

THE EFFECTS OF DIETARY EGYPTIAN PROPOLIS AND BEE POLLEN SUPPLEMENTATION AGAINST TOXICITY OF SODIUM FLUORIDE IN RATS

Fatma A. Khalil and Nora M. El-Sheikh

Biochemistry and Nutrition Department,
Women's College, Ain Shams University, Cairo, Egypt.

Received: 27/12/2010

Accepted: 05/12/2010

SUMMARY

Propolis and bee pollen are substances produced by honey bees. Its components are strong antioxidants and free radical scavengers. The present study aimed to study the protective effects of propolis and bee pollen supplementation against toxicity of sodium fluoride (F) in rats. Rats were divided into six groups each 10 rats and treated for 6 weeks: group 1 as control group; group 2 fed on standard diet with F; groups (3-6) fed on standard diet with F and supplemented with different concentration of propolis or bee pollen at 0.1%, 0.2% and 1%, 2% respectively. After the end of experimental period, the rats were sacrificed and biochemical analyses were carried out. The results showed that the administration of fluoride alone caused a significant increase of malondialdehyde (MDA) level and a significant decrease of antioxidant system

as measured by erythrocyte superoxide dismutase (SOD) activity and reduced glutathione (GSH) levels in blood and brain. Also F caused a significant increase in alkaline phosphatase (ALP) activity, urea, creatinine, sodium and potassium levels. Moreover, a significant decrease in total protein, calcium, magnesium and phosphorus levels as compared to control group ($P < 0.05$) was recorded. The administration of propolis or bee pollen with F led to a significant decrease in MDA level and a significant increase in SOD activity, GSH levels in blood and brain. As well as significant decrease in ALP activity, urea, creatinine, sodium and potassium levels in serum was also observed. The propolis or bee pollen supplementation enhanced total protein, calcium, magnesium and phosphorus levels in serum as compared to F group alone.

In conclusion; supplementation of natural antioxidants (propolis or bee

pollen) during Fluoride administration, facilitates reduction of the toxic effects and enhanced both the antioxidant system, as well as the levels of minerals in the serum.

Keywords:

Propolis, bee pollen, sodium fluoride, rats, antioxidant system, minerals.

INTRODUCTION

Fluoride (F) is a highly electronegative anion with cumulative toxic effects, from prolonged ingestion that can lead to the pathogenesis known as fluorosis which is a condition especially persistent in the third world countries, where populations have little choice as to the main source of their often times-F-contaminated drinking, other sources include private water supplies, dietary ingredients, dental products, industrial emissions, and/or occupational exposure, which can cause an individual's total F intake to exceed safe dose (Ozsvath, 2009).

Fluoride crosses the cell membrane very rapidly and then distributes from the plasma to all tissue and organs (Bouaziz et al., 2006).

Propolis and bee pollen are natural substances collected by honey bees from buds and trees. Propolis is a sticky substance that have bees manufacture by mixing their own waxes with resinous sap (Yoshimi et al., 2009). The main chemical

classes found in propolis are flavonoids, phenolic and various aromatic compounds. Propolis also contains many of B-complex vitamins, important mineral and trace elements, caffeic acid phenethyl ester (CAPE) which is an active component of propolis exhibits antioxidant properties.

Nowadays propolis is used in many medical formulae and food supplements for improving health, preventing and treating infections, inflammatory diseases and effects of toxic substances (Attalla and Owayss, 2008).

Bee pollen is rich in carotenoids, flavonoids and phytosterols. The exact profile varies depending on the plant sources and growing conditions. However, beta-carotene, beta sitosterol, isohammetin, kaempferol, lycopene, quercetin and rutin are consistent (Campos et al., 2003). The antioxidant activity of flavanoids present in propolis and bee pollen has been shown to be capable of scavenging free radical (Survswaran et al., 2007).

The aim of this study was therefore to evaluate the protective effects of dietary Egyptian propolis and bee pollen supplementation against toxicity of sodium fluoride in rats.

