

## MICROBIOLOGICAL EVALUATION OF SOME CHICKEN MEAT AND MEAT PRODUCTS

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### Abstract

One Hundred random samples of chicken meat products (shieshtawook and pattie) and meat products (Scallop and salami) were collected from different supermarkets in Kalubia and Gharbia governorates (25 of each) and subjected to bacteriological and mycological examinations. The result of bacteriological examination proved that the mean values of total Aerobic Plate Count (APC), Enterobacteriaceae count and Staphylococcal count were  $2.29 \times 10^6 \pm 0.36 \times 10^6$ ,  $8.61 \times 10^3 \pm 1.95 \times 10^3$  and  $5.42 \times 10^4 \pm 1.08 \times 10^4$  for shieshtawook samples,  $7.64 \times 10^5 \pm 2.10 \times 10^5$ ,  $2.76 \times 10^3 \pm 1.47 \times 10^3$  and  $1.13 \times 10^4 \pm 0.24 \times 10^4$  for pattie samples,  $2.08 \times 10^5 \pm 0.52 \times 10^6$ ,  $9.96 \times 10^2 \pm 2.37 \times 10^2$  and  $3.34 \times 10^3 \pm 0.60 \times 10^3$  for scallop samples and  $4.88 \times 10^5 \pm 0.48 \times 10^5$ ,  $4.18 \times 10^4 \pm 1.48 \times 10^4$  and  $6.58 \times 10^3 \pm 2.15 \times 10^3$  for salami samples, respectively. Enteropathogenic E.coli were isolated from 24%, 12%, 8% and 8% of examined shieshtawook, pattie, scallop and salami samples, respectively, also, Salmonella species were isolated only from 8% and 4% of shieshtawook and pattie samples, respectively. Mean values of total mould count were  $2.43 \times 10^4 \pm 1.32 \times 10^4$ ,  $4.06 \times 10^3 \pm 1.43 \times 10^3$ ,  $1.63 \times 10^4 \pm 0.88 \times 10^4$  and  $1.18 \times 10^4 \pm 0.42 \times 10^4$  in shieshtawook, pattie, scallop and salami samples, respectively and the most frequently encountered moulds were Aspergillus species. On the other hand, the incidence of toxigenic strains of Aspergillus flavus isolated from examined samples was 8%, 8% and 16% for shieshtawook, scallop and salami samples, respectively. Also, Aflatoxins B1 and B2 were produced from cultivated toxigenic strains on the media at different levels. The hazardous effects of isolated bacteria and mould strains as well as recommendations to improve the quality of these products were discussed.

### INTRODUCTION

Ready to eat meat products are highly demanded for their high biological value, reasonable price, agreeable taste and easily serving. Meat products are considered as an excellent sources of high quality protein, minerals and vitamins (WHO, 1984 and Mosupy et al., 1998).

Meat poultry products play an important role in filling the gap of protein deficiency and they can be considered the best choice in solving the human nutritional problems (Stephan et al., 2003).

Microbiological food safety and food borne infections are important public health concern worldwide. There have been a number of food borne illnesses resulting from the ingestion of contaminated foods such as chicken meat . Most of the pathogens that play a role in food borne diseases have a zoonotic origin (Busani et al.,2006). Insufficient cooking may result in survival of *E.coli* and subsequently causes food poisoning to consumers (Belongia et al.,1991). Also enteropathogenic *E.coli* are well recognized as a cause of infantile diarrhea and /orgastrointestinal illness in adult human (Woodenburn and Raab, 1997).

A wide range of food has been implicated in food borne Salmonellosis. However, as the disease is primarily zoonotic , food of animal origin has been consistently implicated as the main source of human salmonellosis (FAO/WHO,2002). Consumption of food contaminated with *Salmonellae* can cause salmonellosis; one of the most common bacterial food borne illness (FSIS,2003) . *Staph.aureus* plays a great role as a bacterial contamination of cooked meat during preparation and processing of cooked meat that may be eaten without sufficient cooking or heating (Soliman,1988). Meat was considered an ideal media for mould growth as it has an optimum pH (5.6 -6.7), high water content ( $a_w = 0.99$ ), rich supply of nitrogenous substance and a source of carbohydrate (Coni, et al.,1994). Some mould species could be a public health hazard because they have the ability to produce mycotoxins (Scheurlen, 1996). In this respect, some *Aspergillus* species have received a great attention as they can produce aflatoxins which have carcinogenic effects (Montagana ,etal., 2004) . Therefore the aim of this work is to evaluate some meat and chicken products from bacteriological and mycological aspects beside, discussion of health hazards effects of the isolated organisms.

