

## MICROBIOLOGICAL EVALUATION OF FROZEN CHICKEN NUGGETS AND STRIPS

H. A. Abd El-Rahman, Soad A. Soliman, Mona M. Abd ELWahab and  
Amany M. Ahmed

### ABSTRACT

50 samples of frozen chicken nuggets and strips (25 of each) were examined microbiologically. The mean values of microbiological indices in the frozen chicken nuggets and strips for total aerobic plate counts (cfu/g) were  $1.9 \times 10^5 \pm 4.7 \times 10^4$ ;  $7.4 \times 10^4 \pm 1.8 \times 10^4$ ; Enterobacteriaceae counts were  $7.7 \times 10^2 \pm 2.1 \times 10^2$ ;  $1 \times 10^2 \pm 1.3 \times 10$ ; *Staphylococcus aureus* counts were  $5.8 \times 10^3 \pm 1.2 \times 10^2$ ;  $3.4 \times 10^2 \pm 3.8 \times 10$ ; Aerobic spore forming count were  $2.7 \times 10^3 \pm 1.2 \times 10^2$ ;  $4.9 \times 10^3 \pm 7.8 \times 10^2$ , The identified aerobic spore forming bacteria were *B. subtilis*, *B. cereus*, *B. sterothermophilus*, *B. lentus* and *B. coagulans*. The mean value of mould and yeast counts were  $6.5 \times 10 \pm 1 \times 10$ ;  $5 \times 10^2 \pm 1.4 \times 10$ , The identified mould genera were Alternaria, Cladosporium, Rhizopus, Aspergillus, Fusarium, penicillium, Mucor, The Aspergillus and Penicillium genera were identified. The identified yeast genera were Rhodotorula, Deparomyces, Saccharomyces, Candida, Endomyces, Pichia, and Cryptococcus. The mean value of MPN/g. of Coliforms were  $1.1 \times 10 \pm 0.3 \times 10$ ;  $3.8 \times 10 \pm 0.3 \times 10$ , The Incidence of identified Enterobacteriaceae organisms were *E. coli* 5(10.6%), 7(23.3%) *K. pneumoniae* 3(6.3%), 1(3.3%), *C. diversus* 1(2.1%), (0%), *C. freundii* 11(23.4%), 3(10%), *p. mirabilis* 9 (19.1%), (0%), *p. vulgris* (0%), 2(6.6%), *serratia marcescens* 7 (14.9%), 4(13.3%), *Morgenalla morgani* 5(10.6%), 6(20%), *Enterobacter aerogenes* (0%), 4(13.3%), *Enterobacter agglomerans* 4(8.5%), (0%), *Edwardsiella tarda* 1(2.1%), 3(10%), *Shigella* spp. (0%), (0%) and salmonella spp. 1(2.1%), (0%), respectively. The isolated serotype of *E. coli* were *E. coli* O<sub>103</sub>: k 3(12%), 3(12%) and *E. coli* O<sub>119</sub>: K<sub>69</sub> 2(8%), 4(16%). The isolated serotype of salmonella organisms were *S. pullorum gallinarum* 1(4%), (0%) respectively. In all examined samples the *clostridium perfringens* couldn't be detected.

## INTRODUCTION

Chicken nuggets and strips are liable to be contaminated with different kinds of microorganisms such contaminants may be of public health hazard to consumer. The majority of food borne illness result from the under cooking of raw animal products, which allows pathogenic bacteria to survive and cause illness when ingested. *Teo et al., (1996); Noha and Gehad (2005) .El Hoti (2006)* examined chicken nuggets and fingers bacteriologically and found that both coat and meat of them had noticeably high bacterial counts and were contaminated with various food-borne pathogens.

*ICMSF (1980) and National academy of sciences (1985)* reported that salmonella and *staphylococcus aureus* organisms were commonly found on raw poultry throughout the world and inadequate cooking or cross-contamination could account for their presence in ready-to-eat poultry meats.

