J. of Food and Dairy Sci., Mansoura Univ., Vol. 12 (8): 189 -194, 2021

Journal of Food and Dairy Sciences

Journal homepage: <u>www.jfds.mans.edu.eg</u> Available online at: <u>www.jfds.journals.ekb.eg</u>

Characteristics of children Marshmallow Candy Colored by Natural Anthocyanin Extract from Jamun (*Syzygium cumini*) during cold Storage

Dalia M. El-Mesiry¹*; Soad M. El Desoky¹; Rabab H. Abd El-Razek¹ and M. F. Ahmed²

¹Food Science and Technology Department, Faculty of Home Economics, Al-Azhar University ²Food Technology Research Institute, Agricultural Research Center, Giza, Egypt.

ABSTRACT



It is difficult to produce children's marshmallow candy colored with anthocyanin enriched extract (AEE) from jamun (*Syzygium cumini*) without first researching the product's quality. Different levels of anthocyanin enriched extract (AEE) additionally mixed into the standard marshmallow candy (at 3, 5 and 7g into 100g recipe compared with marshmallow sample coloring by synthetic color (Carmine) at 0.10 g/100g. Characterize of (AEE) were analysis. Water activity, color properties and organoleptic characteristics of the products during storage for 90 days were evaluated. Present study reports (AEE) at 0.6, 0.8 and 1 mg/ml was found to exhibit a higher significant DPPH radical scavenging activity ($P \le 0.05$) than that of BHT at concentration 200 and 300 ppm. there were no significant difference (p<0.05) in (AEE) at pH 5 and incubation period 0, 30, 60, 90 and 120 min at 50°C was 2.65, 3.06, 3.03, 3.18 and 3.16 mg/100 ml. Water activity of marshmallow candy tended to decrease with the addition of (AEE). It caused by the ability of (AEE) to prevent moisture absorption and retention. There were no significant difference in (appearance, odor, taste, texture and overall acceptability) between the control sample and marshmallow samples colored with 3% (AEE). Jamun is a dark purplish fruit and its color undoubtedly contributed to the color of marshmallow by decreasing the L* and b* values and increasing a* value so that Jamun extract can be successfully used to improve properties of children marshmallow candy.

Keywords: Candy products, jambolan, stability, water activity and Organoleptic characteristics

INTRODUCTION

Color is an important quality attribute in the food industry, and it influences consumer's choice and preferences (Pathare *et al.*, 2013). It considered one of the most impressive and delightful attributes of foodstuffs, which directly influences preference, selection and eating desires of the consumers (Shim *et al.*, 2011). Synthetic food colors are substances of chemicals that do not synthesis in nature and made by human. These colors are water-soluble which used in foods without any further processing (Bachalla, 2016).

Biocolourants are prepared from renewable sources and majority is of plant origin. The main food biocolourants are anthocyanidins, carotenoids, flavanoids, chlorophyll, crocin and betalain that extracted from several horticultural plants. In addition to food coloring, biocolourants also act as antimicrobials, antioxidants and thereby prevent several diseases and disorders in human beings. Although, biocolourants have several potential benefits, tedious extraction procedures, low color value, higher cost than synthetic dyes, instability during processing etc. hinder their popularity (Rymbai et al., 2011), so that there has been an increased interest in the development of food colorants from natural sources as alternatives to synthetic dyes because of both legislative action and consumer concern (Giusti and Wrolstad, 1996). Anthocyanins as natural colorants used to color a number of non-beverage foods including gelatin desserts, fruit fillings and certain confectionaries (Chattopadhyay et al., 2008).

Jamun (*Syzygium cumini*), mainly known as 'Jaam' in Bengali, black plum in English and 'Indian berry', belongs to the family Myrtaceae. It found in tropical and subtropical countries. Because of its high nutritional value and excellent processing qualities, it is now gaining popularity in subtropics and arid region of the country (Bukya and Madane, 2018). World production of Jamun estimated to 13.5 million tones, out of which 15.4% contributed by India, Maharashtra state is the largest Jamun producer (Anonymous, 2005).

Cross Mark

Jamun fruit is having higher amount of antioxidants when compared to the other seasonal fruits like banana, papaya, guava and sapota (Ghosh *et al.*, 2017). The higher antioxidant activity in the fruit is attributed to the presence of antioxidant vitamins, tannins and anthocyanins (Koley *et al.*, 2011). Antibacterial activity of water extract of jamun by HPLC shows antibacterial activity at various levels against *Escherichia coli, Bacillus cereus and Staphylococcus aureus* because it contains a large amount of phenolic compounds, tannins and flavonoids so that we can use it in folk medicines (Borhade, 2012).

Functional foods consumed in the normal diet and contain biologically active compounds with potential to improve health or to reduce the risk of disease (Butnariul and Sarac, 2019). The functionality of food depends on some of its bioactive compounds (Asghari *et al.*, 2020). The sugarbased products characterized by high nutritional value, pleasant taste and aroma, attractive appearance, so that the demand for this product group is continuously increasing (Ghendov-mosanu, 2018). Marshmallow candy is becoming

more popular, especially among children, due to the variety of its forms, appearances and flavors (Magomedo *et al.*, 2014).

The aim of this research work was to characterize the pigments obtained from jamun (*Syzygium cumin*) and using these pigments as alternative natural red colorant in marshmallow candy. Water activity, color properties and organoleptic characteristics of the products during storage for 90 days were evaluated.