## **MATERIALS AND METHODS**

### **1- Material**

One hundred random samples of chicken and meat products (50 of each ). Chicken meat samples were shieshtawook and pattie (25 of each) and the meat sample were scallop and salamy (25 of each ). Samples were collected from different supermarkets at El-kalubia and El-Gharbia Governorates, the collected samples were frozen and transferred in their original packages directly to the laboratory in an insulated ice box under a complete septic conditions to be examined bacteriologically and mycologically.

## **2- Method**

A- Preparation of samples for microbiological examination according to APHA (2001).

B- Bacteriological examination of chicken and meat products samples:

- 1-Determination of APC according to APHA(2001).
- 2-Determination of total Enterobacteriaceae count according to APHA(2001).
- 3- Isolation and identification of E.coli according to FDA(2002) .
- 4- Determination of total Staphylococcal count and isolation of S.aureus according to FDA(2002).
- 5- Isolation and identification of Salmonellae according to ISO 6579 (2002).

C- Mycological examination of chicken and meat products samples:

- 1- Preparation of the samples according to APHA(2001):
- 2- Determination of total mould count according to ISO 21527/1(2009).
- 3- Identification of mould isolates according to Koneman and Roberts (1985).
- 4- Screening of toxigenic Aspergilli according to Hara, et al.,1994.
- 5- Confirmation of mycotoxins production by cultivation and extraction of Aspergillus toxins according to Pestka (1996) and application of thin layer chromatography (TLC) according to Schuller and Egmond (1991).

The obtained data were statistically evaluated by Analysis of Variance (ANOVA) according to Feldman et al., (2003)

## RESULTS

Table (1): Statistical analytical results of bacterial counts in the examined samples of ready to eat chicken and meat products ( n = 25 ).

Criteria	Chicken products						Meat products					
	Shieshtawook			Pattie			Scallop			Salami		
	Min.	Max.	Mean $\pm$ SE	Min.	Max.	Mean $\pm$ SE	Min.	Max.	Mean $\pm$ SE	Min.	Max.	Mean $\pm$ SE
APC	$9.7 \times 10^4$	$1.1 \times 10^7$	$2.29 \times 10^6$ $\pm 0.36 \times 10^6$	$8.5 \times 10^3$	$6.2 \times 10^6$	$7.64 \times 10^5$ $\pm 2.10 \times 10^5$	$1.9 \times 10^3$	$4.7 \times 10^6$	$2.08 \times 10^5$ $\pm 0.52 \times 10^6$	$1.0 \times 10^4$	$4.0 \times 10^6$	$4.88 \times 10^5$ $\pm 0.48 \times 10^5$
TEC	$3.3 \times 10^2$	$2.6 \times 10^5$	$8.61 \times 10^3$ $\pm 1.95 \times 10^3$	$1.2 \times 10^2$	$4.9 \times 10^4$	$2.76 \times 10^3$ $\pm 1.47 \times 10^3$	$8.0 \times 10^0$	$1.5 \times 10^4$	$9.96 \times 10^2$ $\pm 2.37 \times 10^2$	$1.5 \times 10^2$	$3.0 \times 10^5$	$4.18 \times 10^4$ $\pm 1.48 \times 10^4$
TSC	$4.0 \times 10^2$	$7.0 \times 10^5$	$5.42 \times 10^4$ $\pm 1.08 \times 10^4$	$2.0 \times 10^2$	$3.1 \times 10^5$	$1.13 \times 10^4$ $\pm 0.24 \times 10^4$	$1.0 \times 10^2$	$6.0 \times 10^4$	$3.34 \times 10^3$ $\pm 0.60 \times 10^3$	$1.0 \times 10^2$	$5.0 \times 10^4$	$6.58 \times 10^3$ $\pm 2.15 \times 10^3$

Min. = Minimum, Max. = Maximum, APC = Aerobic Plate Count, TEC = Total Enterobacteriaceae Count,

TSC = Total Staphylococci count, S.E = Standard Error of mean.