*Dominguez and Schaffner (2009)* recorded that frozen chicken products have been identified recently as a cause of Salmonellosis. At least eight Salmonellosis outbreaks from 1998 to 2008 have implicated in undercooked frozen chicken nuggets, strips, and entrees as infection vehicles. Thus, the presence of Salmonella in frozen poultry products may pose an infection risk if the product is improperly cooked. Mould and yeast contamination of

meat and meat products may occur during slaughtering of the animals or birds, transportation, or during further processing of meat products through the use of contaminated equipments or contaminated additives and spices which considered the most important source of mould contamination in meat products *Scott and Kennedy (1973), Flanniga and Hui (1976), Misra (1981) and Abdel-Rahman (1987)*. Moreover, sensory deviation may occur in addition to increase microbial, biological and toxicological contamination resulted from the use of avian skin and mechanically deboned poultry meat in the formulation of the Egyptian meat products *Emara and Nouman (2003)*.

Although some mould species are beneficial and used in the industrial purposes as in ripening of certain types of meat products, other are saprophytic and largely concerned with the decomposition of organic matter and decay of meat products. The senescence of lipase enzyme by many species of moulds particularly *Aspergillus* and *Penicillium* may cause hydrolysis and decomposition of fat which lead to severe economic losses *Frazier (1976)*. Also several species of moulds are responsible for production of toxic metabolites (mycotoxins) which hazard human and animal health *Gracey (1981)*. The objective of this study is to determine the microbial quality of frozen chicken nuggets and strips.

## MATERIALS AND METHODS

The methods applied in bacteriological indices were carried out as recommended by APHA, (1992), ICMSE, (1978) Beerens et al., (1980) and Quinn et al., (1994). While for Total Mould and Yeast Counts were determined according to ARX Von (1967). The genus *Aspergillus* was identified according to Rapper and Fennel (1965) and Samson (1979), the genus *Penicillium* according to Rapper and Thorn (1949) and other mould genera according to ARX Von (1967). The yeast genera were identified according Deak and Beuchat (1996).

## RESULTS & DISCUSSION

The results given in table (1&2) revealed that the mean values of TAPC (cfu/g) in frozen chicken nuggets and strips were  $1.9 \times 10^5 \pm 4.7 \times 10^4$ ;  $7.4 \times 10^4 \pm 1.8 \times 10^4$ , respectively. The obtained results is lower than those recorded by El-Tahan (2006); El-Bassuony (2008); Al-Dughaym and Altabari (2010), and nearly similar to those recorded by Mukprasirt et al., (2001); Osman (2001); El Hoti (2006); Bkheet et al., (2007); Eglezos et al., (2008) but higher than that recorded by Othman (1997); Noha and Gehad (2005) and Qoboory (2008). The mean value of Enterobacteriaceae count in frozen chicken nuggets and strips were 7.7

$\times 10^2 \pm 2.1 \times 10^2$ ;  $1 \times 10^2 \pm 1.3 \times 10$ , respectively. The obtained results are lower than those recorded by Mohamed (1996) but higher than that recorded by Hefnawy and Moustafa (1990); Bkheet et al., (2007). The mean value of *Staphylococcus aureus* count in frozen chicken nuggets and strips were  $5.8 \times 10^3 \pm 1.2 \times 10^2$ ;  $3.4 \times 10^2 \pm 3.8 \times 10$ , respectively. The obtained results is higher than those recorded by Noha and Gehad (2005); El-Tahan (2006); Al-Dughaym and Altabari (2010) but nearly similar to those that recorded by Othman (1997); El-Dosoky (2004); El Hoti (2006). The mean value of Aerobic spore forming count in frozen chicken nuggets and strips were  $2.7 \times 10^3 \pm 1.2 \times 10^2$ ;  $4.9 \times 10^3 \pm 7.8 \times 10^2$ , respectively. The obtained results is lower than those recorded by El-Dosoky (2004) but nearly similar to those that recorded by Nassif (1996); Mira and Abouzied (2006). Frequency distribution of identified aerobic spore forming bacteria in the examined frozen samples of chicken nuggets and strips were *B.subtilis* 14 (40%), 9 (36%); *B.cereus* 11(31.4%), 13 (52%); *B. sterothermophilus* 7(20%), 2(8%); *B. lents* (0%), 1 (4%); *B. coagulans* 3(8.6), (0%) respectively.

