

Effect Of Different Doses Of Cadmium Chloride On Interleukin-2, Lymphocyte Transformation And DNA Fragmentation In Adult Male Albino rats

Kawther A El-Hady, Abdel-Hamid A K and Elgawish R A

Department of Forensic Med. and Toxicology, Faculty of Vet. Medicine, Suez Canal University

ABSTRACT

Forty adult male albino rats were divided randomly into 4 equal groups (one control and three treated groups). The control group received only dis. water, while the treated groups received 5 ppm, 50 ppm and 100 ppm of cadmium chloride (CdCl₂) for consecutive 90 days in drinking water. Blood samples were collected just before scarification of the rats and serum was separated. The level of interleukin-2 in the serum of control and treated male albino rats was measured by ELISA. Lymphocyte transformation and DNA fragmentation assay in blood lymphocyte of rats were evaluated. At Day 90 of the experiment, rats were sacrificed by cervical dislocation and thymus was collected, weighted and stored in 10% neutral buffered formalin for histopathology. Data were analyzed by one-way analysis of variance using GraphPad Prism software. The interleukin-2 level was declined in a dose-dependent manner in treated rats with different doses (5, 50 and 100 ppm) of CdCl₂. Obviously the rats treated with 100 ppm of CdCl₂ showed a significant decrease (P<0.001) in the level of interleukin-2 compared to control. Moreover, rats administrated 100 ppm of CdCl₂ showed a significantly (P<0.05) high rate of lymphocyte transformation compared to control. Rats treated with 5, 50 and 100 ppm of CdCl₂ showed certain levels of DNA strand breaks in blood lymphocyte. Male rats exposed to 100 ppm (P<0.001) and 50 ppm (P<0.05) of CdCl₂ showed a significant decrease in the relative thymus weight than that of control ones. The thymus of rats treated with 50 and 100 ppm of CdCl₂ showed a reduction in the medulla's diameter and interlobular fat globules compared to control. In conclusion, CdCl₂ decreased the production of interleukin-2 in dose-dependent pattern and triggered the lymphocyte transformation particularly in rats exposed to 100 ppm of CdCl₂. While, CdCl₂ at different doses might induce DNA fragmentation in blood lymphocyte and might lead to thymic degeneration.

INTRODUCTION

There have been some conflicting results in the effect of cadmium (Cd) exposure on various immune components. Despite, the well-documented effects of Cd (1-2) some studies demonstrated only negligible influences of Cd on the immune response. Chronic Cd feeding of 5 mg/kg body wt up to 7 weeks in rats (3) and for 2-6 months in the primate Rhesus monkeys (4) did not result in any signs of immunomodulation. Furthermore, proliferation was insignificantly increased in these monkeys. Similarly, chronic exposure of B6C3F1 mice to 10-250µg/ml of CdCl₂ for 90 days, revealed no changes in humoral immunity even though Cd had accumulated in significant quantities in the tissues (5). Cd at a dose of 20 µg/ml preferentially enhances the proliferation of murine TH2 cells activated *in vitro* by concanavalin A (6).

Interleukin-2 is a cytokine released from T-helper lymphocytes plays a central role in the activation and proliferation of lymphocytes. IL-2 plays a pivotal role in the expansion of most T-cells, natural killer cells and B-cells during certain phases of their response. Through these mechanisms, it plays an important role in antitumor immune responses (7).

There are different research results on the effect of cadmium on lymphocyte transformation. In some reports, cadmium did not significantly affect the response of lymphocyte proliferation by concanavalin A (Con A) (8), while in other reports Spleen cells from mice given 200 ppm Cd for 3 to 4 weeks showed increased proliferative responses to Con A (9). The stimulation index of spleen lymphocytes to Con A tended to be higher in cadmium-treated animals than control mice (10).

Apoptosis, or physiological cell death, can be distinguished from necrosis on the basis of a series of morphological and biochemical parameters. Among these parameters, DNA fragmentation is very typical of the apoptotic process. *In vitro*, thymocytes exposure to Cd resulted in a DNA ladder confirming the induction of apoptosis by the heavy metal. The same result has been observed for mouse thymocytes (11), human T cell line (12), human promonocytic cells (13) and normal human mononuclear cells (14).

The objective of this study is to evaluate the effect of different doses of cadmium chloride in drinking water for 90 days on interleukin-2, lymphocyte transformation, DNA fragmentation and thymus histopathology in adult male albino rats.

MATERIALS AND METHODS

Experimental animals

Forty male albino rats weighted (100-140g) were assigned randomly and divided into 4 equal groups, one control and three treated groups. All groups received distilled water for one week. From the second week of the experiment, the control group received only dis. water, while the treated groups received 5 ppm, 50 ppm and 100 ppm of CdCl₂ for 90 days.

Interleukin assay

Blood samples were collected from control and CdCl₂ treated rats at the end of experimental period and serum was separated. Serum Interleukin-2 assay was measured using rat IL-2 ELISA kit (Cat # KA0276, Abnova, USA).

Lymphocyte transformation test

Blood samples were collected from control and treated rats in heparinized tubes just before slaughter of the rats. The sample immediately put on ice bags and transferred immediately to the laboratory for lymphocyte transformation assay by MTT reduction assay (15-19).

