Associative Effect of the Rhizobacteria Streptomyces chibaensis and Commercial Biofertilizers on the Growth, Yield and Nutritional Value of Vicia faba

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ABSTRACT

A pot experiment was carried out to investigate the effect of the commercial biofertilizers Biogen (B) and Potassiomage (PG) as well as the rhizobacteria *Streptomyces chibaensis* on the growth of faba bean (*Vicia faba* CV. Giza 843). The experiment was undertaken through two successive growing seasons (2008/09, 2009/10). In the first season, three different doses of each inoculant were used to select the profitable dose while in the second season; the co-inoculation by the three profitable doses was used as inoculant for faba bean seeds. Data presented as, growth, yield parameters, seed content of minerals (N, P and K), protein; vitamins (ascorbic acid, riboflavin and thiamin) and carbohydrates, were estimated by applying the co-inoculant of the three biofertilizers. The highest values of all these items were obtained from the application of the Biogen + S. chibaensis, followed by Potassiomage + S. chibaensis, which also showed a biological control activity against fungal diseases.

Key words: Biofertilizers, Vicia faba, Streptomyces chibaensis, fungal diseases.

INTRODUCTION

Symbiotic nitrogen fixation is the result of a delicate balance between a higher plant and specific bacteria. Most of nitrogen-fixing prokaryotes are free-living microorganisms found in soil (Elahi et al., 2004). Use of soil microorganisms which can either fix atmospheric nitrogen, solubilize phosphate or stimulate plant growth through synthesis of growth promoting substances by deliver a number of benefits including plant nutrition (Kaci et al., 2005).

Legume seeds are one of the most important sources of food in the world, have made a significant contribution to the diet since ancient times. They are known to be a good and inexpensive source of protein, also rich in carbohydrates, some vitamins (thiamine, riboflavin ,niacin) and certain minerals (phosphorus, potassium and sodium), this was reported by Ejigui et al. (2005).

Faba bean is a legume which is used as a green vegetable dried, fresh or canned in the Middle East, Mediterranean region, China and Ethiopia. It is considered as a cash crop in Egypt and Sudan (El Wakiel and El Sebai, 2007). Several soil-borne fungi attack *Vicia faba* during its various growth stages. Disease control measures which have been applied to minimize infection with fungal disease usually include the use of biological control (Sallam *et al.*, 2008).

Biological nitrogen fixation is the key to sustain agricultural productivity through the application of bio-fertilizers in the field. Application of bio-fertilizers is an acceptable approach for higher crops with good quality and safe to human health.

The objectives of this study were to investigate the effect of bio-fertilizers, with the plant growth promoting rhizobacteria (PGPR) Streptomyces chibaensis, on growth and yield of faba bean as well their role in controlling soil-borne fungal diseases.

MATERIALS AND METHODS

Two pot experiments were conducted in sandy soil at the experimental garden of the Botany Department, Women's College, Ain Shams Univ., Heliopolis, Cairo, Egypt during the two growing seasons 2008/09 and 2009/10. Faba bean seeds, cultivar Giza 843, were obtained from the Agriculture Research Center (ARC), Ministry of Agriculture, Giza. Egypt.

Bio-fertilizers Used

Three types of bio-fertilizers were used; the two commercials Biogen and Potassiomage, obtained from Biofertilizers Unit, General Organization of Agriculture Equalization Fund (GOAEF), ARC, Giza, Egypt, and S. chibaensis which was isolated from Toushka's region Egyptian soil and maintained on starch-nitrate agar medium, characterized by Hewedy (2003).

Bio-fertilization treatments

Faba bean seeds were sown in pots 35cm diameter and 40 cm depth, each contained 5kg loamy-sandy soil. Five seeds/ pot (6 replicates) were used for each treatment.

First pots experiment was carried out during the winter season, 2008/09, consisted of four main groups:

- Un-inoculated seeds (Control).
- Seeds inoculated with Biogen (B), using three doses (B1 0.08g, B2 0.16g and B3 0.32g/pot).
- Inoculation with Potassiomage (PG), using three doses (PG1 0.04g, PG2 0.08g and PG3 0.16g/pot).