The mean value of mould and yeast count in frozen chicken nuggets and strips were  $6.5 \times 10 \pm 1 \times 10$ ;  $5 \times 10^2 \pm 1.4 \times 10$ , respectively. The frequency percentage of

identified mould genera in frozen chicken nuggets and strips were *Alternaria* 44(20.7%) , 30(21.3%) ; *Cladosporium* 35 (16.4%), 20 (14.2%); *Rhizopus* 5 (2.3%), 3 (2.1%); *Aspergillus* 59 (27.7%), 43 (30.5%); *Fusarium* 10 (4.7%), 2 (1.4%); *penicillium* 45 (21.1%), 38 (27%); *Mucor* 15 (7%), 5 (3.5%) respectively.

The frequency distribution of *Aspergillus* in frozen chicken nuggets and strips were *A. flavus* 10 (16.9 %), 11(25.6%); *A. fumigatus* 12(20.3%) ,10(23.3%) ; *A. niger* 12(20.3%) , 8(18.6%); *A. terreus* 11(18.6 %), 6(14 %); *A. oryzae* 9(15.3%) , 5(11.6%) and *A. versicolor* 5(8.5%) , 3(7%) respectively.

The frequency distribution of *Penicillium* in chicken nuggets and strips were *P. cyclopium* 20(44.4%), 15 (39.5%); *P. veredictum* 15 (33.3%), 13 (34.2%) and *P. chrysogenum* 10(22.2%), 10(26.3%) respectively. The Frequency percent of identified yeast genera in the examined Frozen chicken Nuggets and Strips were *Rhodotorula* 20 (20.40%), 10 (20.40%); *Debaromyces* 17(17.35%), 9(18.4%); *Saccharomyces* 17(17.35 %), 8 (16.3 %); *Candida* 15 (15.31 %), 8 (16.3 %); *Endomyces* 12(12.24 %), 7 (14.3%); *Pichia* 9 (9.18 %),5 (10.2%); *Cryptococcus* 8(8.17%), 2(4.1%), respectively. The mean value of MPN/g. of Coliforms in frozen chicken nuggets and strips

were  $1.1 \times 10 \pm 0.3 \times 10$  ;  $3.8 \times 10 \pm 0.3 \times 10$ , respectively. The obtained results is lower than those recorded by *El-Dosoky (2004)*; *El Hoti (2006)*; *Bkheet et al., (2007)*; *El- Bassuony (2008)*; nearly similar to those that recorded by *Othman (1997)*; *Eglezos et al., (2008)*.

The Incidence of isolated Enterobacteriaceae organisms among frozen chicken nuggets and strips *E.coli* 5(10.6%), 7(23.3%) *K. pneumoniae* 3(6.3%), 1(3.3%), *C. diversus* 1(2.1%), (0%), *C. freundii* 11(23.4%), 3(10%), *p. mirabilis* 9 (19.1%) , ( 0%), *p. vulgris* ( 0%) , 2( 6.6%), *serratia marcescens* 7( 14.9%) , 4(13.3%), *Morgenella morgani* 5(10.6%),6(20%), *Enterobacter aerogenes* (0%), 4(13.3%), *Enterobacter agglomerans* 4(8.5 %), ( 0 %), *Edwardsiella tarda* 1( 2.1%) , 3( 10%), *Shigella* spp.

