Studies on Some Bacterial Infections In Quails In Bohira Province

Bkheit A A

Animals Health Res., Inst. Damanhour Bransh

ABSTRACT

A total of 70 quail's samples of variable ages were collected from different private farms in Bohaira province. The birds were suffering from a disease problem bacteriologically investigated and subjected. *E.coli*, *Klebsiella*, *Enterobacter*, *Citrobacter* and *Proteus* species as members of *Enterobacterreaces* were isolated in. an incidence 60.7%, 9.7%, 5.9%, 9.3%, and 9.7% respectively. *Pseudomonus. aerogenosa, Pseudomonus. flourescen*, and.*Pseudomonus aerofascice* were isolated in an incidence of 68%, 23.7 and 7.9 respectively. *Staphylococcus aureus* was also isolated with a percent of 50.6. *Staph. Gallinarum,Staph. Epedermidis,Staph. Haemolyticus, Staph. Intermedius, Staph cohini I,II*, *Staph. capitis* and *Staph. hycus* were isolated with an incidences of 8.6%, 6.6%, 6.6%, 4.6%, 5.3%, 4.6% and 3.2% respeatively. Enterococci was isolated in incidence of 67.3% classified into *Enterococci fecales*, *Enterococci faciun* and *Enterococci avium* in an incidence of 40.4%, 15.4% and 11.5%, in addition to *Streptococci* in an incidence of 32.6% *Strept. gallinarun Strept dysagalactea, Strept. bovis and Strept pneumoni* were isolated in an incidence of 11.5%, 9.6%,3.8% and 7.6% respectively. The prevalence of isolation from different sites of isolation were calculated.

INTRODUCTION

Quails becomes widely distributed in Egypt as a source of meat and egg production, since their meat and eggs are highty preferable to consumers for low content of cholesterol (1). Moreover, quail farmes occupies less space area per bird and consumed less feed with higher metabolic rate (2).

E. coli is a normal inhabitant of intestinal tract (3) found with an incidence of 16 % and 52 % in liver and intestine respectively with an average of 36 % sixty percentage (4). recovery rat from intestine. Coli-septicemia induced experimentally by ser-group O165 in Japanese quail resulted in 90% mortalities (5). Several members of Enterobacteriaceae could be isolated from birds. Citrobacter was isolated from weak chicks and yolk sac infection (6,7). Also Enterobacter is normal inhabitant in digestive tract of avian species (8). It induced omphalities, yolk sac infection (7,9). Klebsiella have been associated with septicemia and respiratory, ocular, reproductive disease in poultry (6). Proteus as genus of family Enterobacteriaceae habitat lower intestinal tract and could induce septicemia in quails (10). Proteus mirabilis was isolated from lung, trachea, kidneys and intestine (11). Pseudomonas aeroginosa is the predomenant in causing infection, however, *Pseudomonus fluorescens* could be recovered as multi causal respiratory disease in chicken (11).

Staphylococcus spp. frequently isolated from poultry as Staph. aureus and Staph. epidermidis, also Staph. gallinarum, Staph. hyicus has been associated with many diseases in poultry (10) Streptococcus spp isolated from avian species and associated with disease include Strept zooepidemicus (occasionally referred as Strept. gallinaram), Strept. bovis Strept. dysaglactea and Moreover. Enterococcus faecales, Enterococcus faecium, Enterococcus durans are naturally and experimentally caused poultry infections, bacterial reslting in endocarditis and septecemia (10) Campylobacteriosis is contagious disease in poultry characterized by low mortality and high morbidity with chronic course (12). Campylobacter jejuni is the main etiologic agent of the disease and has a zoonotic significance in human (13).

The aim of this study was planned to focus on prevalence of *Enterobactereace*, *Pseudomonas*, *Staphylococci and Streptococci* & *Enterococci*, in addition to trials for isolation of *Campylobacter* from Japanese quails

MATERIAL AND METHODS

A total of 70 japanease quails were collected from different private farms at different localities of Bohaira province. The collected samples were at different ages (few days old up to breeders) All were clinically diseased. Sample from lung, heart liver, crop, and intestine, were taken and bacteriologically investigated.