MATERIAL AND METHODS

1. Materials

Sodium fluoride was purchased from El Gomhoriee Chemical Co. The sodium fluoride was added to the standard

diet at 1 g/kg diet to induce toxicity (Bellack and Schoube, 1968).

The propolis and bee pollen used in the present study originated from the hive in Cairo, Egypt. These products were harvested in September 2009. Bee pollen was obtained as yellow granules, while propolis was derived in the form of yellow-brown powder.

2. Experimental animals:

Adult male albino rats weighing (130 ± 13.9 g) were kept in [12:12 h (light: dark) photo period] and temperature ($22 \pm 0.5^\circ\text{C}$) controlled room maintained at constant relative humidity of 65-70% and fed a standard diet and water *ad libitum*.

3. Diet:

The standard diet was prepared according to (Revees et al., 1993).

Table (1): Composition of the standard diet.

Ingredient	g/kg
Protein (Casein)	140
Corn starch	610
Sucrose	100
Soy been oil	50
Fiber	50
Mineral mix.	35
Vitamin mix.	10
L. cystine	1.8
Choline bitartrate	2.5
Tert-butylhydroquinone	0.08

4. Experimental design:

Rats were fed on the standard diet and then they were divided into 6 groups (10 rats in each group):

Group (1): Control rats (without any supplementation).

Group (2): Rats fed on standard diet supplemented with sodium fluoride

(F) alone 1 g/kg diet to induce F toxicity.

Group (3): Rats fed on standard diet supplemented with F and propolis powder in diet 0.1% (Haro et al., 2000).

Group (4): Rats fed on standard diet supplemented with F and propolis powder in diet 0.2%.

Group (5): Rats fed on standard diet supplemented with F and bee pollen in diet 1% (Haro et al., 2000).

Group (6): Rats fed on standard diet supplemented with F and bee pollen in diet 2%.

5. Biochemical analysis:

After the end of the experimental period (42 days), all rats were fasted overnight and sacrificed. Brain sample and two blood samples were collected from each animal, with and without anticoagulant for biochemical analysis. Reduced glutathione (GSH) was measured in blood and brain homogenate according to the method of Beutler et al. (1963). Malondialdehyde (MDA) level was measured in brain according to Satoh (1978).

Erythrocyte superoxide dismutase (SOD) activity was determined in accordance with the method described by Sun et al. (1988). Alkaline phosphatase (ALP) activity in serum was determined by the method of Anon (1974), serum total protein level was determined by the method of Gornall et al. (1949). Serum creatinine and urea were determined by the methods of Bonsens and Taussky (1984) and Patton and Crouch (1977), respectively.

Serum sodium, potassium, calcium, magnesium and phosphorus were

estimated by the colorimetric method of Berry et al. (1988), Sunderman and Sunderman (1958), Sarkar and Chauvan (1967), Teitz (1983) and Drewes (1972), respectively.

6. Statistical analysis:

Results are expressed as mean \pm SD. The data were statistically analyzed following the one way analysis of variance [ANOVA, F test and least significant difference (L.S.D)] at ($P < 0.05$) were carried out using SPSS version 11 (Levesque, 2007) SPSS Chicago / L, USA.

RESULTS

From the results shown in Table (2), it was evident that fluoride administration alone in group (2) caused a significant increase of MDA level in the brain and significant decrease in GSH levels in blood and brain, erythrocyte SOD activity as compared to control group ($P < 0.05$). However, the administration of propolis or bee pollen with fluoride significantly decrease the MDA level in the brain and also significantly increase GSH levels in the blood and brain, erythrocyte SOD activity as compared to F group ($P < 0.05$) and were nearly to control group.

Table (2): Effects of propolis and bee pollen supplementation on lipid peroxide as (MDA) and antioxidant system in rats (mean \pm SD).