( 0%),( 0%) and salmonella spp. 1( 2.1%) , ( 0%), respectively. The isolated serotype of *E.coli* in chicken nuggets and strips were *E.coli* O<sub>103</sub>: k- 3( 12%), 3(12%) and *E.coli* O<sub>119</sub>: K<sub>69</sub> 2( 8%), 4 ( 16 %), respectively. a significant public health importance where the diarrhoeal disease is a major cause of illness and death among infants and young children worldwide *Frenzen et al., (2005)*; *Amela et al.,(2009)*. The isolated serotype of salmonella organisms among chicken nuggets and strips were *S. pullorum gallinarum* 1(4%), (0%)

respectively. The presence of salmonella species in the products reflects the degree of sanitation in the processing plant and will harm the consumer health as Salmonellosis caused by *S. pullorum* has also been reported in humans *Savage and Darre (2008)*; *Saif et al., (2003)*. In all examined samples

of frozen chicken nuggets and strips, the *clostridium perfringens* couldn't be detected, which agree with the *E.S,(2005 - 3493)* ; *Mukprasirt et al., (2001)*; *EL-Tahan (2006)* not agree with *Kessel et al., (2001)* who could detect *CL. Perfringens* with 21% .

**Table (1): Microbiological indices of frozen chicken Nuggets.**

Microbiological indices	Min	Max	Mean	S.E.
Total Aerobic Plate Counts	$1.2 \times 10^4$	$7.4 \times 10^5$	$1.9 \times 10^5$	$4.7 \times 10^4$
Enterobacteriaceae counts	$1.3 \times 10^2$	$4 \times 10^3$	$7.7 \times 10^2$	$2.1 \times 10^2$
<i>Staphylococcus aureus</i> counts	$1.3 \times 10^2$	$6.2 \times 10^3$	$5.8 \times 10^2$	$1.2 \times 10^2$
Aerobic spore forming Counts	$1.1 \times 10^3$	$3.2 \times 10^4$	$2.7 \times 10^3$	$2.7 \times 10^3$
Moulds and yeast counts	$1.3 \times 10$	$5 \times 10^2$	$6.5 \times 10$	$1 \times 10$

**Table (2): Microbiological indices of frozen chicken Strips**

Microbiological indices	Min	Max	Mean	S.E.
Total Aerobic Plate Counts	$2 \times 10^2$	$4.3 \times 10^5$	$7.4 \times 10^4$	$1.8 \times 10^4$
Enterobacteriaceae counts	$4 \times 10$	$5.8 \times 10^2$	$1 \times 10^2$	$1.3 \times 10$
<i>Staphylococcus aureus</i> counts	$6 \times 10$	$5.5 \times 10^3$	$3.4 \times 10^2$	$3.8 \times 10$
Aerobic spore forming Counts	$1 \times 10^2$	$2.6 \times 10^4$	$4.9 \times 10^3$	$7.8 \times 10^2$
Moulds and yeast counts	$2 \times 10^2$	$1 \times 10^3$	$5 \times 10^2$	$1.4 \times 10$

**Table (3): Incidence of isolated Enterobacteriaceae species in examined frozen chicken nuggets and strips**

Microorganisms	Frozen Nuggets		Frozen Strips	
	No.	%	No.	%
<i>Escheichia coli</i>	5	10.6	7	23.3
<i>Klebsiella pneumoniae</i>	3	6.3	1	3.3
<i>Citrobacter diversus</i>	1	2.1	-	-
<i>Citrobacter freundii</i>	11	23.4	3	10
<i>Proteus mirabilis</i>	9	19.1	-	-
<i>Proteus vulgaris</i>	-	-	2	6.6
<i>Serratia marcescens</i>	7	14.9	4	13.3
<i>Morgenalla morganii</i>	5	10.6	6	20
<i>Enterobacter aerogenes</i>	-	-	4	13.3
<i>Enterobacter agglomerans</i>	4	8.5	-	-
<i>Edwardsiella tarda</i>	1	2.1	3	10
<i>Shigella spp.</i>	-	-	-	-
<i>Salmonellae spp.</i>	1	2.1	-	-
<b>Total</b>	<b>47</b>	<b>100</b>	<b>30</b>	<b>100</b>

**Table (5): Serotypes of E.coli and salmonella in chicken nuggets and strips.**

Serotypes	Frozen Nuggets		Frozen Strips	
	No.	%	No.	%
<i>E.coli</i> O <sub>103</sub> : k.	3	12	3	12
<i>E.coli</i> O <sub>119</sub> : K <sub>69</sub>	2	8	4	16
<i>S. pullorum gallinarum</i>	1	4	0	0