MATERIALS AND METHODS

Materials

Fully ripe Jamun Fruit: (Syzygium cumini) obtained from the Farm of Horticulture Research Institute at El-Qanatir El-khayriyah city, Qalyubiyah Governorate, Egypt, during September 2019.

Glucose syrup (30-32DE) purchased from National Co. for Maize Products, 10th of Ramadan City, Egypt.

Sugar: Commercial grade granulated sugar cane (sucrose) was obtained from the local market, a product of Egyptian Sugar and Integrated Industries Company, Hawmdia City, Egypt.

Other ingredients: Orange flavor (BC 101), gelatin powder, sodium chloride (food grade) and corn starch were obtained from local market, Tanta City, El-Gharbia Governorate, Egypt.

Chemicals: Synthetic antioxidants namely butylated hydroxyl toluene (BHT) and synthetic red colorants namely carmine (Alum Lake of carminic acid C.I. 75470), Potassium Ferric cyanide, Ferric Chloride, Trichloroacetic acid, Ethanol and 2, 2-Diphenyl-1-Picryl-Hydrazyl (DPPH) all were obtained from sigma-Aldrich Chime, Steinheim, Germany. **Methods**

Extraction of Anthocyanin enriched Extract (AEE) from Jamun (*Syzygium cumini*):

Fresh jamun fruit cleaned and washed. The seeds removed from edible part. Pulp and skin were mashed into particles by an electric mixer (Barun, Combimax 700, Germany). An aqueous extraction was prepared from jamun pulp. The obtained crude extracts were collected and centrifuged at 6000 rpm for 30 min (Hermle Z206A, Germany), and evaporated at 40°C to a small volume (Rotary evaporator, Stuart, UK) . The extract was freeze-dried (Snijders, Netherlands), at an absolute chamber pressure of 0.04 Pa, freeze-drying began at -40° C (shelf temperature). The collected powder processed until it examined in a hermetically sealed glass container.

Preparation of marshmallow candy

Marshmallow samples were prepared using the recipe shown in Table 1. The gelatin was soaked with warm water (45°C). The sugar, glucose syrup, sodium chloride and the remaining water heated. The brix (°B) value of the mixture adjusted to 68-75°B by adding a portion of water. The control sample was coloring by synthetic color (Carmine) at 0.01 g/100g. Different levels of the (AEE) were additionally mixed into the standard marshmallow candy (additional levels at 3, 5 and 7g into the 100g recipe of the marshmallow candy), The mixture was then poured into cylindrical shaped to the required thickness, cooled overnight and cut to desired shape (3cm). Stored in sealed polypropylene plastic bags at $5\pm1^{\circ}$ C and analyzed after 0, 30, 60 and 90 days.

Table 1. Quantity	of ingredients	used in	the formu	lae of
marshm	allow candy			

Ingredients	Weight (g/100g)
Gelatin	10.30
Water to soak gelatin	12.74
Sugar	45.09
Water	28.81
Glucose syrup (30-32 DE)	1.93
Sodium chloride (food grade)	0.13
Orange flavor	1
Total	100

DE= Dextrose Equivalent

Analytical Methods:

Antioxidant Activity of Anthocyanin Enriched Extract from Jamun (*Syzygium cumini*):

The free radical scavenging activity measured according to the DPPH method as reported by Brand-Williams *et al.*, (1995). The FRAP (Ferric reducing antioxidant power) assay was conducted following the method described by Yen and Chen, (1995).

Stability of anthocyanin enriched extract from Jamun (*Syzygium cumini*):

The stability was determined according to the methods of Stintzing and Carle, (2004).

Water activity of marshmallow candy samples:

Water activity (a_w) measured using a Lab Swift-a_w (Novasina, Switzerland) hygrometer (Piga *et al.*, 2005).

Color parameter of marshmallow candy samples:

Color measured by a colorimeter (Minolta CR 400, Minolta Camera, Co, Osaka, Japan) and recorded in the L*, a^* , b^* color system (Hunt, 1991).

Organoleptic characteristics of marshmallow candy samples:

Organoleptic characteristics was carried out (Watts *et al.*, 1989) by panelists from Food Science and Technology Department, Faculty of Home Economic, Al-Azhar University, Tanta, Egypt.

Statistical Analysis

The results were analyzed statistically to establish mean values, standard deviation $(\pm SD)$. The data were statistically evaluated according to the 5 percent level of probability procedure by the least significant differences (L.S.D) (Snedecor and Cochran, 1980) using Costat program Version of (6.451).

RESULTS AND DISCUSSION

Antioxidant activity of anthocyanin enriched extract from jamun (*Syzygium cumini*):

The free radical–scavenging capacity of anthocyanin enriched extract at different concentrations was measured by DPPH assay and ferric reducing antioxidant power (FRAP) is given in Table 2. Data showed the ability of (AEE) to scavenge DPPH radicals demonstrated as the percentage of DPPH inhibition. At the concentration of (1, 0.8, 0.6 and 0.4 mg/ml) (AEE) caused the highest scavenging of the DPPH radical (97.9, 93.9, 76.07 and 51.03%), respectively. While the lowest inhibition values of DPPH radical were (36.72 and 25.06%) at concentration (0.2 and 0.1mg/ml), respectively. Anthocyanin enriched extract at 0.6, 0.8 and 1mg/ml was found to exhibit a higher significant DPPH radical scavenging activity ($P \le 0.05$) than that of BHT which caused scavenging of the DPPH radical (59.14 and 82.45), respectively at concentration 200 and 300 ppm. The IC₅₀ value of the extract studied was found to be 3.60 mg/ml and was equivalent to the synthesis antioxidant BHT (IC₅₀ = 139.5 ppm). This result was accordance with (Menaka and Venkatasubramanian, 2018) found that the antioxidant activity of Lyophilized Jamun Pulp (22.54%) Inhibition and was better than Jamun Pulp (18.5%).