Table (5): Total mould counts/gm of the examined chicken and meat products samples: ( N= 25 of each ):

Samples	Positive samples		Minimum	Maximum	Mean $\pm$ S.E
	No.	%			
Shieshtawook	19	76%	$5.0 \times 10^2$	$3.0 \times 10^5$	$2.43 \times 10^4 \pm 1.32 \times 10^4$
Pattie	22	88%	$2.0 \times 10^2$	$3.0 \times 10^4$	$4.06 \times 10^3 \pm 1.43 \times 10^3$
Scallop	20	80%	$3.0 \times 10^2$	$2.1 \times 10^5$	$1.63 \times 10^4 \pm 0.88 \times 10^4$
Salami	21	84%	$5.0 \times 10^2$	$5.0 \times 10^4$	$1.18 \times 10^4 \pm 0.42 \times 10^4$

S.E = Standard Error of mean.

Table (6): Incidence of mould species isolated from examined chicken and meat products samples: ( n = 25 )

Criteria	Chicken products				Meat products			
	Shieshtawook		Pattie		Scallop		Salami	
	NO.	%	NO.	%	NO.	%	NO.	%
Aspergillus	5	20%	15	60%	10	40%	8	32%
Cladosporium	-	-	5	20%	-	-	5	20%
Fusarium	15	60%	3	12%	15	60%	-	-
Penicillium	15	60%	7	28%	-	-	-	-
Trichoderma	10	40%	-	-	-	-	5	20%
Sporotricum	5	20%	4	16%	10	40%	15	60%
Alternaria	5	20%	-	-	5	20%	10	40%
Thamnidium	-	-	-	-	5	20%	10	40%

Table (7): Incidence of toxigenic strains of Aspergillusflavus(A. flavus) isolated from examined chicken and meat products samples ( n = 25 ):

Samples	Total isolated A.falvus		Toxins producer of A.flavus	
	No	%	No	%
Shieshtawook	3	12%	2	8%
Pattie	5	20%	-	-
Scallop	5	20%	2	8%
Salami	5	20%	4	16%

Table (8): Levels of Aflatoxins B1 and B2(ppp/ml of media) extracted from toxigenic strains of *A.flavus* isolated from the examined chicken and meat products samples.

Samples	Aflatoxin B1			Aflatoxin B2		
	Minimum	Maximum	Mean±SE	Minimum	Maximum	Mean±SE
Shieshtawook	41.4	69.4	55.4±19.80	6.8	14.1	10.45±5.16
Scallop	17.6	30.2	23.9±8.91	2.3	6.6	4.45±3.04
Salami	19.9	25.4	18.13±3.10	1.6	6.4	3.4±1.21

ppp = Part Per Pillion, S.E = Standard Error of mean.

## DISCUSSION

Meat and chicken products are the most common food vehicles of human infection with enteropathogens throughout the world (Abd-El-Aziz *et al.*,2001). Meat exposed to different types of treatments from the point of slaughtering until it is ready for consumption,these will be added to the bacterial load of this meat. thus it may be contaminated with several types of organisms through long chain of preparation,handling of raw meat, processing, distribution,storage and retailing.

The result in table (1) recorded that the APC/g of examined samples of chicken and meat products were ranged from  $9.7 \times 10^4$  to  $1.1 \times 10^7$  with an average of  $2.29 \times 10^6 \pm 0.36 \times 10^6$  for shieshtawook,  $8.5 \times 10^3$  to  $6.2 \times 10^6$  with an average of  $7.64 \times 10^5 \pm 2.10 \times 10^5$  for pattie,  $1.9 \times 10^3$  to  $4.7 \times 10^6$  with an average of  $2.08 \times 10^5 \pm 0.52 \times 10^6$  for scallop and  $1.0 \times 10^4$  to  $4.0 \times 10^6$  with an average of  $4.88 \times 10^5 \pm 0.48 \times 10^5$  for salamy. Shieshtawook samples were higher in microbial load than other products and this may be due to the adding of spices and vegetables (as green piper and tomatoes) during the preparation of shieshtawook. Nearly the same result was obtained by Purabi and Joshi (2010) in chicken meat products.