Parameters Groups	Brain MDA (nmol/ g tissue)	Blood GSH (mg/dL)	Brain GSH (mg/g tissue)	Erythrocyte SOD activity (U/mL)
Group (1) (control)	^d 0.70 \pm 0.11	^a 28.09 \pm 1.91	^b 14.99 \pm 0.99	^a 277.75 \pm 5.70
Group (2) (F group)	^a 3.08 \pm 0.23	^e 16.27 \pm 0.94	^e 8.04 \pm 0.27	^d 177.88 \pm 10.13
Group (3) (F + propolis 0.1%)	^e 0.51 \pm 0.08	^c 22.16 \pm 1.20	^c 13.25 \pm 0.88	^b 265.75 \pm 6.78
Group (4) (F + propolis 0.2%)	^b 1.10 \pm 0.12	^d 20.13 \pm 1.18	^d 9.75 \pm 0.46	^{bc} 260.25 \pm 4.68
Group (5) (F + bee pollen 1%)	^c 0.93 \pm 0.10	^a 27.70 \pm 2.13	^b 14.76 \pm 0.91	^a 279.38 \pm 10.15
Group (6) (F + bee pollen 2%)	^c 0.88 \pm 0.08	^b 25.38 \pm 1.07	^a 16.14 \pm 1.43	^b 270.75 \pm 7.98
L.S.D.	0.135	1.49	0.91	7.93

n=10 rats each group.

Values in the same column with different superscripts vary significantly ($P < 0.05$).

Table (3) shows that there was a significant increase in ALP activity, urea and creatinine, and a significant decrease in total protein in F group as compared to control group ($P < 0.05$). However, propolis or bee pollen improved the liver

and kidney function which manifested by decrease the ALP activity, urea and creatinine and increase the total protein in treated groups as compared to F group and were nearly to the control group.

Table (3): Effects of propolis and bee pollen supplementation on serum ALP activity, total protein, urea and creatinine in rats (mean \pm SD).

Parameters Groups	ALP activity (IU/L)	Total protein (g/dL)	Urea (mg/dL)	Creatinine (mg/dL)
Group (1) (control)	^e 167.56 \pm 5.23	^a 5.71 \pm 0.52	^b 33.93 \pm 2.96	^b 1.51 \pm 0.08
Group (2) (F group)	^a 323.13 \pm 11.63	^d 3.68 \pm 0.11	^a 63.57 \pm 6.20	^a 2.06 \pm 0.14
Group (3) (F + propolis 0.1%)	^c 258.00 \pm 11.14	^c 4.21 \pm 0.18	^b 36.68 \pm 3.40	^d 1.05 \pm 0.04
Group (4) (F + propolis 0.2%)	^b 269.75 \pm 7.74	^c 4.27 \pm 0.11	^b 34.68 \pm 2.61	^d 1.09 \pm 0.08
Group (5) (F + bee pollen 1%)	^{bc} 266.63 \pm 11.39	^c 4.36 \pm 0.10	^b 35.19 \pm 2.01	^c 1.22 \pm 0.10
Group (6) (F + bee pollen 2%)	^d 235.25 \pm 9.21	^b 4.83 \pm 0.15	^b 35.33 \pm 2.32	^d 1.08 \pm 0.07
L.S.D.	9.76	0.25	3.64	0.09

n=10 rats each group.

Values in the same column with different superscripts vary significantly ($P < 0.05$).

Table (4) and Figs. (1-5) show the levels of serum cations in control group (G 1), fluoride group (G 2) and treated group (G 3 - G 6).

There was a significant increase in serum sodium and potassium levels and significant decrease in serum calcium,

magnesium and phosphorus in F group as compared to control group ($P < 0.05$). Whereas the administration of propolis and bee pollen improved the levels of cations in treated groups as compared to F group ($P < 0.05$) and were nearly to the control group.

Table (4): Effects of propolis and bee pollen supplementation on serum cations in rats (mean \pm SD).