Inoculation with the rhizobacteria S. chibaensis (S1 10⁻⁴, S2 10⁻⁵ and S3 10⁻⁶ CFU/ ml) (20ml/pot). Biofertilizers inoculation treatments were undertaken post planting. The plants were irrigated weekly using tap water; supported dose was given after 2 weeks later for each pot.

Data were recorded at 60 days post sowing, 3 plants were randomly uprooted from each pot to determine: shoot fresh weight (SFW), shoot dry weight (SDW), number of flowers (NF) and number of pods (NP)/plant. At harvest (120 days after sowing), a random sample of 3 plants was taken from each treatment to determine: number of pods (NP), pods length (PL), fresh and dry weight of pods (FWP, DWP) and dry weight of seeds (DWS). In the second growing season (2009/10), the best doses of the previous treatments (B:0.32g and PG 0.16g.) were mixed and used as co-inoculant for faba bean seeds, in addition to the best conc. 10⁴ of rhizobacteria (S. chibaensis) to determine the following growth parameters after 50 days: shoot and root dry weight (SDW and RDW), number of nodules (nN), dry weight of nodules (DWN) and acetylene reduction (ARA) (Hardy et al., 1973). At plant maturity (120 days), the following data were determined: number of pods (NP), dry weight of pods (DWP), dry weight of seeds (DWS), and number of seeds (NS) all by ten plants.

Chemical analysis

Seed contents of N, P and K were determined according to AOAC (1975). Crude protein% of seeds was obtained by multiplying N% X 6.25, extraction and determination of total carbohydrates content were carried out according to Dubois *et al.* (1956).

Quantification of vitamins

Concentration of three vitamins (riboflavin, thiamine and ascorbic acid) was determined by using KNAUER Smartline HPLC system in the ARC, Plant Pathology Research Institute, Center Laboratory of Biotechnology, Giza, Egypt.

Statistical analyses

Obtained data were subjected to Analysis of Variance (ANOVA). L.S.D. test was used to compare the treatment means according to the procedures outlined by Snedecor and Cochran (1980) using MSTAT computer program software program (MSTAT Ver., 1.42).

RESULTS AND DISCUSSION

This work showed that application of bio-fertilizers, either single or mixed inocula, gave positive response to the studied plant parameters. Highest concentration at each treatment gave the best results; shoot fresh weight (SFW), shoot dry

weight (SDW), number of flowers (NF) and number of pods (NP) at 60 days (Table 1) and NP, pod length (PL), fresh weight of pods (FWP), dry weight of pods (DWP) and dry weight of seeds (DWS) after 120 days (Table 2).

Nitrogen deficiency, indicated by brown pigment of leaves, was not observed at the high concentrations (0.33g of Biogen and 0.16g of Potassiomage and 10⁻⁴ CFU/ ml of S. chibaensis) but it was observed clearly on the untreated plants and lightly at the other plants treated with other concentrations of both bio-fertilizers and S. chibaensis.

A small round, slightly raised, cream colored spots occurred in late season only on leaves of untreated plants (control). The plants treated with different concentrations of S. chibaensis were found resistant to these fungal spots. Data illustrated in fig. (1) showed that all the applied concentrations of Biogen, Potassiomage and S. chibaensis led to decrease the number of non-healthy leaves resulted from soil-borne plant pathogens and highest values were obtained from the application of S. chibaensis 10⁻⁴, while Biogen was the least effective. Thus, it is clear that S. chibaensis might play an important role in reducing chances of survival of pathogens and primary infections in the field. This is in accordance with the results obtained by Hewedy (2003) which indicated that S. chibaensis produced chitinase enzyme and HCN which played a major role against Fusurium oxysporum which significantly improved plant health and promoted the growth and yield of Lupinus termis.

Plants treated with different concentrations of S. chibaensis surpassed Biogen and Potassiomage and gave significant high results in the majority of parameters. Growth parameters of V. faba, treated with the best concentration of each treatment individually or in combination, gave promotive effect compared to the control in the two seasons. These results are on line with Gomaa et al. (2002) and Hewedy et al. (2006) who cleared that the biofertilizers were used to stimulate plant growth by producing plant growth regulators.