Total Enterobacteriaceae count in examined samples were ranged from  $3.3 \times 10^2$  to  $2.6 \times 10^5$  with an average of  $8.61 \times 10^3 \pm 1.95 \times 10^3$  in shieshtawook,  $1.2 \times 10^2$  to  $4.9 \times 10^4$  with an average of  $2.76 \times 10^3 \pm 1.47 \times 10^3$  in pattie,  $8.0 \times 10^2$  to  $1.5 \times 10^4$  with an average of  $9.96 \times 10^2 \pm 2.37 \times 10^2$  in scallop and  $1.5 \times 10^2$  to  $3.0 \times 10^5$  with an average of  $4.18 \times 10^4 \pm 1.48 \times 10^4$  in salamy, The most highest count were found in salami samples may be due to its more frequently handling in

meat products were obtained by Elwi,1994 and Enas(2011). Meanwhile, total staphylococcal count in examined samples were ranged from  $4.0 \times 10^2$  to  $7.0 \times 10^5$  with an average of  $5.42 \times 10^4 \pm 1.08 \times 10^4$  in shieshtawook, from  $2.0 \times 10^2$  to  $3.1 \times 10^5$  with an average of  $1.13 \times 10^4 \pm 0.24 \times 10^4$  for pattie , from  $1.0 \times 10^2$  to  $6.0 \times 10^4$  with an average of  $3.34 \times 10^3 \pm 0.60 \times 10^3$  in scallop and from  $1.0 \times 10^2$  to  $5.0 \times 10^4$  with an average of  $6.58 \times 10^3 \pm 2.15 \times 10^3$  in salamy. Nearly similar results of salamy and lower results of scallop were obtained by Enas(2011)in meat products while she recorded lower results in case of chicken products.

Table (2) indicates the incidence of E.coli isolated from chicken and meat samples in which 24% of sheishtawook samples were contaminated with E.coli that identified as O26:K6 (B6) (4%) , O119: K69 (B19) (8%) , O124 :K72 (B17) (8%) and untypable(4%)serovars.E. coli could also isolated from 12% of pattie samples and identified as 8% serovar O124:K72(B17) and 4% serovar O128:K67(B12). Meanwhile E. coli could be isolated from 8% of scallop "4% each of O26:K60(B6) and O119:k69 (B19)", and from 8% of salamy samples "4% each of O111:K58(B9) and Untypable serovars".

Such enteropathogenic E.coli were previously isolated from chicken products by Abou-Hussein –Reham (2007), as well as from different ready to eat meat products by Soliman and El-Tabiy (2006), El-Rayes (2008) and Enas (2011). According to Bryan (1982), O128 serotype of E.coli is called enterotoxigenic E. coli (ETEC), while strains causing desentry like syndrome (O124) were known as Enteroinvasive E. coli (EIEC) and the strains causing haemorrhagic colitis (O111) were recognized as Enterohaemorrhagic E.coli (EHEC).

Table (3) revealed that the incidence of Salmonella species isolated from chicken products samples were 8% from shieshtawook (4% each of S.entritidis and S. typhimurium) and 4% from pattie samples identified as S.typhimurium but we couldn't isolate any Salmonellae from meat product samples (scallop and salamy). Salmonella spp. were previously isolated from ready to eat meat products by Soliman et al., 2002; Richardson and Stevens (2003) and Enas (2011) while Ehab(2003) and Reham(2007) couldn't isolate salmonella from chicken and meat products. Practically, all food of animal origin may be a vehicle for transmission of salmonella to man. Meat and chicken products may be contaminated with human excreta at any step of processing chain; during handling of raw material or preparation of such food in kitchen (Fathi et al.,1994). Our obtained results showed that E.coli is more prevalent in examined samples than salmonella spp. This is agree



with Zaho et al., (2001) and Purabi and Joshi (2010) whose reported high prevalence of *E. coli* than salmonella spp. in poultry products as well as in retail meat market had reported by Kumar et al., (2001).

Table(4) showed the incidence of *Staph.aureus* in the examined chicken and meat products samples. Results revealed that 16% of salami samples contaminated with *Staph. aureus* while all other samples were free from *Staph. aureus*. The presence of *Staph.aureus* in salami samples only may be due to the excessive handling in supermarket during retail.

The negative samples results were similar to those obtained by Tolba(1994) and Mohamed (2000); they couldn't detect or isolate *Staph .aureus* in any of finished heat treated meat products they were examined. On the other hand, Nesreen(2003), Reham(2007) and Enas (2011) could isolate *Staph.aureus* from meat and chicken products.

*Staph.aureus* enters the food through the ex-charges of nasal passage and the infected wounds of many persons that may act as a common source of infection (Frazier and Wethoff, 1983).