DNA fragmentation assay

DNA extraction

Blood samples were collected from control and treated rats in heparinized tubes just before

slaughter of the rats. The sample immediately put on ice bags and transferred immediately to the laboratory for DNA fragmentation assay. Lymphocyte were pelleted by centrifugation at 200 *xg* for 10 min and the pellet was lysed with 0.5 ml lysis buffer (10mM Tris-HCl, pH7.5, 20mMEDTA and 0.5% Triton X-100) on ice for 30 min. The DNA in lysed solution was extracted with phenol/chloroform and precipitated with 3M sodium acetate (pH 5.2) and cold ethanol. After repeated washings, the DNA was dissolved in TE buffer (10mM Tris-HCl, pH 8.0, and 1mM EDTA). The purity of DNA at 260 and 280 nm absorbance ratio was between 1.7 and 1.9.

DNA fragmentation test

The internucleosomal fragmentation pattern (DNA ladder) was carried out by Agarose gel electrophoresis as DNA (2 μ g) was loaded on 0.7% agarose gel and electrophoresis carried out then the bands were visualized by ethidium bromide staining under UV light (20-21).

Organ relative weights

At the end of the study period, rats were sacrificed by cervical dislocation and thymus was dissected. Thymus were removed and weighed. The organ relative weight (organ weight / body weight X 100) was measured for each treated and control groups.

Histopathological examination

Immediately after scarification of the rats, the rats were necropsied and all organs and tissue were subjected to gross examination. Whole thymus was fixed in 10% neutral buffered formalin. They were gradually dehydrated and embedded in paraffin; 5- μ m sections were stained with hematoxylin and eosin (H&E) for histopathological examination (22).

Statistical analysis

Data were analyzed by one-way analysis of variance followed by Tukey's posttest using GraphPad Prism version 3 for Windows (GraphPad Software, San Diego California, USA). A p-value below 0.05 was considered statistically significant.

RESULTS

Interleukin-2

The interleukin-2 level was declined in a dose-dependent manner in treated rats with different doses (5, 50 and 100 ppm) of CdCl₂. Obviously the rats treated with 100 ppm of CdCl₂ showed a significant decrease ($P < 0.001$) in the level of interleukin-2 compared to control and rats treated with 5 ppm of CdCl₂ (Table 1 and Fig. 1).

Lymphocyte transformation assay

Rats received 100 ppm CdCl₂ in drinking water for 90 days showed a significant increase

in the lymphocyte transformation level. However, rats treated with 5 and 50 ppm of CdCl₂ showed a relatively decline in the level of lymphocyte proliferation than that of control rats (Table 1 and Fig. 2).

Thymus weight

The relative weight of the thymus was decreased in a dose dependent pattern as male rats exposed to 100 ppm ($P < 0.001$) and 50 ppm ($P < 0.05$) of CdCl₂ showed a significant decreased in the relative thymus weight than the control ones (Table 1).

Table 1. Effect of CdCl₂ (5, 50 and 100 ppm in drinking water) on interleukin-2, lymphocyte transformation and thymus relative weight (mean \pm SE) in treated rats for 90 days

	Control	5 ppm	50 ppm	100 ppm
Interleukin-2 (pg/ml)	30.6 \pm 1.9 ^a	14.9 \pm 1.3 ^b	7.4 \pm 0.4 ^{cd}	2.7 \pm 0.3 ^d
Lymphocyte transformation (OD)	0.6 \pm 0.05 ^a	0.4 \pm 0.04 ^a	0.4 \pm 0.02 ^a	0.9 \pm 0.01 ^b
Thymus relative weight (gm)	0.05 \pm 0.01 ^a	0.04 \pm 0.003 ^{ab}	0.03 \pm 0.01 ^b	0.02 \pm 0.004 ^b

Superscripts with dissimilar values are significantly different within the same line

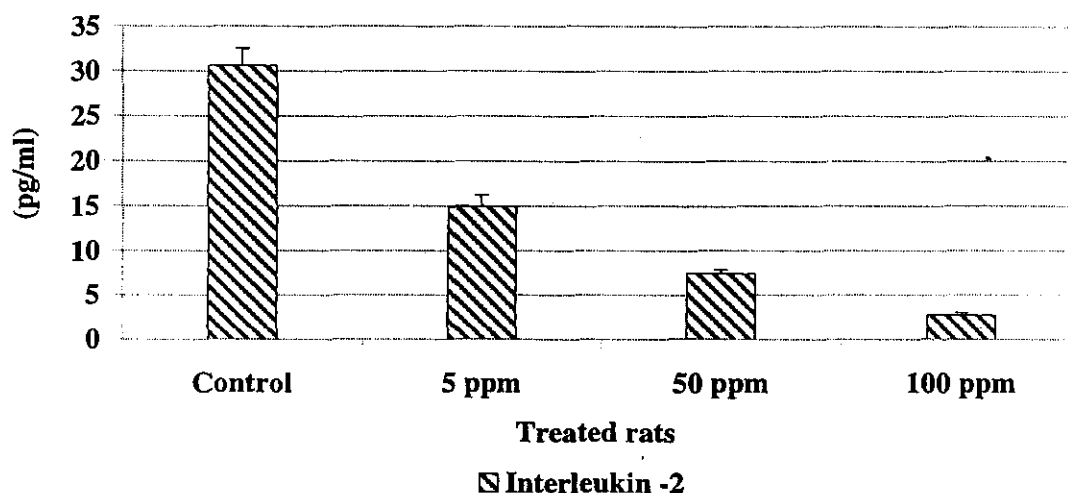


Figure 1. Effect of CdCl₂ (5, 50 and 100 ppm in drinking water) on interleukin-2 (mean \pm SE) in treated male albino rats at Day 90