Nodules were formed on all plants (either inoculated or not), which indicated the presence of the native rhizobia. Root nodulation was characteristically promoted in plants treated with B+S. chibaensis, followed by plants treated with Biogen, respectively (Figs. 2a, b). These treatments gave also the highest results for shoot and root dry weights (Fig. 3).

The competitiveness among S. chibaensis, Biogen and Potassiomage gave results in which B + S. chibaensis treatment was the best, followed

Table (1): Effect of individual inoculants of Biogen, Potassiomage and Streptomyces chibaensis on certain growth parameters with different concentrations after 60 days from planting

Treatments	Shoot fresh wt./3 plants (g)	Shoot dry wt./3 plants (g)	No. of flowers/3 plants	No. of pods/3plants
C	112.8h	18.1e	27g	20f
B1	134.0fg	20.1d	30fg	27de
B2	149.8eg	20.9cd	47e	33cd
B3	159.8de	29.4a	70c	37bc
PG1	218.0bc	21.4cd	37f	0g .
PG2	176.3d	24.7b	57d	23ef
PG3	202.2c	26.2b	103a	40b
S1	256.4a	25.6b	103a	53a
S2	232.4b	22.0c	80b	30d
S3	188.5cd	12.3f	50de	27de
LSD at 5%	20.5	1.8	9.5	6.2

Values in the same column with the same letter are not significantly differed at p<0.5%.

Table (2): Effect of individual inoculants of Biogen, Potassiomage and Streptomyces chibaensis on the yield of Vicia faba plants after 120 days

Treatments	No. of pods/3 plants	Pod length (cm)	Fresh wt. of pods/3 plants	Dry wt. of pods/3 plants	Dry wt. of seeds/3 plants
C	23f	6.3c	16.8e	9.44e	7.05e
B1	3 0e	7.0bc	16.2e	15.18d	11.89bc
B2	33 de	7.2b	20.2d	15.75d	11.98bc
B3	47c	7.3b	33.2b	20.43b	14.84a
PG1	30e	7.3b	11.2f	7.02f	4.27f
PG2	37 d	8.2a	20.1d	14.96d	9.56d
PG3	53b	8.5a	27.2c	18.78c	14.42a
S1	77a	9.0a	37.5a	22.96a	15.71a
S2	53b	8.7a	30.5b	21.28b	13.78ab
S3	47c	9.0a	20d	15.77d	10.45cd
LSD at 5%	5.6	0.8	3.2	1.5	2.0

Values in the same column with the same letter are not significantly differed at p<0.5%.

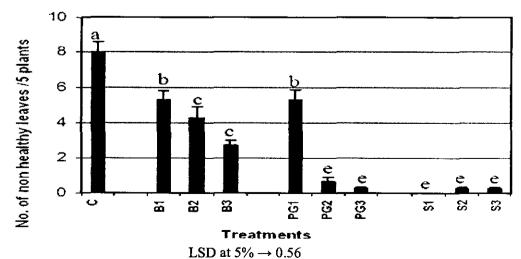


Fig. (1): Effect of different concentrations of Biogen, Potassiomage and Streptomyces chibaensis on the presence of non-healthy leaves in Vicia faba.

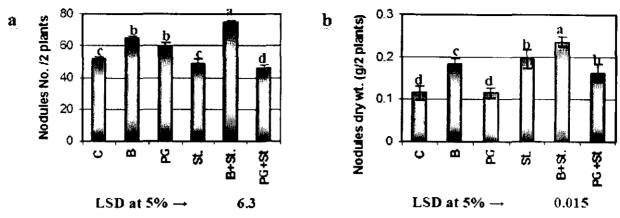


Fig. (2): Effect of individual and mixed inoculants of Biogen, Potassiomage and Streptomyces chibaensis on shoot and root dry weight of Vicia faba.

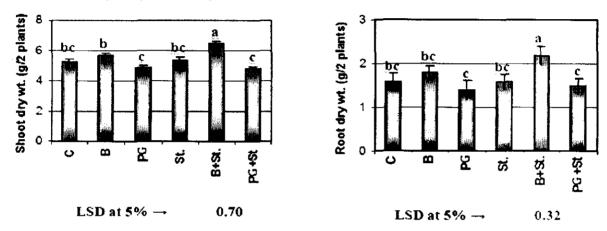


Fig. (3): Effect of individual and dual inoculants of Biogen, Potassiomage and Streptomyces chibaensis on shoot and root dry weight of Vicia faba.