**Table (6): Conformity of the microbiological indices of frozen Chicken Nuggets with the Egyptian Standardization (ES: 2005 -- 3493).**

Microbial indices	E.S. limit	Within limit		Exceed limit	
		No.	%	No.	%
Aerobic plate counts	< 10 <sup>4</sup>	0	0	25	100
Enterobacteriaceae counts	< 10 <sup>2</sup>	7	28	18	52
<i>Staphylococcus aureus</i> Counts	0	11	44	14	56
Aerobic spore forming counts	< 10 <sup>2</sup>	0	0	25	100
Mould and yeast counts	0	16	64	9	36
<i>Cl. Perfringens</i>	0	25	100	0	0
<i>E. coli</i>	0	7	28	18	52
<i>Salmonella .spp</i>	0	24	96	1	4

**Table (7): Conformity of the microbiological indices of examined frozen Chicken Strips with the Egyptian Standardization (2005 - 3493).**

Microbial indices	E.S. limit	Within limit		Exceed limit	
		No.	%	No.	%
Aerobic plate counts	$< 10^4$	12	48	13	52
Enterobacteriaceae counts	$< 10^2$	14	56	11	44
<i>Staphylococcus aureus</i>	0	13	52	12	48
Aerobic spore forming counts	$< 10^2$	0	0	25	100
Mould and yeast counts	0	14	56	11	44
<i>Cl. Perfringens</i>	0	25	100	0	0
<i>E. coli</i>	0	10	40	15	60
<i>Salmonella spp.</i>	0	25	100	0	0

## CONCLUSION:

From the above results we can concluded that the chicken nuggets and strips were subjected to contamination during processing; from the raw materials or unsanitary measuring during manufacturing, also it may be due to unsuitable environmental condition during storage. The large percent of samples which are not conformed to the Egyptian standards reflect the unsatisfactory hygienic standard and the quantity and quality of additives used during processing of Chicken Nuggets and Strips.

## REFERENCES

Abdel-Rahman, H.A.; Youssef, H.; and Hefnawy, Y. (1987): Mycological quality of meat products in Egypt. *Asiut Vet. Med. J.*, 12 (24) 153-159.

Al-Dughaym, A. M.; Altabari, G. F. (2010): Safety and quality of some chicken meat products in Al-Ah-sa markets - Saudi Arabia. *Saudi J. of Biological science* (2010) 17, 37-42.

Amela, D.; Mirsada, H.; Daria, B. and Amra Z. (2009): Frequency and Distribution of Diarrhoeagenic *Escherichia Coli* Strains Isolated From Pediatric Patients With Diarrhea In Bosnia And Herzegovina. *Bosnian J. of basic medical sciences*; 9 (2): 148-155.

APHA (1992): Compendium of methods for microbiological examination of food. 3<sup>rd</sup> Ed. American public health association, Washington DC.

Arx, J. A. Von (1967): *Pilzkunde von J. Cramer*. In der A.R.Ganter Verlag. Kommanditgesellschaft. F1-9490 Vaudz.