Bacteriological examinations

1.Culture methods

Collected samples were inoculated onto nutrient broth, for *Staphylococci*, *Streptacocci*, *Pseudomonus*, and on Seleniet F. broth for culture *Enterobactereaceae*. In addition to thiol broth for culture *Campylobacter*. All were incubated overnight at 37°C then nutrient broth was cultured on nutrient agar, blood agar, Barid parker agar, Manitol salt agar and K.F.agar for *Staphylococci*, *Streptococci* and *Pseudomonus*, Seleniet -F- broth was cultured on Mackonky agar for *Enterobactereaceae*. Thiol broth was cultured on to T.C.B.S. agar. All plates except T.C.B.S. were incubated at 37°C for 24 hr.T.C.B.S were cultured by candle jar at 37°C for 48-72 hrs

2-.Identification of bacterial isolates

Bacterial colonies were identified morphologically and biochemically (14-16)

RESULTS

Results of prevalence and distribution of all bacterial isolates from diseases quails collected from different farms are presented in Tables (1-4).

Organism	L	ung H		eart	rt Liver		Сгор		Intestine		Total	
Organ	N	%	N	%	N	%	N	%	N	%	N	%
E coli	18	12.5	25	16.4	45	31.3	13	9	45	29.9	144	60.7
Kllaebsiel	2	0.8	1	0.4	4	1.6	1	0.4	10	4.2	18	7.6
Kelb. pnumonis	2	22.2	1	11.1	3			33.3	3	33.3	9	3.8
Kleb.pnuozaene					2		1		2		5	2.1
Oxytocia					1	25	<u> </u>		3	75	4	1.7
Enterobacter					4	1.7	3	1.3	7	2.9	14	5.9
Ent . arogens					3	30	3	30	4	40	10	4.2
Ent. Cloaca					1	25			3	75	4	1.7
Citrobacter	3	1.3	2	0.8	7	2.9			10	4.2	22	9.3
Cit. frundii	3	18.8	1	6.3	5	13.3			7	43.7	16	6.8
Cit diverus	. <u></u>		1	16.6	2	33.3			3	50.0	6	2.5
Proteus	2	0.8	2	0.8	5	2.1	5	2.1	7	3.1	23	9.7
Prot. vulgaris	-		_	<u> </u>	2	25	2	25	4	50	8	3.4
Prot. merabillis	2	15.4	2	15.4	3	23	3	23	3	23	13	5.5
TOTAL	25	10.5	30	12.6	65	27.4	22	10.1	79	33.3	237	100

Table 1. Prevalence and sit distribution of Enterobacteriaceae in Japanese Quails.

N = Number of positive samples

-- = Negative

% = Number of positive per total number of positive samples from different organs

Zag. Vet. J.

Isolata	Lung		Heart		Liver		Сгор		Intestine		Total	
Isoute	Ν	%	Ν	%	N	%	N	%	N	%	N	%
Pseud. aerogenosa	4	15.4	4	15.4	4	15.4	5	19.2	9	346	26	68.4
Pseud. florec III	1	25	1	25	1	25			1	25	4	10.5
Pseud florec IV			1	20			1	20	2	40	4	10.5
Pseud. aerofassiencel							1	33	2	66	3	7.9
Total	5	13.2	6	15.8	5	13.2	_ 7	18.4	14	36.8	37	100
N = Number of positive same	ples			= Ne;	gative							

Table 3 .Prevalence and sit distribution of Pseudomonus spp in Japanese Quails.

N = Number of positive samples

% = Number of positive per total number of positive samples from different organs

Table 4. Prevalence and sit distribution of Staph. Cocci in Japanese Quails.

Icolato	L	ung	H	eart		iver	C	rop	Intesting		Total	
	Ν	%	Ν	%	Ν	%	N	%	N	%	N	%
Staph. aureus	22	28.6	16	20.8	18	23.4	7	9.1	14	18.2	77	50.6
Staph. gallinarun	5	38.5	2	15.4	2	15.4	1	7.7	3	23.1	13	8.6
Staph. epedermidis	2	15.4	3	23.1	3	23.1	4	30.8	1	7.7	15	8.6
Staph. heam0lytius	2	20	2	20	3	30	2	20	1	10	10	6.6
Staph. intermedius	2	20	3	30	3	30	1	10	1	10	10	6.6
Staph. cohini 1	1	14.3	2	28.6	2	28.6	1	14.3	1	14.3	7	4.6
Staph. cohini II	1	12.5	3	37.5	2	25	1	12.5	1	12.5	8	5.3
Staph. capitis	1	14.3	2	28.6	2	28.6	1	14.3	1	14.3	7	4.6
Staph. cornosus	—					_	1	50	1	50	2	1.3
Staph. hycus	—		1	20	2	40	1	20	1	20	5	3.2
Total	36	23.7	34	22.4	37	24.3	20	13.1	25	16.4	152	100%