Table (5) revealed the results of total mould count /gm of examined chicken and meat samples. It ranged From  $5 \times 10^2$  to  $3.0 \times 10^5$  with an average of  $2.43 \times 10^4 \pm 1.32 \times 10^4$  in shieshtawook, from  $2.0 \times 10^2$  to  $3.0 \times 10^4$  with an average of  $4.06 \times 10^3 \pm 1.43 \times 10^3$  in pattie, from  $3.0 \times 10^2$  to  $2.1 \times 10^5$  with an average of  $1.63 \times 10^4 \pm 0.88 \times 10^4$  in scallop and from  $5.0 \times 10^2$  to  $5.0 \times 10^4$  with an average of  $1.18 \times 10^4 \pm 0.42 \times 10^4$  in salami samples. Nearly similar results in chicken products recorded by Purabi and Joshi (2010). Meanwhile, nearly similar results in meat products were recorded by Hegazi et al., (1992), Maha and Sohad (2005) and El-shazly (2006), while, higher results were recorded by Shaltout (1996), Shahenda (2002) and Hanan and Marionette (2008).

Fungal growth can produce toxic metabolites which lead to mutagenic, carcinogenic and teratogenic effects to the human health (EL-Shinawy et al., 1994). Also mould contamination of food may cause acute and chronic intoxication to man resulting in serious clinical symptoms such as jaundice and vomiting (Sayed et al., 2002).

Table (6) showed the incidence of mould species isolated from the examined samples of chicken and meat products, which were comprised; *Aspergillus*, *Cladosporium*, *Fusarium*, *Penicillium*, *Trichoderma*, *Sporotricum*, *Alternaria* and

Thamnidium. The highest percentage of *Aspergillus* species (60%) was isolated from pattie samples, while, the highest percentage of *Penicillium* was found in shieshtawook samples. This high percentage of *Aspergillus* may be high enough to produce toxins and constitute a dangerous hazard to consumers. Similar to our results, Alalhi and Alboshan (2004) had also reported contamination of chicken products with *Penicillium* sp. and *Aspergillus* sp.

The incidence of toxigenic strains of *Aspergillus flavus* isolated from examined samples of chicken and meat products was shown in table (7). The toxin producing *A. flavus* was found in highest percentage in examined salami samples (16%) followed by shieshtawook and scallop samples (8% in each). In contrast, in pattie samples, we could not detect toxigenic strains of *Aspergillus flavus* in such product. Nearly the same results of *Aspergillus* spp. incidence were recorded by Hassan (2004); Ouf (2004) and Hanan and Marionette (2008), in meat product samples they were examined.

Regarding the results in Table (8), the average amount of Aflatoxins B1 and B2 produced by *A. flavus* (on cultivated media) isolated from shieshtawook was the highest contents among our results followed by salami and scallop isolates. These results indicate that; the risk of production of aflatoxins B1 and B2 in these products is present if subjected to suitable growth conditions for the toxigenic strains they were harbored. The current results agreed with those recorded by many investigators; Hassan (2004); Hanan and Marionette (2008) as they could isolate aflatoxigenic strains of *A. flavus* from meat products.

We can conclude that; the organisms like *Salmonellae*, *E. coli*, *Aspergillus* spp., *Penicillium* spp. and *Fusarium* spp. were the most prevalent contaminants in the examined samples from various markets of our study. Also, our study confirms the prevalence of toxigenic strains of *Aspergillus flavus* in most examined products except pattie samples. These contaminants can pose health hazards to consumers in consumption of such meat products.

We can recommend that; contamination should be controlled through strict precautions, to minimize the risk of hazards, such as accurate control of the relative humidity and temperature for the storage of chicken and meat products. Also, addition of preservatives and/or food additives should be under control and examined periodically for bacterial, fungal contamination and mycotoxins production as well as suitable storage conditions to avoid microbial or/and mycological growth.

