organisms and plant health enhancement. Meded Rijksuniv Gentfak Landbouwkd Toegep Biol wet.; 66(2b): 609-617.
- Padares, D. F., J. Hockenbull, D. F. Jensen and S. B. Mathus (1992). In vivo screening of *Trichoderma* isolates for antugonism against *Sclerotium rolfsii* using rice seedlings. Bulletin oil 15 (1): 33-35. (C.P. Rev. PL. Pathol., vol. 72 No. 1, 208
- Sahebani N. and N. Hadavi (2008). Biological control of the root-knot nematode *Meloidogyne javanica* by *Trichoderma harzianum*. Journal of Soil Biology &Biochemistry, 40: 2016-2020.
- Sharon, E., M. Bar-Eyal, I. Chet, A. O. Herrera Estrella, Kleifeld and Y. Spiegel (2001).Biological control of the root-knot nematode *Meloidogyne javanica* by *Trichoderma harzianum*.Phytopathology.,91:7, 687-693.
- Shawky, Samaa M. and Maisa L. Abd El- Moneim (2005). Non chemical control of root knot nematode, *Meloidogyne incognita* on grape vines. Minufiya J. Agric. Res., 30(6): 1685- 1708.
- Siddiqui, I. A. and S. S. Shaukat (2004). *Trichoderma harzianum* enhances the production of nematicidal compounds *in vitro* and improves biocontrol of *Meloidogyne javanica* by *Pseudomonas fluorescens* in tomato. Letters in Applied Microbiology ,38(2): 169-175.
- Tsai, B.Y. (2008). Effect of peels of lemon, orange, and grapefruits against *Meloidogyne incognita*. Plant Pathology Bulletin 17: 195-201.
- Vasyukova, N. I., S. V. Zinova, L. Ilinskaya, E. A. Perkhod, G. I. Chalenko, N. G. Gerasimova, A. V. Ilina, V. P. Varlamova and O. L. Ozeretskovskaya (2001). Modulation of plant resistance to diseases by water-soluble chitosan. Applied Biochemistry and Microbiology volume 37, Number 1,103-109

# كفاءة استخدام بعض الكائنات الحية و مركبات الشيتوزان و مستخلص قشر البرتقال في مكافحة نيماتودا ميلودوجين هابلا على الفراولة المصابة بمرض تعقد الجذور في مصر

مايسة لطفي عبد المنعم' وسماء محمود شوقي ' ومنال محمد سليمان'

١ - المعمل المركزي للزراعة العضوية - مركز البحوث الزراعية -وزارة الزراعة - الجيزة - مصر.

٣- قسم بحوث الامراض النيماتودية - معهد بحوث أمراض النباتات - مركز البحوث الزراعية -وزارة الزراعـة - الجيزة - مصر

#### الملخص العربي

تم اختبار خمسة معاملات و هي: بكتريا باسيلس ساتلس و قطسر تريكودرما هارزيانم ومركب شيتوكير و مركب السوفت جارد ومستخلص قشر البرتقال. استخدمت ثلاثة تركيان مختلفة من المعاملات لمقاومة نيماتودا تعقد الجذور من النوع ميلودوجين هابلا في المعسل و الصوبة و الحقل على شتلات الفراولة (صنف كاماروزا). كذلك تم دراسة تاثير عدد مسرات الاضافة من المعاملات على مقاومة نيماتودا تعقد الجذور تحت ظروف الصوبة.

كانت اكثرالمعاملات تأثيرا تحت ظروف كلا من المعمل و الصوبة في مقاومة نيماتودا تعقد الجذور هو مركب السوفت جارد بينما كانت المعاملة بمستخلص قشر البرتقال اقلهم تأثيرا لذلك تم استبعادها في التجربة الحقلية. أظهرت المعاملة بالسوفت جارد أعلى كفاءة في مقاومية نيماتودا تعقد الجذور بينما كانت المعاملة بفطر تريكودرما هارزياتم اقلهم كفاءة تحت ظروف الحقل.

تحت ظروف المعمل أظهرت جميع المعاملات كفاءة في زيادة نسبة موت اليرقات خاصـة عند التركيزات العالية و بعد ٧٢ ساعة.

اظهر استخدام كلا من المعاملتين السوفت جارد و بكتريا باسيلس ساتلس أعلى كفاءة فسي خفض أعداد العقد النيماتودية و الإناث و الأطوار الغير مكتملة النمو و كتل البيض و عدد البيض على النبات الواحد في الجذور و كذلك تعداد البرقات من العمر الثاني في التربة بينما كانت المعاملة بمستخلص قشر البرتقال اقلهم تأثيرا تحت ظروف الصوبة.

أظهرت جميع المعاملات المستخدمة تحت ظروف الصوبة زيادة في الوزن الخضري سواء المجموع الخضري أو الجذري للشتلات خاصة عند التركيزات العالية. أدى استخدام الجرعات العالية بمعدل ثلاثة مرات إلى كفاءة عالية في خفض تعداد النيماتودا في كلا من التربة والجذور.

أظهرت جميع المعاملات تحت ظروف الحقل كفاءة فى خفض تعداد النيماتودا فى كسلا مسن التربة و الجذور بالاضافة الى زيادة في وزن محصول الفراولة عند إضافة المعساملات ثلاثسة مرات كل اسبوع باستخدام التركيز العالي.