- Beerens, H.; Remond, C.H.; Lepage, C. and Criquelion, J. (1980):** A direct method for the enumeration of *Clostridium Perfringens* in foods and faeces. Proc. 2<sup>nd</sup>. World congress of food borne infection and intoxication. Berlin-west.
- Bkheet, A. A.; Rezk, M. SH. and Mousa, M.M. (2007):** Study on the microbiological content of local manufactured poultry meat products in El-Bahira governorate. Assuit Vet. Med. J. 53 no. 112:115-125.
- Deek, T. and Beuchaut, L. R. (1996):** Hand book of food spoilage yeasts. CRC Press, Boca Raton, Fl. USA.
- Dominguez, S. A. and Schaffner, D. W. (2009):** Survival of *Salmonella* in Processed Chicken Products during Frozen Storage. Journal of Food Protection, 72, (10), 2088-2092.
- Eglezos, S.; Dykes, G. A.; Huang, B.; Fegan, N.; Stuttard, E. (2008):** Bacteriological profile of raw, frozen chicken nuggets. J. Food Prot.; 71(3): 613-615.
- Egyptian organization for standardization and quality control (E. S (2005):** Egyptian standards for poultry meat products treated with heat.
- El-Bassuony, R. A. (2008):** Bacterial evaluation of frozen poultry and some poultry products in Luxor city. J. Egypt Vet. Med. Assoc. 68(4):295-307.
- El-Dosoky, H. F.A. (2004):** Hazards of ready to eat chicken meat. ph.D. thesis for fac. of Vet. Med. Zagazig University.
- El-Hoti, F.A.I. (2006):** Quality improvement of battered and breaded formed poultry meat. M. V. Sci., Thesis., Fac. of Vet. Med., Cairo Uni.
- El-Tahan, F.H.; Abd El-Salam, A.F. and El Tahan, M.H. (2006):** Microbiological and chemical properties in chicken products collected from local market. J. Agric. Scinc. Mansoura Univ. 31(2) 989-997.
- Emara, M.M. and Nouman, T. (2003):** Detection of avian skin in meat products. (Personal communication)
- Flanniga, B. and Hui, S. (1976):** The occurrence of aflatoxin producing strain of *Aspergillus Flavus* in the mould flora of ground spices. J. Food Bacteriol., 41:411-418.
- Frazier W.C. (1976):** Food microbiology: Manual for identification of fungi. 2<sup>nd</sup> Ed., Brogyers A.W. (ed.), OSIO Norway.
- Frenzen, P. D., Drake, A., Angulo, F. J. and Emerging infections program foodnet working group. (2005):** Economic cost of illness due to *E.coli* O157 infections in the United States. J. of food protection, 68 (12): 2623 – 2630.
- Gracey, J.F. (1981):** Meat microbiology and food poisoning, Thronton's Meat Hygiene. 7<sup>th</sup> Ed., Ch. 13, English language Book Society / Bailliere Tindall .UK.



- Hefnawy, J. and Moustafa, S. (1990):** Quality evaluation of ready to eat poultry in Assiut city. Assiut Vet. Med. J. 73 (46): 119 – 125.
- ICMSF (1978):** Micro-organisms in foods: their significance and methods of enumeration. Univ. of Toronto press, Toronto ,Canada.
- ICMSF (1980):** Microbial Ecology of foods vol. I factors affecting life and death of microorganisms and vol. 2 food commodities. Acad. press, NY.
- Kessel, A. S.; Gillespie, I. A. ;O' Brien S. J. ; Adak, G. K.; Humphrey, T. J. and Ward, L.R. (2001):** General outbreaks of infectious intestinal diseases linked with poultry England Wales , 1992-1999. Communicable disease and public health, 4 :171 -177.
- Mira, E. K. I. and Abouzied, S. M. A. (2006):** Prevalence of *Bacillus cereus* and its enterotoxin in some cooked and half cooked chicken products. Assiut Vet. Med. J. 52 (109): 70 – 78.
- Misra, N. (1981):** New records of fungi from the bark of *cinnamon* in storage. Science and culture, 49 (5) 133-135.
- Mohamed , A.S. (1996):** Hygienic quality of some poultry products. M.V. Sci., vet. Med., Alex. Univ.
- Mukprasirt, A.; Herald, T. J.; Boyle, D. I. and Boyle, E. A. (2001):** Physicochemical and microbiological properties of selected rice flour – based batters for fried chicken drumsticks. J. poultry Science -80:988-996.
- Nassif , M. R. M (1996):** Occurrence and significance of aerobic spore forming microorganisms in some meat products with special reference to *bacillus cereus*. Ph. D. V. Sci., Fac. of vet. Med., Suez Canal Univ.,
- National academy of sciences (1995):** An evaluation of the role of microbiological criteria for foods and food ingredients. National academy press, Washington, DC.
- Noha, R. M. and Gehad, F. A. (2005):** Bacteriological status of some chicken products.J. Egypt Vet. Med. Assoc. 65 (3): 295- 306.
- Othman., E.M.S. (1997):** Bacteriological evaluation of heated chicken and chicken products with special reference to enterotoxigenic *staphylococcol* isolates.Thesis for M.V. Sci., Fac. of vet. Med. Cairo Univ..
- Osman , E. (2001):** Quality assurance of locally dressed broilers and their products. Ph. D. Thesis. , Faculty of Vet. Med, Cairo University.
- Qoboory, H.A.H. (2008):** Study the changes in quality attributes of semi fried chicken during home freezing. A thesis Submitted to the Degree of M.Sc. in Home Economics Department: Nutriion & Food science. KSA.