## REFERENCES

1. Abd-El-Aziz, A.T.N.; Hassan,M.K.; Shabaan A.I. and EL-Moneim, K.M.A. 2001. Prevalence of Salmonellae and Listeria in Cairo – poultry Abattoir and broiler carcasses. *J.Egypt Vet.Med. ASS.* 61(60): 209-218.
2. Altalhi, A. and M. Albashan. 2004. Mycological study on fresh and frozen meat in Taif city, Saudi Arabia. *Assiut Vet. Med. J.*, 50:22-32.
3. APHA "American Public Health Association". 2001. Compendium of methods for the microbiological examination of foods. 2nd Ed. Speck, H.L. (ed.). APHA. Washington D.C.
4. Belongia, E.A.; Macdonald K.L.; Parham, G.A.; White, K.E.; Karlton, J.A.; Lobot, M.N.; Busani, L., G.; Scavia, I. Luzzi and A. Caprioli. 1991. Laboratory surveillance for prevention and control of food borne zoonoses. *Ann. Ist super Sanita*, 42:401-404.
5. Bryan, F.L. 1982. Disease transmitted by foods. Textbook, 2nd Ed. HHS. Publ. Services, public health services, center for disease control, Atlanta, Georgia 30333 USA.
6. Busani, L.; G. Scavia, I. Luzzi and A. Caprioli. 2006. Laboratory surveillance for prevention and control of food borne zoonoses. *Ann. Ist Supersanita*, 42:401-404.
7. Coni, W., Costilow, R. and Blumer, T. 1994. Occurrence of fungi in meat and their properties with regard to food hygiene. *Excerpta Medica - Veterinaria* 23:73-76.
8. CSIRO, T. and AFISCA, A. 1993. Food Science Australia, handling food in Home – a Joint Venture of CSIRO and AFISCA, Australia P.O. Box 52, North Ryde NSW 1670.
9. Ehab, S.Y. 2003. Aerobic and anaerobic enterotoxigenic bacteria in ready to eat food. Ph.D. Thesis, Fac. Vet. Med., Moshtohor, Zagazig Univ. Banha branch.
10. El-Rayes, A.M.A. 2008. Incidence of pathogenic E. coli in fast food. M.V.Sc. thesis, Fac. Vet. Med., Banha University.
11. Elshazly A.M. 2006. Mycological investigations of some meat products. M.V.Sc. Thesis Fac. Vet. Med. Cairo Univ.
12. El-Shinawy, S.H.; Abd-El-Aziz, A.M. and Hady, H.A. 1994. Microbiological quality of infant powdered milk. *J. Egyptian Vet. Med. Ass.* 55 (2,1):147-157.
13. Elwi, E.M. 1994. Sanitary improvement of meat meals in governmental hospitals in Assiut city. Ph.D. thesis, Meat Hygiene, Fac. of Vet. Med., Assiut University.
14. Enas A.A.M. 2011. Microbial and chemical evaluation of fast food. M.V.Sc. thesis, Fac. Vet. Med. Moshtohor, Zagazig Univ. Banha branch.
15. FAO/WHO. 2002. Risk Assessment of Salmonella in eggs and broiler chickens. Food and Agriculture Organization / World Health Organization, Geneva, Switzerland.

16. Fathi, S.; El-Khateib, T.; Moustafa, S. and Hassanein, K. 1994. Salmonellae and enteropathogenic E.coli in some locally manufactured meat products. *Assuit Vet. Med. J.* 31(61) : 190-199.
17. Feldman, D.; Ganon, J.; Hffman, R. and Simpsn, J. 2003. The solution for data Analysis and presentation Graphics , 2nd Ed., Abacus Lancripts, Inc., Berkeley, CA, USA.
18. FDA "Food and Drug Administration". 2002. Isolation and identification of E.coli.
19. FDA "Food and Drug Administration". 2002. Isolation and identification of Staph.aureus.
20. Food Safety and Inspection Service "FSIS" United States Department of Agriculture. 2003. Meat Preparation : Beef from farm to Table .Washington . DC.20250 – 3700.
21. Frazier, W.C. and Westhoff, D.C. 1983. Microbiology, 4th Ed. Tata Megraw Hill Publishing Co. Limited New york USA.
22. Hanan, M.Lamada and Marionette, Z.Nassif. 2008. Mycological profile of meat products , with special reference to aflatoxins. 9th Vet.Med.Zag.Conference (20-22 August) Port-said, 201-208.
23. Hara, S.; Fennel, D. and Hesseltine, C. 1994. Aflatoxin producing strains of A.falvus detected by fluorescence of agar medium under U/V light .*Appl. Microbiol.* 47(6): 118.
24. Hassan, M.A. 2004. Control of mycological hazards in sheep carcasses with special reference to Aflatoxins. *Suez canal Vet.Med.J.VII (2)*, 359-366.
25. Hegazi, S.M.; El-Far, F.A.M. and Aziz N.A. (1992): Studies of fungal contamination of meat products .*Vet.Med. J. Giza* 31:40.
26. ISO 6579. 2002. Microbiological of food and animal feeding stuffs- Horizontal method for the detection of salmonella spp.
27. ISO 21527/1. 2009. Microbiological of food and animal feeding stuffs- Horizontal method for enumeration of yeast and mould part 1 colony count technique in products with water activity more than 0.95.
28. Koneman, E. and Roberts, C. 1985. Practical Laboratory Mycology. 3rd., Williams and Wilkins . Baltimore, London.
29. Kumar, H.S., S.Ottu, I. Karunasagar and I. Karunasgar. 2001. Detection of Shiga – Toxigenic E.coli (STEC) in Fresh sea food and meat marketed in Mangalore, India by PCR *Lett. Applied Microbiol.*, 32:334-338.
30. Maha, I.A.E. and Sohad, H.E.E. 2005. Some chemical and mycological examination of meat and fish products .*Vet.Med. J.Giza* (55:941).

