

ASSOCIATION BETWEEN A31G SNP OF THE MYOSTATIN GENE AND SERUM LEVELS OF ALP, GOT, P AND CA IN EGYPTIAN BUFFALO (*BUBALUS BUBALIS*)

Michel Fahmy Saad^a, Shawky Mahmoud^a, Mohammed Abu El-Magd^b
and Rasha Alsaed Alwakeel^a

^aDepartment of Physiology, Fac. of Vet. Med., Kafrelsheikh Univ.

^bDepartment of Anatomy & Embryology, Fac. of Vet. Med., Kafrelsheikh Univ.

ABSTRACT

Myostatin, a member of the transforming growth factor β (TGF- β) superfamily, is a negative regulator of skeletal muscle growth in mammals. The loss or inhibition of myostatin signaling dramatically increases muscle mass. This study is aimed to detect single nucleotide polymorphisms (SNPs) in the myostatin (Mstn) gene and to associate measure their effect on mineral (Ca and P) and enzymatic (GOT and ALP) serum levels in high and low meat producing Egyptian buffaloes. Mstn gene was amplified by PCR and subsequently, subjected to sequence analysis to identify different allelic patterns. The nucleotide sequence analyses revealed presence of a novel A31G SNP in Mstn1 locus at nucleotide number 50 before exon2. This SNP was synonymous as it is located in the non coding sequence and so no amino acid was changed. However, the sequences of the other three loci showed 100% identity among the Egyptian buffaloes and as compare to the published sequences of buffalo (GenBank accession number, DQ091762). To identify whether A31G SNP is accompanied by change in serum levels of GOT, ALP enzymes, and Ca and Ph, the levels of these enzymes and minerals were estimated in the serum of high and low meat producing Egyptian buffaloes at age of 2 year. The serum levels of ALP, GOT, P, and Ca were significantly increased in high meat producing buffaloes as compared to low meat producing animals. The results of this study can be enable breeders to do an accurate and earlier selection of high meat producing buffaloes on a genotype basis.

Keywords: Mstn gene, Egyptian buffalo, PCR, sequencing, SNPs.

INTRODUCTION

Buffalo meat is well comparable to beef in many of the physicochemical, nutritional, functional properties and palatability attributes (*Anjaneyulu et al., 1990*). Egyptian buffalo represents an important part of animal production in Egypt (about 41% of total red meat production). However, because their growth rate and meat production are very poor when compared to foreign breeds, their meat production is insufficient for our needs (*El-Magd et al., 2013; El-Nahas et al., 1998*).

Myostatin (*Mstn*), a member of the transforming growth factor β (TGF- β) superfamily, is expressed in skeletal muscle (throughout life, from the early stages to late adulthood) and to some extent in fat tissue, heart and mammary tissues (*Kambadur et al., 1997; McPherron and Lee, 1997*). *Mstn* is a negative regulator of skeletal muscle growth in mammals (*McPherron et al., 1997*). Therefore, inhibition of this gene either naturally as in some cattle breeds or experimentally will cause sever increase in muscle growth (double-muscling). Myostatin knockout doubles the muscle mass of mice through both hypertrophy and hyperplasia of muscle fibers (*Amthor et al., 2009*).

Muscle metabolism can be assessed by biochemical techniques such as using aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), glutamic-oxaloacetic transaminase (GOT), lactate dehydrogenase (LDH) and creatine phosphokinase (CPK) enzymes activity (*Latimer et al., 2006*). Serum GOT content is normally low in level, but after extensive destruction of cardiac, hepatic, or skeletal tissues, this enzyme is librated into the blood at high levels (*Harper, 1971*). The estimation of serum GOT is widely used as diagnostic tool for liver injuries, myocardial infarctions, and skeletal muscle sympathies (*Ogita and Markert, 1989*).

Ca is an important regulator of several body functions: intracellular signalling, muscle contraction, functioning of the nervous system, hormone and enzyme secretion and blood clotting. Calcium ions are believed to be critical for the fusion of myoblasts, as this process can be activated by calcium ionophores that carry calcium ions across cell membranes (*David et al., 1981*). Phosphorus (P) is an essential nutrient for the skeleton and Ph deficiency causes rickets in children (*Pettifor, 2008*) and osteomalacia in adults (*Genant, 1993*). Nevertheless, P deficiency is seldom due to a low dietary intake of Ph, but rather to a genetic disorder.