- Quinn, P.J.; Carter, M.E.; Markey, B.K. and Carter, G.R.(1994):** Clinical veterinary microbiology. Wolfe publishing, Mosby-year book - Europe Limited London WCIH, LB England.
- Rapper, K.B. and Fennel, D.I. (1965):** The genus *Aspergillus*. Williams & Wilkins., Co., Baltimore Reprint Hafner, New York., USA.
- Rapper, K.B. and Thorn, C. (1949):** A manual of the penicillia . Williams & Wilkins., Co., Baltimore.
- Saif, Y. M.; Barnes, H. J. ; Glisson, J. R.; Fadly, A. M.; McDougald, L. R. and Swayne, D.E. (2003):** Diseases of poultry. 11<sup>th</sup> ed. Iowa State University Press. Blackwell pub. Company. Section II, P: 568.
- Savage, T and Darre, M.J. (2008):** Fact sheet by Tina savage, university of new Hampshire Cooperative Extension Agricultural Resources Educator in collaboration with Michael J. Darre, University of Connecticut. <http://extension.unh.edu/Agric>.
- Samson, R. A. (1979):** A compilation of the *Aspergilli* described since 1965 Studies in Mycology. 18: 1-38.
- Scott, P.M. and Kennedy, B.P. (1973):** Analysis and survey of ground black, white, and red peppers for aflatoxins. J.A.O.A.C., 56 (6)1452.
- Teo, Y.; Rayonor, T.J.; Ellajosyula, K.R. and Knabel, S.J. (1996):** Synergistic effect of high temperature and high pH on destruction of salmonella enteritidis and E.coli O157:H7. J. of food protection 59; (10) 1023 – 1030.

### الملخص العربي

#### دراسات على صلاحية بانية وناجتس الدجاج المجمد

أ.د. حسنى عبد اللطيف عبد الرحمن، د. منى محمد عبد الوهاب، د. سعاد أحمد سليمان،  
أماني مأمون أحمد

تم جمع 50 عينة من ناجتس وبانية الدجاج المجمد ( 25 من كل نوع) حيث تم تقييم المنتج بالفحص الميكروبيولوجي، وكان متوسط العدد الكلي للبكتريا الهوائية في ناجتس و بانية الدجاج المجمد  $10 \times 1.9 \pm 10 \times 4.7$ ؛  $10 \times 7.4 \pm 10 \times 1.8$  و متوسط العدد الكلي للميكروبات القولونية  $10 \times 7.7 \pm 10 \times 2.1$ ؛  $10 \times 1 \pm 10 \times 1.3$  و متوسط العدد الكلي لميكروب العنقود الذهبى  $10 \times 5.8 \pm 10 \times 1.2$ ؛  $10 \times 3.4 \pm 10 \times 3.8$  و متوسط العدد الكلي للبكتريا الهوائية المتجرثة  $10 \times 2.7 \pm 10 \times 1.2$ ؛  $10 \times 4.9 \pm 10 \times 7.8$  على التوالي. كانت النسبة المنوية لعزل البكتريا الهوائية المتجرثة كالتالى باسيلس ساتلس 14(40%)، 9(36%)؛ باسيلس سيريس 1(31.4%)، 13(52%)؛ باسيلس ستيروثيرموفيلس 7(20%)، 2(8%)؛ باسيلس لينتس 0(%)، 1(4%)؛ باسيلس كواجيلنس

3 (8.6%)، (0%) على التوالي. كان متوسط العدد الكلى للفطريات والخمائر  $10 \times 6.5 \pm 1$  ×  $10 \times 5 \pm 1.4$  على التوالي.