Table 2. Antioxidant activity of Anthocyanin Enriched
Extract (AEE) from Jamun (Syzygium cumini)
compared with butylated hydroxytoluene (BHT)

	Antioxida	nt activity			
Concentrations	DPPH (%)	FRAP (Absorbance at			
0.1 / 1	05.05	700 nm)			
0.1 mg/ml	25.06	0.166			
0.2 mg/ml	36.72	0.254			
0.4 mg/ml	51.03	0.402			
0.6 mg/ml	76.07	0.553			
0.8 mg/ml	93.9	0.700			
1 mg/ml	97.9	0.855			
(AEE) IC ₅₀ (mg/ml)	3.60				
BHT (100ppm)	36.28	0.306			
BHT (200ppm)	59.14	0.789			
BHT (300ppm)	82.45	0.988			
(BHT) IC ₅₀	139.60				

DPPH=2, 2-diphenyl-1-picrylhydrazyl radical. FRAP= ferric reducing antioxidant power. IC_{30} = (Half maximal inhibitory concentration)

Data in the same table showed that in ferric reducing antioxidant power (FRAP), the yellow shade of the test arrangement changes to green or blue shading relying upon the reducing power of antioxidant samples. A higher absorbance indicates a greater reducing power of the ferric. An increase in FRAP was observed in the anthocyanin enriched extract with increasing concentration. In general, the natural antioxidant extract (AEE) would be preferred over synthetic antioxidant (BHT) to minimize the adverse health effects. Anthocyanin, as well as other phenolic compounds could acts as antioxidants by donating hydrogen to highly reactive radicals thereby preventing further formation of oxidation products (Fu-kumoto and Mazza, 2000).

Stability of Anthocyanin Enriched Extract (AEE) from Jamun (*Syzygium cumini*):

The stability of Anthocyanin Enriched Extract (AEE) dissolved in buffer solution at various pH values of buffer ranging from 2.5 to 8.0 and water at pH 6.65. The results in Table 3 presented that, there is no significant difference (p<0.05) in (AEE) at pH 5 and incubation period 0, 30, 60, 90 and 120 min at 50°C was 2.65, 3.06, 3.03, 3.18 and 3.16 mg/100 ml. Meanwhile, anthocyanin enriched extract was more stable at pH 2.5, 3 and 8, which was 21.92, 8.90 and 10.87 mg/100 ml, respectively. However, by diminishing the pH underneath 3 or expanding the pH over 8, (AEE) content was significantly decreased.

The statistical analysis in Table 4. show non a significant difference (p<0.05) in (AEE) content by the raising of time from 0: 120 min on buffer solutions of anthocyanin pigment at different pH and temperatures resulted led to no significantly decreased from 7.85 to 7.89 mg / 100 ml, respectively. However, there was not a vast difference in retention between the 50 and 70 °C treatments, this could indicate imply that most of the pigments were degraded at 90 °C and no further degradation could occur as reported by (Saira and Kamran, 2017). Our results were according to Troise and Fogliano, (2013) noticed that the stability of anthocyanin and all pigments found in foods decreased with increases in temperature.

Table 3. Effect of temperature, time and pH level on stability of Anthocyanin Enriched Extract (AEE) (mg /100 ml) from Jamun (Syzygium cumini)

Temperature	Time	pH							
(°C)	(min)	2.5	3	4	5	6	7	8	Water
	0	23.07 ^a	8.51 mnop	3.54 yzABCD	2.65 ^D	2.94 ^{BCD}	4.48 vwxyz	14.69 ^e	2.90 ^{CD}
	30	22.82 ^a	8.98 ^{jklmno}	3.72 xyzABCD	3.06^{ABCD}	3.31 zabcd	5.12 ^{tuvw}	10.31 ^{fghi}	3.10 ^{ABCD}
50	60	22.77 ^{ab}	9.95 ^{fghijk}	3.70 xyzABCD	3.03 ABCD	3.60 yzabcd	4.25 ^{wxyzA}	8.98 ^{jklmno}	3.25 ^{zABCD}
	90	22.71 ^{ab}	9.07 hijklmno	3.91 wxyzABCD	3.18 ABCD	3.56 yzabcd	5.71 stuv	8.87 ^{jklmno}	3.49 ^{zABCD}
	120	22.62 ^{ab}	9.12 hijklmn	4.00 wxyzABC	3.16 ABCD	3.73 xyzABCD	5.85 stu	8.79 ^{jklmno}	3.29 ^{zABCD}
	0	23.07 ^a	8.51 mnop	3.54 yzabcd	2.65 ^D	2.94 ^{BCD}	4.48 vwxyz	14.69 ^e	2.90 ^{CD}
	30	22.62 ^{ab}		3.86 wxyzABCD	3.20 zabcd	3.98 wxyzABC	6.60 qrs	9.25 hijklmn	3.26 zabcd
70	60	22.57 ^{av}	8.92 ^{jklmno}	3.99 wxyzABC	3.32 zabcd	4.31 wxyzA	7.28 ^{pqr}	10.01 ^{fghij}	3.57 yzabcd
	90	22.51 ^{ab}	9.16 ^{hijklmn}	4.11 wxyzABC	3.52 zabcd	4.82 ^{uvwxy}	7.80^{opq}	9.87 ^{fghijkl}	3.37 ^{zABCD}
	120	22.21 ^{ab}	9.04 ^{ijklmno}	4.14 wxyzABC	3.62 xyzABCD	4.83 ^{uvwxy}	7.81 ^{opq}	9.95 ^{fghijk}	3.39 zabcd
	0	23.07 ^a	8.51 mnop	3.54 yzabcd	2.56 ^D	$2.94 ^{\text{BCD}}$	4.48 vwxyz	14.69 ^e	2.90 ^{CD}
	30	21.48 ^{bc}	8.73 ^{jklmno}	4.03 wxyzABC	3.56 yzabcd	4.92 tuvwx	8.07 nop	10.61 ^{fg}	3.30 zabcd
90	60	20.65 ^c	8.66 klmno	4.08 wxyzABC	4.08 wxyzABC	5.71 stuv	8.25 mnop	10.34^{fgh}	3.49 ^{zABCD}
	90	18.80 ^d	8.51 mnop	4.23 wxyzAB	4.19 wxyzABC	6.02 rstu	8.58 Imno	10.96 ^f	3.58 yzabcd
	120	17.80 ^d	8.39 mnop	4.26 ^{wxyzA}	4.26 wxyzA	6.15 rst	8.39 mnop	11.06 ^f	3.57 yzabcd
-	LSD _{0.05} (temperature*time * pH level) = 1.29								