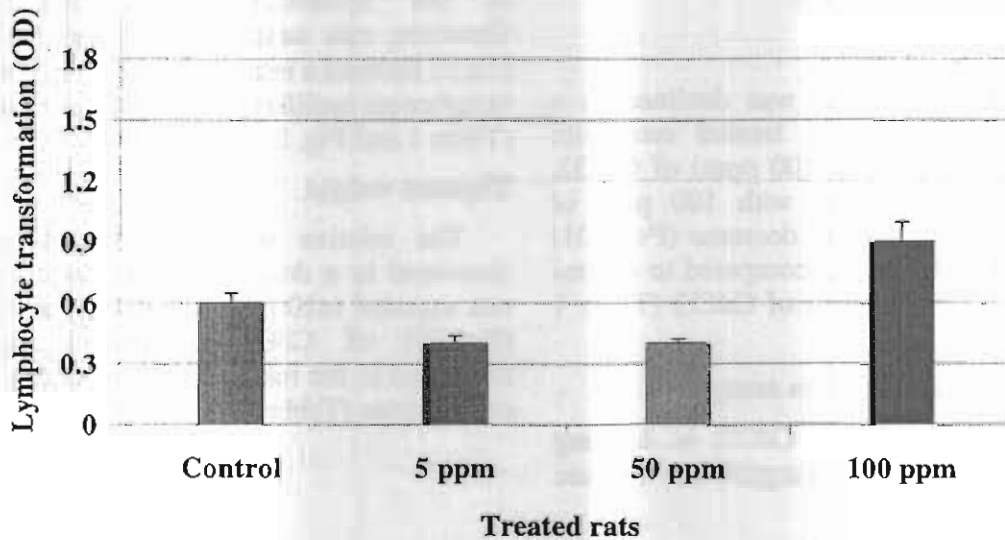


Figure 2. Effect of CdCl₂ (5, 50 and 100 ppm in drinking water) on lymphocyte transformation test (mean ± SE) in treated rats at Day 90

DNA fragmentation test

The DNA fragmentation assay indicated the level of DNA strand break due to the CdCl₂ exposure. Moreover, it indicated the level of apoptosis in the lymphocyte of cadmium chloride treated rats. The incidence

of DNA break and apoptosis increased where the dose of CdCl₂ increased. The high level of DNA fragmentation (23.7%) is mostly observed in rats exposed to 100 ppm CdCl₂ for 90 days (Table 2 and Fig. 3).

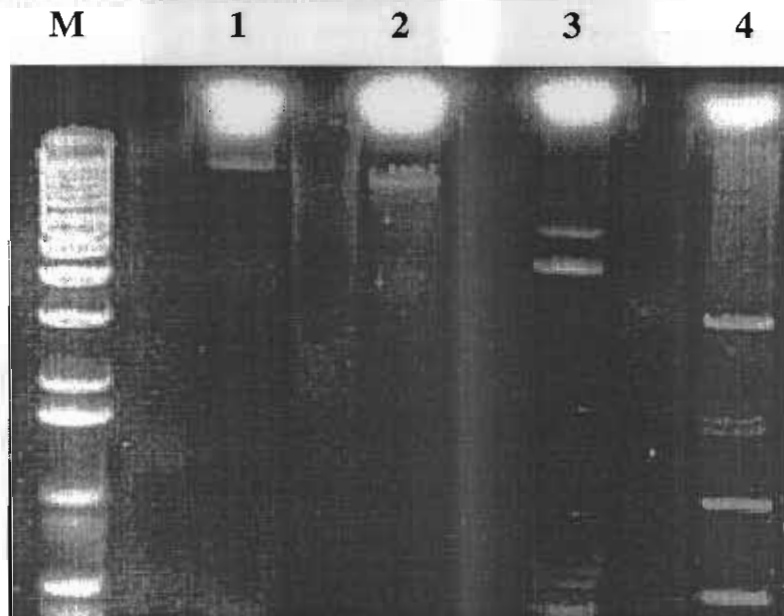


Figure 3. DNA fragmentation test of blood lymphocyte of control and treated male albino rats collected just before scarification. Ladder marker 100- 3000 bp (M), control (Lane no. 1), rats treated with 5 ppm of CdCl₂ (Lane no. 2), rats treated with 50 ppm of CdCl₂ (Lane no. 3) and rats treated with 100 ppm of CdCl₂ (Lane no. 4)

Table 2. DNA fragmentation test of blood lymphocyte collected just before scarification of treated adult male albino rats treated with different doses of CdCl₂