امكن عزل 213 عترة تنتمي الى 7 انواع من الفطريات هي ألتيرنيريا 44 (20.7%)، 30 (21.3%) ؛ كلاوسبوريم 35 (16.4%)، 20 (14.2%)؛ ريزوبس 5 (2.3%)، 3 (2.1%)؛ اسبيرجلس 59 (27.7%)، 43 (30.5%)؛ فيوزاريم 10 (4.7%)، 2 (1.4%)؛ بينسليوم 45 (21.1%)، 38 (27%)؛ ميوكرا 15 (7%)، 5 (3.5%) . كانت نسبة جنس الاسبرجلس موزعه كالتالى اسبرجلس فلافس 10 (16.9%)، 11 (25.6%)؛ اسبرجلس فيوميچيتس 12 (20.3%)، 10 (23.3%)؛ اسبرجلس نيجر 12 (20.3%)، 8 (18.6%)؛ اسبرجلس تيريس 11 (18.6%)، 6 (14%)؛ اسبرجلس اوريازى 9 (15.3%)، 5 (11.6%)؛ اسبرجلس فيريزىكلر 5 (8.5%)، 3 (7%) . كانت نسبة البنسليوم موزعه كالتالى بنسليوم سيكلوبيوم 20 (44.4%)، 15 (39.5%)؛ بنسليم فيرنديكاتوم 15 (33.3%)، 13 (34.2%)؛ بنسليوم كريزوجينم 10 (22.2%)، 10 (26.3%) على التوالي.

امكن عزل 98 عترة من الخمائر تنتمي الى رودوتريولا 20 (20.4%)، 10 (20.4%)؛ دييروميسيز 17 (17.35%)، 9 (18.4%)؛ سكاروميسز 17 (17.35%)، 8 (16.3%)؛ كانديدا 15 (15.31%)، 8 (16.3%)؛ ايندوميسز 12 (12.24%)، 7 (14.3%)؛ بيشيا 9 (9.18%)، 5 (10.2%)؛ كريبتوكوكس 8 (8.17%)، 2 (4.1%) على التوالي. كان متوسط العد الاكثر احتمالية للميكروبات القولونية (MPN /g)  $10 \times 3.6 \pm 10 \times 0.9 \pm 2$  ،  $10 \times 2.7 \pm 2$  على التوالي. تم عزل الميكروبات القولونية بالنسب الاتية ايشريشيا كولى 5 (10.6%)، 5 (10.6%)؛ كليسيلا نيمونيا 3 (6.3%)، 1 (3.3%)؛ سيتروبيكتر دايفرسس 1 (2.1%)، (0%)؛ سيتروبيكتر فريندى 11 (23.4%)، 3 (10%)؛ بروتيس ميرابيلس 9 (19.1%)، (0%)؛ بروتيس فالجريس (0%)، 2 (6.6%)؛ سيراتيا مارسينس 7 (14.9%)، 4 (13.3%)؛ مورجينلا مورجاني 5 (10.6%)، 6 (20%)؛ انتيروبيكتر ايروجينز (0%)، 4 (13.3%)؛ انتيروبيكتر اجلوميرنز 4 (8.5%)، (0%)؛ ادورديلا تاردا 1 (2.1%)، 3 (10%)؛ شيجلا (0%)، (0%)؛ سالمونيللا 1 (4%)، (0%) على التوالي. تصنيف الايشريشيا كولاى :  $O_{103} : k$  بنسبة 3 (12%)، 3 (12%)؛  $O_{119} : k_{69}$  بنسبة 2 (8%)، 4 (16%) على التوالي. تم عزل سالمونيللا باللورم جالينيرم بنسبة 1 (4%)، (0%) على التوالي.