N = Number of positive samples -- = Negative

% = Number of positive per total number of positive samples from different organs

Isolate	Lung		Heart			Liver		Сгор		Intestine		'otal
	N	%	N	%	Ν	%	Ν	%	N	%	N	%
Enterococci	4	7.7	3	5.8	9	17.3	6	11.5	13	25	21	40.4
Ent. foecales	- 3	14.3	2	9.5	4	1 9	3	13.3	9	42.9	21	40.4
Ent. facian					4	50	2	25	2	25	8	15.4
Ent. avium	1	16.6	1	1 6.6	1	16.6	1	16.6	2	33.3	6	11.5
Streptococci	4	7.7	2	3.8	6	11.5	3	5.8	2	3.8		17
Strept. gallinaram	1	16.6	1	16. 6	2	33.3	1	16.6	1	16.6	6	11.5
Strept dysagalactea		-			2	40	2	40	1	20	5	9.6
Strept. bovis	. 1	50		-	1	50		-		-	2	3.8
Strept. pneumoni	2	50	1	25	1	25		-		-	4	7.6
Total	8	15.4	5	9.6	15	28.8	9	17.3	15	28.8	52	100%

Table 5. Prevalence and sit distribution of Enterococci isolated from Japanese Quails .

N = Number of positive samples-- = Negative

% = Number of positive per total number of positive samples from different organs

DISCUSSION

Quail farms have gained popularity during the last decade and the best known breed in Egypt is Japanese quail which breeds for egg and meat production due to low cholesterol content and valuable protein source (I).

The contact of quail farms & broiler possibility of farms facilate the pathogens transmisson regardless the species occured. This work was planed to focus some light on microbial state of 70 diseased Japanese quails of variable ages collected from different The prevalence farms. of Enterobactereocea as show in Table 1 indicated that. E.coli was the predominant isolate forming 60.7% among the Enterobactereaceae. The site distribution of E coli was the lung, heart, liver, crop and intestine which revealed 12.5%, 16.4%, 31.3% 9% and 29.9% respectively, E.coli was recovered from liver and intestine as 16% and 52% respectively with total percent 34, (3,4,17). Also. infection rate was 34% of examined liver and a higher rate (18) was (19). Who found recovered an incidence 55.3% of E. coli among diseased or dying quails. The variation of isolation rate among different organs revealed that, the lowest rate was in crops which denote the oral contamination via water and ration and growth rates, followed scanty by respiratory origin. E. coli gains assess to circulation following damage to the respiratory mucosa by infectious or noninfectious agents (10,20) .Heart and liver isolation rates are higher where isolation colisepticeimia in The increased. intestine has high incidence because this is the normal habitat of the organism.

Klebsiella is an environmental contaminant that occasionally cause mortality in young birds in addition respiratory signs ocular lesions and septicemia, Kelbsiella pneumonia had the highest mortalites (21).

In the present study Klebsiella was isolated from lung, heart liver, crop and intestine (Table1). The overall incidence of Klebsiella species reached 7.4 % Klebsiella pnemonia, Klesiella pnemonia sub spp. ozaena and Kebsiella oxytoca were in incidences of 3.8 %, 2.1 % and 1.7% respectively.

Enterobacter is normal inhabitant of the avian digestive tract (8) in common with other Enterobacteriacea and could infect eggs and young chick causing mortalities (7). In the present study Enterobacter aerogeneous and Enterobacter cloacae were isolated with incidence of 4.2% and 1.7% respectively. The total incidence of Enterobacter spp.was 5.9% they were recovered from liver, intestine and crop, the higher rate of recovery was from intestine as the normal habitat of the organism.