It was hypothesized that mutation of *Mstn* can increase meat production in animals and this may be accompanied with increase enzymatic activity of GOT and AP and Ca and Ph. Thus, we conducted this experiment to confirm this hypothesis.

MATERIALS AND METHODS

Animal source:

Egyptian buffalo either male or female with different age and weight were used in this study. Animals were maintained at El-Nataff El-Gidid Experimental Stations (Mahalet Mousa in Kafrelsheikh Governorate, Egypt) belonging to Animal Production Research Institute, Ministry of Agriculture. All records of growth traits (body weight, body height and average daily gain) for different growth periods (birth, 3, 6, 9, 12, 18 and 24 months) in Beheiry buffaloes were collected from farm records to be used for statistical analysis. For more details see our recent published paper (*El-Magd et al., 2013*).

DNA extraction:

Blood samples from the jugular veins were collected, each sample was divided into two tubes, EDTA- tube (for DNA extraction) and plain tube (for measure blood parameters in serum). Blood samples were collected from jugular vein puncture into two vacutainer tubes, the first containing an anticoagulant (disodium EDTA) and kept in ice box. The genomic DNA was extracted using Gene JET genomic DNA extraction kit following the manufacturer protocol (Fermentas, #K0721/USA) .The second tube is a plain tube to measure blood parameters in serum.

Polymerase chain reaction (PCR)

A partial sequence of myostatin was amplified by PCR using primers (Table 1) designed by Primer 3.0 software based on the published sequences of Indian buffalo (GenBank accession number, AY495587). To ensure primer sequence is unique for the template sequence, we checked similarity to other known sequences with BLAST (www.ncbi.nlm.nih.gov/blast/Blast.cgi).

Table (1): Primer sequences, PCR product size and localization of the selected three loci of myostatin gene.

Locus	Forward primer (5' — 3')	Reverse primer (5' — 3')	Size (bp)	Localiz--ation
Mstn1	ATTCACTGGTGTGGCAAG TTGTCTCTCAGA	CCCTCCTCCTTACATACAA GCCAGCAG	620	5' UTR, exon1, intron1
Mstn2	GTTCATAGATTGATATGG AGGTGTTCG	ATAAGCACAGGAACTGG TAGTTATT	550	Intron1, exon2, intron3
Mstn3	TGAGGTAGGAGAGTGTTT TGGG	TCGAAATTGAGGGGAAGA CC	500	Intron3, exon3, 3UTR

The PCR was carried out in a reaction volume of 50 μ L, containing 4.0 μ L DNA template (approximately 100 ng), 1.0 μ L (0.20 mM) dNTP, 5.0 μ L buffer, 3.0 μ L (2.5 mM) MgCl₂, 2.0 μ L 10 μ mol/L forward primer, 2.0 μ L 10 μ mol/L reverse primer, 1.0 μ L 10X Taq DNA

polymerase (5 U/ μ L, Fermentas, #K1071, European Union), and 32 μ L nuclease free water. Thermal cycling parameters were as follows: initial denaturation at 94°C for 5 min, 35 cycles of amplification (94°C for 40 s for DNA denaturation, annealing 56°C for 1 min, extension at 72°C for 1 min) and final extension at 72°C for 10 min. The samples were held at 4°C. PCR products were resolved by electrophoresis on 2% agarose gel in 1X TBE, stained with ethidium bromide and visualized with UV light of gel documentation system (BiometraBiomedizinischeAnalytik, GmbH).

DNA sequencing:

PCR products with expected size were purified using PCR purification kit following the manufacturer protocol (Jena Bioscience # pp-201 \times s) to remove primer dimmers, primers, nucleotides, proteins, salt, agarose, ethidium bromide and other impurities (El-Magd et al., 2013). The purified PCR products were sequenced in automated sequencer (Applied Biosystem, USA). The Sequences were analyzed using the Chromas Lite 2.1 program (http://technelysium.com.au/?page_id=13) and the identity of the sequenced PCR product was examined using Blast search against GenBank database of Indian buffalo (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>). The alignments, annotations and assembly of the sequences were performed using Geneious 4.8.4 software <http://www.geneious.com/web/geneious/home>.