	Ladder		Control		5 ppm		50ppm		100 ppm	
	bp	%	bp	%	bp	%	bp	%	bp	%
1	3000	6.6	3820.3	100	3861.1	90.3	3957.2	80.3	3809.3	76.3
2	2000	5.3			2975.3	6.4	1021.4	6.2	971.1	1.4
3	1500	6.3			1942.4	3.3	813.1	3.2	790.4	4.2
4	1200	7.8					360.1	3.0	720.4	1.2
5	1000	7.3					180.4	7.3	540.3	4.8
6	900	7.3							360.2	3.2
7	800	5.9							210.4	2.8
8	700	7.2							180.4	2.8
9	600	9.7							125.5	3.3
10	500	5.1								
11	400	8.4								
12	300	8.9								
13	200	8.3								
14	100	5.9								
Sum		100		100		100		100		100

Thymus histopathology

The thymus of rats treated with 50 ppm of CdCl₂ showed a reduction in the medulla's diameter compared to control (Fig. 4 & 5). Furthermore, the thymus of rats treated with 100 ppm of CdCl₂ showed fat globules between the lobes of the thymus. The cortex

showed lymphocyte depletion. The medulla replaced with C.T with a reduction in its diameter with low number of lymphocytes. This might indicate degenerative changes in the thymus of rats particularly treated with 100 ppm of CdCl₂ (Fig. 6 & 7).

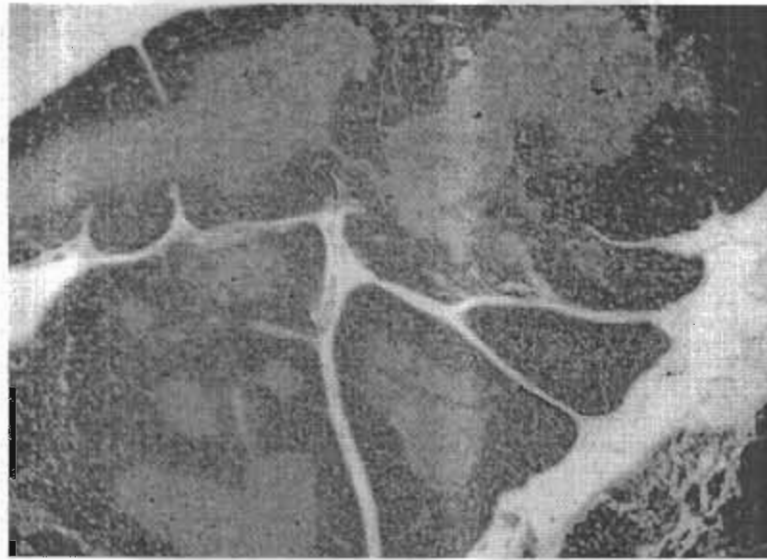


Figure 4. Section in thymus of control rats. The medulla showed normal diameter. H&E 40 X

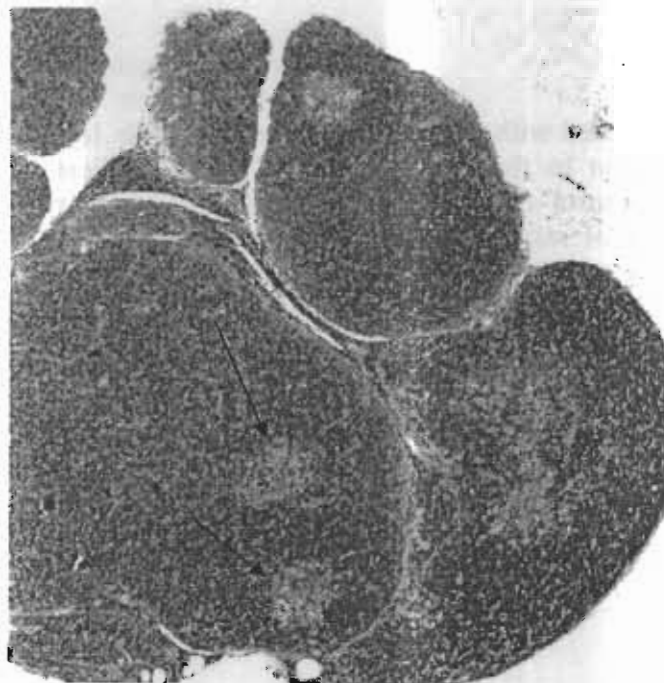


Figure 5. Section in thymus of rat treated with 50 ppm of CdCl₂. The medulla showed a reduction in its diameter compared with control rats. H&E. 40 X