31. Mohamed, E.N. 2000. Quality investigation into beef frankfurter produced in Egypt .M.V.SC., Thesis, Fac.Vet.Med., Cairo Univ.
32. Montagna, M., Santacroce, M., Spilotros, G., Napoli, C., Papa, A. and Dragoni, I. 2004. Investigation of fungal contamination in sheep and goat in Southern Italy. *Mycopathologia* 158(2) : 245 -247.
33. Mosupy, F.M.; Arntzen, L. and Van Holy, A. 1998. Microbiological survey of street – Vended foods in the Johannesburg metropolitan area of South Africa. *Food Sci.*, 63(7) :842-846.
34. Nesreen, E.Z.H. 2003. Effect of chemical preservatives on food poisoning bacteria in some locally manufactured meat products. Ph. D. Thesis, Fac. Vet. Med., Zagazig University.
35. Ouf, J.M. 2004. Microbiological evaluation and mycotoxin residues in some frozen camels meat products. *Vet.Med.J.Giza* .52,(2):213-230.
36. Pestka, J.J. 1996. Fungi and Mycotoxins in meat . In: *Advances in Meat Research*, (eds.A.M.Pearson and T.R. Dutson ), Mac Millan, Basingstoke, pp.227-309).
37. Purabi Saikia and Joshi S.R. (2010): Retail market poultry meat of North- east Indian –A microbiological survey for pathogenic contaminants. Dep. of Biotechnology and Bioinformatics , Microbiology Lab., North – Eastern Hill Univ., Shillong, Meghalaya, India. *Research Journal of Microbiology* 5(1) : 36-43.
38. Reham, A.A. 2007. Detection of food mediated pathogens in some meat and chicken products by using recent techniques . Ph.D., Thesis , Fac.Vet.Med., Moshtohor, Zagazig Univ. Banha.
39. Richardson, I.R. and Stevens, A.M. 2003. Microbiological examination of ready to eat stuffing from retail premises in the north east of England . The "Get Staffed" survey . *J. Appl. Microbiol*, 94(4) :733-737 .
40. Sayed, M.A. ; Mahmoud, E.A. and Abo El-Alla, A.A. 2002. Microflora and natural occurrence of mycotoxin in meat of imported bulls, poultry and some meat products. *Assuit Vet.Med.J.* 43(89):189-200.
41. Scheurlen, M. 1996. Pathogenicity of fungi in the intestine *fortschr. Med.* 114: 319 -321.
42. Schuller, P. and Egmod, H. 1991. Detection and determination of mycotoxins in food and feed. *J.Food Protect.* 44 (8):227-231.
43. Shahenda, N. 2002. Microbial evaluation of food additives in some meat products . M.V.SC. Thesis (Meat Hygiene) Fac.Vet.Med. Zagazig University.
44. Shaltout, F.A. 1996. Mycological and mycotoxicological profile of some meat products. Ph.D. Thesis , Fac.Vet. Med. Zagazig Univ. Moshtohor, Banha branch.