Spectrophotometric method for measurement of mineral level and enzymatic activities:

Calcium was measured using calorimetric method following the manufacturer protocol (BioMerieux[®]- SA-English 3) (*Mundy and Guise 1999*). Phosphorus was measured using photometric method following

the manufacturer protocol (Human Gessellschaft fur Biochemica and Diagonstic GmbH, D-65205 Wiesbaden - Germany) (*Gamset et al.,1980*). ALP was measured using kinetic method following the manufacturer protocol (EliTech) (*Berth and Delanghe 2004*). GOT was measured using kinetic method following the manufacturer protocol Human Gessellschaft fur Biochemica and Diagonstic GmbH, D-65205 Wiesbaden - Germany) (*Schumann and Klauke 2003*).

RESULTS

Three loci of the myostatin gene were amplified using PCR (Fig. 1).

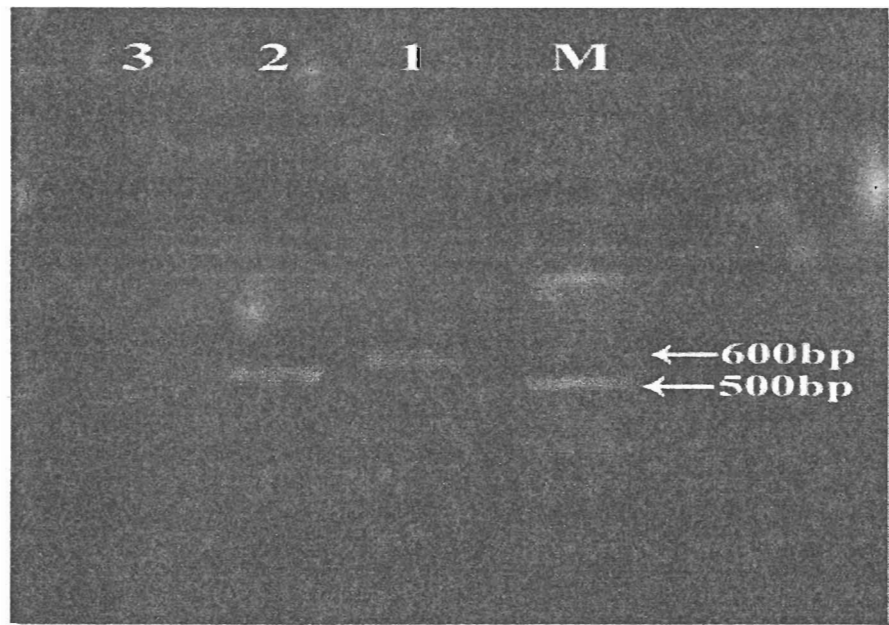


Fig.(1): Ethidium bromide stained agarose gel of PCR products representing amplification of the three loci of *Mstn* gene in Egyptian buffaloes: *Mstn1* with size of 620bp (lane 1), *Mstn2* with size of 550bp (lane 2), *Mstn3* with size 500bp (lane 3). M represents 1kb ladder.

After purification of these loci, sequencing was carried out in order to verify the identity of the PCR product and to detect any SNPs. This sequence was submitted to the GenBank database with accession number KJ123755. The nucleotide sequence analyses of these four loci revealed presence of a novel A31G SNP in *Mstn1* locus at nucleotide number 50 before exon2 (Fig. 2). This SNP was synonymous as it is located in the non coding sequence and so no amino acid was changed. However, the sequences of the other three loci showed 100% identity among the Egyptian buffaloes and as compare to the published sequences of buffalo (GenBank accession number, DQ091762). In all examined sequences, we found only homozygous genotype which either AA or GG and did not find the heterozygous (AG) genotype.