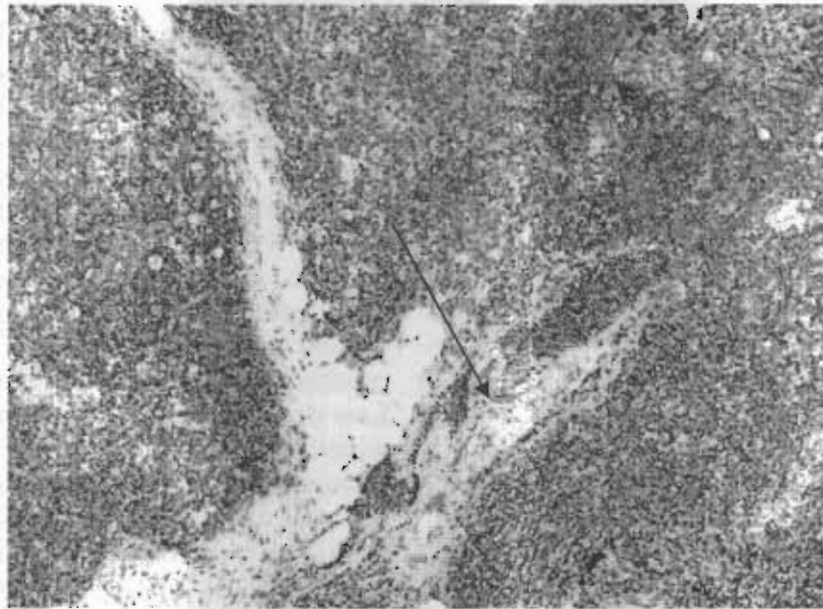


Figure 6. Section in thymus of rat treated with 100 ppm of CdCl₂ showed lymphocyte depletion in the cortex and the medulla replaced with connective tissue (black arrow). H&E 100 X

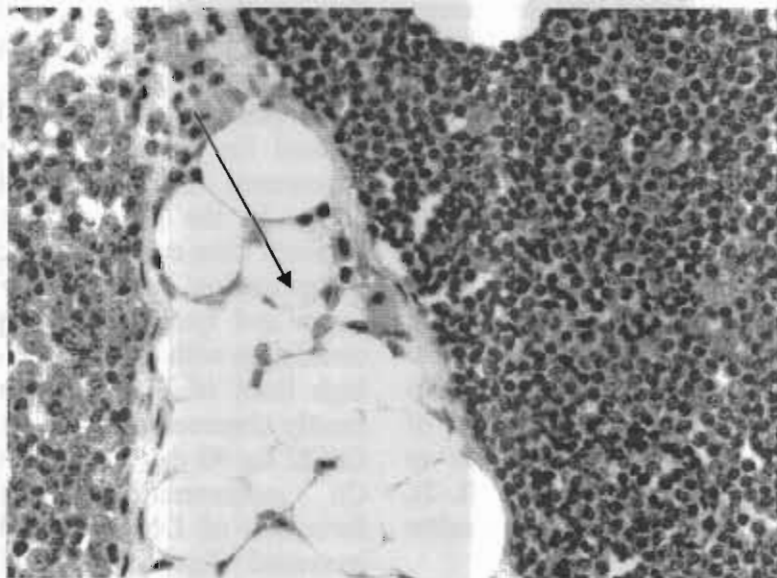


Figure 7. Section in thymus of treated rats with 100 ppm of CdCl₂ showed fat globules in the interlobular space. H&E 400 X

DISCUSSION

The immunotoxic effects of cadmium remain controversial. Exposure of humans or animals to cadmium is still of major toxicological importance. One site of attack of cadmium in the body is considered to be the immune system (23). Although immunomodulatory activities have been investigated in several *in vivo* and *in vitro* model systems, conflicting results of Cd immunotoxicity have emerged. Data about the immunotoxicity of cadmium are contradictory; in some cases it stimulates the immune response, in others Cd acts as an immunosuppressant, while yet other studies find no effect, even if seemingly identical aspects of immunity were investigated (24).

In the present study, the interleukin-2 (IL-2) level was declined in a dose-dependent manner in treated rats with different doses (5, 50 and 100 ppm) of CdCl₂. In agreement to our result, a decrease in IL-2 production in the presence of 10⁻⁴ M Cd²⁺ by human peripheral blood lymphocytes *in vitro* was found (25). Moreover, it has been estimated that the IL-2 staining in cadmium-treated spleen lymphocyte cells removed from male mice was markedly reduced. Generally, a cadmium-induced decrease in intracellular IL-2 content (26). Failure of IL-2 secretion and reduction of IL-2R expression in the *in vitro* Cd-treated cells were reported for human lymphocytes (27). Cadmium led to a significant increase in the level of some cytokines such as IL-1a and IL-10, however, the production of IL-2, on the other hand, was significantly depressed (6). Exposure of activated T-cells to low Cd doses leads to suppression of early T_H1 cytokine events (IL-2), and stimulation of the T_H2 cytokines IL-4 and/or IL-10 (28).

In the present study, the lymphocyte transformation is significantly increased in rats treated with 100 ppm of cadmium chloride. In agreement to our result, Spleen cells from mice given 200 ppm Cd for 3 to 4 weeks showed increased proliferative responses to Con A. The authors conclude that Cd enhances the proliferative response to T-cell mitogens (9).

Furthermore, an increased proliferative response of T-cells to PHA mitogens after 4 weeks of oral cadmium treatment was observed (9). Moreover, in other study, cadmium at the dose of 50 ppm in drinking water tended to increase the proliferative response of spleen cells to T-cell mitogen-PHA, but with no significant difference from the non-treated controls (29). CdCl₂ at very low concentrations were able to activate murine lymphocytes activated *in vitro* by Con A resulting in increased rate of DNA synthesis (6). Low doses of cadmium treatment, at 1 µM level, stimulated sheep lymphocytes proliferation (30). In contrary, cadmium did not significantly affect the response of lymphocyte proliferation by Con A (8). Mouse strain variations in Cd-mediated suppression of lymphocyte proliferation are not based on intrinsic lymphocyte sensitivities, but likely are due to differences in the metabolism of Cd, which is under genetic control (31).