Columns with the same letter are not significantly differed at p < 0.5%.

Table (3): Effect of individual and dual inoculants of Biogen, Potassiomage and Streptomyces chibaensis on the yield of Vicia faba plants

Treatments	No. of pods /10 plants	Dry wt. of pods (g/10 plants)	No. of seeds/10 plants	Dry wt. of Seeds g/10 plants)
C	70c	7.10c	100d	4.36c
В	80a	11.81a	190a	9.47a
PG	77a	7.01c	100 d	5.72c
St.)	73bc	10.14b	100d	7.83b
B+St.	80a	11.56a	153b	9.55a
PG+St	73bc	6.94c	135c	7.41b
LSD at 5%	5.0	1.4	15.0	1.5

Values in the same column with the same letter are not significantly differed at p < 0.5%.

by Biogen. This result improved the ability of S. chibaensis to compete for V. faba nodulation and exhibition for acetylene reduction than in the other treatment (Fig. 4), which means that S. chibaensis had an indication to promote N₂ fixation. Hewedy (2003) indicated that a neutralism relationship between S. chibaensis and N₂ fixing bacteria, which are widely distributed in the Egyptian soil, might be the result of positive dual treatments.

Concerning yield parameters in the present study, the used biofertilizers could improve the growth of the economic crop, faba bean (Table 3), but in the presence of S. chibaensis. This improvement might be due to the inoculants filament bacteria which degraded organic matter and supplied to the plants, also converted atmospheric nitrogen into available form, and stimulated plant growth directly through the production of phyto-stimulating compounds (Hewedy, 2006 and Zaied et al., 2007). This may be also due to their ability to produce antibacterial and antifungal compounds (Hewedy, 2003 and Hosseny & Ahmed 2009).

NPK uptake in seeds was significantly affected by different treatments. The dual inoculations gave

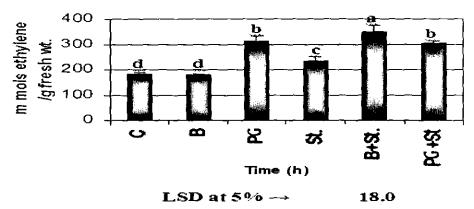


Fig. (4): Effect of individual and dual inoculants of Biogen, Potassiomage and Streptomyces chibaensis on acetylene reduction activity of Vicia faba.

Columns with the same letter are not significantly differed at p < 0.5%.

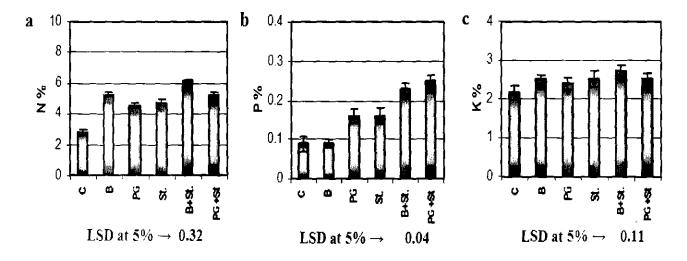


Fig. (5): Effect of individual and dual inoculants of Biogen, Potassiomage and Streptomyces chibaensis on the NPK of Vicia faba seeds.

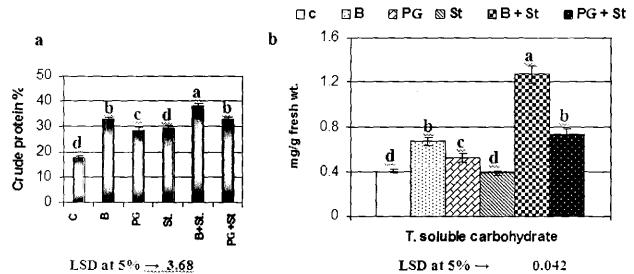


Fig (6): Effect of individual and dual inoculants of Biogen, Potassiomage and Streptomyces chibaensis on crude protein and total soluble carbohydrate of Vicia faba plants.