W: Water at pH 6.65 Means with different letters (a, b, c ...K, J) indicate significant differences (P < 0.05) LSD= Least significant difference

Water activity of marshmallow candy samples during storage period at 5±1°C:

Table 5 demonstrate that water activity values as affected by supplementing with Anthocyanin Enriched

Extract (AEE) during cold storage period .The data showed that, control sample a_w recorded as the highest value (0.846) followed by marshmallow candy colored with (3%) (AEE) (0.830) at zero time. The lowest a_w value (0.806) was

recorded for marshmallow candy colored with 7% (AEE), followed by marshmallow candy colored with 5% (AEE) (0.807) and marshmallow candy colored with 0.1% synthetic red color (SRC) (0.808) was the next at zero time. Water activity of marshmallow candy tended to decrease with the addition of (AEE). It caused by the ability of (AEE) to prevent moisture absorption and retention.

Table 4. Significant trends on influence means of
temperature, time and pH level on
Anthocyanin Enriched Extract (AEE) (mg
/100 ml) from Jamun (Syzygium cumini)

Temperature (°C)	Total anthocyanin	Time (min)	anthoevanin	рН	Total anthocyani	c in T
50	7.60 ^b	0	7.85 ^a	2.5	21.92 ^a	
70	7.98 ^a	30	7.81 ^a	3	8.90 ^b	
90	8.04 ^a	60	7.87 ^a	4	3.91 ^e]
		90	7.94 ^a	5	3.34 ^e	
		120	7.89 ^a	6	4.25 ^e	(
LSD _{0.05} =0.20				7	6.48 ^d	S
		тс	-1.07	8	10.87 ^b	3
		LO	$D_{0.05}=1.07$	Water	: 3.29 ^e	5
				LS	SD _{0.05} =1.36	7

W: Water at pH 6.65 LSD= Least significant difference Means with different letters (a, b, c ... K, J) indicate significant differences (P < 0.05)

Table 5. Water activity values of marshmallow candy supplemented with different levels of (AEE) during cold storage periods at 5+1⁰C

during cold storage periods at 5±1°C							
	Storage period						
Treatments	Zero	After 1	After 2	After 3	at		
Treatments	time	month	months	months	0.05		
Control	0.846 ^{aA}	$0.792^{aB} \pm$	$0.758^{aC}\pm$		0.022		
Colluor	± 0.006	0.013	0.004	± 0.003	0.022		
synthetic	0.808 ^{cA}	$0.801^{aA}\pm$	$0.747^{aA} \pm$	0.663 ^{cB}	0.072		
color 0.10%	± 0.004	0.011	0.050	±0.000	0.072		
20/ AEE	0.830 ^{bA}	0.802^{aAB}	0.756^{aAB}	0.660 ^{cB}	0 142		
3%AEE	± 0.003	± 0.095	±0.037	± 0.014	0. 143		
5% AEE	0.807 ^{cA}	0.776^{aAB}	0.767^{aAB}	0.742^{aB}	0.050		
5% AEE	± 0.005	± 0.035	± 0.009	± 0.003	0.050		
	0.806 ^{cA}	$0.773^{aB}\pm$	$0.715^{aC}\pm$	0.618^{dD}	0.012		
7%EAE	± 0.008	0.002	0.001	± 0.003	0.012		
LSD at0.05	0.014	0.118	0.073	0.017			

Synthetic red color Prepared with 0.10% (carmine). (AEE) = Anthocyanin Enriched Extract. In a column, means having the same superscript small letters are not significantly different at 5% level for marshmallow samples. In a row, means having the same superscript capital letters are not significantly different at 5% level for cold storage period. LSD = Least significant differences.

Regularly, recorded data of the same Table illustrated significance decrease of the a_w values for the control sample and all treatment by increasing the storage period from zero time up to three months at cold storage. Our results were agreement with Opriș *et al.*, (2020) found that the numerical value of the water activity varies in the range of (0.779) for control (fondant candy), (0.713) for fondant colored with grape extract and (0.724) for fondant colored with aronia powder after the first day of storage and reduced to (0.765) for control, (0.676) for grape extract sample and (0.705) for aronia powder sample after the 35 day of storage.