DNA fragmentation is one of the hallmarks of apoptosis in Cd-exposed rats. Cadmium triggers an apoptosis like form of cell death in many cell types including T lymphocytes (12). Extensively fragmented double-stranded DNA can be separated from chromosomal DNA upon centrifugal sedimentation. Agarose gel electrophoresis of cellular DNA is the usual method for demonstrating apoptosis, with the appearance of a characteristic ladder pattern (32). In the present study, the lymphocyte obtained from control rats didn't show any DNA fragmentation. However, the incidence of DNA break and lymphocyte apoptosis increased in correlation with the increase of CdCl₂ dose. The high level of DNA fragmentation (23.7%) is mostly observed in rats exposed to 100 ppm of CdCl₂ for 90 days. In the presence of the lower Cd concentrations, the dose-dependent formation of DNA strand breaks in the cells increased significantly (33). However, high concentration of Cd destabilizes DNA causing DNA strand breaks and other DNA damage. It was found that internucleosomal cleavage of DNA from apoptotic cells gave products showing a 'ladder' pattern on agarose gel electrophoresis (34). These morphological and biochemical features are accepted to be important criteria for confirmation of apoptosis (35). Activation of endogenous endonuclease,

resulting in DNA degradation, is thought to be responsible for the morphological and biochemical alterations seen in apoptosis (36). Furthermore, a significant increase in DNA fragmentation in the presence of 10 mM Cd in a Ca^{+2} free system was observed (37). *In vitro* Cd exposure induced apoptotic features in mouse thymocytes (11).

In the present study, The adult male albino rats exposed to 50 and 100 ppm of CdCl₂ showed a significant decline in the relative thymus weight than that of control. Thymus, an important primary lymphoid organ, the place where successive stages of cell development and selection generate functionally competent T cells from immature precursor cells is a target organ of Cd-induced immunotoxicity (38). For instance, cadmium exposure resulted in thymic damage and modified the proliferation rate of thymocytes in rats (39). In agreement to our result, a number of *in vivo* experiments have demonstrated that cadmium is able to cause marked weight decrease of thymus or thymus atrophy (40-41).

Thymus of rats treated with 50 and 100 ppm of CdCl₂ showed fat globules between the lobes of the thymus. The medulla showed a reduction in diameter with low number of lymphocytes. This might indicate degenerative changes in the thymus of rats particularly treated with 100 ppm of cadmium chloride. Our result was supported with the finding in male Wister rats that exposed to sc injections of CdCl₂ at doses of 0.5, 1, and 2 mg Cd/kg, 3 days/week, for 4 weeks and showed histological changes of fibrous tissue proliferation in the thymus (42).

In conclusion, the results indicated that cadmium chloride affected the immune system in adult male albino rats through decreasing the production of interleukin-2 in dose-dependent pattern, triggering the lymphocyte transformation and inducing DNA fragmentation in blood lymphocyte particularly in rats exposed to 100 ppm of cadmium chloride for 90 days.

REFERENCES

1. **Satoh M, Kaji T and Tohyama C (2003):** Low dose exposure to cadmium and its health effects. Part 3. Toxicity in laboratory animals and cultured cells. *Nippon Eiseigaku Zasshi*; 57: 615-623.
2. **Satarug S and Moore M R (2004):** Adverse health effects of chronic exposure to low-level cadmium in foodstuffs and cigarette smoke. *Environ Health Perspect*; 112: 1099-1103.
3. **Chopra R K, Sehgal S and Nath R (1984):** Cadmium an inhibitor of lymphocyte transformation and stimulator of antibody-dependent cell-mediated cytotoxicity (ADCC) in rats: the role of zinc. *Toxicology*; 33: 303-310.
4. **Chopra R K, Sharma M and Nath R (1985):** A nonimmunomodulatory effect of chronic cadmium exposure on lymphocyte transformation response in rhesus monkeys (*Macaca mulatta*). *Exp Cell Biol*; 53: 19-23.
5. **Thomas P T, Ratajczak H V, Aranyi C, Gibbons R and Fenters J D (1985):** Evaluation of host resistance and immune function in cadmium-exposed mice. *Toxicol Appl Pharmacol*; 80: 446-456.
6. **Krocova Z, Macel A, Kroca M and Herychovah L (2000):** The Immunomodulatory effect(s) of lead and cadmium on the cells of immune system *in vitro*. *Toxicology in Vitro*; 14: 33-40.
7. **Gong F L (2003):** *Medical Immunology*. 1st ed. Beijing, China: Science Publication; pp. 238-252.
8. **Koller L D, Roan J G and Kerkvliet N I. (1979):** Mitogen stimulation of lymphocytes in CBA mice exposed to lead and cadmium. *Environ Res*; 19: 177-188.
9. **Malave l and DeRuffino D T (1984):** Altered immune response during cadmium administration in mice. *Toxicol Appl Pharmacol*; 74: 46-56.
10. **Chowdhury B A, Friel J K and Chandra R K (1987):** Cadmium-induced immunopathology is prevented by zinc administration in mice. *J Nutr*; 117: 1788-1794.