Citrobacter is а member of Enterobacteriaceae commonly colonized mucous membrane of respiratory and digestive tract in normal but could bird be an opportunistic pathogen and could be isolated from weak chick and yolk-sac infection (6,7). Citrobacter frundii and Citrobacter diversus could be isolated in an incidence of 6.8 % and 2.5% respectively (Table 1) Citrobacter frundii was isolated from intestine, liver, lung.and.heart, in a percent.of 43.7, 31.3, 18.8 and 6.3 respectively. 9.3% Citrobacter forms of Enterobacteriaceae isolates similar results were previously recorded (6,7).

Proteus spp Isolation as shown in Table 1 were 3.4% and 5.5% for *Proteus vulgares* and *Proteus mirabilis*. They was *Proteus* spp isolated from 8-90% of dead young quails and japese quails (3,22). This variation could be attriutd to that, the isolation was carried out on died young quails .On the other hand *Proteus* merabillis was isolated from lung trachea and parenchymatus organs (23). This finding agreed with the present study where *Proteus* merabillis were isolated from lung, heart liver, crop and intestine in an incidence of 15.4%, 15.4%, 23% and 23% respectively from total *Proteus* isolates.

Pseudomonus infections in birds are of great importance for its rapid spread, mortalities in all ages and difficulty in treatments. In the study **Psudomaus** current aerogensa, floresence Pseudomonus biovar Ш floresence Psudomouis biovar IVand Pseadomonus aerofascience were isolated in distribution of 68.4 %, 10.5%, 13.2% and 7.2% respectively. These results agreed with literatures which emphasized those Pseudonus aerogenosa is the predominant species causing infection among quails (24). Pseudomonus aerogenosa could be isolated from intestinal content, cloacal swab, nasal swab, lung, liver and yolk sac in diseased. This study coincided with the previous records (25,26) which showed that Pseudomones aerogenosa could be isolated from lung, heart, liver, intestine in an incidence of 15.4%, 15.4%, 15.4% and 34.6% respectively as distribution of the positive isolates. This study revealed isolates of Pseudomonus florescence in lower incidences as 10.5% from gained isolates Psudomonus floresence has been associated with multicosal respiratory diseases of chick and turkey (11).

Genus Staphylococcus contains 32 species are associated with a wide variety of diseases in chicken and other avian species. They include septicemia, arthritis. yolk sac infection and many disease conditions (27). In the present Study as in Table 3 Staplylococci members were isolated in incidences of 50.6 %, 8.6 %, 8.6 %, 6.6 %, 4.6%, 5.3%, 4.6%, 1.3% and 3.2% for both of Staph. aureus Staph. gallenarum, Staph. epedermidis, Staph. haemolyticus, Staph. entermedius, Staph. choini I&II, Staph. capitis, Staph. cornosus and Staph. hycus respectively. Staph. aureus was most prevalent strain with an of 27% followed Staph incidence by epedermides (11.76%) (28). Higher incidences

of isolated *Staph.aureus* were recorded (27) as 62%. On the other hand, lower incidences were recorded (3) and an incidence of 6% in each of liver and intestine of Japanese quails was recorded. *Staph. aureus, Staph. epedermidis, Staph. gallenarum* and *Staph. hycus* are frequently isolated from poultry which agreed with the present study.

Concerning Enterococci, the prevalence and site distribution are illustrated in Table 4 Ent. foecales, Ent. facium, Ent. avian. Strept. gallinarum, Strept. dysagalacted, Strept. bovis and Strept. pneumoni were isolated from quail in distribution of 40.4 % 15.4%, 11.5 %, 11.5%, 9.6%, 3.8 and 7,6 respectively. Streptococci was isolated from avian species and associated with disease conditions include Strept. zooepedemicus occasionally refaird as Strept. gallinarum), Strept. bovis and Strept. dysagalactea, moreover, (10,29). Experimentaly and naturally infection of Strept. bovis causing acute septicemia in pigeons. Moreover racing bacterial endocarditis is commonly associated with Strept. gallinarum, Ent. Faecales and Entero. faecuim (30) .

Ent. fecales was the predominant isolate among Enterococci and Stereptococci where it could be detected in 40.4% Table 4. Other Enterococcus species isolated from avian species and associated with disease were Ent. fecales, Ent. facium Ent. avium (31). These results agreed with those recovered in the present study Table 4.