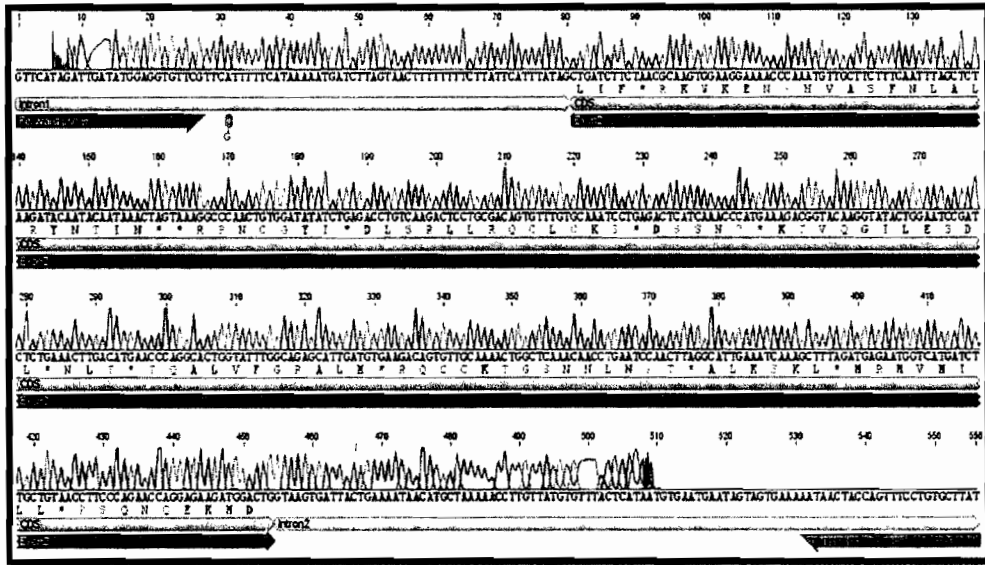


Fig.(2): Nucleotide sequences of *Mstn1* locus in Egyptian buffalo showed anovel synonymous A31G SNP in *Mstn1* locus. The small yellow box indicates the position of SNP.

Association between A31G SNP and serum levels of GOT, ALP, Ca and P:

To identify whether A31G SNP is accompanied by, change in serum levels of GOT, AP enzymes, and Ca and Ph, the levels of these enzymes and minerals were estimated in the serum of high (GG) and low (AA) meat producing Egyptian buffaloes at age of 2 year.

The serum levels of both ALP and GOT enzymes were significantly increased in GG genotype buffaloes with higher meat production as compared to AA genotype animals with lower meat production (Table 2 and Fig.3).

Table (2): The effect of A31G SNP on ALP and GOT serum levels in 24 month old Egyptian buffaloes with high (GG) and low (AA) meat production.

Enzyme	Animal genotype	Range		Mean \pm Standard error of mean
		Minimum	Maximum	
ALP	AA (control)	105	205	159.5 \pm 5.87 ^d
	GG	207	339	263.2 \pm 8.10 ^a
GOT	AA (control)	37	144	92.41 \pm 6.10 ^c
	GG	144	230	197.1 \pm 4.90 ^b

LSD 5% = 0.128, LSD 1% = 0.153

Means with different letters in each column are significantly differed at ($p \leq 0.05$).

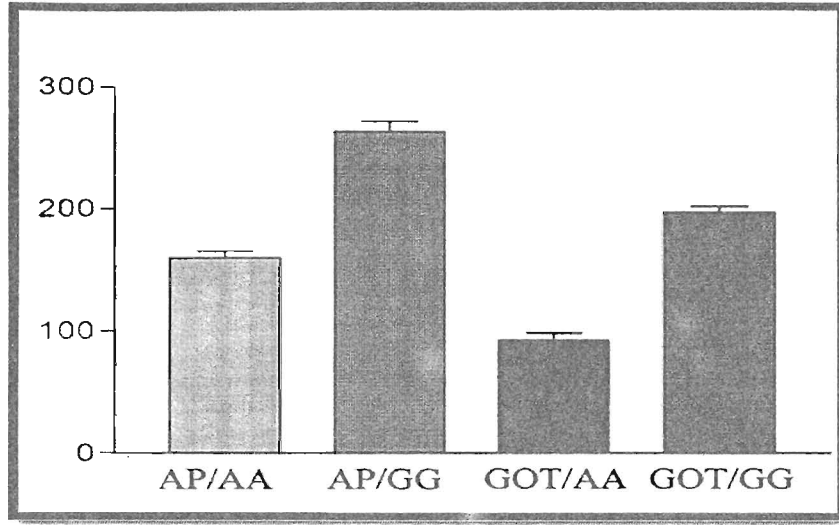


Fig. (3): Serum levels of ALP and GOT enzymes in high (GG) and low (AA) meat producing buffaloes.