Color parameter of marshmallow candy samples during storage period at 5±1°C:

From statistical analysis of data given in Table 6 it could be noticed that control sample was the highest L* value (91.12) followed by marshmallow candy colored with 0.1% synthetic red color (SRC) (75.25) indicating that they were the clearest samples, on the other hand, Marshmallow candy colored with 7% (AEE) (57.47) was the darkest samples. It was illustrated that, increasing the levels of (AEE) showed a tendency to decrease lightness (L*) at zero time. Finally, decrease in L* value suggests darkening of the product and can be attributed to the occurrence of non-enzymatic browning.

 Table 6. Color measurement of marshmallow candy supplemented with different levels of (AEE) during cold storage period at 5±1°C

Treatments	L*	a*	b*	C*		
Zero time						
Control	91.12	-1.39	12.29	12.36		
Synthetic color	75.25	8.27	4.20	9.27		
3%AEE	74.68	1.22	12.30	12.3		
5%AEE	64.92	2.55	5.22	5.08		
7%AEE	57.47	2.65	5.71	6.29		
	After montl	n				
Control	87.76	-0.17	10.69	10.69		
Synthetic color	74.63	10.92	3.69	11.52		
3%AEE	68.08	3.77	7.40	8.5		
5%AEE	49.79	6.06	4.67	7.65		
7%AEE	57.68	6.95	5.01	8.56		
A	After 2 mont	hs				
Control	81.68	0.12	2.44	2.44		
Synthetic color	70.38	12.21	-2.07	12.38		
3%AEE	66.11	3.81	5.50	6.69		
5%AEE	46.95	6.32	-0.73	6.36		
7%AEE	54.72	8.11	-0.26	8.11		
A	After 3 mont	hs				
Control	78.34	0.19	2.08	2.08		
Synthetic color	54.97	13.16	-3.99	13.75		
3%AEE	53.35	4.14	4.87	6.39		
5%AEE	30.19	9.49	-1.37	9.58		
7%AEE	34.22	11.09	-0.80	11.11		
		b) =yellown		= Chroma.		
Synthetic red color F	'repared with	0.10%	(carmin	e) (AEE)		

Synthetic red color Prepared with 0.10% (carmine) (AEE) =Anthocyanin Enriched Extract.

There were increased in redness (a*) with increasing the supplemented (AEE) ratio for the control sample and all treatments at zero time. This increase in red color was due to the anthocyanin activity as the red pigment in (AEE). Similar result was found by Ekie and Evanuarini, (2020) found that the average value of redness (a*) in milk candy was 12.68 to 19.10 where along with the increase of rosella powder percentage, this due to the anthocyanin activity as the red pigment giver in rosella where the different treatments; (12.68) for control treatment increased to (15.00, 17.15 and 19.10) for (1, 3 and 5%) rosella powder. These data showed that, there were increase in a* value among the control samples and all treated samples at zero time up to three months. (b*) values indicating yellowness, ranged from 4.20 to 12.30 in marshmallow samples at zero time. There were decrease in (b*) values among control sample and all other treatments at the end of storage period (3 months) comparing to zero time. The yellowness (b*) should be considered as the physical parameters to describe the visual color degradation of the candy during storage. Changes in color intensity presumably caused by the presence of the enzyme. Change in color parameters of candy can be due to the occurrence of non-enzymatic browning reactions that also took place together with oxidation (Zhou *et al.*, 2013).

It could be noticed that, there were general decrease in (C*) among the control sample and all treatments supplemented with (AEE) at zero time. Overall, the results showed that the marshmallow supplemented with (AEE) were redder and less yellow than control marshmallow. Jamun is a dark purplish fruit and its color undoubtedly contributed to the color of marshmallow by decreasing the L* and b* values and increasing a* value so that Jamun extract can be successfully used to improve color properties of candy.

Organoleptic characteristics of marshmallow candy samples during storage period at 5±1°C:

The sensory panelists were recorded comparable appearance, odor, taste, texture and overall acceptability scores for marshmallow samples that supplemented with anthocyanin enriched extract (AEE) stored at $5\pm1^{\circ}$ c for three months

(Table 7). From tabulated data, score of appearance was 9.9 for control sample followed by samples colored with (3 and 5 % AEE) which recorded (9.8 and 9.7) which was the best appearance samples. On the other hand, sample colored with 7% (AEE) and synthetic red color samples recorded the same lowest score (9.5) at zero time. Score of appearance at all samples decreased during storage period. Furthermore, the highest appearance value found in control sample and 3% (AEE) (7.8), while the lowest (6.1) for synthetic red color samples at the end of storage period (three months).

The value of odor was highest at control samples (9.9) followed by samples treated with 3% (AEE) (9.8) but the lowest score recorded by 7% (AEE) (9.1) at zero time. On the other hand, there were general decrease in odor at all storage period ranged from 7.9 for control sample and 3% (AEE) to 7.1 for synthetic red which was the lowest at the end storage periods (three months).

The data showed that the highest score of taste was 3% (AEE) (9.9) which was the best taste, while the worst taste recorded by synthetic color (9.1) at zero time. Control samples, 3 and 5% (AEE) was the highest score at texture and over all acceptability (9.8, 9. 7 and 9.7) (9.9, 9.9 and 9.8), respectively and was the best at zero time and continues as the best at the end of storage (three months) (8.1, 7.9 and 7.6) (7.8, 7.8 and 7.7), respectively.