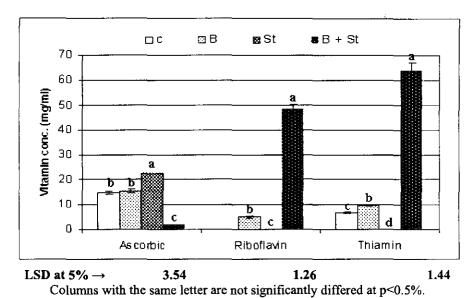


Fig. (7): Effect of single and dual inoculants of Biogen and Streptomyces chibaensis on vitamin concentration of Vicia faba seeds.

the greatest of N, P and K% values. Plants treated with B. + S. chibaensis gave the highest uptake of N & K, followed by PG + S. chibaensis, which gave the highest uptake of P. (Fig. 5). This could be explained as both biofertilizers and PGPR filamentous bacteria might improve the rhizosphere conditions which are mainly due to the bacterial production of N, P and indole 3-acetic acid (Wu et al., 2005 and Rothballer et al., 2006). Also, it might be due to the capability of S. chibaensis to dissolve P and its affectivity to fix N₂ (Hewedy, 2003 and Hewedy et al., 2006).

It is obvious from the data recorded in Figs. (6 a and b) that the seeds produced from the plants treated with PGPR, S. chibaensis individually or combined with Biogen and Potassiomage had significant increase in seeds protein and total carbohydrates content. Biogen + S. chibaensis gave the highest protein content, followed by Potassiomage+ S. chibaensis. This result is in agreement with Abu-Zekry (2000) and Hewedy et al. (2006) who recorded that soybean treated with biofertilizers gave the highest yield and seed protein and carbohydrates.

Because of its promising results, treatment of S. chibaensis + Biogen was chosen for studying its effect on the content of vitamins in seeds (Fig. 7). This combination increased the production of both thiamine and riboflavin than in the control. On the contrary, ascorbic acid production was increased in the presence of S. chibaensis individually, followed by Biogen > control > Biogen + S. chibaensis. Atta et al. (2008) reported that the spectroscopic analysis (UV, IR and HPLC spectrum) was used to perform comparative studies between vitamin B12 and two mixed cultures of S. halstedii

AZ-8A and Bacillus firmus AZ-78B, which showed an increase in the mixed culture (peak was at 2.503) than the standard (peak was at 2.496). Isabel et al. (1988) found that thiamine and riboflavin were found in high abundance, early in the development of lupine and pea, but their contents decreased during maturation The thiamine content of faba bean seeds was increased from 33 to 47 days after flowering (DAF), then a decrease was observed until 61 DAF, the riboflavin was increased after 54 DAF.

In conclusion, this work may prompt further screenings of Actinomycetes, especially Streptomycetes as potential plant growth promoters for use in nutrient impoverished soils. The isolate of S. chibaensis examined in this study, in addition to being a native of Egypt soils, is the most suited for seed inoculation because of its high levels of rhizosphere competency.

Therefore, this work recommends the utilization of the commercial inoculants in combination with S. chibaensis, which gave a good response as a PGPR commercially for improving production of faba bean with its competitive ability to survive and affect growth of inoculated plants at the presence of indigenous microflora to protect the environment from chemical pollution and its harmful effect on nature. Also, S. chibaensis may be utilized as a biocontrol agent to help in protecting plants against non-induced fungal infection and to minimize the risk and hazard of toxic fungicides. This might help in modern agriculture through the increase use of microorganisms. which an advantageous is alternative to chemical treatments and may contribute substantially to the goal of environmental friendly agriculture.

ACKNOWLEDGMENT

Thanks are due to assistant Professor Dr. Hisham Abd-Elmonaem, Plant Pathology Institute, ARC, Giza, for his technical assistance with vitamin analysis.