Parameters Groups	Sodium (mmol/L)	Potassium (mmol/L)	Calcium (mmol/L)	Magnesium (mmol/L)	Phosphorus (mmol/L)
Group (1)	d 159.38 \pm 10.16	d 4.94 \pm 1.12	a 3.48 \pm 0.20	bc 1.41 \pm 0.20	a 4.36 \pm 0.11
Group (2)	a 272.75 \pm 9.79	a 12.19 \pm 1.40	d 2.34 \pm 0.14	d 1.06 \pm 0.08	d 3.01 \pm 0.14
Group (3)	c 203.75 \pm 18.10	c 6.49 \pm 0.30	b 3.09 \pm 0.12	bc 1.49 \pm 0.08	b 3.80 \pm 0.09
Group (4)	b 237.38 \pm 11.65	b 8.05 \pm 0.28	bc 2.84 \pm 0.14	bc 1.49 \pm 0.08	c 3.55 \pm 0.16
Group (5)	d 158.13 \pm 7.74	c 7.06 \pm 0.61	b 2.98 \pm 0.19	b 1.54 \pm 0.05	a 4.28 \pm 0.13
Group (6)	d 160.13 \pm 9.82	b 8.33 \pm 0.86	c 2.83 \pm 0.09	a 1.66 \pm 0.04	a 4.26 \pm 0.14
L.S.D.	11.79	0.87	0.15	0.1	0.13

n=10 rats each group.

Values in the same column with different superscripts vary significantly ($P < 0.05$).

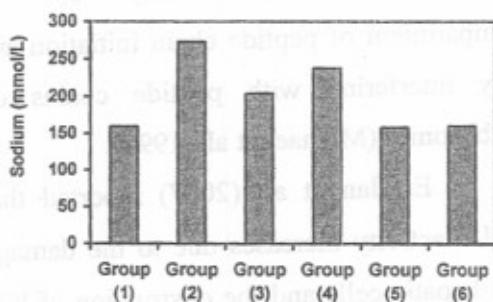


Fig. (1): Effects of propolis and bee pollen on serum sodium in rats.

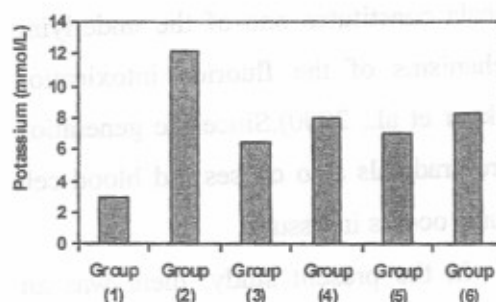


Fig. (2): Effects of propolis and bee pollen on serum potassium in rats.

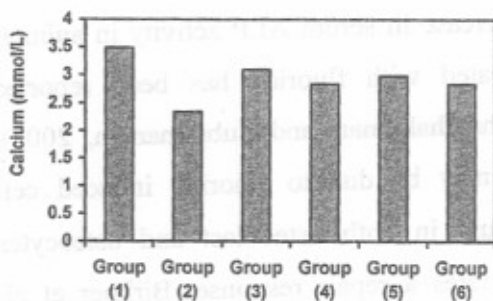


Fig. (3): Effects of propolis and bee pollen on serum calcium in rats.

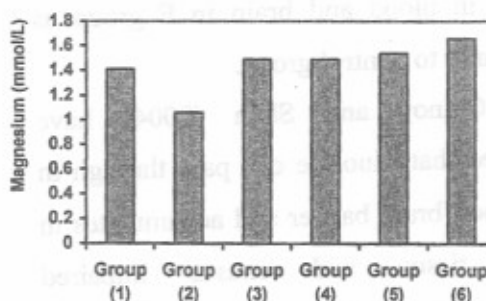


Fig. (4): Effects of propolis and bee pollen on serum magnesium in rats.

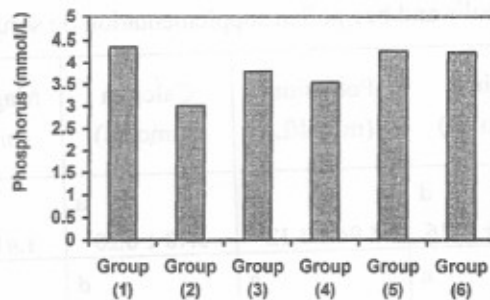


Fig. (5): Effects of propolis and bee pollen on serum phosphorus in rats.