The serum levels of both P and Ca were significantly increased in GG genotype buffaloes with higher meat production as compared to AA genotype animals with lower meat production (Table 3 and Fig.4).

Table (3): The effect of A31G SNP on P and Ca serum levels in 24 month old Egyptian buffaloes with high (GG) and low (AA) meat production.

mineral	Animal genotype	Range		Mean ± Standard error of mean
		Minimum	Maximum	
P	AA (control)	3.57	4.61	4.08±0.05 ^d
	GG	4.66	6.77	5.53±0.13 ^c
Ca	AA(control)	7.1	11.06	9.15±0.26 ^b
	GG	11.22	15.55	13.49±0.27 ^a

LSD 5% = 0.126; LSD 1% = 0.158

Means with different letters in each column are significantly differed ($p \leq 0.05$).

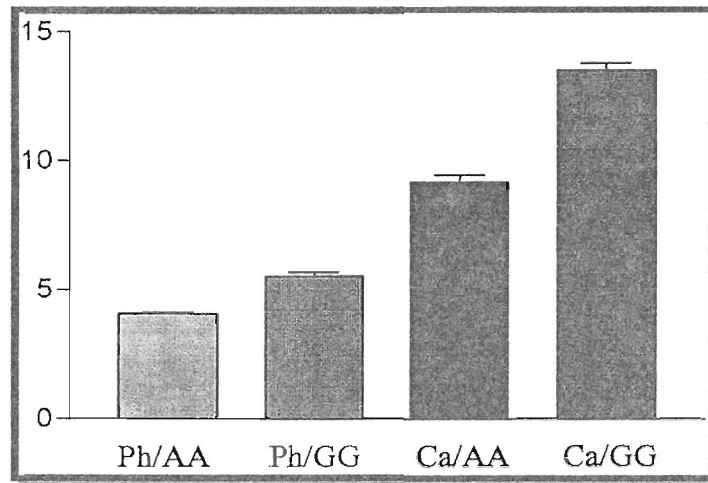


Fig. (4): Serum levels of P and Ca in higher (GG) and lower (AA)meat producing buffaloes.

DISCUSSION

Myostatin (*Mstn*) limits the growth and development of all skeletal muscle in the mammals and so its inhibition dramatically increases muscle mass, due to hypertrophy and hyperplasia of muscle fibers, and produce double muscled animals (*Amthor et al., 2009*). Muscle metabolism in these animals is significantly higher than animals with low growth rate, and subsequently low meat production. Because muscle metabolism accompanied by altered activity of alkaline phosphatase (ALP), glutamic-oxaloacetic transaminase (GOT) (*Latimer et al., 2006*) and Ca and P (*David et al., 1981*), it is possible that the activities of these enzymes and minerals can be elevated in high meat producing animals. Therefore, this study is aimed to detect mutations [such as single nucleotide polymorphisms (SNPs)] in the *Mstn* gene and to associate their effect on enzymatic (GOT and ALP) and mineral (Ca and P) serum levels in high and low meat producing Egyptian buffaloes.

To achieve this, *Mstn* gene was amplified by PCR and subsequently, subjected to sequence analysis to identify different allelic patterns. The nucleotide sequence analyses revealed presence of a novel A31G SNP in *Mstn1* locus at nucleotide number 50 before exon 2. This SNP was synonymous as it is located in the non coding sequence and so no amino acid was changed. However, the sequences of the other three loci showed 100% identity among the Egyptian buffaloes and as compare to the published sequences of buffalo (GenBank accession number, DQ091762).