REFERENCES

- 1.KaraKaya M and Aktumsek A (1996) An invostigation of the fatty acid Compositions of the meat, liver and egg fats of quails Gida, 21, 4: 239 - 241.
- 2.Abd El Aziz S A, Ragheb S and Sheshtawy E (2000): Bacterial and parasitis carried by migratory ducks in damietta and port said province. J Egy.Vet.Med.Ass. 61 (6): 257-266.
- 3.Medani G G, Mahmoud MA, Hussian, MM and Sabry (2002): Comparative study on bacterial infection of Enterobactereaceae in

japanese and migratory quials J. Egg. Vet. Med. Asc. 62 "N" I: 93 - 102.

- 4.Sherief M A, Ahmad A H and Nasef S A (1998): Micro biological studies on quail carcasses Suez Canal Vet. Med. J. (1): 233 241.
- 5.Arenas A S, Vicent I L Cgomaz-Villamandos R, Astorga, A.M and Tarkdas (1999): Outbrak of septicemic colibacellosis.in Japanese. quails, zentralbl Veteriner. Med. B., 46: 399-404.
- 6.Lin JA, Shyu and Shyu CL (1996): Detection of garm negative bactereal flora from diarrahia on shell chieken. J. Chinease Soc. Vet Sci 22 : 361 – 366.
- 7. VenKangouda G Krishnappa N and Upadhaye R (1996): Bacterial etiology of early chick mortality. Indian Vet. J. 73: 256 - 256.
- 8.Binak M W, Berzemeska R, Pisariski B, Blaszezk G, Kosowrska H and Karpenisk (2000): Evaluation of the efficacy of feed providing of gastrointestinal flora of newly hatched broiler chickens Archeiv fur geffug elkwmde 64: 147-151.
- 9. Welsh R D, Neman SL, Vanhooser and Dye LB (1997): Bacterial infection in ratites. Vet . Med. 92: 992 – 998.
- 10.Saif Y M, Barnes H J, Glison R J, Fadly A M, Mc-Dougald LR and Swayne B E (2001): Diseases of poultry 11 th Ed. Ed.Board Am. Asso. Avian Pathology, Lowa Stat Press U.S.A
- 11.Lin MY Cheng MC, Huang KJ and Tsai WC (1993): Classification, pathogenicity and drag Ruseptahlity of heamoly tie gram negtatvie bacteria isolated from sick or dead chicken Avian dis 37:6-9
- 12.Pekham M C (1984): Avian vibrionic hepatitis in Hofstade, H.J, Barnes, B. w., calnk. W. M Reid and yada, H.W. 8th Ed: 221-231
- 13.Skirrow M B (1490): Food borne illness caused by Campylobacter. Lancet . 336 : 921 – 923 Smith
- 14. Edward P R and Ewing WH (1972): Identification of Enterobactreaceae 3 rd Ed

. Burgeos publish Minnepolis Ahanta ,USA . 208 – 337

- 15.Cruickshank R Duguid T P Mormon B P and Swain R H M (1975): Practice of medical Micro biology12 th Ed .V. 11. Churchilliungs tirn, London.
- 16.MaC Faddian J (1980): Biochemical tests for identification of Med. Bacteriology 2nd Ed.Willings & Wilken Cop, USA
- 17.Elsheshtawy E A and Shaker M M (1999): Some studies on Mycoplasma and bacterial infection In Quails J. Egy Vet. Med. Ass. 59 (6):1575 – 1584.
- 18.Morishita, TY, Aye PP, Elizabeth, LC and Harr BS (1999) : Survay of the pathogens and Blood parites in free living passerines.Avian disease 43 : 349 – 352.
- 19.Eissa AA, El-Shorbagy M and Mossad A (2001):Colibacellosis in quails ad trials for vaccination J. Egypt. Vet. Met. Ass.61 No 2 : 395-403
- 20.Ginns C A, Brouning F, Benhan l and whi ther KG (1998):Devlopment and application of an aerosol challing method of reproduction of avian colibacollosis.Avian Path.27:505-511.
- 21.Dessauky M I, Moursy Z U, Niazy and Abd All (1982):Experimental Klebsiella infecteon in baby chicks. Archiv. Gefluglk undl 46: 145 – 150
- 22.Sah M P Mal and Mohanty G C (1983): Septicemic proteus infection in Japanese quails chick Avian disdease Vol 27 (1): 296 – 300
- 23.Ye S Xu, Huang Zhang Z, Wong W, Jiang SZ and Jiang M (1995): Investion of proteus inffection in chickens Chineas J. Vet. Sci. Tech 25: 14 15.
- 24.Elsharbagy M A M Ossaaol A A Boshra M Hussain M and Meruat M (2000): Studies on Pseudomomas aerogenosa infection in Quails. J. Egypt . Vet . Med. Aes. 61 (1):205-221
- 25. Younes J S, Yousef H, Abd El-Karim S and Hassanin (1975): Epedeniological Studies of Psaudomonus aerogenosa in chicken, fish