DISCUSSION

Reactive oxygen species (ROS) play key roles in many physiologic and pathogenic processes. In fact, many ophthalmologic and neurodegenerative diseases seem to be mediated, at least in part, by oxidative stress (Finkel and Halbrook, 2000). The generation of free radicals constitutes one of the underlying mechanisms of the fluoride intoxication (Birkner et al., 2000). Since the generation of free radicals also causes red blood cell damage occurs in tissues.

In the present study, there was an increase in the level of MDA in brain and decrease of SOD activity and glutathione levels in blood and brain in F group as compared to control group.

Chinoy and Shah (2004) have reported that, fluoride can pass through in the blood brain barrier and accumulates in brain tissue and causes impaired antioxidant defense system. Furthermore, other researchers obtained similar results of increase in MDA level in chronic

fluoride intoxication (Kumari and Rao, 1991).

In the present study, the obtained results revealed that a decrease in total protein and increase in ALP activity, urea and creatinine in F group as compared to control group. Fluoride is known to inhibit protein synthesis, mainly due to impairment of peptide chain initiation and by interfering with peptide chains on ribosomes (Michael et al., 1996).

Eraslan et al. (2007) reported that ALP activity increases due to the damage of hepatic cells and the obstruction of bile ducts. ALP is the marker enzyme of fluoride toxicosis and bone pathology. The increase in serum ALP activity in animals treated with fluoride has been reported (Shanthakumari and Subramanian, 2007), it may be due to fluoride induced cell injury in both osteoblast and osteocytes initiates a repair response. Birkner et al. (2000) have also reported an increase in the serum urea level of rats with acute fluoride intoxication. The administration of

fluoride suggests failure of excretion in the kidney.

In the present study, the F group showed a significant increase in the serum potassium and sodium levels, calcium, magnesium and phosphorus decreased significantly as compared to control group. The results are similar to Chinoy et al. (1993) who have reported that demonstrated rats fed with sodium fluoride, it may lead to alteration in adrenergic function. Fluoride interacts and alters the metabolism of calcium and magnesium, the decrease in serum calcium related to decrease of intestinal absorption of calcium by fluoride (Xin et al., 2006).

Propolis and bee pollen are apicultural products which are composed of nutritionally valuable substances and contain considerable amounts of polyphenol substances which may act as potent antioxidant (Teixeira et al., 2008). Flavonoids and phenolic acids are major classes of polyphenolic compounds, whose structure-antioxidant activity (Gardjeva et al., 2007). Mechanisms of antioxidant action may include suppression of ROS formation, removal or inactivation of oxygen reactive species and up-regulation or protection of antioxidant defenses (Montoro et al., 2005).

The results of the present study revealed that, supplementation of propolis or bee pollen to rats with F toxicity led to a significant decrease in MDA level in brain

and a significant increase in antioxidant system as SOD activity and GSH levels. The propolis or bee pollen significantly decrease the ALP activity, urea, creatinine, potassium and sodium levels. Also propolis or bee pollen enhanced total protein, calcium, magnesium and phosphorus levels.

Caffeic acid phenethyl ester (CAPE) is an active component of propolis and has been used in traditional medicine to treat a number of diseases, CAPE treatment have been shown to protect tissues from ROS mediated oxidative stress and reduce lipid peroxidation in ischemia and toxic injuries. The antioxidant activity of CAPE is due to the presence of two hydroxyl groups in its structure (Sud'ina et al., 1993).

Twelve different flavonoids, pinocembrin, acacetin, chrysin, rutin, catechin, naringenin, galangin, luteolin, kaempferol, a pigenin, myricetin and quercetin, two phenolic acids, cinnamic and caffeic acid are present in propolis (Volpi, 2004).

Propolis contain acid derivatives such as benzoic-4-hydroxy benzoic which improves the digestive utilization of calcium, phosphorus and magnesium (Haro et al., 2000).