The serum level of ALP and GOT, can be used as a marker of the functional status of muscle tissue and varies widely in both pathological and physiological conditions. Previous studies have shown a conflict regarding to the relationship between muscle metabolic enzymes and both muscle growth and damage. There are two controversial points of view: the first stated that the metabolic enzymes are located inside the muscle fibers and their serum levels are only elevated when the muscle damaged or diseased, whereas the second theory denoted that elevation of metabolic enzymes in blood is mainly due to increase the muscle mass and activity. Several studies were in consistence with the first point of view. In human, an increase in metabolic enzymes may represent an index of cellular necrosis and tissue damage following acute and chronic muscle injuries (*Brancaccio et al., 2007; Szumilak et al., 1998*). The patients with progressive muscular dystrophy often show elevated serum levels of ALP and GOT (*Srivastava et al., 2007*). Subsequently, measurement of serum ALP and GOT is an important indicator of the occurrence of muscle cell necrosis and tissue damage due to disease or trauma (*Brancaccio et al., 2007*).

On the other hand, the second points of view revealed that the large body animals as increasing in meat production can cause increasing in the serum levels of muscle metabolic enzymes. In agreement with this point of view, (*Szabo et al., 2005*) have found that the increased growth rate, mainly due to the muscular hypertrophy, changed the dynamics and the serum levels of some muscle metabolic enzymes such as ALP and GOT. In birds, the activity of muscle metabolic enzymes increased significantly in the heavy body weight compared with the light body weight. For example; serum levels of muscle metabolic enzymes was higher in domestic turkeys than wild turkeys (*Bounous et al., 2000*). This fact may be a direct consequence of the highly intensive selection for muscle development of the domestic turkey, leading to a significantly higher muscle mass paralleled with increased muscle fiber anaerobic capacity (*Yost et al., 2002*). The strongly increased muscle growth compromises skeletal muscle membrane integrity, leading to elevated serum activities of intracellular enzymes, such as ALP and GOT. Thus, the muscular metabolic activity and the physiological state of growth can be assessed by measuring serum levels of these muscle metabolic enzymes. The significant differences in the serum levels of these muscle metabolic enzymes among low and high body weight may be attributed to the genetic make-up of each weights (*Shata, 2001*).

Ca is an important regulator of several body functions: intracellular signalling, muscle contraction, functioning of the nervous system, hormone and enzyme secretion and blood clotting. Calcium ions are believed to be critical for the fusion of myoblasts, as this process can be activated by calcium ionophores that carry calcium ions across cell

membranes (*David et al., 1981*). It is therefore can be elevated in animals with higher growth rate and meat production. In contrast, elevated Ca levels may be due to increased muscle activities rather than increased muscle mass. This hypothesis means that high growth rate and meat producing animals which are usually lazy should have a lower Ca level in their blood.

As a trial to find an answer for this paradox, we measured serum levels of ALP, GOT, Ca and P in A31G SNP of *Mstn* gene in Egyptian buffalo high and low growth rate and meat production. The serum levels of AP, GOT, P, and Ca were significantly increased in high meat producing buffaloes as compared to low meat producing animals. Because all examined animals were healthy, our study supports the second theory and disagrees with the first theory. In consistence, higher muscle metabolic enzymes was shown in a rapidly growing turkey line, as compared to a slower growing one (*Hassaan et al., 2008*). Our results were in contrast to *Wilson et al.,(1988)* who found that serum muscle metabolic enzymes isn't increased in parallel with the total body or muscle mass (*Wilson et al., 1988*).

The results of this study can enable breeders to do an accurate and earlier selection of high meat producing buffaloes on a genotype basis.

ACKNOWLEDGMENTS

This study was funded by Science Technology Development Fund (STDF), Ministry of Scientific Research, Egypt. All experiments were conducted in the Biotechnology Lab, Faculty of Veterinary Medicine, Kafrelsheikh University, Egypt.