45. Soliman, M.R. 1988. Sanitary status of ready to eat meat products and fish. M.V.Sc. Thesis (Meat hygiene) Fac. of Vet. Med. Cairo Univ.
46. Soliman, M.R.; Abd El-Moneim, K.M. and Saad, S.M. 2002. Microbiological quality of ready to eat meat products and fish in Urban and Rural areas. *J. Egypt. Vet. Med. Assoc.* 62 (6):39-51.
47. Soliman, Z.I. and El-Tabiy, A.A. (2006): A study on the occurrence of E.coli in some beef products with special references to E.coli O157 :H7 Assiut Vet. Med. J., 52(110):75-87.
48. Stand, S.M.; Kasale, K.A. and Osterholm, M. T. 1991. An outbreak of E.coli O157 :H7 colitis. Associated with consumption of pre-cooked patties. *J. Infection Disease* , 164:338-343.
49. Stephan, R.; Schumacher, S. and Zychowska, M.A. 2003. The VITR technology for rapid detection of listeria monocytogenes and other Listeria spp. *International Journal of Food Microbiolog.*, 287-290.
50. Tolba, K.S. 1994. Microflora in locally processed frozen meat. *Vet. Med. J. Giza*, 42(2) : 99.
51. Woodenburn, M.J. and Raab, C.A. 1997. House hold food preparers, Food safety knowledge and practice following widely publicized outbreaks of food borne illness. *J. Food Port.* 60 (9) : 1105-1109.
52. World Health Organization "WHO 1984". The role of food safety in health development .Report of Joint FAO/WHO Expert Committee on food safety, Geneva.
53. Zaho C., B. Ge, J.D.Villene, R.Sudler and E.Yeh et al., 2001. Prevalence of Campylobacter spp. ,E.coli and Salmonella serovares in retail chicken, turkey, pork and beef from the Greater Washington D.C. , Area. *Applied Environ. Microbial.*, 67: 5431-5436.

## التقييم الميكروبي لبعض منتجات الدواجن واللحوم

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وأجريت هذه الدراسة على عدد مائة (١٠٠) عينة من منتجات الدواجن (الشيخ طاووك والباتيه) ومنتجات اللحوم (الاسكالوب والسلامى) ٢٥ عينة من كل منتج والتي تم تجميعها بصورة عشوائية من المحلات السوبر ماركت بمحافظة القليوبية والغربية لفحصها بكتيريا وميكولوجيا. أوضحت نتائج الدراسة على ان متوسط الجعد الكلى للميكروبات الهوائية، الجعد الكلى للميكروبات المعوية والجعد الكلى للمكورات العنقوديات:  $1.6 \times 2.69 \pm 1.6 \times 0.36$ ،  $1.03 \times 1.95 \pm 1.03 \times 8.61$ ،  $1.04 \times 1.08 \pm 1.04 \times 0.42$ ، لعينات الشيخ طاووك،  $1.05 \times 2.1 \pm 1.05 \times 7.64$ ،  $1.03 \times 1.47 \pm 1.03 \times 2.76$ ،  $1.04 \times 0.24 \pm 1.04 \times 1.13$ ، لعينات الباتيه،  $1.05 \times 0.48 \pm 1.05 \times 4.88$ ،  $1.04 \times 1.48 \pm 1.04 \times 4.18$ ،  $1.03 \times 2.15 \pm 1.03 \times 6.08$ ، لعينات السلامى. كما تم عزل ميكروب الايشريشيا كولاي بنسبة ٢٤%، ١٢%، ٨%، ٨% لعينات الشيخ طاووك، الباتيه، الاسكالوب والسلامى على التوالي. كذلك تم عزل ميكروب السالمونيلا بنسبة ٨% و ٤% لعينات الشيخ طاووك والباتيه على التوالي، كما تم عزل ميكروب المكورات العنقودية الذهبية بنسبة ١٦% من عينات السلامى فقط. وكذلك تم الفحص الميكولوجى للعينات وقد وجد ان المتوسط الجعد الكلى للفطريات هو  $1.04 \times 2.43 \pm 1.04 \times 1.32$ ،  $1.03 \times 1.43 \pm 1.03 \times 1.63$ ،  $1.04 \times 1.03 \pm 1.04 \times 0.88$ ،  $1.04 \times 1.88 \pm 1.04 \times 0.42$  لكل من الشيخ طاووك، الباتيه، الاسكالوب والسلامى على التوالي. وقد تم عزل وتصنيف فطر الاسبرجيلس وتحديد عترات الاسبرجيلس فلافس التي لها قدرة على إفراز الافلاتوكسين وكذلك تم استخلاص سموم الافلاتوكسين B1، B2 بكميات مختلفة من هذه العترات المعزولة من عينات الشيخ طاووك، الاسكالوب والسلامى. وقد تمت المناقشة الصحية للميكروبات المعزولة من البكتيريا والفطريات وكذلك تم مناقشة بعض التوصيات اللازم تطبيقها للحد من تلوث منتجات الدواجن واللحوم والتي تؤدي إلى تحسين جودتها وسلامتها.