Propolis has an anabolic effect and bee pollens are rich in essential amino acids, protein, unsaturated fatty acids and also contains many vitamins, minerals and trace elements which contribute to the

health effects (Campos et al., 2003). Pollen is extremely rich in rutin and may have the highest content than other source. Bee pollen has been shown to improve immune system and remove toxins from our bodies (Campos et al., 1997).

The recent investigations indicated that bee pollen contain a significant amount of polyphenolic substances, mainly flavonoids. The polyphenols also have metal chelation properties and free radical scavenging activity (Abdella et al., 2009).

The conclusion of the present study suggests that the propolis or bee pollen and its components, are strong antioxidants and free radical scavengers. Ameliorated the liver, kidney and brain from toxicity with sodium fluoride as well as enhanced the levels of minerals in serum.

REFERENCES

- Abdella, E.M.; Tohamy, A. and Ahmad, R.R. (2009): Antimutagenic activity of Egyptian propolis and bee pollen water extracts against cisplatin-induced chromosomal abnormalities in bone marrow cells of mice. *I.J.C.P.* 2, (4).
- Anon (1974): Recommended methods for the determination of four enzymes in blood. *Scand. J. Clin. Lab. Invest.* 33. 291-306.
- Attalla, F. El-Kott and Owayss, A.A. (2008): Protective effects of propolis against the amitraz hepatotoxicity in mice. *J. Pharmacol. Toxicol.* 3(5): 402-408.
- Bellack, E. and Schoube, P.J. (1968): Rapid photometric determination of fluoride with SPADNS-zirconium lake. *Anal. Chem.* 30: 2032-2034.
- Berry, M.N.; Mazzachi, R.D.; Pejakovic, M. and Peake, M.J. (1988): Enzymatic determination of sodium in serum. *Clin. Chem.* 34: 2295-2298.
- Beutler, E.; Duron, O. and Kelly, M.B. (1963): Improved method of estimation of blood glutathione. *Lab. Clin. Med.* 61(5): 882.
- Birkner, E.; Grucka, E.; Machoy, Z.; Tarnawski, R. and Polaniak, R. (2000): Disturbance of protein metabolism in rats after acute poisoning with sodium fluoride. *Fluoride*, 33: 182-186.
- Bonsens, K.E. and Taussky, S. (1984): Determination of serum creatinine. *J. Ch. Inv.* 27: 648-660.
- Bouaziz, H.; Ketata, K.; Jammoussi, T.; Boudawara, F.; Ayedi, F. and Ellouze, N. (2006): Effects of sodium fluoride on hepatic toxicity in adult mice and their suckling pups. *Pestic. Biochem. Physiol.* 86. 124-130.
- Campos, M.; Markham, K.; Mitchel, K.; Proena Dacunha, A. (1997): An approach to the characterization of bee pollen via their flavonoid/phenolic profiles. *Photochem . Anal .* 8: 181-185.
- Campos, M.G.; Webby, R.F. and Markhan, K.R. (2003): Age-induced diminution of free radical scavenging capacity in bee pollens and the contribution of constituent flavonoids. *J. Agric. Food Chem.* 51(3): 742-745.
- Chinoy, N.J. and Shah, S.D. (2004): Biochemical effects of sodium fluoride and arsenic trioxide toxicity and their reversal in the brain of mice. *Fluoride*, 37(2): 80-87.
- Chinoy, N.J.; Sharma, N.I. and Michael, M. (1993): Beneficial effects of ascorbic acid and calcium on reversal of fluoride toxicity in male rats. *Fluoride*, 26(1): 45-56.