11. **Fujimaki H, Ishido M and Nohara K (2000):** Induction of apoptosis in mouse thymocytes by cadmium. *Toxicology Letters*; 115: 99-105.
12. **El Azzouzi B, Tsangaris G T, Pellegrini O, Manuel Y, Benveniste J and Thomas Y (1994):** Cadmium induces apoptosis in a human T cell line. *Toxicology*; 88: 127-139.
13. **Galan A, Garcia- Bermejo L, Troyano A, Vilaboa N E, Fernandez C, De Blas E and Aller P (2001):** The role of intracellular oxidation in death induction (apoptosis and necrosis) in human promonocytic cells treated with stress inducers (cadmium, heat, X-rays). *Eur J Cell Biol*; 80: 312.
14. **de la Fuente H, Portales-Perez D, Baranda L, Diaz-Barriga F, Saavedra-Alanis V, Layseca E and Gonzalez-Amaro R (2002):** Effect of arsenic, cadmium and lead on the induction of apoptosis of normal human mononuclear cells. *Clinical Experimental Immunology*; 129: 69-77.
15. **Boyum A (1968):** Isolation of mononuclear cells and gametocytes from human blood. *Scand J Lab Invest*; 21:77.
16. **Burrells S and Well P W (1977):** In-vitro stimulation of ovine lymphocytes. *Res Vet Sci*; 23: 84-86.
17. **Hudson L and Hay F C (1980):** *Immunology*, 2nd ed. Blackwell Scientific Publication, Oxford, London, Edinburgh, Boston, Melbourne.
18. **Rai El Balhaa G, Pellerine J L, Bodin G, Abdullah M L and Hiron H (1985):** Lymph plastic transformation assay of sheep peripheral blood lymphocytes: A new rapid and easy to read technique. *Comp Immune Microbiol Infec Dis*; 8: 311-318.
19. **Denise I, Bounous A, Raymond Q, Campagnoli A and John Brown B (1992):** Comparison of MTT colorimetric assay and tritiated thymidine uptake for lymphocytes proliferation assay using chicken splenocytes. *Avian Diseases*; 36: 1022-1027.
20. **Sellins K S and Cohen J J (1987):** Gene induction by gamma-irradiation leads to DNA fragmentation in lymphocytes. *J Immunol*; 139: 3199.
21. **Pathak N and Khandelwal S (2006):** Influence of cadmium on murine thymocytes: potentiation of apoptosis and oxidative stress. *Toxicol Letters*; 165: 121-132.
22. **Bancroft J P and Stevenes A (1990):** Theory and practice of histological techniques, 3rd edition, Clurechill Livigston, Edinburgh, London.
23. **Descotes J (1992):** Immunotoxicology of cadmium. *IARC Sci. Publ.* 385-390.
24. **Haase H and Rink L (2009):** Immunotoxicology of cadmium. In: Parvau, R.G. (Ed.), *Cadmium in the environment*. Nova Science Publishers, New York, pp. 447- 460.
25. **Theocharis S, Margeli A and Panayioditis P (1991):** Effect of various metals on DNA synthesis and lymphokines production by human peripheral blood lymphocytes *in vitro*. *Comp Biochem Physiol*; 99: 131-133.
26. **Payette Y, Lachapelle M, Daniel C, Bernier J, Fournier M and Krzystyniak K (1995):** Decreased interleukin-2 receptor and cell cycle changes in murine lymphocytes exposed *in vitro* to low doses of cadmium chloride. *Int J Immunopharmac*; 17 (3): 235-246.
27. **Cifone M G, Alesse E, Procopio A, Paolini R, Morrone S, Eugenio R D, Santoni G and Santoni A (1988):** Effects of cadmium on lymphocyte activation. *Biochem biophys Acta*; 1011: 25-32.
28. **Hemdan N Y, Emmrich F, Sack U, Wichmann G, Lehmann J, Adham K and Lehmann I (2006):** The *in vitro* immune modulation by cadmium depends on the way of cell activation. *Toxicology*; 222 (1-2): 37-45.
29. **Blakley B R (1985):** The effect of cadmium chloride on the immune response in mice. *Can J Comp Med*; 49: 104-108.