Maiia *et al.*, (2017) found that marshmallow with Sudanese rose and black chokeberry extracts possesses a regular shape with clear outlines without deformation. Almost all of the organoleptic characteristics (shape, consistency, taste, smell, color) of the new products with natural anthocyanin dyes exceed the results of the control sample, manufactured without them.

Finally, consumer perception has been that natural food colorant ingredient would be healthful and considered as potential food colorants for preparing hard candy and Jellies these findings are in accordance with that of El-Gharably (2005).

Table 7. Organoleptic characteristics of marshmallow candy supplemented with different levels of (AEE) during cold storage period at 5±1°C

	(ILL) u	ur ing cor	u stor age	per lou at	5±1 C
Treatment	Appearance	Odor	Taste	Texture	Overall acceptability
-		Ze	ero time		
Control	9.9 ^a ±0.32	$9.9^{a}\pm0.48$	$9.8^{a} \pm 0.42$	9.8 ^a ±0.52	9.9 ^a ±0.53
Synthetic color	9.5 ª±0.74	9.6 ^{ab} ±0.74	9.1 ^b ±0.85	$8.7^{\rm b}\pm\!0.85$	8.6 ^b ±0.48
3% AEE	9.8ª±0.57	$9.8^{a} \pm 0.57$	9.9 ^a ±0.57	9.7 ^a ±0.74	9.9 ^a ±0.94
5%AEE	9.7 ^a ±0.57	9.2 ^b ±0.32	9.6 ^{ab} ±0.82	9.7 ^b ±0.63	9.8 ^a ±0.70
7%AEE	9.5 ^a ±0.67	9.1 ^b ±0.57	9.4 ^{ab} ±0.78	$9.6^{a}\pm0.44$	8.9 ^b ±0.67
LSD at 0.05	0.45	0.51	0.52	0.50	0.54
		Afte	er month		
Control	8.9 ^a ±0.53	8.5 ^{ab} ±0.67	$8.6^{a} \pm 1.03$	8.5 ^a ±0.74	8.9 ^a ±0.53
Synthetic color	7.5 ^b ±0.79	7.9 ^{bc} ±0.48	7.6 ^b ±0.57	$8.1^{ab}\pm0.48$	$8.4^{ab}\pm\!0.79$
3% AEE	$8.7^{a} \pm 0.79$	$8.7^{a}\pm0.84$	$8.8^{a} \pm 0.67$	7.9 ^{ab} ±0.57	8.7 ^a ±074
5%AEE	$7.8^{b} \pm 0.63$	$7.8^{\circ} \pm 0.82$	7.8 ^b ±0.71	$7.8^{b}\pm0.57$	$7.8^{b} \pm 0.79$
7%AEE	$7.8^{b}\pm0.78$	7.9 ^{bc} ±0.33	$7.5^{b} \pm 0.60$	$7.9^{ab} \pm 0.93$	7.9 ^b ±0.60
LSD at 0.05	0.68	0.66	0.59	0.67	0.64
		After	r 2 months		
Control	$8.4^a \pm 0.88$	$8.8^{a}\pm0.70$	7.6 ^b ±0.94	8.4 ^a ±0.82	8.4 ^a ±0.52
Synthetic color	$7.4^b \pm 0.52$	7.6 ^b ±0.82	$7.8^{b}\pm\!0.79$	6.6° ±0.63	7.7 ^b ±0.70
3% AEE	$7.7^{b} \pm 1.08$	7.9 ^b ±0.63	$8.9^{a} \pm 0.82$	7.7 ^b ±0.74	$7.8^{ab} \pm 0.82$
5%AEE	7.9 ^{ab} ±0.74	$7.7^{b} \pm 0.57$	7.7 ^b ±0.71	7.6 ^b ±0.85	7.7 ^b ±0.71
7%AEE	7.4 ^b ±0.53	$7.4^{b}\pm0.60$	$7.4^{b}\pm 0.53$	$7.4^{b}\pm0.73$	$7.4^{b}\pm0.97$
LSD at 0.05	0.69	0.67	0.68	0.59	0.67
		After	r 3 months		
Control	$7.8^{a}\pm0.70$	7.9 ^a ±0.63	$7.5^{\rm a}\pm0.92$	8.1ª ±0.57	$7.8^{a} \pm 0.59$
Synthetic color	$6.1^{b}\pm 0.67$	7.1 ^b ±0.85	7.3 ^a ±0.63	6.1° ±0.84	6.5° ±0.54
3% AEE	$7.8^{a} \pm 0.74$	$7.9^{a} \pm 0.85$	$7.6^{a} \pm 0.63$	7.9 ^{ab} ±0.84	$7.8^{a} \pm 0.84$
5%AEE	7.7 ^a ±0.52	7.8 ^a ±0.70	7.5 ^a ±0.67	7.6 ^{ab} ±1.03	7.7 ^{ab} ±0.48
7%AEE	7.5 ^a ±0.71	7.4 ^{ab} ±0.67		7.4 ^b ±0.92	7.2 ^b ±0.71
LSD at 0.05	0.54	0.59	0.52	0.61	0.56

Synthetic red color Prepared with 0.10% (carmine) (AEE) =Anthocyanin enriched extract. Means having the same superscript letters are not significantly different at 5% level for marshmallow samples LSD = Least significant differences.