- Drewes, P.A. (1972): Direct colorimetric determination of phosphorus in serum and urine. *Clin. Chim. Acta.* 39: 81-88.
- Eraslan, G.; Kanbur, M. and Silici, S. (2007): Evaluation of propolis effects on some biochemical parameters in rats treated with sodium fluoride. *Pestic. Biochem. Physiol.* 88: 273-283.
- Finkel, T. and Halbrook, N.H. (2000): Oxidants, oxidative stress and biology of aging. *Nature*, 408: 239-247.
- Gardjeva, P.A.; Dimitrova, S.Z.; Kostadinov, I.D.; Murdjeva, M.A.; Peyche, L.P.; Lukanov, L.K.; Stanimirova, I.V. and Alexandrov, A.S. (2007): A study of chemical composition and antimicrobial activity of Bulgarian propolis. *Folia Med. (Plovdiv)* 49(3-4): 63-69.
- Gornall, A.G.; Bardawill, C.S. and David, M.M. (1949): Determination of serum protein by means of the biuret reaction. *J. Biol. Chem.* 177: 751-760.
- Haro, A.; Aliaga, L.I.; Lisbona, F.; Barrionuevo, M.; Alferez, M. and Campos, M. (2000): Beneficial effect of pollen and/or propolis on the metabolism of iron, calcium, phosphorus and magnesium in rats with nutritional ferropenic anemia. *J. Agric. Food Chem.* 48: 5715-5722.
- Kumari, D.S. and Rao, R.P. (1991): Red cell membrane alterations in human chronic fluoride toxicity. *Biochem. Int.* 23: 639-648.
- Levesque, R. (2007): Programming and data management: A Guide for SPSS and SAS users. 4th ed. SPSS Inc. Chicago, III.
- Michael, M.; Barot, V.V. and Chinoy, J.N. (1996): Investigations of soft tissue functions in fluorotic individuals of North Gujarat. *Fluoride*, 29(2): 63-71.
- Montoro, P.; Braca, A.; Pizz, C. and De Tomasi, N. (2005): Structure-antioxidant activity relationships of flavonoids isolated from different plant species. *Food Chem.* 92: 349-355.
- Ozsvath, D.L. (2009): Fluoride and environmental health: a review. *Rev. Environ. Sci. Biotechnol.* 8: 59-79.
- Patton, C.J. and Crouch, S.R. (1977): Determination of serum urea. *Anal. Chem.* 49: 464.
- Reeves, P.; Nielson, F. and Fahmy, G. (1993): Purified diets for laboratory rodents: Final report of the American Institute of Nutrition on the reformation or rodent diet. *J. Nutr.* 123: 1939-1951.
- Sarkar, B.C.R. and Chauvan, U.P.A. (1967): A new method for determining microquantities of calcium in biological materials. *Anal. Biochem.* 20: 155-166.
- Satoh, K. (1978): Serum lipid peroxide in cerebrovascular disorders: determined by a new colorimetric method. *Clinica. Chimica Acta.* 90: 37-43.
- Shanthakumari, D. and Subramanian, S. (2007): Effect of fluoride intoxication on bone tissue of experimental rats. *Res. J. Environ. Sci.* (3): 82-92.
- Sud'ina, G.F.; Mirzoeva, O.K.; Puskareva, M.A.; Korshunova, G.A.; Sumbatyan, N.V. and Varfolomeev, S.D. (1993): Caffeic acid phenethyl ester as a lipoxygenase inhibitor with antioxidant properties. *FEBS Lett.* 329: 21.
- Sunderman, F.W. Jr. and Sunderman, F.W. (1958): A rapid reliable method for serum potassium using tetraphenylboron. *Am. J. Clin. Pathol.* 29: 95.
- Sun, Y.; Oberley, L.W. and Li, Y. (1988): Simple method for clinical assay of superoxide dismutase. *Clin. Chem.* 343: 497-500.