REFERENCES

- **Amthor, H., Otto, A., Vulin, A., Rochat, A., Dumonceaux, J., Garcia, L., Mouisel, E., Hourdé, C., Macharíá, R., Friedrichs, M., Relaix, F., Zammit, P. S., Matsakas, A., Patel, K., Partridge, T., (2009).** Muscle hypertrophy driven by myostatin blockade does not require stem/precursor-cell activity. *Proceedings of the National Academy of Sciences of the United States of America*. 106, 7479-7484.
- **Anjaneyulu, A., Lakshmanan, V., Sharma, N., Kondaiah, N., (1990).** Buffalo meat production and meat quality. *Indian Food Packer*. 44, 21-31.
- **Bounous, D. I., Wyatt, R. D., Gibbs, P. S., Kilburn, J. V., Quist, C. F., (2000).** Normal hematologic and serum biochemical reference intervals for juvenile wild turkeys. *J Wildl Dis*. 36, 393-6.
- **Brancaccio, P., Maffulli, N., Limongelli, F. M., (2007).** Creatine kinase monitoring in sport medicine. *Br Med Bull*. 81-82, 209-30.
- **David, J. D., See, W. M., Higginbotham, C. A., (1981).** Fusion of chick embryo skeletal myoblasts: Role of calcium influx preceding membrane union. *Dev. Biol*. 82, 297-307.
- **El-Magd, M. A., Abbas, H. E., El-kattawy, A. M., Mokhbatly, A., (2013).** Novel polymorphisms of the IGF1R gene and their association with average daily gain in Egyptian buffalo (*Bubalus bubalis*). *Domestic animal endocrinology*. 45, 105-110.
- **El-Nahas, S., Abdel-Tawab, F., Zahran, M., Soussa, S., Rashed, M., Ali, S., (1998).** Gene Mapping of River Buffalo by Somatic Cell hybridization. *Egypt J genet cytol*. 27, 171-179.

- **Genant, H. K., (1993).** Radiology of osteoporosis. In: Favus MJ (ed.) Primer on the metabolic bone diseases and disorders of mineral metabolism. New York, US: Raven Press, Ltd. 229-240.
- **Harper, H. A., (1971).** Review of physiological Chemistry Lange Medical Publications, los Altos,CA.
- **Hassaan, S. F., El Salmony, A. E., Fathi, M. M., (2008).** relationship between triiodothyronine (T3) and insulin like growth factor(IGF1) hormone in egyptian local chicken during growth period. *Poult. Sci.* 28, 251-263.
- **Kambadur, R., Sharma, M., Smith, T. P., Bass, J. J., (1997).** Mutations in myostatin (GDF8) in double-muscled Belgian Blue and Piedmontese cattle. *Genome Res.* 7, 910-6.
- **Latimer, K. S., Mahaffey, E. A., Prasse, K. W. Eds.), (2006).** Clinical pathology.
- **McPherron, A. C., Lee, S. J., (1997).** Double muscling in cattle due to mutations in the myostatin gene. *Proc Natl Acad Sci U S A.* 94, 12457-61.
- **Ogita, Z., Markert, C. L., (1989).** Isozymes Structure, Function and Use in Biology and Medicine. John Wiley and Sons, New York. 853 – 875.
- **Pettifor, J. M., (2008).** What's new in hypophosphataemic rickets? *Eur J Pediatr* 167, 493-499.
- **Shata, I. A. E., (2001):** Physiological and biochemical studies on the skeletal muscles of Japanese quail. fac.of agric., Vol. PH.D. Ain Shams Univ., Cairo.

- **Srivastava, T., Chosdol, K., Chattopadhyay, P., Sarkar, C., Mahapatra, A. K., Sinha, S., (2007).** Frequent loss of heterozygosity encompassing the hMLH1 locus in low grade astrocytic tumors. *J Neurooncol.* 81, 249-55.
- **Szabo, A., Mezes, M., Horn, P., Suto, Z., Bazar, G., Romvari, R., (2005).** Developmental dynamics of some blood biochemical parameters in the growing turkey (*Meleagris gallopavo*). *Acta Vet Hung.* 53, 397-409.
- **Szumilak, D., Sulowicz, W., Walatek, B., (1998).** [Rhabdomyolysis: clinical features, causes, complications and treatment]. *Przegl Lek.* 55, 274-9.
- **Wilson, B. W., Abplanalp, H., Buhr, R. J., Entrikin, R. K., Hooper, M. J., Nieberg, P. S., (1988).** Inbred crosses and inherited muscular dystrophy of the chicken. *Poult Sci.* 67, 367-74.
- **Yost, J. K., Kenney, P. B., Slider, S. D., Russell, R. W., Killefer, J., (2002).** Influence of selection for breast muscle mass on myosin isoform composition and metabolism of deep pectoralis muscles of male and female turkeys. *Poult Sci.* 81, 911-7.