and human. Assiut vet. Med 23 (45): 48 – 56

- 26.Raja Rajesward, K Dhananjay, Reddy B J R and Dhanalakshmi K (1992): Normal airobic bacterial flora of respiratory, intestinal trect and oviduct of japanese quails. Indian J. Vet. Med 46: 547 – 551
- 27.Mostafa F A and Hussain S Z (1999): Gram positive cocci causing septicemia in chicken in Assiut Governorate Assint Vet. Med. J. 41 (82) 227 – 237
- 28.Sohad M Dorgham (2003): Staphylococcus infection in poultry. M. Vet. Sc. Alex. Univ .Vet. Microbiology

- 29.De Herdt P R, Deukatella Desmidate F Haesebrouck IA Groote B De Roels (1994): An unusual, Outbreak of Streptococcus bovis septicema, In racing pigeons. Vet . Rec . 134:42-43
- 30.Domermuth C and Gross (1169): Medium for isolation and Tentative identification of *Fecal Strept.*, and their rule in avian pathgenesis J.Avian dis.13:394-399
- 31.Collens O, Jones J, Farrow R, Kilpper B and Schleifer (1984): Enterococcusavium nom rev., E.caleseflavis,E durans,E galinarum ada E maldoratus Int J. System. Bacteriol (34): 220 – 223.

الملخص العربى دراسة عن بعض الإصابات البكتيرية في السمان بمحافظة البحيرة أحمد أبو المجد بخيت معهد صحة الحبو ان – محافظة البحير ة - دمنهو ر

أجريت هذه الدراسة على عدد ٧٠ سبعون عينة مختلفة الأعمار من مزارع خاصة بمحافظة البحيرة وكانت كل عينة ممثلة لمشكلة مرضية تم إجراء الدراسة البكتريولوجية لهذه العينات على الأنسجة المختلفة التى شملت الرئة والقلب والكبد والحوصلة والأمعاء وقد تم عزل الايشريشيا كولاى القولونية والكليبسيلا والأنتيروباكتر والستروباكتر والعصيات الزائفة ممثلة لعائلة البكتريا المعوية وكانت نسبة تواجدها على التـوالى ٢٠,٧ %و ٩,٧% و ٩,٩% و ٩,٣% و ٩,٩% تم عزل ميكروب السودموناس ايروجينوسا

تم عزل الميكروب العنقودى الذهبى بنسبة ٢, ٥ % من سلالات الميكروب العنقودى المعزولة إضافة إلى العديد من باقى المجموعة وشملت العنقودى الداجنى والمتوسط والكوهينى ٢,١ والعنقودى الرأس إضافة إلى العنقودى مجموعة هيكص بنسب ٢,٨% و ٦,٦% و ٦,٦% ٦,٦% و ٣,٥% و ٣,٥% و ٣,٥ على التوالى تم عزل الميكروب السبحى بشقيه البرازى متمثلاً فى المكورات المعوية البرازية والمكورات نوع فيكم و الداجنى بنسب ٤٠,٤ % و ١٥,٤% و ١٥,٥% من إجمالى السبحيات المعزولة المقدرة ٢ عتره هذا إضافة إلى المكورات السبحية نوع جالينيرم ونوع دييس أجلاكتيا و المكورات المعوية البرازية والمكورات المؤرة من إحمال الميدين بنسب ٤٠,٤ شوع فيكم و ١٥,٥ % من إجمالى السبحيات المعزولة المقدرة ٢ متره هذا إضافة إلى المكورات السبحية نوع جالينيرم ونوع دييس أجلاكتيا و المكورات المعوية البورية المؤرة من المكورات السبحية المؤلية مع من المكورات المعوية المقدرة ٢٠ والمكورات السبحية الرؤية ١١,٥ % و ٩,٦ % و ٣,٥ % و ٢,٠ %