- Surveswaron, S.; Cai, Y.Z.; Carke, H. and Sun, M. (2007): Systematic evaluation of natural phenolic antioxidants from 133 Indian medicinal plant. *Food Chem.* 102: 938-953.
- Teitz, N.W. (1983): *Clinical guide to laboratory tests.* Sanders Co.
- Teixeira, E.W.; Message, D.; Negri, G.; Salatino, A. and Stringheta, P.C. (2008): Seasonal variation, chemical composition and antioxidant activity of Brazilian propolis samples. *eCMA*.177: 1-9.
- Volpi, N. (2004): Separation of flavonoids and phenolic acid from propolis by capillary zone electrophoresis. *Electrophoresis*, 25: 1872-1878.
- Xin, T.; Zi Rong, X.U. and Yi Z.W. (2006): Effects of dietary fluoride levels on growth, serum indexes and antioxidant system in growing pigs. *Turk. J. Vet. Anim. Sci.*, 30: 65-70.
- Yoshimi, N.; Kazuhiro, T.; Masamitsu, S.; Satoshi, M. and Hiseaki, H. (2009): Comparison of bee products based on assays of antioxidant capacities. *Bio. Med. Central: Complementary and Alternative Medicine*, 9: 4.

تأثير المتناول الغذائي من صمغ العسل وحبوب اللقاح ضد سمية فلوريد الصوديوم في الجرذان

فاطمة عبد الحميد خليل ونورا محمد الشيخ

قسم الكيمياء الحيوية والتغذية - كلية البنات - جامعة عين شمس
القاهرة - جمهورية مصر العربية

صمغ العسل وحبوب اللقاح مواد من منتجات عسل النحل وهي مواد مضادات أكسدة قوية وتخلص الجسم من الشقوق الحرة .

تهدف الدراسة الحالية إلى معرفة التأثير الوقائي ضد السمية المحدثة بواسطة فلوريد الصوديوم في الجرذان . قسمت الجرذان إلى ست مجموعات كل مجموعة تحوى عشرة حيوانات ، المجموعة الأولى تناولت الغذاء المتوازن بدون أي إضافات ، المجموعة الثانية تناولت الغذاء المتوازن بالإضافة إلى فلوريد الصوديوم ١ جم/كيلو جم ، المجموعات من ٣-٦ تناولت غذاء متوازن بالإضافة إلى فلوريد الصوديوم وتم تدعيمها بصمغ العسل وحبوب اللقاح بتركيزات ٠.١% ، ٠.٢% ، ١% ، ٢% على الترتيب و التوالي . بعد انتهاء فترة التجربة (٦ أسابيع) تم تشريح الجرذان وكانت النتائج كالتالي: المجموعات التي تناولت فلوريد الصوديوم بمفرده بدون أي إضافات ظهرت بها ارتفاع معنوي في مستوى المألون داي ألدهيد بينما ظهر انخفاض معنوي في مستوى مضادات الأكسدة بالجسم وهي تتمثل في انخفاض قيم الجلوتاثيون المختزل وكذلك انخفاض نشاط إنزيم السوبر أوكسيد ديسميوتيز في كل من الدم والمخ .

أيضاً تناول فلوريد الصوديوم بمفرده أدى إلى ارتفاع في نشاط الفوسفاتيز القاعدي ومستوى اليوريا ، الكرياتينين ، الصوديوم والبوتاسيوم كما تسبب في انخفاض مستوى البروتين الكلي ، الكالسيوم ، الماغنسيوم والفوسفور مقارنة بالمجموعة الضابطة (الكنترول) . بينما إعطاء صمغ العسل وحبوب اللقاح كمكمل غذائي أدى إلى تقليل تأثير السمية المحدثة بواسطة فلوريد الصوديوم وظهر في ارتفاع نشاط السوبر أوكسيد ديسميوتيز ومستوى الجلوتاثيون المختزل في الدم والمخ . كما أظهر تحسن في مستوى البروتين الكلي ، الكالسيوم ، الماغنسيوم والفوسفور وحدث انخفاض في مستوى الصوديوم والبوتاسيوم.

وخلص هذا البحث إلى أن تناول صمغ العسل بتركيز ٠.١% و ٠.٢% وحبوب اللقاح بتركيز ١% و ٢% كمكمل غذائي إلى الوجبات يؤدي إلى تقوية الجهاز المناعي ويقلل من تأثير السموم كما يرفع مستوى الكالسيوم والفوسفور ويحسن مستوى البروتين في الجسم ويرفع مستوى مضادات الأكسدة في الجسم لكونهم مضادات أكسدة طبيعية .