30. *Stec J (2003)*: Effect of cadmium and lead on [³H]- thymidine uptake in sheep lymphocytes infected with bovine leukaemia virus. *Bull Vet Inst Pulawy*; 47: 77-87.
31. *Ohsawa M, Masuko-Sato K and Takahashi K (1986)*: Strain differences in cadmium-mediated suppression of lymphocyte proliferation in mice. *Toxic Appl Pharmac*; 84: 379-388.
32. *Compton M M and Cidowski J A (1986)*: Rapid in vivo effects of glucocorticoid on the integrity of rat lymphocyte genomic deoxyribonucleic acid. *Endocrinology*; 118: 38.
33. *Mikhailova M V, Littlefield N A, Hass B S, Poirier L A and Chou M W (1997)*: Cadmium-induced 8-hydroxydeoxyguanosine formation, DNA strands breaks and antioxidant enzyme activities in lymphoblastoid cells. *Cancer Letters*; 115: 141-148.
34. *Wyllie A H, Beattie G J and Hargreaves A D (1981)*: Chromatin changes in apoptosis. *Histochem J*; 13: 681-692.
35. *Walker N I, Harmon B V and Gobe G C (1988)*: Patterns of cell death. *Methods Achiev Exp Pathol*; 13: 18-54.
36. *Hamada T, Tanimoto A and Sasaguri Y (1997)*: Apoptosis induced by cadmium. *Apoptosis*; 2: 359-367.
37. *Lohmann R D and Beyersman D (1993)*: Cadmium and zinc mediated changes of the Ca²⁺ dependent endonuclease in apoptosis. *Biochem Biophys Res Commun*; 190: 1097-1103.
38. *Kisielow P and Von Boehmer H (1995)*: Development and selection of T cells: Facts and puzzles. *Adv Immunol*; 58: 87-209.
39. *Morselt A F W, Leene W, DeGroot C, Kipp J B A, Evers M, Roelofsen A M and Bosch K S (1988)*: Differences in immunological susceptibility to cadmium toxicity between two rat strains as demonstrated with cell biological methods. Effect of cadmium on DNA synthesis of thymus lymphocytes. *Toxicology*; 48: 127-139.
40. *Mackova N O, Lenikova S, Fedorocko P, Brezani P and Fedorockova A (1996)*: Effects of cadmium on haemopoiesis in irradiated and non-irradiated mice: 2. Relationship to the number of circulating blood cells and haemopoiesis. *Physiol Res*; 45: 101-106.
41. *Liu J, Liu Y, Habeebu S S and Klaassen C D (1999)*: Metallothionein-null mice are highly susceptible to the hematotoxic and immunotoxic effects of chronic CdCl₂ exposure. *Toxicol Appl Pharmacol*; 159: 98-108.
42. *Yamano T, Shimizu M and Noda T (1998)*: Comparative effects of repeated administration of cadmium on kidney, spleen, thymus, and bone marrow in 2-, 4-, and 8-month-old male wistar rats. *Toxicological Science*; 46: 393-402.

الملخص العربي

تأثير جرعات مختلفة من كلوريد الكاديوم علي الانترلوكين-٢ ومعدل تكسير الحامض النووي الديوكسي
lymphocyte transformation,

كوثر عبدالواحد الهادي, ايمن كمال عبدالحميد, رانيا عبد الرحمن الجاويش
قسم الطب الشرعي والسموم- كلية الطب البيطري -جامعة قناة السويس

أجريت هذه الدراسة على عدد (٤٠) من ذكور الفئران البالغة البيضاء يتراوح أوزانها بين ١٠٠ جم و ١٤٠ جم. تم استخدام كلوريد الكاديوم في معالجة ذكور الفئران البالغة بجرعات مختلفة, تعرضت الفئران لهذه الجرعات من خلال مياه الشرب و لمدة تسعون يوما مدة إجراء البحث. قسمت الفئران إلى ٤ مجاميع كل مجموعة عشرة فئران: المجموعة الأولى: هي المجموعة الضابطة و التي تجرعت الماء المقطر فقط. المجموعة الثانية: هي المجموعة التي تعرضت إلى جرعة من كلوريد الكاديوم تعادل (٥) أجزاء في المليون. المجموعة الثالثة: هي المجموعة التي تعرضت إلى جرعة من كلوريد الكاديوم تعادل (٥٠) جزء في المليون. المجموعة الرابعة: هي المجموعة التي تعرضت إلى جرعة من كلوريد الكاديوم تعادل (١٠٠) جزء في المليون. في اليوم الأخير من التجربة تم أخذ عينات من الدم جزء منها سحب علي الهيارين لإجراء lymphocyte transformation assay and DNA fragmentation و الآخر استخدم لاستخلاص السيرم لقياس مستوى Interleukin-2. تم ذبح الفئران في نهاية التجربة و أخذت الغدة الصعترية و تم وزنها و فحص أنسجتها لمعرفة مدى التأثير السمي لكلوريد الكاديوم بجرعاته المختلفة.

و قد أسفرت نتائج هذا البحث عن الأتي: كان هناك انخفاض معنوي في مستوى Interleukin-2 في المجموعات التي عولجت بكلوريد الكاديوم مقارنة بالمجموعة الضابطة. حدث ارتفاع معنوي في معدل تكسير الحامض النووي الديوكسي ريبوز في المجموعة التي اعطيت (١٠٠) جزء في المليون كلوريد الكاديوم كما انه حدث ارتفاع معنوي في معدل Lymphocyte transformation في المجموعة التي تلقت (١٠٠) جزء في المليون كلوريد الكاديوم. كان هناك نقص معنوي في وزن الغدة الصعترية في المجموعة التي تلقت (١٠٠) جزء في المليون كلوريد الكاديوم.

و في الختام فان استخدام كلوريد الكاديوم في مياه الشرب لمدة تسعون يوما في ذكور الفئران البيضاء البالغة بجرعات مختلفة (١٠٠، ٥٠، ٥٠) جزء في المليون يقلل من انتاج الانترلوكين-٢ ويساعد على ضمور الغدة الصعترية, و أيضا يمكن لكلوريد الكاديوم بجرعاته المختلفة الحث علي تفتيت الحمض النووي في الخلايا للمفاوية وزيادة معدل Lymphocyte transformation.