CONCLUSION

This study provides clear evidence it is economic, proper and successful applicable to produce children marshmallow candy colored by natural color (anthocyanin extract from *Syzygium cumini*) as a substitute of the artificial colors, hence must resort to natural sources which are very suitable to be taken as a good natural food or natural-food additive with many categories of healthy foodstuffs.

REFERENCES

- Anonymous (2005). Jamun production trends in India and world. Market updates fruits.
- Asghari, A. A., Mokhtari-Zaer, A., Niazmand, S., Mc Entee, K. and Mahmoudabady, M. (2020). Anti-diabetic properties and bioactive compounds of *Teucrium polium* L. Asian Pacific Journal of Tropical Biomedicine, 10(10): 433-441.
- Bachalla, N. (2016). Identification of synthetic food colors adulteration by paper chromatography and spectrophotometric methods. International Archives of Integrated Medicine, 3(6):182-191.

- Borhade, S. (2012). Antibacterial activity, phytochemical analysis of water extract of *Syzygium cumini* and analytical study by HPLC. Asian Journal of Experimental Biological Sciences, 3(2): 320-324.
- Brand-williams, W., Culvelier, M.E. and Berset, C. (1995). Use of free radical method to evaluate antioxidant activity. LWT-Food Science and Technology, 28(1): 25-30.
- Bukya, A. and Madane, L. P. (2018). Preparation and standardization of jamun jam (*Syzygium cumini L.*) its chemical and storage studies. World Journal of Pharmacy and Pharmaceutical Sciences, 7(4): 876-885.
- Butnariu1, M. and Sarac, I. (2019). Functional Food. International Journal of Nutrition, 3(3): 7-16.
- Chattopadhyay, P., Chatterjee, S. and Sen, S. K. (2008). Biotechnological potential of natural food grade biocolorants. African Journal of biotechnology, 7(17):2972-2985.
- Ekie, M. A. B. and Evanuarini, H. (2020). The quality of milk candy using rosella powder (*Hibiscus sabdariffa L.*) addition as natural food colouring. In IOP Conference Series: Earth and Environmental Science, 492(1):1-7.
- El-Gharably, A. M. A. (2005). Characterization of anthocyanin pigments extracted from grape skins and its potential uses as antioxidant and natural food colorants. Journal of Home Economic. Minufiya University, 15(4): 51-70.
- Fu-kumoto, L. R. and Mazza, G. (2000). Assessing antioxidant and prooxidant activities of phenolic compounds. Journal of Agricultural and Food Chemistry, 48(8): 3597-3604.
- Ghendov-Mosanu, A. (2018). The use of dog-rose hips (*Rosa canina*) fruits in the production of marshmallow-type candy. Food and Environment Safety, 17(1): 59-65.
- Ghosh, P. P., Radha, R. C., Mishra, S., Patel, A. S. and Kar, A. (2017). Physicochemical and nutritional characterization of Jamun (*Svzygium cuminii*). Current Research in Nutrition and Food Science Journal, 5(1): 25-35.
- Giusti, M. and Wrolstad, R. E. (1996). Radish anthocyanin extract as a natural red colorant for maraschino cherries. Journal of Food Science, 61(4): 688-694.
- Hunt, R. W. G. (1991). Measuring color, 2nd Ed., Ellis Horwood, New York, 75-76.
- Koley, T. K.; Barman, K. and Asrey, R. (2011). Nutraceutical properties of Jamun *Syzygium cumini* (L.) and its processed products. Indian Food Industry, 30(4):34-37.
- Magomedo, G., Plotnikova, I., Zhuravlev, A., Shevjakova, T. and Popova, A. (2014). Perfection of gelatin mini-zephyr (marshmallow) technology (in Russian), Konditerskoe i Hlebopekarnoe Proizvodstvo,11(12): 6-9.
- Maiia, A., Inna, P., Olga, S., Natalia, M., Oleksii, K., Iryna, F. and Anthelia, G. (2017). A study of properties of marshmallow with natural anthocyanin dyes during storage. Technology and Equipment of Food Production, 3(11):23-30.