REFERENCES

- Abu Zekry, S. H. 2000. Effect of biological fertilization on the productivity of some cereals and legumes. Ph. D. thesis, Botany Department, Girl's College, Ain Shams Univ., 153pp.
- Association of Official Agricultural chemists (A.O.A.C.) 1975. Official Methods of Analysis 12th ed. Washington, D.C., USA.
- Atta, H. M.; R. A. Arafa; M. S. Salem and M. A. El-Meleigy 2008. Microbiological studies on the production of vitamin B12 from two mixed cultures under solid state fermentation condition. J. Appl. Sci. Res., 4(11): 1463-1470.
- Dubois, M.; K. A. Gilles; J. K. Hamilton; P. A. Roberts and P. Smith 1956. Colorimetric method for determination of sugar and related substances. Analytical Chemistry, 28(3):350-356.
- Ejigui, J.; L. Savoie and J. Martin 2005. Influence of traditional processing methods on the nutritional composition and antinutritional factors of red peanuts (*Arachis hypogea*) and small red kidney beans (*Phaseolus vulgaris*). Biol. Sci., 5(5):597-605.
- El Wakeil, N. E. and T. N. El Sebai 2007. Role of biofertilizer on faba bean growth, yield and its effect on bean aphid and the associated predators. Research J. Agric. and Biolo. Sci., 3(6): 800-80.
- Elahi, N. N.; Akhter, W. and J. I. Mirza 2004. Effect of combined nitrogen on growth and nodule of two Mungbean (Vigna radiata L. Wilczek) cultivar. Journal of Research (Science), Bahauddin Zakarya University, Multan, Pakistan. 15(1): 67.
- Gomaa, A. M.; A. A. Bahr and M. E. El. Kramany 2002. The bio-organic forming and its effect on nodular growth and yield parameters of *Vicia sativa L.* Egyptian. J. Agron.. 24: 79-92.
- Hardy, R. W. F.; R. C. Burns and R. D. Hols!

 1973. Applications of the acetylene ethyle...
 assay for measurement of nitrogen fixation. Soil

- Biol. Biochem., 5:47-81.
- Hosseny, M. H. and M. M. Ahmed 2009. Effect of nitrogen, organic and biofertilization on productivity of lettuce (cv. Romaine) in sandy soil under Assiut conditions. Ass. Univ. Bull. Environ. Res. Vol. 12. pp. 79 93.
- Isabel, S.; C. Vidal-Valverde and H. Kozlowska 1998. Effect of ripening stage on thiamin and riboflavin levels in lupin, pea and faba bean seeds. Springer Berlin, Heidelberg. pp: 126-129.
- Kaci, Y.; A. Heyraud; B. Mohomed and T. Heulin 2005. Isolation and identification of an EPS-producing rhizobium strain from arid soil (Algeria): characterization, its EPS and the effect of inculcation on wheat rhizosphere soil structure. Res. Microbiol., 156: 522-531.
- Hewedy, Maha, A. 2003. Effect of local microbial isolate as biocontrol agent against *F. oxysporum* root rot. Bull. Fac. Sci., Assiut Univ. 32(2-D) P. 383-395.
- Hewedy, Maha, A.; A. S. Sheteawi and M. K. Tawfik 2006. Beneficial interactions between *Strepomyces chibaensis*, Okadein, *Bradyrlizobium japonicum* and soybean. Assiut Univ. J. Botan. 35(2), 405-422.
- Rothballer, M.; M. Schmid; A. Fekete and M. Blumnel 2006. Comparative in situ analysis of ipdC-gfpmut3 promoter fusions of Azospirillum brasilense strains Sp7 and Sp245. Plant and Cell Physiol., 46: 1848-1854.
- Sallam N. A.; K. A. Abo-Elyousr and M. A. Hassan 2008. Evaluation of *Trichoderma* species as biocontrol agents for damping-off and wilt diseases of *Phaseolus vulgaris* L. and efficacy of suggested formula, Egypt. J. Phytopathol. 36: 81-93.
- Wu, S. C.; Z. H. Cao; Z. G. Li; K. C. Cheung and M. Wong 2005. Effects of biofertilizer containing N-fixer, P and K solubilizers and A M fungi on maize growth: a greenhouse trial. Geoderma, 125: 155-166.
- Zaied, K. A.; A. H. Abd El-Hady; A. E. Sharief; E. H. Ashour and M. A. Nassef 2007. Effect of horizontal DNA transfer in *Azospirillum* and *Azotobacter* strains on biological and biochemical traits of non legume plants. J. Appl. Sci. Res., 3(1): 73-86.