- Menaka, M and Venkatasubramanian, Ch. (2018). Nutrient content and Antioxidant Profile of Raw and Lyophilized Jamun (*Syzygium Cumin*) Fruit Pulp. International Journal of Chemtech Research, 10(2): 968-974.
- Opriş, O. I., Lung, I., Soran, M. L., Sturza, R. and Ghendov-Moşanu, A. (2020). Fondant candies enriched with antioxidants from aronia berries and grape marc. Revista de Chimie (Rev. Chim.), 71(2): 74-79.
- Pathare, P. B., Opara, U. L. and Al-Said, F. A. J. (2013). Colour measurement and analysis in fresh and processed foods: a review. Food and Bioprocess Technology, 6(1):36-60.
- Piga, A., Catzeddu, P., Farris, S., Roggio, T., Sanguinetti, A. and Scano, E. (2005). Texture evolution of "Amaretti" cookies during storage. European Food Research and Technology, 221(3): 387-391.
- Rymbai, H., Sharma, R. R. and Srivastav, M. (2011). Biocolorants and its implications in health and food industry–a review. International Journal of Pharmacological Research, 3(4): 2228-2244.
- Saira, S. and Kamran, K. M. (2017). Stability of anthocyanins from *Syzygium cumini* (Jamun) at Different Processing Condition. Journal of Food Technol Pres, 2 (1):1-5.
- Shim, J. H., Kim, J. Y., Park, M., Park, J. and Cho, D. W. (2011). Development of a hybrid scaffold with synthetic biomaterials and hydrogel using solid freeform fabrication technology. Biofabrication, 3(3): 1-9.
- Snedecor, G.W. and W.G. Cochran. (1980). Statistical methods. Oxford and J. B. H publishing Com. 7th edition, 224-308.
- Stintzing, F. C. and Carle, R. (2004). Functional properties of anthocyanins and betalains in plants, food, and in human nutrition. Trends in Food Science and Technology, 15(1): 19-38.
- Troise, A. D. and Fogliano, V. (2013). Reactants encapsulation and Maillard reaction. Trends in food science and technology, 33(1): 63-74.
- Watts, B. M., Ylimaki, G. L., Jeffery, L. E. and Elias, L. G. (1989). Basic sensory methods for food evaluation (No. TX 546. B37 1989). International Development Research Centre (Canada).
- Yen, G. C. and Chen, H. Y. (1995). Antioxidant activity of various tea extracts in relation to their antimutagenicity. Journal of Agricultural and Food Chemistry, 43(1): 27-32.
- Zhou, C. L., Liu, W., Zhao, J., Yuan, C., Song, Y., Chen, D. and Li, Q. H. (2013). The effect of high hydrostatic pressure on the microbiological quality and physical–chemical characteristics of Pumpkin (*Cucurbita maxima* Duch.) during refrigerated storage. Innovative Food Science and Emerging Technologies, 21: 24-34.

خصائص حلوى مارشميلو الأطفال الملونة بمستخلص الأنثوسيانين الطبيعي من ثمار البامبوزيا أثناء التخزين المبرد داليا محمد المسيرى1،سعاد محمود الدسوقي1،رباب حسن عبد الرازق1 و محسن فرغلي أحمد 2 1علوم وتكنولوجيا الأغذية كلية الإقتصاد المنزلي. جامعة الازهر 2تكنولوجيا الحاصلات البستانية .معهد بحوث تكنولوجيا الأغذية.مركز البحوث الزراعية

من الصعب إنتاج حلوى الأطفال (المار شيملو) الملونة بمستخلص الأنثوسيانين الطبيعي من شرة البامبوزيا دون البحث أو لأعن صفات المنتج لذلك فقد تم إضافة مستويات مختلفة من مستخلص الصبغة بنسب 3, 5 و 7 جرام لكل 100 جرام وخلطها مع حلوى المار شميلو الكنترول ومقارنتها مع عينة مار شميلو تم تلوينها باللون الصناعي (كارمين) بنسب إضافة 10.0جم لكل 100 جرام. تم تحليل خصائص المنتج من حيث تقدير النشاط الماتي وتقييم خصائص اللون والخصائص الحسبة للمنتجات عند التخزين المبرد لمدة 90 بوما. في هذه الدر اسه أيضا أظهر المستخلص نشاط أكبر في إز الة الشقوق الحرة عند تركيز 0.6, 0.8 و 1 مجم / مل عند مقارنته بمضد الأكسده الصناعي عد تركيز 200 و 300 جزء في المليون . أيضا الم يكن هناك فروق معنوية للمستخلص الغني بالصبغة عند الأس الهيدروجني 5 وكانت فترة التحضين 0 , 30, 00 و 200 دقية حرف قرق 1.5 مينا لم يكن هناك فروق معنوية للمستخلص الغني بالصبغة عند الأس الهيدروجني 5 وكانت فترة التحضين 0 , 30, 00 و 200 دقية حرف المليون . أيضا الم 3.16 ملجم / 100 مل، قيم النشاط الماتي لحلوى المار شميلو الجهت إلى الإخفاض مع إضافة المزيد من ساب المستخلص لين المرد لمنة 3.16 ملجم / 100 مل، قيم النشاط الماتي لحلوى المار شميلو البهدر وجني 5 وكانت فترة التحضين 0 , 30 م 30 و 200 دقية 2.5 م 3.5 م 3.5 م 3.6 ملجم / 100 مل، قيم النشاط الماتي لحلوى المار شميلو التجهت إلى الإنخفاض مع إضافة المزيد من نسب المستخلص بسبب قدرته على منع امتصاص الرطوية والاحتفاظ بهاءلم يكن هناك فروق معنوية في الخواص الحسية من حيث (المظهر ، الرائحة، الطعم، والمامس والقبول العام) بين الحياة المنترول وعينات المار شميلو المونية والاحقاظ بهاءلم . البامبوزيا فكور في منوية هذا ساهم بلا شك في تلوين حلوي المار، شيلو عنهم الميان والاصغرار وزيادة قيم اللون الأحمر بحيث يمكن بنجاح استخلص . البامبوزيا فكنهة أرجو الية دالون المونه هذا ساهم بلهم على عنه المعان والمونية والمونية بنسبة إضافة لائم م . البامبوزيا فاكهة أرجو الية داكلة اللون ولونها هذا ساهم بلا شك في تلوين حلوي المار، شيلو قيم المعان والاصغرار وزيلاية قيم اللون الأحمر بحيث يمكن بنجاح استخدام مستخلص الثمره في تحسين خصائص حلوي المار شرك موي المار شميلو عن طريق تقليل قيم اللمعان والاصغرار وزيلاة قيم الون الأحمر بحيث يمكن بنجاح استخدام

الكلمات الاسترشائية: منتجات الحلوى ، الجامبولان ، الثبات ، النشاط المائي والخصائص الحسية